Supplementary Material and Methods

Homing of CTO-stained dDCs

EpCAM negative dDCs were isolated from wt and Mrp14 -/- mice 48h after challenge with DNFB and stained with cell tracker orange (CTO; 5µM; Molecular Probes, Darmstadt, Germany). $9x10^6$ of the stained wt and Mrp14 -/- cells were injected in the tail veins of two identical groups of DNFB-sensitized wt mice prior to challenge. After 48h the immigration of CTO positive cells into ear-draining LNs was determined by flow cytometry.

FITC application

Ears of naïve C57BL/6 wt and Mrp14 -/- mice were painted with 30µl 0.5% FITC in an acetone/dibutylphthalate mixture (Sigma-Aldrich, Munich, Germany). After 18h ear-draining LNs were analyzed for immigrated FITC positive cells by flow cytometry.

Supplementary Figure legends

Figure S1. Histological analyses of wt and Mrp14 -/- ear sections from control ears, treated with acetone/olive oil only. Sections were stained for immune cells ($CD11b^+$ myeloid cells, $Gr\text{-}1^+$ granulocytes, $F4/80^+$ macrophages) as well as for Mrp8 and Mrp14. Scale bar: 250µm.

Figure S2. Identical cellular homing capacities of wt and Mrp14 -/- dDCs. **(A)** EpCAM negative dDCs were isolated from wt and Mrp14 -/- mice 48h after challenge with DNFB and stained with CTO. Subsequently, two identical groups of DNFB-sensitized wt mice were injected intravenously with these CTO-stained wt or Mrp14 -/- dDCs ($9x10^6$ dDCs per mouse) immediately before challenge with 0.4% DNFB. After 48h the immigration of CTO positive dDCs into ear-draining LNs was determined by flow cytometry. **(B)** Ears of naïve C57BL/6 wt and Mrp14 -/- mice were painted with 30µl 0.5% FITC. After 18h ear-draining LNs were analyzed for immigrated FITC positive cells by flow cytometry.

Figure S1

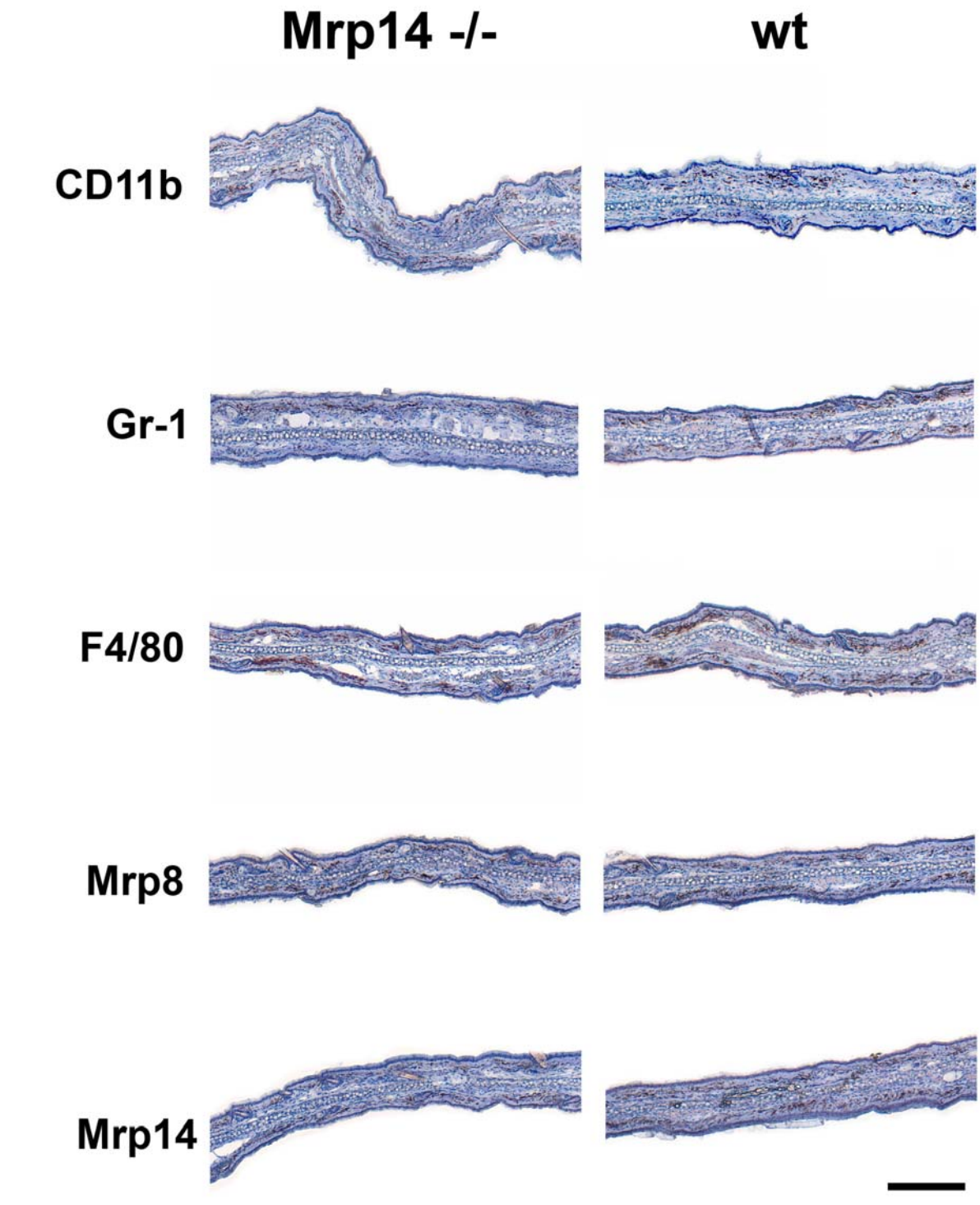
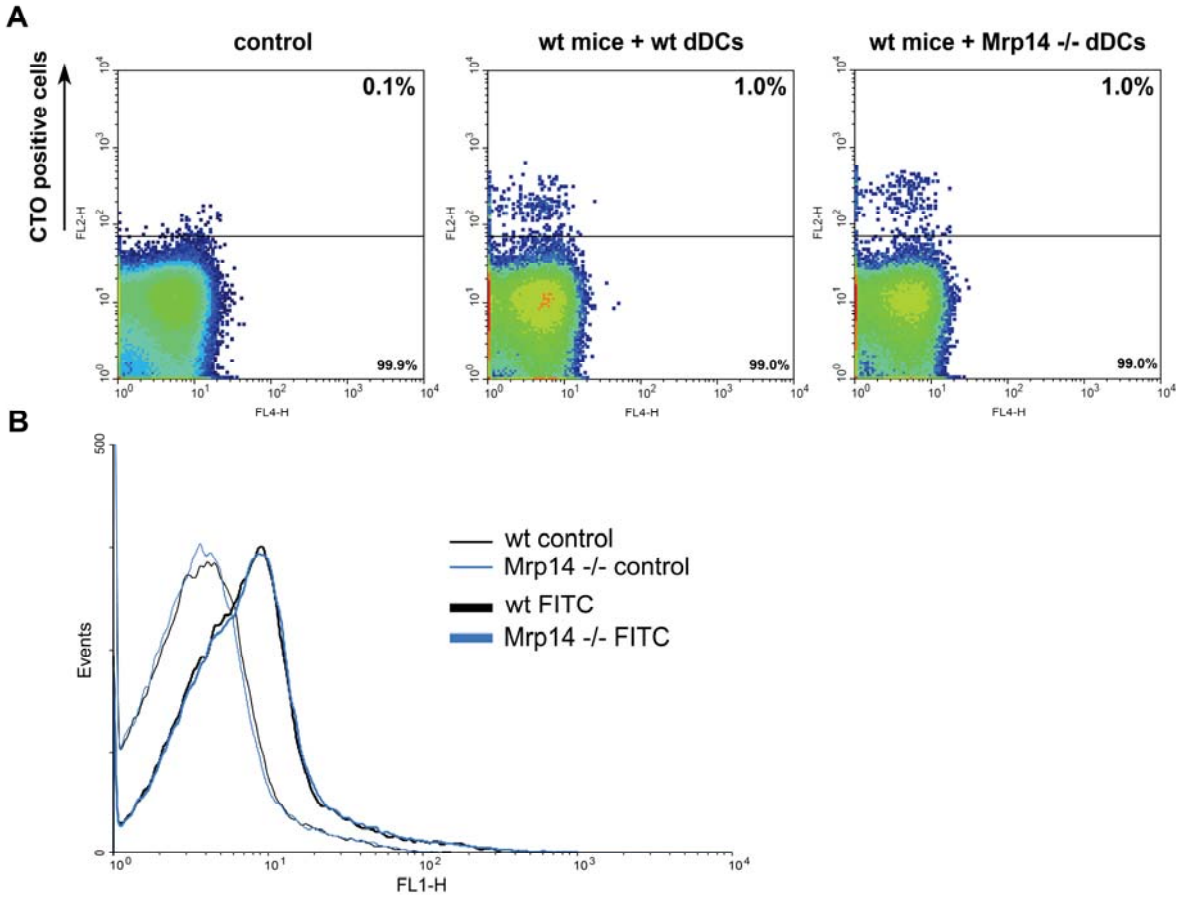


Figure S2


A
control
wt mice + wt dDCs
wt mice + Mrp14 -/- dDCs
CTO positive cells
0.1%
1.0%
1.0%
99.9%
99.0%
99.0%
FL2-H
FL4-H
B
wt control
Mrp14 -/- control
wt FITC
Mrp14 -/- FITC
Events
FL1-H