

Supplementary Materials for

Extracellular Ca^{2+} Acts as a Mediator of Communication from Neurons to Glia

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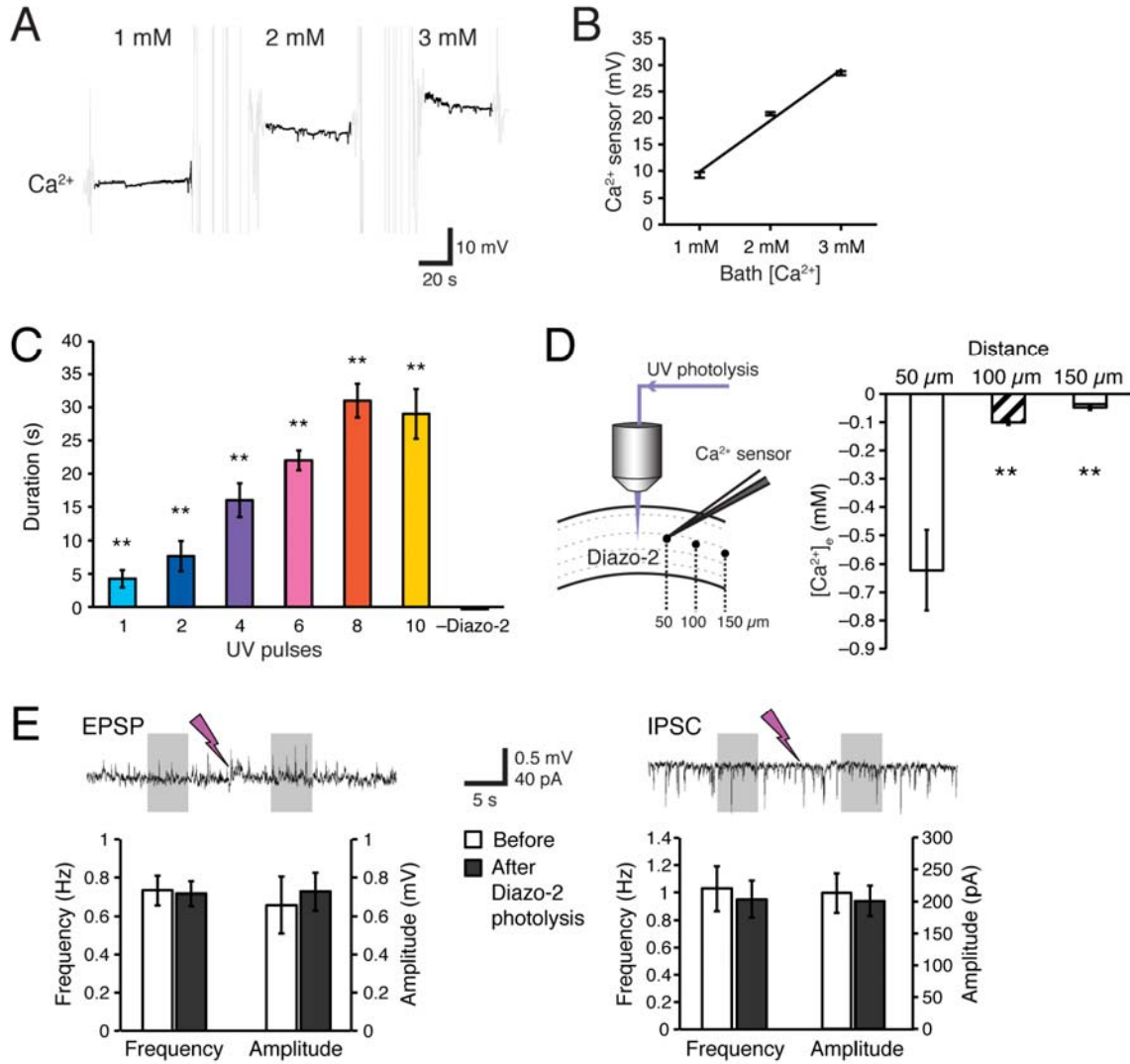


Fig. S1. Calibration of Ca^{2+} -sensitive microelectrode and effects of diazo-2 photolysis on $[\text{Ca}^{2+}]_e$

(A) Representative trace of calibration of a Ca^{2+} -sensitive microelectrode. Extracellular Ca^{2+} in the bath was successively increased from 1, to 2, to 3 mM Ca^{2+} in aCSF. (B) Calibration of Ca^{2+} sensitive microelectrodes in aCSF ($n=3$ microelectrodes, $r^2=0.986$). The electrodes used in experiments typically showed voltage responses of 5-10 mV per mM increase in Ca^{2+} concentration in the concentration range studied. Electrode calibration was done before and after each experiment. (C) Duration of diazo-2-induced decrease in $[\text{Ca}^{2+}]_e$ plotted as a function of stimulus intensity ($n=3$ photolysis events, UV pulses 1-10, and 10 pulses without diazo-2, **, $p<0.01$). (D) Diagram showing recordings obtained by placing the electrode at increasing distances from the photolysis site. Diazo-

2-induced reduction in $[Ca^{2+}]_e$ detected with the Ca^{2+} -sensitive microelectrode placed 50, 100, or 150 μm from the site targeted by photolysis. (n=8 photolysis events, **, $p < 0.01$ compared to 50 μm). (E) Traces show representative recordings of spontaneous EPSPs and IPSCs in hippocampal CA1 neurons. Bar histograms compare the frequency and amplitude of EPSPs and IPSCs 5 s before or 5 s after photolysis (no significant differences in EPSP or IPSC frequency and amplitude was identified using paired t-test, before vs. after, $p > 0.05$, n = 7).

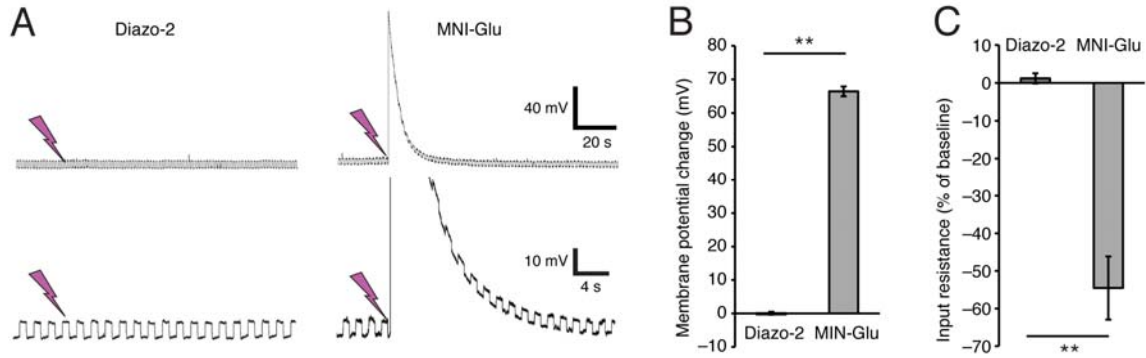


Fig. S2. CA1 pyramidal neurons depolarize and decrease their input resistance in response to photolysis of MNI-glutamate, whereas photolysis of diazo-2 is not associated with changes in the membrane properties

(A) Representative current-clamp recordings from a hippocampal CA1 pyramidal cell located $\sim 70 \mu\text{m}$ from the site of photolysis of diazo-2 (left panel) or MNI-glutamate (right panel). Lower traces show expanded timescale. (B) Comparison of changes in plasma membrane potential in response to photolysis of diazo-2 and MNI-glutamate in CA1 pyramidal neurons ($n=4-11$ photolysis events, **, $p<0.001$). (C) Comparison of relative changes in input resistance in response to photolysis of diazo-2 and MNI-glutamate in CA1 pyramidal neurons ($n=4-5$ photolysis events, **, $p<0.001$).

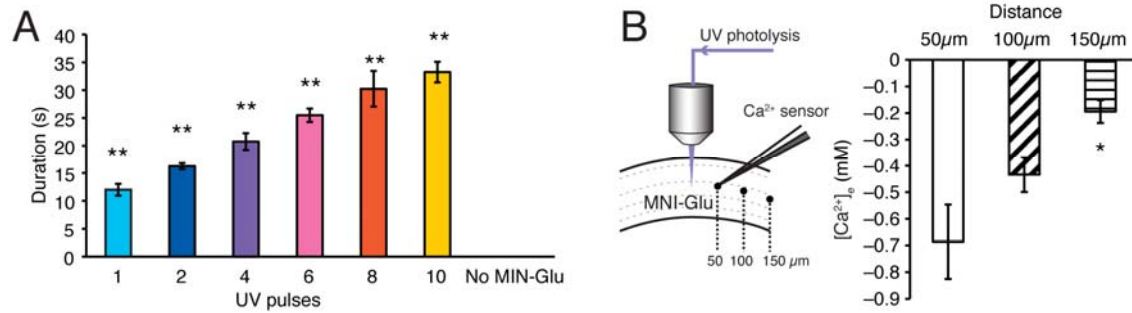


Fig. S3. Photolysis of MNI-glutamate

(A) Duration of the decrease in $[Ca^{2+}]_e$ produced by photolysis of MNI-glutamate plotted as a function of number of UV pulses used for photolysis ($n=5$ photolysis events, **, $p<0.01$). **(B)** Diagram of recordings obtained by placing the electrode at increasing distances from the photolysis site. MNI-glutamate-induced reduction in $[Ca^{2+}]_e$ detected with the Ca^{2+} -sensitive microelectrode placed 50, 100, or 150 μm from the site targeted by photolysis ($n=4-15$ photolysis events. *, $p<0.05$ compared to 50 μm). If MNI-glutamate was omitted, UV pulses failed to decrease $[Ca^{2+}]_e$.

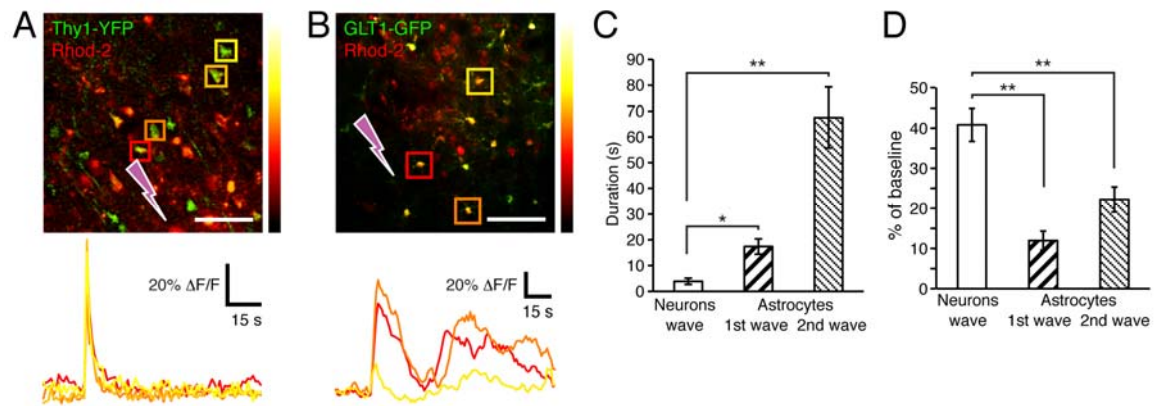


Fig. S4. Transgenic reporter mice reveal distinct responses to MNI-glutamate photolysis of neurons and astrocytes in hippocampal slices

(A) Hippocampal slice from a Thy1-YFP reporter mouse loaded with the Ca²⁺ indicator rhod-2 am. YFP+ neurons show an increase in Ca²⁺ during the 1st, but not the 2nd Ca²⁺ wave. Scale bar, 50 μ m. (B) Hippocampal slice from a GLT1-GFP reporter mouse loaded with the Ca²⁺ indicator rhod-2 am. GFP+ astrocytes show an increase in Ca²⁺ during both the 1st and the 2nd Ca²⁺ waves. Scale bar, 50 μ m. (C) Duration of the increase in Ca²⁺ in astrocytes and neurons during the 1st and the 2nd Ca²⁺ wave (n=12-13 photolysis events, *, p<0.05, **, p<0.01). (D) The amplitude of the increases in Ca²⁺ in astrocytes or neurons during the 1st or the 2nd Ca²⁺ wave (n=10 photolysis events, **, p<0.01).

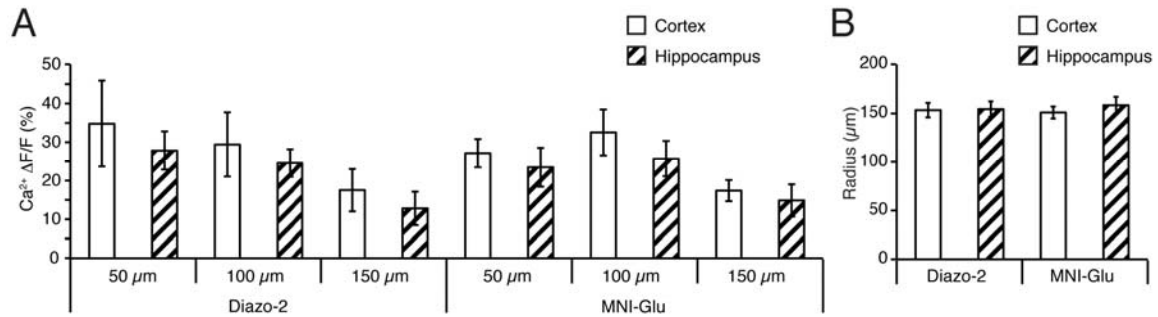


Fig. S5. Comparison of Ca²⁺ waves in hippocampus and cortex evoked by photolysis of MNI-glutamate or diazo-2

(A) Comparison of Ca²⁺ increases in astrocytes during the 2nd Ca²⁺ wave evoked by photolysis of MNI-glutamate and the Ca²⁺ wave evoked by photolysis of diazo-2 at 50, 100, and 150 μm in cortex and hippocampus (n=9 photolysis events, p>0.05). (B) Comparison of maximal radius of the Ca²⁺ waves in astrocytes evoked by photolysis of MNI-glutamate and diazo-2 in cortex and hippocampus (n=10 photolysis events, p>0.05).

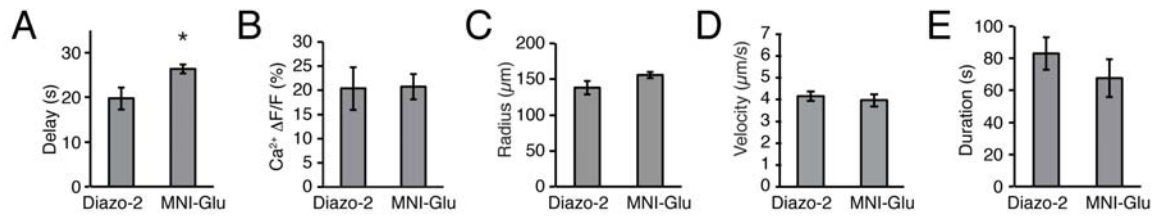


Fig. S6. Comparison of Ca²⁺ waves evoked by diazo-2 or MNI-glutamate photolysis

All comparisons are made between Ca²⁺ waves evoked by diazo-2 photolysis and the 2nd Ca²⁺ wave evoked by MNI-glutamate. **(A)** Histogram comparing the delay between photolysis and the first increase in Ca²⁺ evoked by diazo-2 or MNI-glutamate photolysis (n=20-22 photolysis events, *, p<0.05). **(B)** Ca²⁺ increase at 75 μm evoked by diazo-2 or MNI-glutamate photolysis (n=16 photolysis events). **(C)** Radius of Ca²⁺ waves evoked by diazo-2 or MNI-glutamate photolysis (n=26 photolysis events). **(D)** Velocity of Ca²⁺ waves evoked by diazo-2 or MNI-glutamate photolysis (n=18 photolysis events). **(E)** Duration of Ca²⁺ increases in astrocytes evoked by diazo-2 or MNI-glutamate photolysis (n=12 photolysis events).

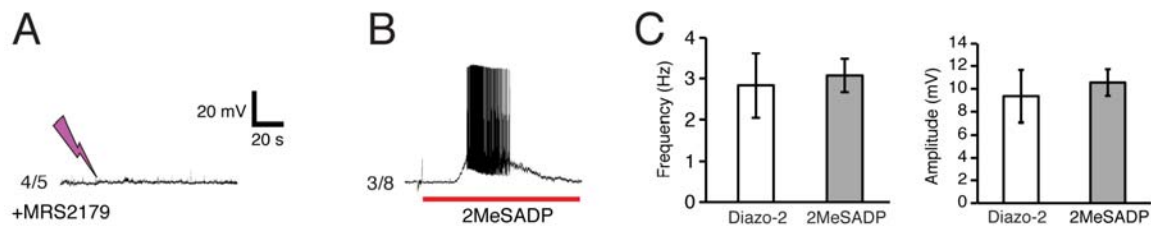


Fig. S7. Effect of manipulation of P2Y1 receptors in hippocampal interneurons

(A) The P2Y1 receptor antagonist, MRS2179 (50 μ M) blocked depolarization and bursting activity induced by diazo-2 photolysis in 4 of 5 interneurons. One of the five interneurons exhibited a transient minor depolarization, but no action potential firing. (B) Exposure to the P2Y1 receptor agonist 2MeSADP (100 μ M) induced depolarization in a total of 8 interneurons tested, whereof 3 exhibited bursting activity. (C) Histograms compare the frequency of interneuronal action potential firing, as well as the amplitude of membrane depolarization induced by photolysis of diazo-2 with or without exposure to 2MeSADP (n = 6-8 photolysis events).