

Supplementary Data for

THE HEME BINDING PROPERTIES OF GLYCERALDEHYDE-3-PHOSPHATE DEHYDROGENASE

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Fig. S1. Gel filtration of the rGAPDH-heme complex. Panel A: Chromatogram of the GAPDH-heme complex. Fractions were collected between 8 and 65 min and analyzed by UV-vis spectroscopy. Panel B: UV-visible analysis of the fractions. The ratio A_{280}/A_{415} of the complex prior to injection was 3.5. The ratio A_{280}/A_{415} of the eluted fractions containing the highest concentration of heme ranged from 5 to 8.

Fig. S2. In-gel heme staining of apo- and heme-bound GAPDH using the o-dianisidine/ H_2O_2 peroxidase assay. Panel A shows the heme-staining bands corresponding to rGAPDH-heme (5:1) and apo-GAPDH (negative control). Panel B shows different doses of GAPDH-heme, and iNOSoxy as a positive control for specific heme staining.

Fig. S3. Far-UV CD spectra of apo- and heme-bound GAPDH. Addition of heme to apo-GAPDH produced mild changes in the far-UV CD spectrum, suggesting that heme binding is accompanied by modest conformational changes.

Fig. S4. Spectrophotometric titrations of rabbit GAPDH with increasing doses of hemin. Panel A. Titration of 10 μ M GAPDH with hemin (0-10 μ M). Panel B: UV-visible spectra of free hemin in buffer.

Fig. S5. Transfer of ferrous heme from GAPDH to apomyoglobin at 25 °C. Heme transfer to apomyoglobin is accompanied by a decrease in absorption at 425 nm (GAPDH-heme) and a concomitant increase at 434 nm (reduced myoglobin). The inset shows the initial and final spectra.

Fig. S6. Reaction of native, anaerobic GAPDH with ferrous heme (Panels A and B), and reaction of GAPDH pre-treated with dithionite and ferrous heme (Panel C). The reaction of GAPDH with ferrous heme (5:1) was slow ($k_{\text{obs}} = 0.0091 \text{ s}^{-1}$ at 10 °C) and incomplete; only 19% of the total available ferrous heme formed a complex with GAPDH after 10 min. No measurable spectral changes were detected for the reaction between dithionite-treated GAPDH and ferrous heme.

Fig. S7. Kinetics of CO binding to two major subpopulations of heme-bound to rGAPDH (2:1, GAPDH:heme, 10 °C). Formation of a Fe(II)-CO complex with a Soret absorption maximum at 410 nm occurs ~4-fold faster than the formation of the 6-coordinate Fe(II)-CO-GAPDH with a characteristic band at 420 nm. The nature of the species absorbing at 410 nm is currently unknown.

Fig. S8. UV-visible spectra of purified human GAPDH (Panel A) or *S. suis* (Panel B) GAPDHs from cultures grown in the presence or absence of the heme synthesis precursor δ -amino levulinic acid.

Fig. S9. Amino acid sequence alignment of *H. sapiens* (Accession P04406.3), *O. cuniculus* (Accession P46406.3) and *S. suis* (Accession AAN86058.1) GAPDHs. The alignment was generated with ClustalW 2.1 software (<http://www.ebi.ac.uk/Tools/msa/clustalw2/>)

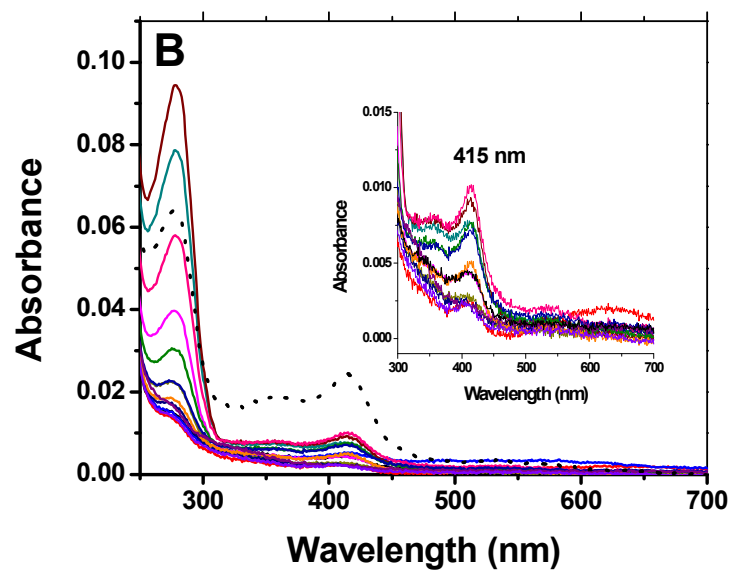
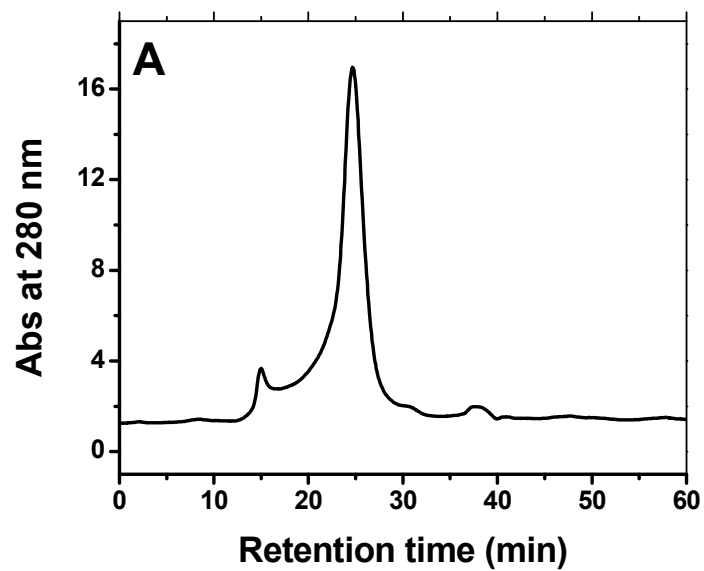
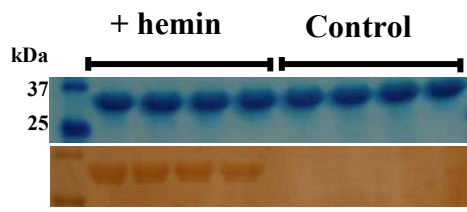


Figure S1

A



B

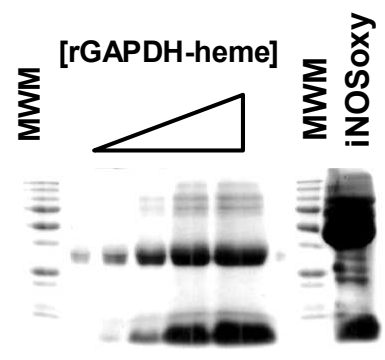


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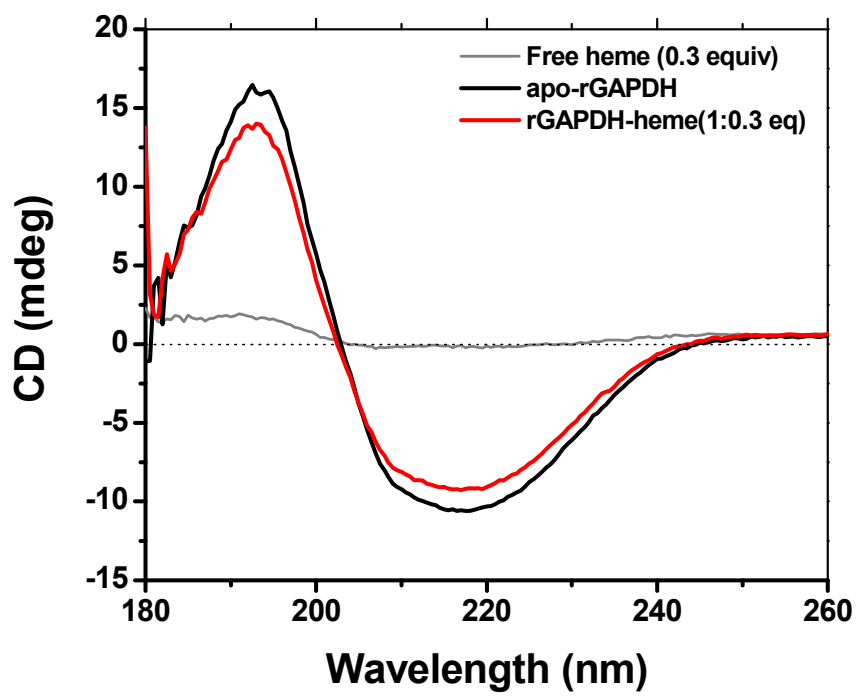


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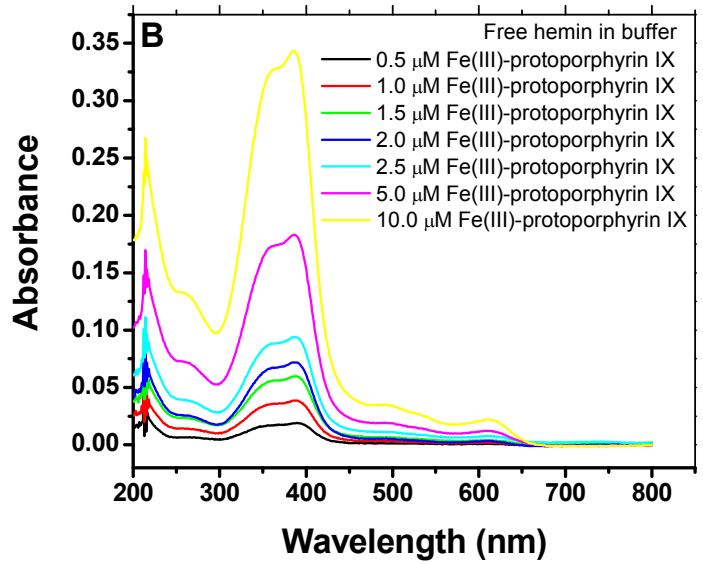
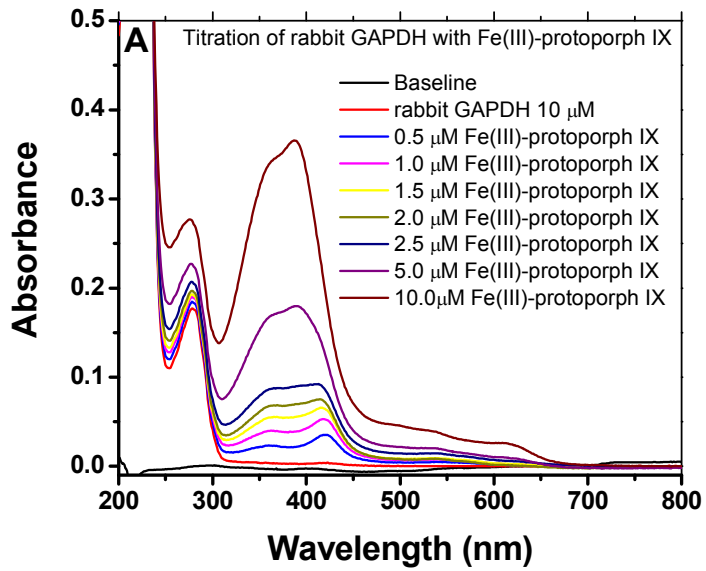


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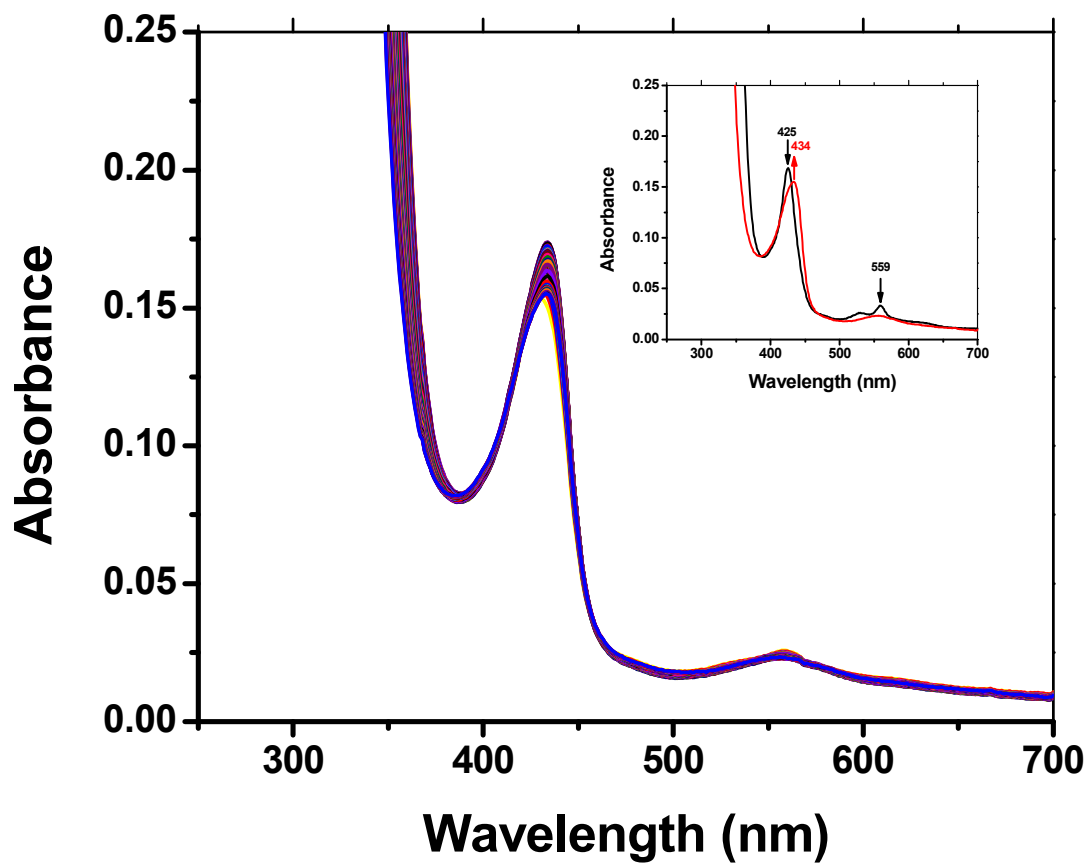


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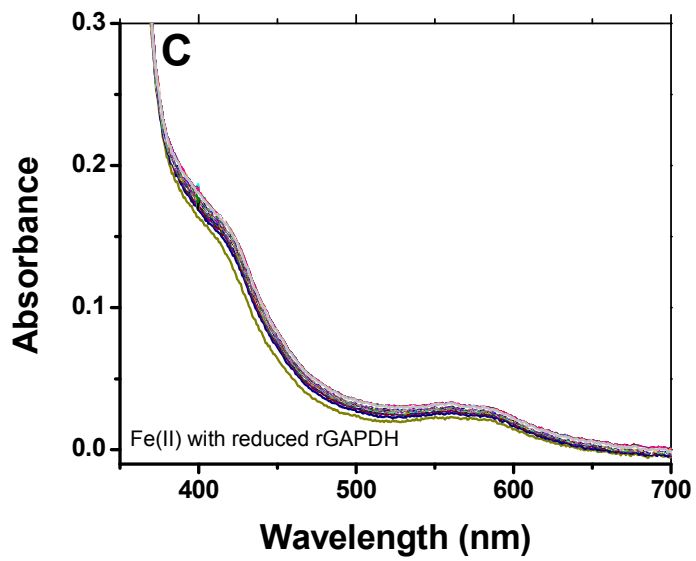
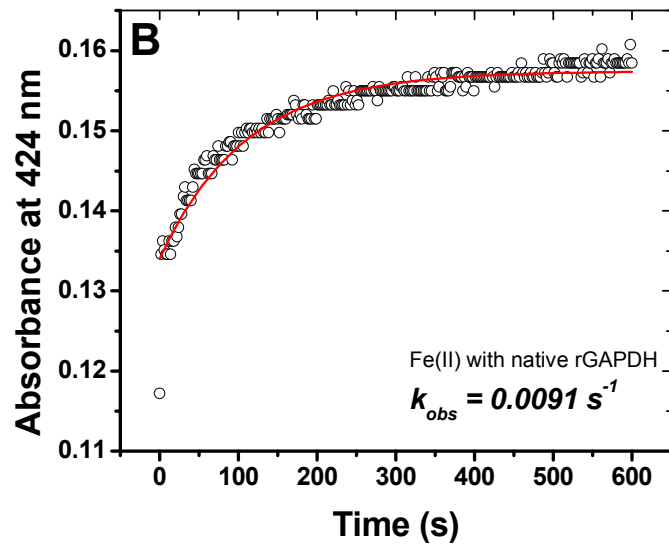
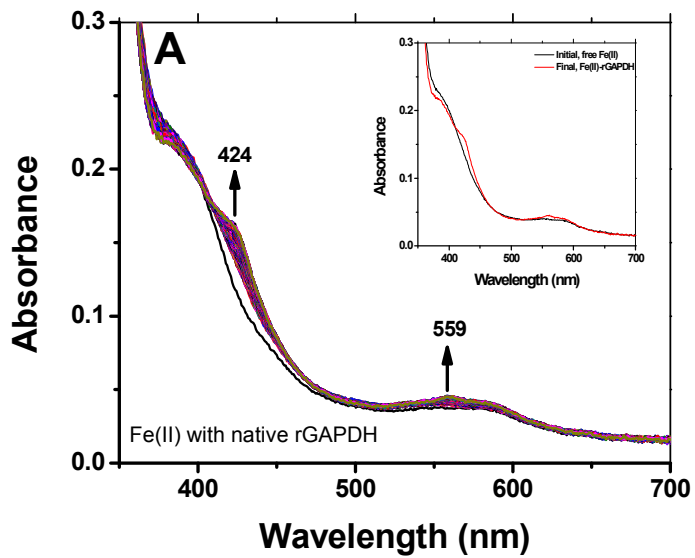


Figure S6

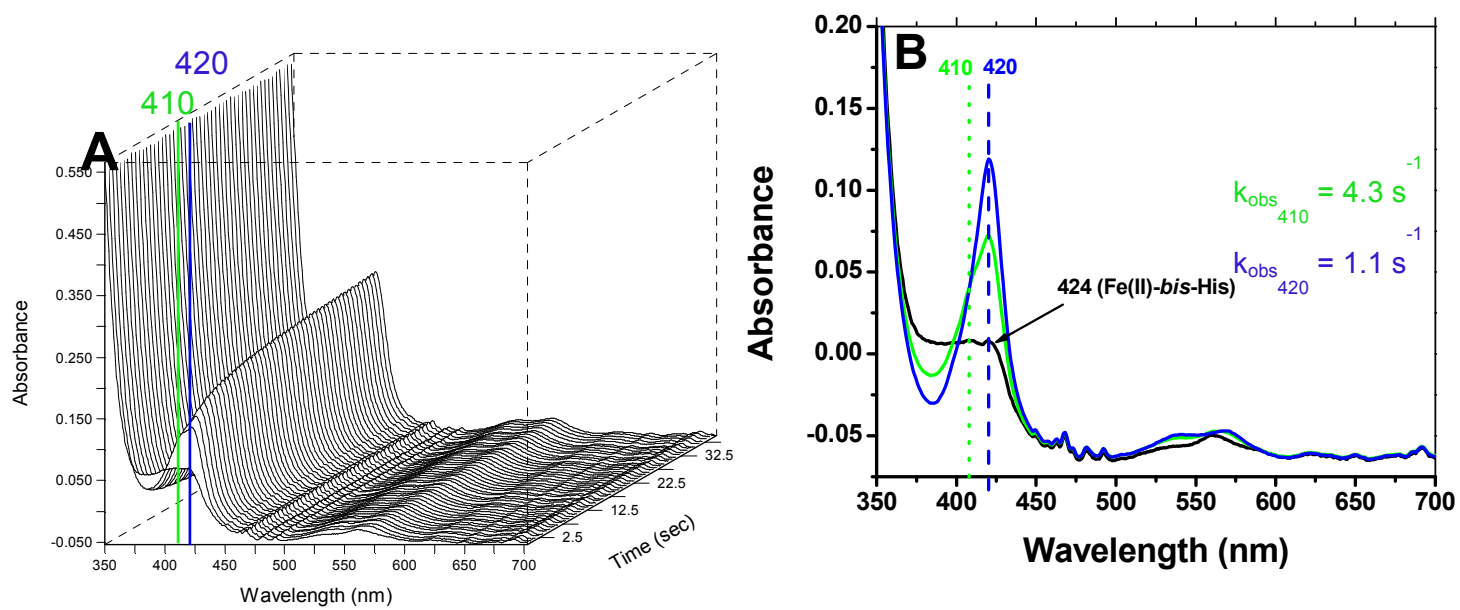


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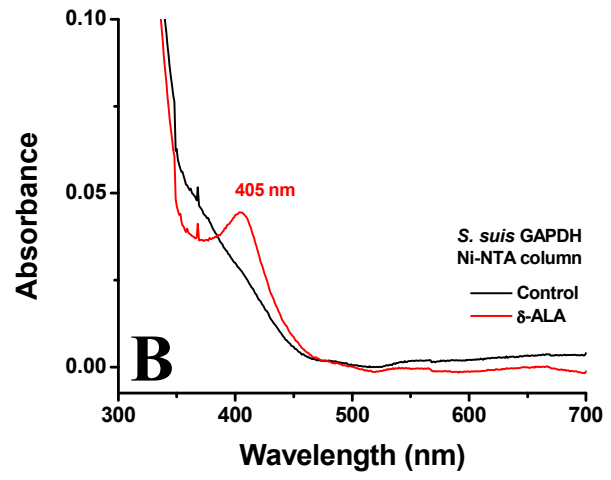
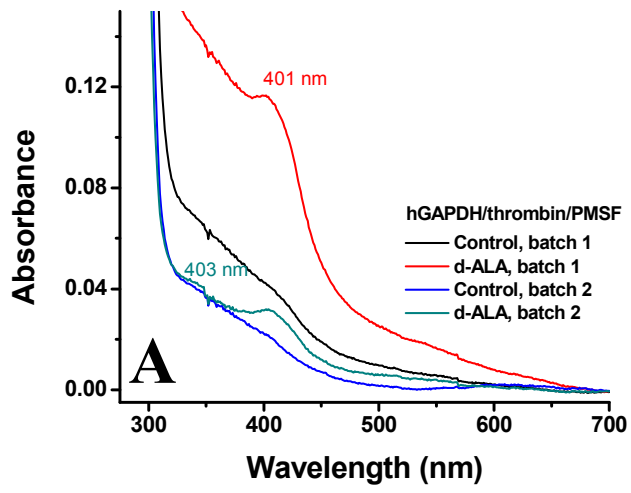


Figure S8

CLUSTAL 2.1 multiple sequence alignment for *H. sapiens*, *O. cuniculus* and *S. suis* GAPDHs (in this order).

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                                     *::  **:*:*:*:*  *::  **:*:*:*:*  ***  *:::  ***.*

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Figure S9