Copper Coordination to the Membrane Bound Form of α -Synuclein

Supplemental Figures

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Figure S1



Supplemental Figure 1. X-Band EPR spectra of membrane-bound α -syn (50 μ M α -syn) as a function of added Cu²⁺. Copper is increased at a fixed α -syn and lipid composition. With greater than 1.0 equivalent of copper, the spectra exhibit features of a superposition of membrane bound α -syn and lipid copper complexes, consistent with a single membrane-bound α -syn copper binding site with affinity sufficient to outcompete the lipids.

Figure S2



Supplemental Figure 2. Three-pulse ESEEM spectra of α -syn (50µM α -syn, 15mM lipids) with up to 10 equivalents of Cu²⁺, both in solution and in the presence of lipid SUV. The spectra of the samples in solution and in the presence of SUV with greater than 1 equivalent of Cu²⁺ reveal the expected quadrupolar transitions associated with the imidazole remote nitrogen and demonstrate coordination by His50 whereas the sample with stoichiometric concentrations of α -syn and Cu²⁺ in the presence of SUV do not. While the ¹⁴N quadrupolar features of the samples containing lipid SUV with 2, 5 and 10 equivalents of copper suggest a Cu²⁺-imidizole interaction, these peaks saturate only at copper concentrations well in excess of 250 µM – 500 µM copper, suggesting a very low affinity interaction.

Figure S3





Deconvolution (above) reveals the expected mass of 14283 as the major species (>90%) of the sample. Wild-type α -syn has a mass of 14456.

Figure S4



Supplementary Figure 4: CD spectra of wild-type α -syn, α -syn N-trunc, and α -syn H50A.

Figure S5



Supplementary Figure 5: Transmission Electron Microscopy images of

70%POPC:30%POPG SUVs both without α -syn (left) and with α -syn (right, lipid: α -syn ratio 300:1). The black bar at lower left is equivalent to 100nM. Most SUVs are in the range 20 – 50 nm, with no indication of tubes or other unusual structures. Samples for electron microscopy were prepared by spotting 3µl onto glow-discharged carbon-coated copper grids followed by staining with 2% (w/v) uranyl acetate solution. Samples were analyzed using a JEOL 1230 microscope operating at 120kV.