

**Supporting Information for:**

**Alternative Pathways of Human Islet Amyloid Polypeptide Aggregation Distinguished by  $^{19}\text{F}$  NMR-Detected Kinetics of Monomer Consumption**

Yuta Suzuki<sup>†</sup>, Jeffrey R. Brender<sup>‡</sup>, Kevin Hartman<sup>‡</sup>, Ayyalusamy Ramamoorthy<sup>‡†\*</sup> and E. Neil G. Marsh<sup>#†\*</sup>

*Departments of Chemistry<sup>†</sup>, Biological Chemistry<sup>#</sup> and Biophysics<sup>‡</sup>, University of Michigan, Ann Arbor, MI 48109*

Running title:  $^{19}\text{F}$  NMR measurements of amyloid fibrillogenesis

\*Corresponding authors

Dr. Neil Marsh

Department of Chemistry

University of Michigan

Ann Arbor MI 48109-1055

Tel 734 763 6096

FAX 734 615 3790

e-mail: nmarsh@umich.edu;

Dr. A. Ramamoorthy

Department of Chemistry

University of Michigan

Ann Arbor MI 48109-1055

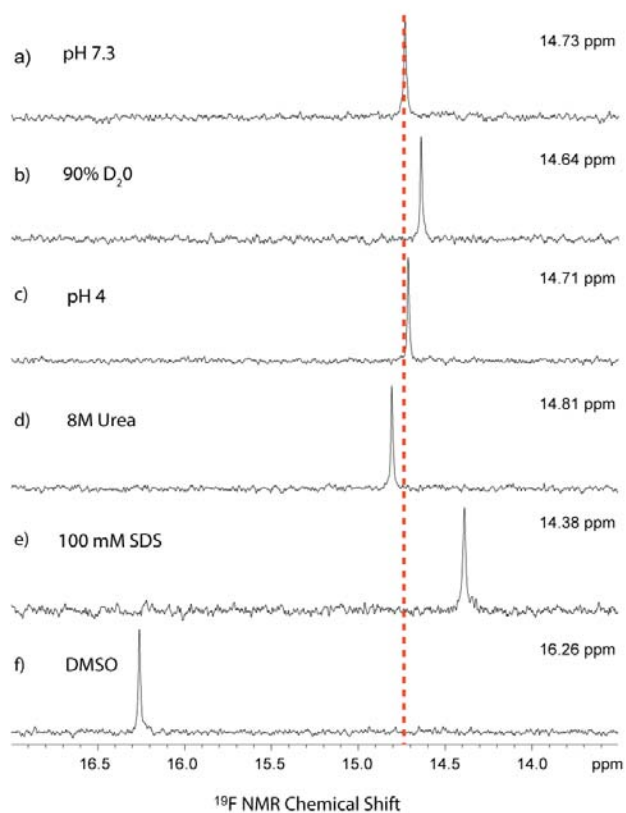
734 647 6572

734 615 3790

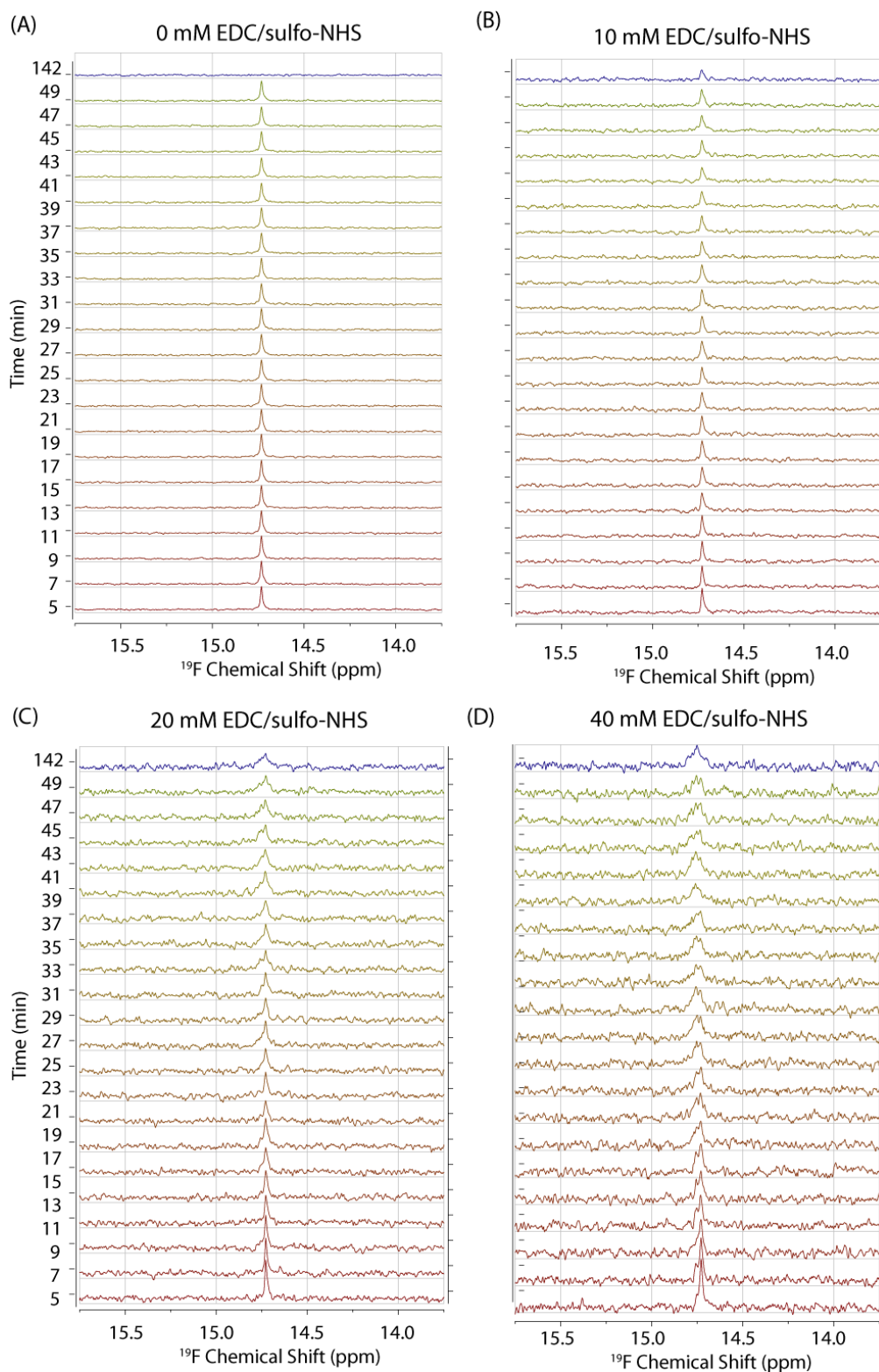
ramamoor@umich.edu

**Complete reference 78:**

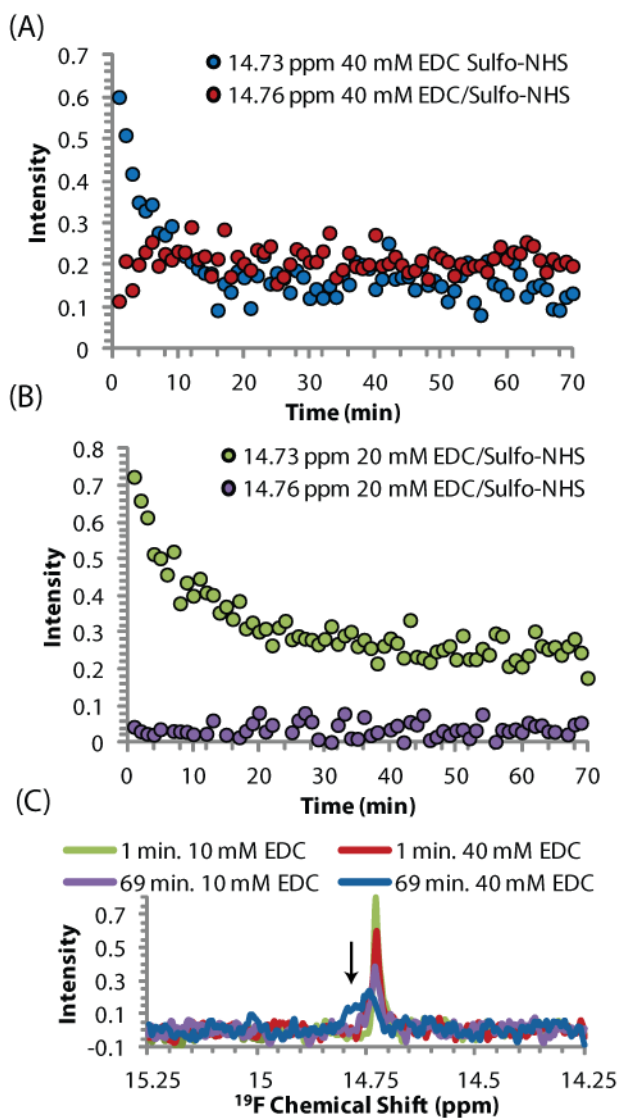
78. Miyata, M.; Sato, T.; Kugimiya, M.; Sho, M.; Nakamura, T.; Ikemizu, S.; Chirifu, M.; Mizuguchi, M.; Nabeshima, Y.; Suwa, Y.; Morioka, H.; Arimori, T.; Suico, M. A.; Shuto, T.; Sako, Y.; Momohara, M.; Koga, T.; Morino-Koga, S.; Yamagata, Y.; Kai, H. (2010) The Crystal Structure of the Green Tea Polyphenol (-)-Epigallocatechin Gallate-Transthyretin Complex Reveals a Novel Binding Site Distinct from the Thyroxine Binding Site. *Biochemistry* 49, 6104-6114.

**Additional supporting figures referred to in the main text**

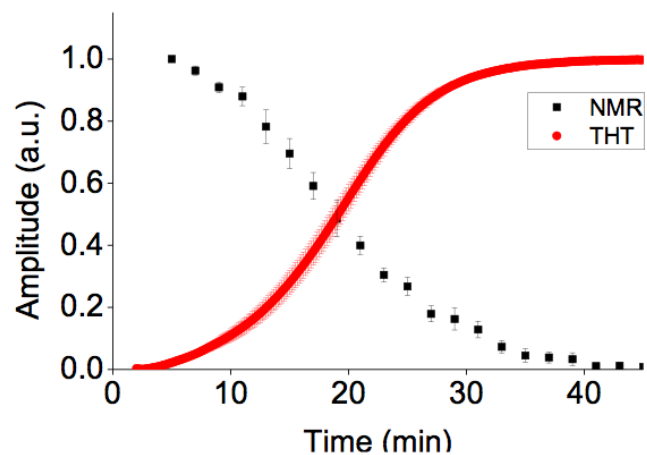
**Figure S1. Environmental sensitivity of the tfmF<sub>23</sub> group.**  $^{19}\text{F}$  NMR spectra of IAPP-tfmF<sub>23</sub> were acquired under various conditions after 5 minutes of incubation at 25 °C: **a)** 20 mM sodium phosphate, 20 mM NaCl, 10% D<sub>2</sub>O, pH 7.3 **b)** 90% D<sub>2</sub>O, 20 mM sodium phosphate, 20 mM NaCl, pH 7.3 **c)** 100  $\mu\text{M}$  HCl, 10% D<sub>2</sub>O, pH 4 **d)** 8M Urea, 10% D<sub>2</sub>O **e)** 100 mM SDS, 20 mM sodium phosphate, 20 mM NaCl, 10% D<sub>2</sub>O, pH 7.3, **f)** 90% DMSO, 10% D<sub>2</sub>O. Spectra were referenced to an external TFE standard (in solution of 20 mM Pi, 50 mM NaCl, pH 7.3) at 0 ppm.



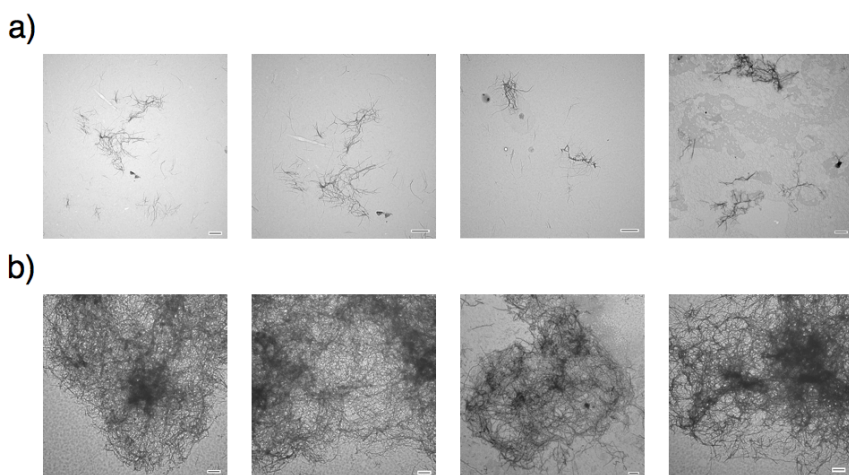
**Figure S2.**  $^{19}\text{F}$  NMR detection of small oligomer formation during cross-linking by EDC/Sulfo-NHS. Intensity change with time of IAPP-tfmF23 when incubated with 0 (A), 10 (B), 20 (C) and 40 mM EDC/Sulfo-NHS (D). A second peak at 14.76 ppm next to the main peak at 14.73 ppm can be seen developing in the 40 mM EDC/Sulfo-NHS sample with time.



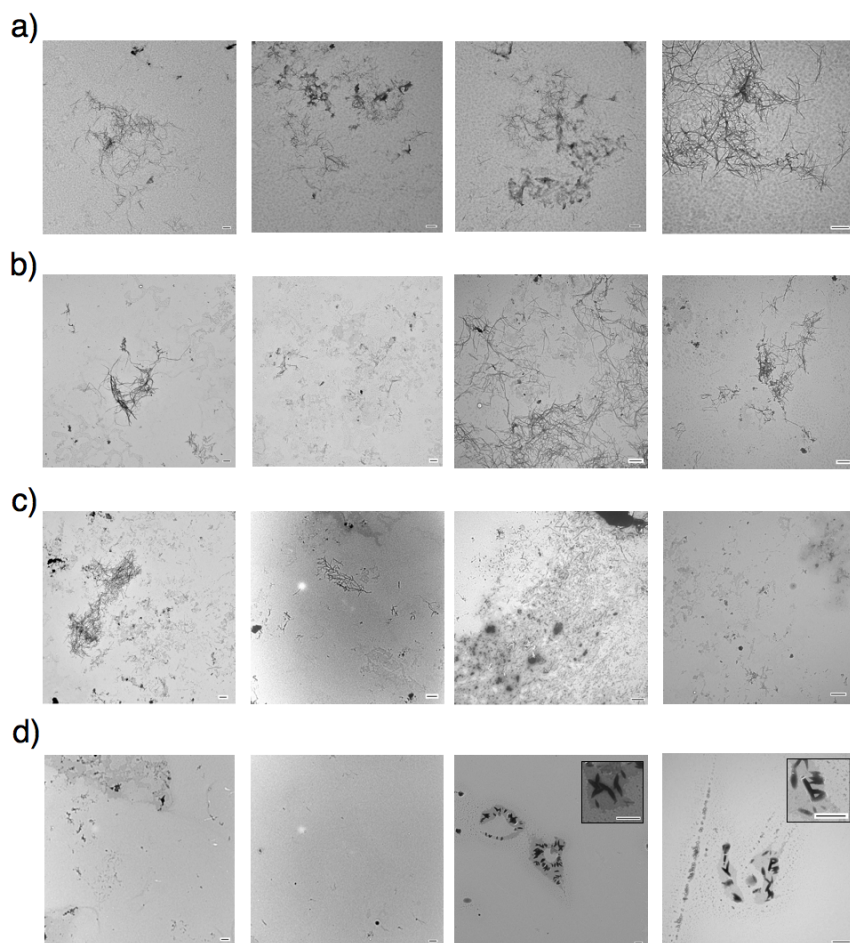
**Figure S3. Changes in the  $^{19}\text{F}$  spectra of IAPP-tfmF23 upon cross-linking.** **A and B:** Intensity change with time of the main peak at 14.73 ppm and the second peak at 14.76 ppm with 40 mM (A) and 10 mM (B) EDC/Sulfo-NHS. (C) Superposition of the first and final spectra of IAPP-tfmF23 with 40 and 10 mM EDC/Sulfo-NHS. The position of the second peak at 14.76 ppm is indicated by an arrow.



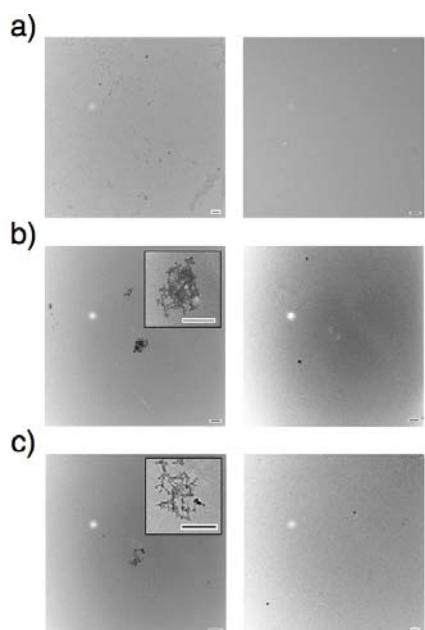
**Figure S4. Monomer consumption and fiber formation by IAPP-tfmF<sub>23</sub> at 37 °C.** Overlay of kinetic traces from <sup>19</sup>F NMR (black), ThT fluorescence (red). Error bars indicate S.E.M. (n=4). The close correspondence between the curves suggests fiber formation closely follows monomer consumption at 37 °C, similar to the measurements at 37 °C shown in Figure 2.



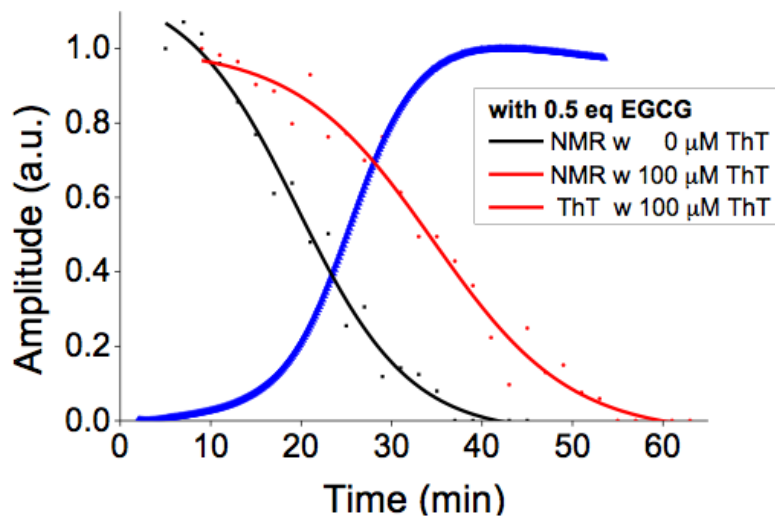
**Figure S5. Additional TEM images of IAPP-tfmF<sub>23</sub> fibers.** TEM images after ½ (c) and complete (d) depletion of the <sup>19</sup>F signal intensity at 25 °C. Scale bars represent 500 nm.



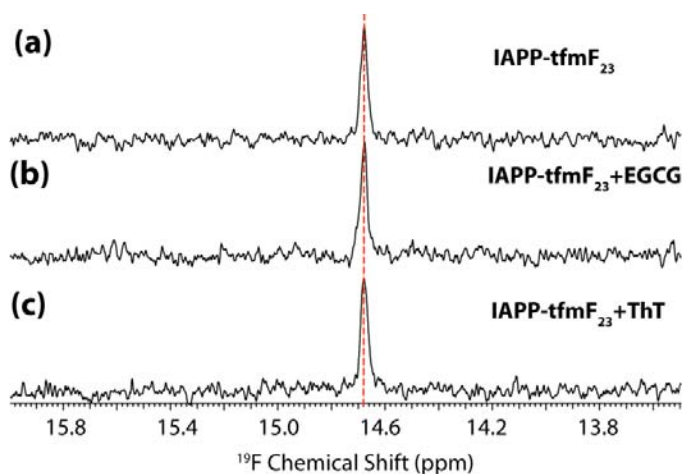
**Figure S6. Additional TEM images of IAPP-tfmF<sub>23</sub> aggregation as a function of EGCG concentration.** TEM images after the complete depletion of the <sup>19</sup>F signal for IAPP-tfmF<sub>23</sub> in the presence of (a) 0.2 eq, (b) 0.5 eq, (c) 1.0 eq, and (d) 5.0 eq of EGCG in the absence of ThT. Scale bars represent 500 nm.



**Figure S7. Aggregate formation by EGCG in the absence of IAPP-tfmF<sub>23</sub>.** TEM images for (a) phosphate buffer alone (20 mM Pi, 50 mM NaCl, pH 7.3), (b) 425  $\mu$ M EGCG in phosphate buffer, (c) 425  $\mu$ M EGCG with 100  $\mu$ M THT in phosphate buffer. Samples were incubated for 3 hours at 37 °C before acquisition. Scale bars represent 500 nm.

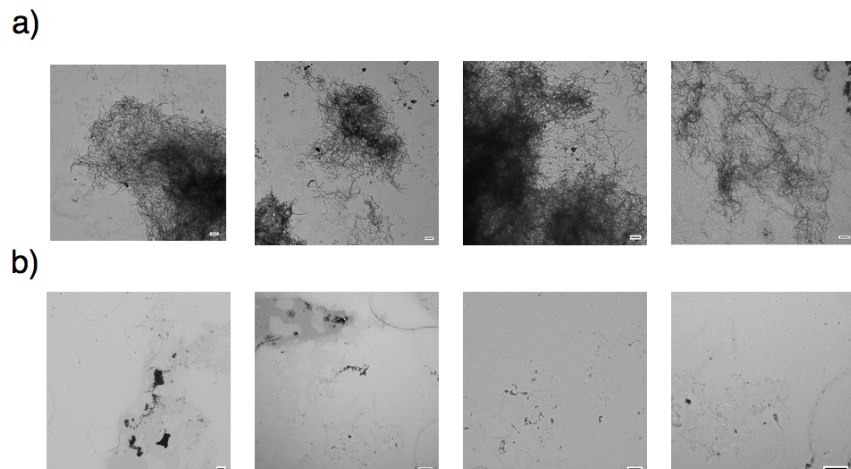


**Figure S8. Competition for amyloid binding between ThT and EGCG.** Overlay of kinetic traces from  $^{19}\text{F}$  NMR (black: 0  $\mu\text{M}$  ThT and red: 100  $\mu\text{M}$  ThT) and ThT fluorescence (blue: 100  $\mu\text{M}$  ThT) in the presence of 0.5 eq. of EGCG.



**Figure S9. EGCG and ThT do not likely interact with monomeric IAPP-tfmF<sub>23</sub>.**  $^{19}\text{F}$  NMR spectra of (a) IAPP-tfmF<sub>23</sub> alone, (b) IAPP-tfmF<sub>23</sub> with 85  $\mu\text{M}$  EGCG, (c) with 100  $\mu\text{M}$  ThT (c). Spectra were acquired at pH 7.3, 37°C after 5 minutes of incubation.





**Figure S10. EGCG disrupts preformed fibers of IAPP-tfmF<sub>23</sub>.** Additional TEM images acquired just before the addition of 5 equivalents of EGCG to preformed fibers of IAPP-tfmF<sub>23</sub> (a) and after complete depletion of ThT fluorescence intensity . Scale bar represents 500 nm.

Table S1

NMR/CD	ThT
$I = \frac{1}{1 + e^{\frac{x - t_{1/2}}{dx}}}$	$I = 1 - \frac{1}{1 + e^{\frac{x - t_{1/2}}{dx}}}$

<b>25°C</b>	<b>t<sub>1/2</sub></b>	<b>dx</b>
ThT	64.3±6.9	17.4±1.3
NMR	91.9±3.1	24.9±1.4
CD	91.8±6.3	22.8±1.4

<b>37°C</b>	<b>t<sub>1/2</sub></b>	<b>dx</b>
ThT	18.8±0.8	4.2±0.2
NMR	16.8±1.6	6.1±0.7