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Supplemental Information

Innate Sensing of HIV-1 Assembly by Tetherin Induces NFkB-Dependent Proinflammatory Responses

Rui Pedro Galão, Anna Le Tortorec, Suzanne Pickering, Tonya Kueck, and Stuart J.D. Neil

Supplementary Figure Inventory

Figure S1: Control Panels demonstrating parallel virus release from experiments in **Figure 1**.

Figure S2: Controls in cell lines and primary cells to demonstrate the potential source of IFN responses observed in **Figure 2** and the specificity of shRNA constructs.

Figure S3 Further virological and signaling phenotypes of tetherin mutants described in **Figure 3**

Figure S4: Interactions of some of the tetherins from Figures 5 with TRAF6

Figure S5: Virological phenotypes, expression levels and TRAF6 interactions of primate tetherins and chimeric proteins from **Figure 6**.

Supplementary Figures



Figure S1, related to Figure 1: Restriction of viral particle release in 293THN cells. (A) Transfection of an HIV-1 Vpu-A14L provirus enhanced NFkB reporter gene activation in 293THN cells. Data are represented as mean +/- SEM of 3 independent experiments **(B)** Parallel infectious virus release from **(A)** and Figure 1D on HeLa-TZM indicator cells. **(C)** Ebola virus-like particles released from 293 and 293THN cells from Figure 1E were analyzed by western blotting of cell lysates and pelleted supernatants. **(D)** MLV infectious virus release by titration of supernatants from Figure 1G on 293T and analyzed by flow cytometry 48h later.



Figure S2, related to Figure 2: Tetherin does not directly activate IFN expression and IRF3 phosphorylation, but does so indirectly through a TRIF-dependent mechanism in infected T cells. IFNß-luciferase reporter activation (left panels), induction of IFNß mRNA (central panels) or phosphorylation of IRF3 (right panels) are not observed following crosslinking with a rabbit anti-tetherin antibody (A), tetherin overexpression (B) or Vpu(-) HIV-1 assembly (C). (D) Purified CD4+ T cells were infected with the indicated virus at an MOI of 5. 48h later IRF-3 phosphorylation and viral infection was analyzed by western blot of cell lysates. (E) Infected CD4+ T cells were cultured in the presence of 40µM of control, TRIF and MyD88 inhibitory peptides. Total RNA from 3 biological replicates of parallel cultures were analyzed for *Cxcl10*, and *lfnb* mRNA levels relative to *Gapdh* by gRT-PCR. Fold changes relative to non-infected cells treated with control peptide. * P>0.05, ** P>0.01 and *** P>0.001 as determined by two-tailed ttest. (F) Activation state of IRF3 in cells cultured in the presence of TRIF inhibitory peptides was assessed by western blotting cell lysates from (E). (G) Activation of NFkB signaling by anti-tetherin antibody crosslink in 293THN cells is not affected by TRIF or MyD88 inhibitory peptides. (H) CD4+ T cells were transduced with lentiviral vectors encoding shRNAs against GFP (shcontrol) or tetherin (sh-THN) and infected with indicated virus at MOI 5 as in Figure 2E. Cell surface staining for endogenous tetherin was analyzed by flow cytometry 48h post-transduction. The solid peak represents the binding of the isotype control and the lines represent tetherin levels in non-treated cells (NT, dotted lines) or transduced cells (solid lines). (I) Parallel infectious virus release from (H) on HeLa-TZM indicator cells. All error bars represent means +/- SEM of 3 independent experiments.



Figure S3, related to Figure 3: NF-kB reporter gene activities and antiviral function of tetherin mutants. (A) NF-kB reporter gene activities of tyrosine, cysteine and membrane proximal ubiquitin-accepting-lysine residue mutant tetherins determined by transient transfection as in Figure 3A. **(B)** NFkB reporter gene activity in 293 cells expressing the indicated tetherin extracellular mutant with wildtype or Vpu(-) provirus as in Figure 3B, and parallel infectious virus release determined on HeLa-TZM cells (right panel).



Figure S4, related to Figure 5: Co-IP of further tetherin mutants with TRAF6. Co-immunoprecipitation of tetherin, tetherin cytoplasmic tail mutants with HA-TRAF6 from transiently transfected 293 cells as in Figure 5C.



Figure S5, related to Figure 6: Antiviral function of species-specific tetherin mutants. Tetherins from Figure 6 were tested for their ability to restrict Vpu(-) virus release (A) and protein expression (B). (C) Chimpanzee and human tetherins with or without DDIWK motif were assessed for their ability to be co-precipitated with HA-TRAF6. Control blot of single transfectants reproduced from Figure S4.