

Supporting Information

of

Optimization of Brush-like Cationic Copolymers for Non-viral Gene Delivery

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1. Table S1 & S2

2. Figure S1-S5

Table S1. Summary of P(GMA)₅₀-*b*-P(HPMA) prepared at different polymerization time using P(GMA)₅₀ as a macro-CTA.

Condition	No.	Time (min)	Conv. (%)	Real structure determined by NMR
	1	40	5.8	P(GMA) ₅₀ - <i>b</i> -P(HPMA) ₁₈
[M]:[Macro-CTA]:[I] = 300:1:0.33	2	90	9.9	P(GMA) ₅₀ - <i>b</i> -P(HPMA) ₃₃
[M] = 1.0 M, T = 70 °C	3	280	16.8	P(GMA) ₅₀ - <i>b</i> -P(HPMA) ₄₉
	4	480	27.2	P(GMA) ₅₀ - <i>b</i> -P(HPMA) ₈₃

Table S2. Summary of P(GMA)₅₀-*b*-P(OEGMA) prepared at different polymerization time using P(GMA)₅₀ as a macro-CTA.

Condition	No.	Time (min)	Conv. (%)	Real structure determined by NMR
	1	10	3.0	P(GMA) ₅₀ - <i>b</i> -P(OEGMA300) ₆
[M]:[Macro-CTA]:[I] = 200:1:0.33,	2	20	5.5	P(GMA) ₅₀ - <i>b</i> -P(OEGMA300) ₁₁
[M] = 1.0 M, T = 70 °C	3	30	7.5	P(GMA) ₅₀ - <i>b</i> -P(OEGMA300) ₁₅
	4	40	11.5	P(GMA) ₅₀ - <i>b</i> -P(OEGMA300) ₂₃

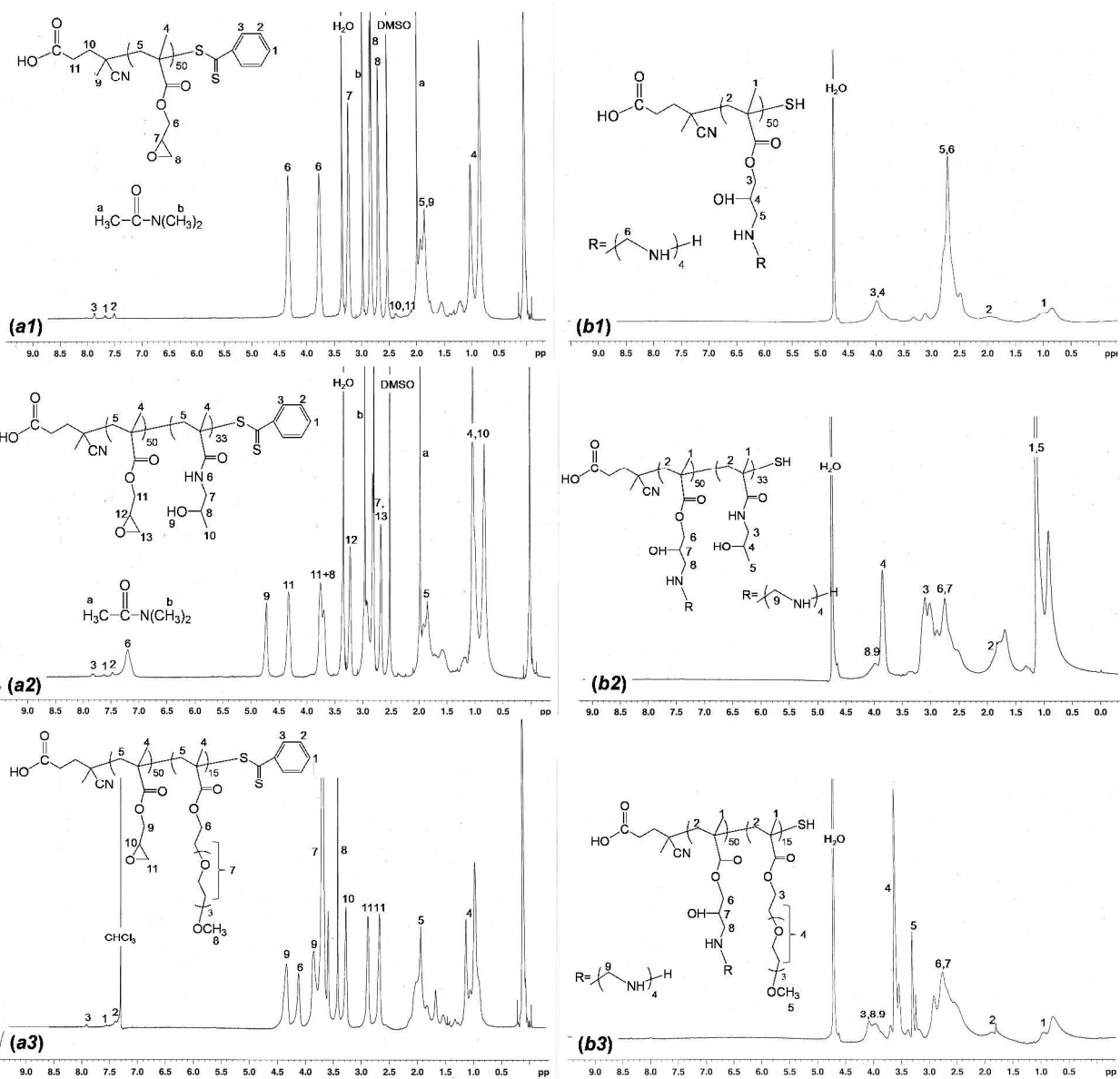


Figure S1. ^1H NMR spectra of (a1) P(GMA)₅₀ in DMSO; (a2) P(HPMA)₃₃-b-P(GMA)₅₀ in DMSO; (a3) P(OEGMA)₁₅-b-P(GMA)₅₀ in CDCl₃; (b1) P(GMA-TEPA)₅₀ in D₂O; (b2) P(HPMA)₃₃-b-P(GMA-TEPA)₅₀ in D₂O; (b3) P(OEGMA)₁₅-b-P(GMA-TEPA)₅₀ in D₂O.

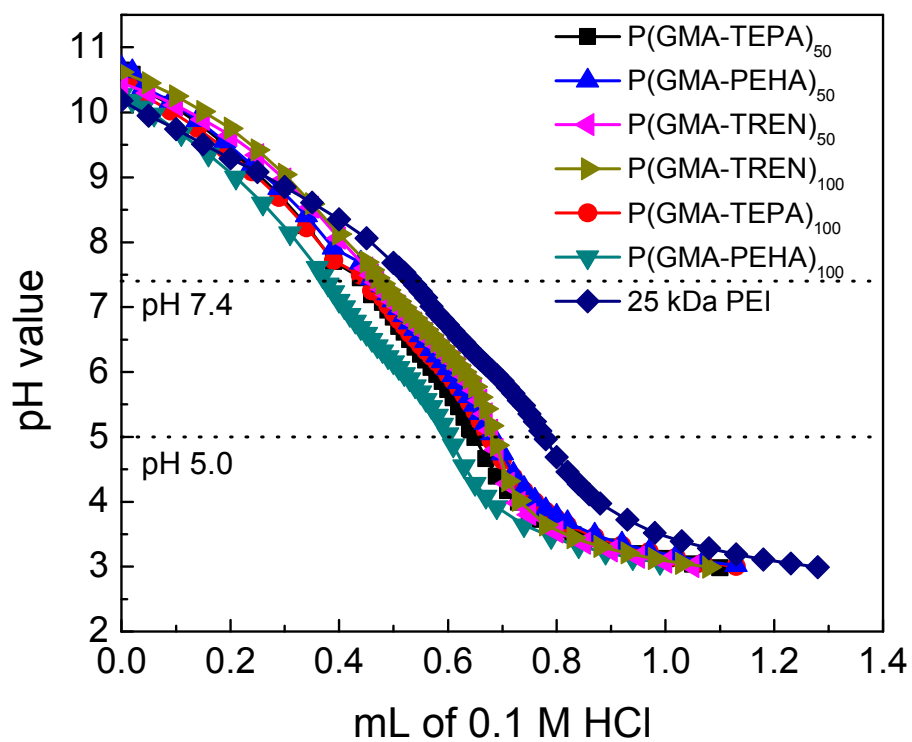


Figure S2. Buffering capacity of various P(GMA-oligoamine)_{50/100} obtained by titrating polymer aqueous solution (0.2 mg/mL) in 0.15 M aqueous NaCl (pH 10, adjusted with NaOH) with 0.1 M HCl. As a reference, the titration curve of 25 kDa bPEI is also presented.

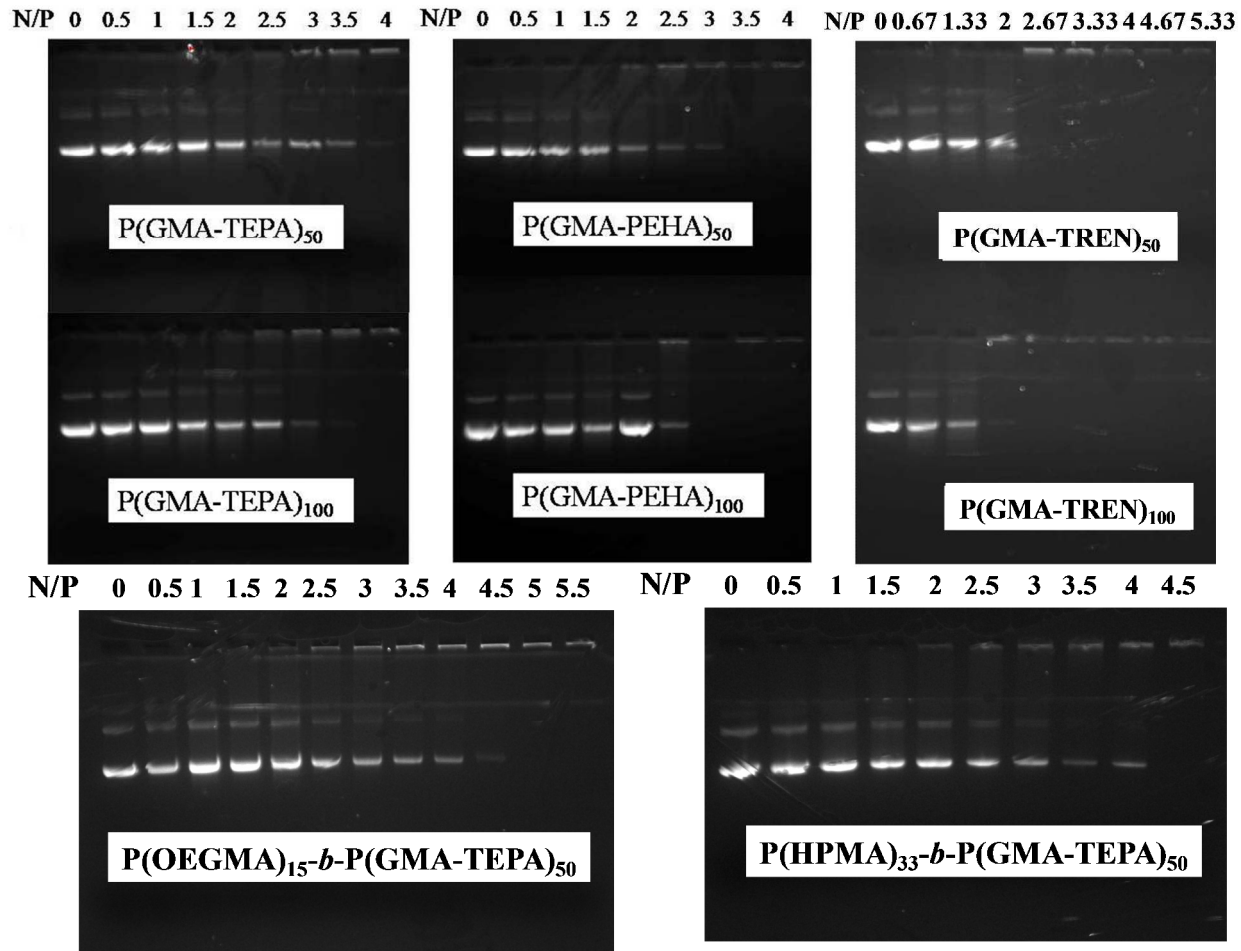


Figure S3. Agarose gel electrophoresis of various P(GMA-oligoamine) polyplexes formed by complexation with plasmid DNA at N/P ratios ranging from 0/1 to 5.5/1.

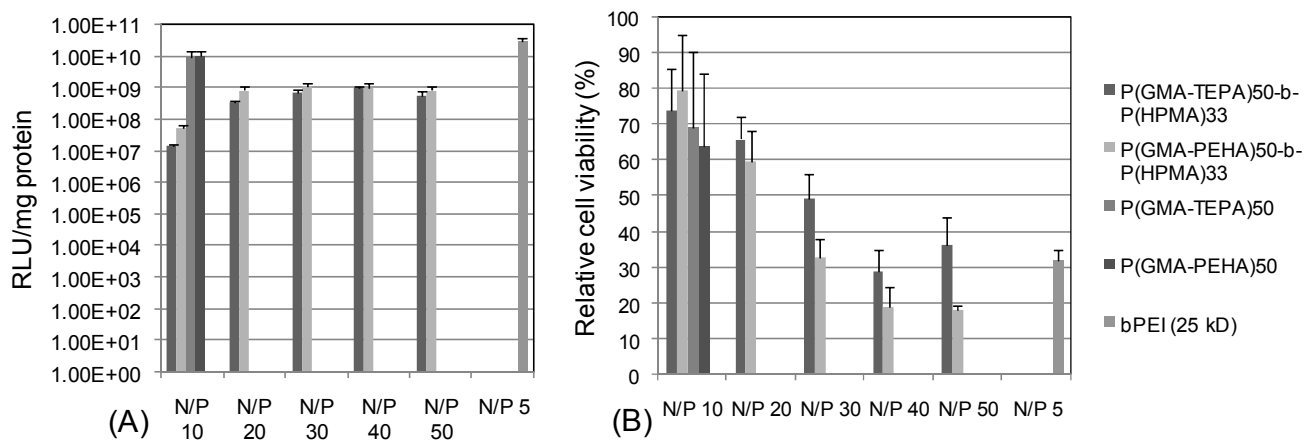


Figure S4. Transfection efficiency (A) and relative cell viability (B) of polyplexes formed by P(HPMA)₃₃-*b*-P(GMA-TEPA/PEHA)₅₀. Data are shown as mean ± SD (n = 3).

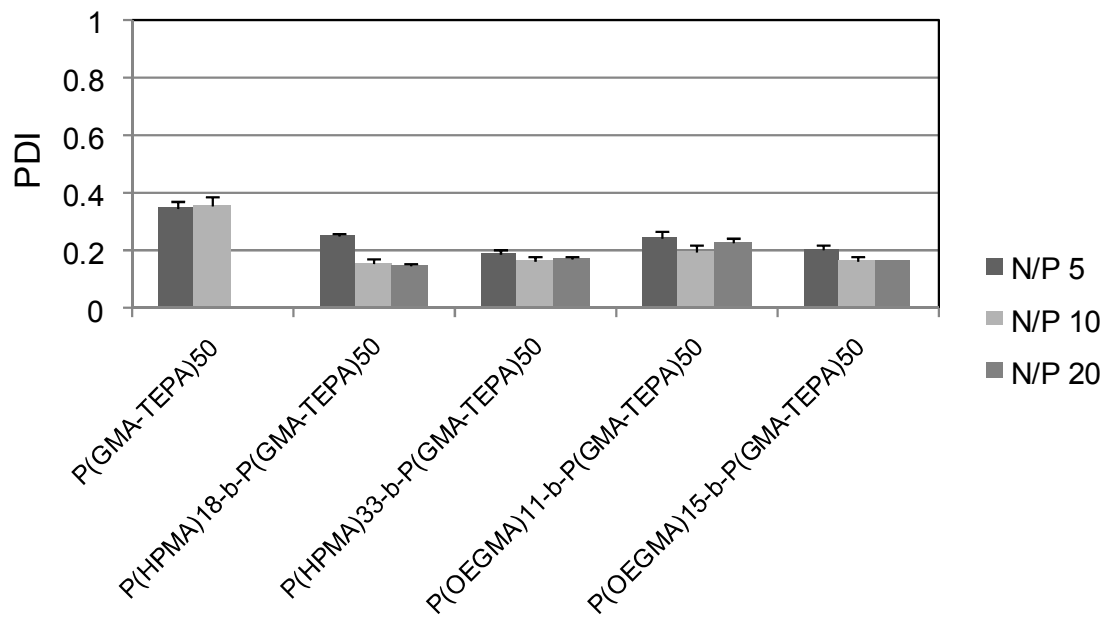


Figure S5. Average size distributions (PDIs) of various polyplexes formed at different N/P ratios in 150 mM PBS.