

1 **Supplemental Material**

2

3 **Characterization of a newly discovered symbiont in the whitefly *Bemisia tabaci***

4 **(Hemiptera: Aleyrodidae)**

5

6 **Running title:** Novel *Orientia* like symbiont in *Bemisia tabaci*

7

8 Xiao-Li Bing¹, Jiao Yang¹, Einat Zchori-Fein², Xiao-Wei Wang¹, Shu-Sheng Liu^{1#}

9

10 ¹Ministry of Agriculture Key Laboratory of Agricultural Entomology, Institute of Insect
11 Sciences, Zhejiang University, Hangzhou 310058, China; and ²Department of Entomology,
12 Newe-Ya'ar Research Center, Agricultural Research Organization, Ramat-Yishay 30095,
13 Israel.

14

15 [#]Correspondence: Shu-Sheng Liu, Email: shshliu@zju.edu.cn.

16

17 **Section:** Invertebrate microbiology

18

19

20 **Appendix 1**

21 **FISH protocol**

22 Specimens were collected directly into Carnoy's fixative (ethanol: chloroform: glacial acetic
23 acid, 6:3:1) and fixed overnight. After fixation, the samples were decolorized in 6% H₂O₂ in
24 ethanol for 2 h and then hybridized overnight in hybridization buffer (20 mM Tris-HCl (pH
25 8.0), 0.9 M NaCl, 0.01% sodium dodecyl sulfate, 30% deionized formamide) containing 10
26 pmol of fluorescent probes. Stained samples were viewed under a Leica TC Sp5 confocal
27 microscope.

28

29

30 **Appendix 2**

31 **Q-PCR protocol**

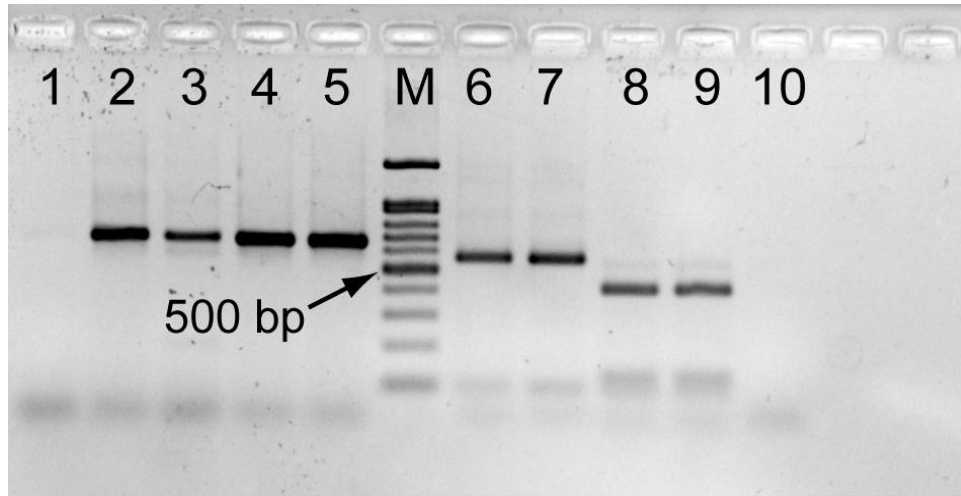
32 Quantitative PCRs were finished mainly by the SYBR[®] Premix Ex Taq[™] II and Bio-Rad
33 CFX96[™] Real-Time System. Each of the PCR mixtures consisted of 10.0 µl of 2×SYBR[®]
34 Premix Ex Taq[™] II buffer (Takara, Dalian, China), 0.8 µl forward and reverse primer solution
35 (10 µM each), 2.0 µl of DNA sample solution and 6.4 µl of double distilled water. The PCR
36 temperature profile was 2 min at 95 °C, 40 cycles of 5 s at 95 °C and 34 s at 60°C. Standard
37 curves for each of the genes were drawn using standard plasmid samples at concentrations of
38 10², 10³, 10⁴, 10⁵, 10⁶, 10⁷ and 10⁸ gene copies per µl of the target gene.

39

40

41 **Fig. S1**

42



43

44

45 **Fig. S1 RFLP pattern of PCR products of 16S rRNA gene of the OLO and *Rickettsia* in *B.***

46 ***tabaci* corresponding to *HphI* digestion.** The different profiles were obtained from two

47 individuals representing each of the OLO and *Rickettsia* in *B. tabaci*. The bands shown on the

48 lower left are primer dimers. Lane 1,10, no template controls; lane 2-3, undigested OLO; lane

49 4-5, undigested *Rickettsia*; lane 6-7, digested OLO, resulting in fragments of 566 bp and 101

50 bp, respectively; lane 8-9, digested *Rickettsia*, resulting in fragments of 411 bp, 128 bp, 99 bp

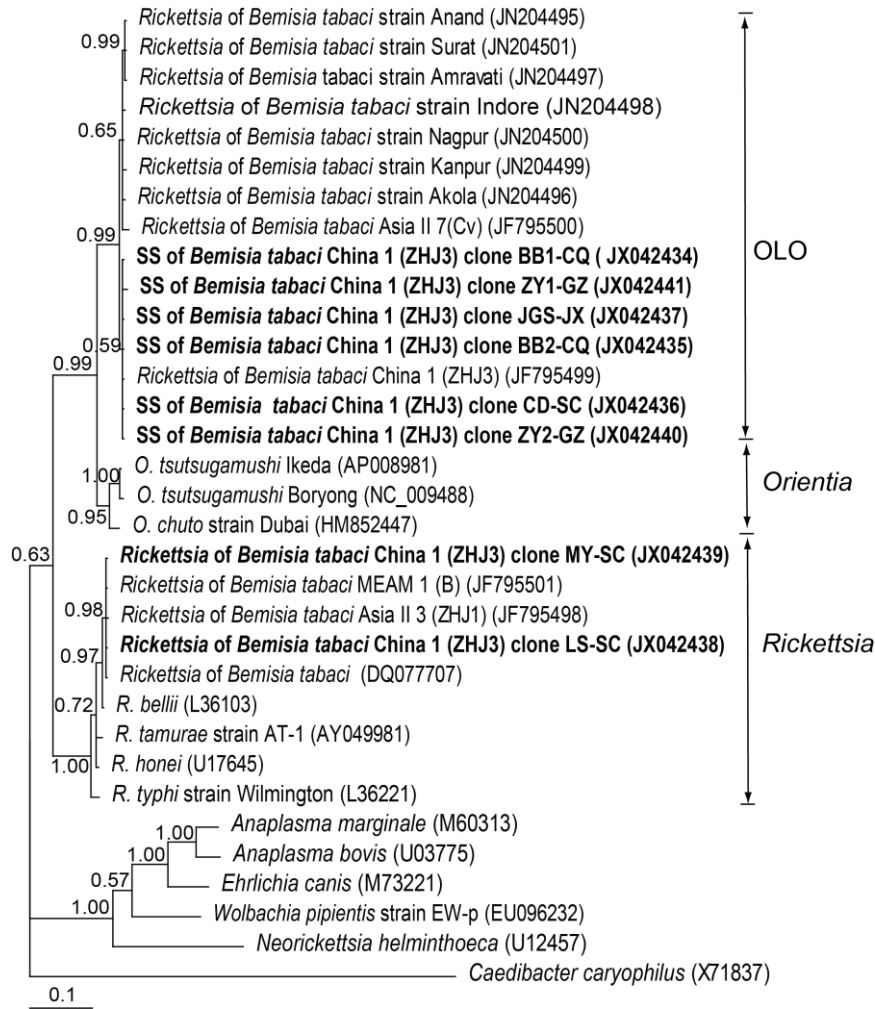
51 and 27 bp (although the three lower bp bands are blurred), respectively; lane M, DNA size

52 markers (0.1, 0.2, 0.3, 0.4, 0.5, 0.6, 0.7, 0.8, 0.9, 1.0 and 1.5 kb from bottom to top).

53

54

55 **Fig. S2**



56

57

58 **Fig. S2** Phylogenetic analysis of the OLO identified from different *Bemisia tabaci* populations
 59 based on bacterial 16S rRNA gene sequences (644 sites). The tree was constructed using a
 60 TPM3 + G substitution model for Bayesian analysis. Bayesian posterior probabilities (>0.50)
 61 are shown at the nodes. The names and sequence accession numbers are shown in parentheses.
 62 Sequences obtained in this study are shown in bold.

63

64 **Fig. S3**

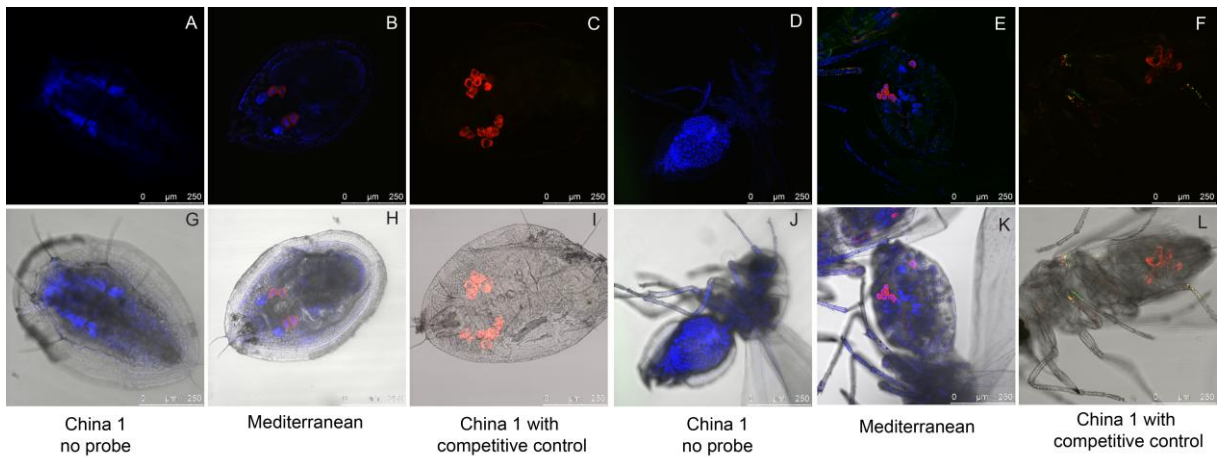


Fig. S3 FISH controls. China 1: no probe control; Mediterranean: OLO-free control; China 1 with competitive control. A-F: Overlay of channels of DAPI (blue), “*Ca. Portiera aleyrodidarum*” (red) and OLO (green); G-L: Overlay of channels of DAPI, “*Ca. Portiera aleyrodidarum*”, OLO and white light. Competitive controls lack DAPI for the break of DAPI detector. Signals on legs and wings are chitin autofluoresence.

73 **Table S1** Statistics of three-factor ANOVA of effects of development time, host sex and symbionts on symbiont density

Source of variation	Bacterial densities in terms of 16S rRNA gene copies per insect			Bacterial densities in terms of 16S rRNA gene copies per β -actin gene copy		
	df	<i>F</i>	<i>P</i>	df	<i>F</i>	<i>P</i>
	Time	2	0.716	0.493	2	2.402
Sex	1	33.179	<0.001	1	24.151	<0.001
Symbiont	1	0.274	0.602	1	0.960	0.331
Time \times sex	2	0.214	0.808	2	1.193	0.311
Time \times symbiont	2	2.255	0.114	2	1.742	0.184
Sex \times symbiont	1	0.037	0.848	1	0.338	0.563
Time \times sex \times symbiont	2	1.533	0.224	2	1.085	0.345
Error	58			58		
Total	69			69		

74

75 **Table S2** AT contents of 16S rRNA gene of OLO, the primary and secondary symbiont of various insects (*B. tabaci* included), and
 76 free-living bacteria representing the Alpha-subclass of the *Proteobacteria*.

Symbiont	Host insect	AT content (%)	GenBank accession no.
OLO of <i>Bemisia tabaci</i> China 1	<i>Bemisia tabaci</i> (whitefly)	48.5	JX042442
Primary symbiont of various insects			
<i>Portiera aleyrodidarum</i>	<i>Bemisia tabaci</i> (whitefly)	52.3	JN204485
<i>Buchnera aphidicola</i>	<i>Baizongia pistaciae</i> (aphid)	51.8	NC_004545
<i>Wigglesworthia glossinidia</i>	<i>Glossina brevipalpis</i> (tsetse fly)	51.3	NC_004344
Secondary symbiont of various insects			
<i>Arsenophonus</i> symbiont	<i>Bemisia tabaci</i> (whitefly)	45.8	JN204476
<i>Arsenophonus</i> symbiont	<i>Stomaphis quercus</i> (aphid)	46.2	FJ655543
<i>Cardinium</i> symbiont	<i>Bemisia tabaci</i> (whitefly)	50.8	JN204480
<i>Cardinium</i> symbiont	<i>Aspidiotus nerii</i> (buckler scale)	52.2	GQ455437
<i>Hamiltonella defensa</i>	<i>Acyrtosiphon pisum</i> (aphid)	46.2	AY907546
<i>Regiella insecticola</i>	<i>Sitobion avenae</i> (aphid)	46.3	FJ357498
<i>Rickettsia</i> symbiont	<i>Bemisia tabaci</i> (whitefly)	49.5	DQ077707
<i>Rickettsia</i> symbiont	<i>Curculio hilgendorfi</i> (weevil)	48.7	AB604668
<i>Serratia symbiotica</i>	<i>Cinara tujafilina</i> (aphid)	45.5	EU348323
<i>Wolbachia</i> symbiont	<i>Bemisia tabaci</i> (whitefly)	53.0	JN204502
<i>Wolbachia pipientis</i>	<i>Drosophila</i> sp. (fruit fly)	52.2	EU096232
Free living bacteria			
<i>Acetobacter aceti</i>		44.6	D30768
<i>Agrobacterium tumefaciens</i>		45.2	D01256
<i>Caulobacter crescentus</i>		44.4	AJ227756
<i>Rhodospirillum rubrum</i>		43.2	D30778