

***Polg2* is essential for mammalian embryogenesis and is required for mtDNA maintenance**

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MATERIALS AND METHODS

Genotyping PCR

PCR primers for genotyping were designed to target the *loxP*-sites, *Polg2*, *cre*-recombinase, and the shortened *Polg2* gene due to the excision of exons 5 through 7. The 5' *loxP*-site forward primer located upstream of exon 5 sequence was 5'-TGGTCCACAGAGCAAGTTCCAGG-3', and the reverse primer sequence was 5'-GCCGCGTGGATCTTGTGTTG-3'. The 3' *loxP*-site forward primer downstream of the NEO cassette sequence was 5'-AAGTACACCGTAGCTGTCCTCAGAC-3', and reverse primer was 5'-TCACCATGTGGTGGTGGACTGGAT-3'. Exon 5 of *Polg2* was targeted using the forward primer 5'-TTGCGTTCTGTCTGTAAGTGGGGACG-3', and

the reverse primer 5'-TAGCAACTGGTGGCAAGATGGCTCAGAGG-3'. The *cre*-recombinase forward primer was 5'-CGCAGAACCTGAAGATGTTTCGCGATTA-3' and reverse primer was 5'-TCTCCCACCGTCAGTACGTGAGATATC-3'. The primer set detecting the knockout of *Polg2* exons 5 - 7 was the 5' *loxP* forward primer and the 3' *loxP* reverse primer. Amplitaq Gold hot start polymerase (Applied Biosystems) was used according to the manufacture's recommendations in a 25 µl reaction with 1 µl of the ear punch lysate. Cycling conditions for all primers sets were: 10 minutes at 95°C, 32 cycles at 95°C for 30 seconds, 57°C for 30 seconds, and 72°C for 1 minute, followed by an extension at 72°C for 7 minutes, and hold at 4°C. All PCR products were run on a 1.25% agarose gel stained with ethidium bromide (Invitrogen).

Clinical Pathology

The hematopoietic system of the 2 y.o. aging mouse population was evaluated using whole blood collected with EDTA as the anticoagulant that was analyzed for a complete blood count (CBC). CBC's were performed on a Hemavet 1700 (Drew Scientific Inc.) using reagents obtained from the instrument's manufacturer. Blood collected without anticoagulant was allowed to clot, centrifuged and serum harvested for evaluation of several serum biomarkers. Serum chemistries included: alanine aminotransferase (ALT), sorbitol dehydrogenase (SDH), aspartate aminotransferase (AST), alkaline phosphatase (ALP), total bile acids (TBA), cholesterol (Chol), triglycerides (Trig), high-density lipoprotein (HDL), low-density lipoprotein (LDL), calcium (Ca), creatinine (Creat) and creatine kinase (CK). All clinical chemistry assays were performed using an Olympus

AU400e Analyzer (Beckman Coulter Inc.). In general, reagents were purchased from the instrument's manufacturer; reagents for TBA, HDL, and LDL analyses were purchased from Sekisui Diagnostics, LLC, and reagents for SDH came from Sigma/Aldrich Inc. Statistical analyses were performed using an exact Mann-Whitney test to compare genotype groups and the p-values are 2-sided.

Histopathology

E8.5 embryos were used for COXI staining. Embryos were collected intact within the deciduum, embedded in Optimal Cutting Temperature compound (Tissue-Tek) and frozen in the vapor phase of liquid nitrogen. 10 µm cryostat sections were applied to charged slides (Erie Scientific, LLC), and were stained for COXI activity, washed in water, counterstained with hematoxylin, and mounted in Advantage Permanent mounting media (Axell). Staining protocols were adapted from Chen *et al.* (1), and the Washington University Neuromuscular Disease Center website (<http://www.neuro.wustl.edu/neuromuscular/index.html>).

Evaluation of mtDNA deletions

Mouse mtDNA deletions were detected using Expand Long Range PCR reagents (Roche), and all amplifications were carried out with a Gene Amp PCR System 9700 (Applied Biosystems). The sequences of the primers and positions correspond to mtDNA sequence submitted by Bibb *et al.* (2), and primers were designed for the three most prevalent deletions in mice as previously described (3). Primer L1 8858 forward was 5'-TCTATTCATCGTCTCGGAAG-3' 8877, L2 12883 forward was 5'-

TACCATTCCTAACAGGGTTC-3' 12902, and H 13354 reverse was 5'-TTTATGGGTGTAATGCGGTG-3' 13335 (3). Cycling conditions were 92°C for 2 minutes, 12 cycles at 92°C for 10 seconds, 56°C for 15 seconds, and 68°C for 5 minutes, followed by a second cycling step consisting of 30 cycles of 92°C for 10 seconds, 56°C for 15 seconds, and 68°C for 5 minutes, 68°C extension for 7 minutes, and a final hold step at 8°C. Primer pair L2/H amplified a 0.47kb fragment from undeleted wild type mtDNA and served as an internal control. L1/H primer pair resulted in three fragments: 1) 262 bp corresponding to a 4236 bp deletion, 2) 630 bp indicative of a 3867 bp deletion, and 3) 772 bp representing a 3726 bp deletion. The wild type band lacking deletions yielded a band of 7.5 kb, but was only seen in 10 month old brain tissue due to a change in stocks of PCR reagents. The deletions in mouse mtDNA are a result of repeats within the sequence as previously described by Tanhauser and Laipis (4).

Mouse mtDNA deletions were quantitated to determine mtDNA heteroplasmy. qPCR primers and the FAM/ TAMRA labeled probes targeting the rarely deleted mitochondrial gene, *Nd1*, and the more frequently deleted mitochondrial gene, *Nd4*, were all designed by Applied Biosystems. qPCR amplifications were carried out on an ABI PRISM 7900HT Sequence Detector (Applied Biosystems) with a cycling protocol consisting of 95°C for 10 minutes, and 40 cycles at 95°C for 15 seconds and 60°C for 1 minute. All reactions were done in triplicate, and 1 µl of diluted (1:50) tissue lysate was used in each 25 µl reaction using TaqMan 2X Universal Mix (Applied Biosystems). To calculate the relative copy number (*R*) the following equation was used: $R = 2^{-\Delta Ct}$, where ΔCt is the $Ct_{Nd1} - Ct_{Nd4}$ (5).

RNA isolation and cDNA generation

Mouse liver, brain, heart, kidney, and muscle tissues were collected and stored at 4°C in RNAlater solution (Ambion). Isolation of total RNA was obtained by using the RNeasy mini-kit (Qiagen) according to manufacture's recommendations. Tissues were disrupted using the TissueLyser apparatus (Qiagen) with the lysis buffer provided in the RNeasy kit. Reverse transcription was performed using a High Capacity RT (Applied Biosystems) with 150 ng of total RNA in a 60 µl reaction. cDNA generation was performed in a Gene Amp PCR System 9700 (Applied Biosystems) with cycling parameters of 25°C for 10 minutes, 37°C for 90 minutes, 85°C for 5 seconds, and held at 4°C to synthesize cDNA.

Transcript analysis of the mtDNA replication fork

All primers and fluorescently labeled probes were predesigned and sold as TaqMan Gene Expression Assay 20X mixes (Applied Biosystems), and all sets spanned intron/exon boundaries to avoid DNA amplification. The TaqMan primers and probes targeted the genes: *Polg2*, *Polg*, *Twinkle*, *Tfam*, *Tfb1*, *Tfb2*, *mtRnap*, and *mtSsb*. All probes were labeled at the 5' end with FAM (reporter dye) and TAMRA (quencher dye) at the 3' end. 1 µl of cDNA was used in each 25 µl reaction with TaqMan 2X Universal Mix with UNG (Applied Biosystems), and all reactions were run in triplicate. QPCR amplifications were carried out on an ABI PRISM 7900HT Sequence Detector (Applied Biosystems) with a cycling protocol consisting of 95°C for 10 minutes, and 40 cycles at 95°C for 15 seconds and 60°C for 1 minute. The standard Comparative C_T Method ($\Delta\Delta C_T$) was used for relative quantification calculations as described in the ABI technical

guide on performing relative quantitation of gene expression using real-time PCR (http://www3.appliedbiosystems.com/cms/groups/mcb_support/documents/generaldocuments/cms_042380.pdf), and *Actin* was used as the endogenous reference. Data were expressed as a change from the wild type, and the Mann-Whitney statistical *U*-test was applied to determine statistical significance.

RESULTS

Characterization of adult *Polg2* heterozygous mice

Histopathology results for the aged *Polg2*^{+/-} mice

Aged *Polg2*^{+/-} mice at 2 years were euthanized and comprehensively examined for any alterations in phenotype associated with the *Polg2* heterozygous deletion. There were no differences microscopically between *Polg2*^{+/-} and WT groups at 2 years of age. There was however a moderate to marked microvesicular steatosis within the livers of all the animals (*Polg2*^{+/-} and WT). In addition, several livers also exhibited macrovesicular steatosis with occasional lipid granulomas and minimal to mild inflammatory cell infiltration. Further, there was a sex related effect in the severity of the skeletal muscle lesions. Within the male animals (from both *Polg2*^{+/-} and WT groups), there were moderate to marked degenerative changes within the skeletal muscle (rectus femoris and gastrocnemius) characterized by loss of cross striations, amorphous slightly hypereosinophilic sarcoplasm, nuclear internalization, and non-uniform muscle fiber diameter. In addition, several muscle fibers had a central pale amphophilic amorphous material devoid of cross striations and in some cases the skeletal muscle fibers are replaced by adipose tissue. In comparison, the skeletal muscle lesions within the female

mice (from both *Polg2*^{+/-} and WT groups) were minimal to mild. There was a sporadic low incidence of various neoplasms such as lymphoma, histiocytic sarcoma and hepatic adenoma that is most likely attributed to age and strain, but it was not related to the loss of one copy of the *Polg2* gene.

Heart Electron Microscopy

No differences in mitochondrial ultra-structural features of 1-year old *Polg2*^{+/-} (n=4) and wild-type animals (n=4) were observed. The myofibrils were uniform with evenly spaced striations. The Z-bands of the myofibrils were also evenly spaced. Small lipid droplets, some containing slender myelin figures, were seen randomly scattered throughout the sarcoplasm along with randomly scattered lysosomes, myelin figures, and glycogen particles in both genotypes (Supplemental fig. 2). While few in number, we identified some randomly scattered mitochondria with cristae that were slightly separated indicative of subtle mitochondrial swelling which were found in both genotypes.

Clinical pathology results for the aged *Polg2*^{+/-} mice

In general, there were no changes in the hematology variables that were considered pathognomonic and attributable to the loss of one copy of the *Polg2* gene in the 2-year old heterozygous population (Supplemental tables 1 - 4). Further, there was no consistent change in the clinical chemistry that would help characterize a phenotype for the *Polg2*^{+/-} genetically modified mice. However, clinical chemistry data of a few individual animals demonstrated changes that suggested pathology in the liver and/or muscle. For example, one female *Polg2*^{+/-} animal (ID 38-8) demonstrated ALT, SDH, AST, and CK activities that were ≥ 4 -fold higher compared to the mean values for the wild-type females

(Supplemental tables 1 and 2). This animal also had an exceptionally low serum calcium concentration. Increases in serum ALT, SDH, and AST activities are consistent with hepatocellular injury resulting in leakage of the enzymes into circulation. While the increases are not exceedingly high, they were high enough to consider the possibility of hepatocellular pathology. Based on previous work showing a high concordance of increases in ALT and SDH to predict histopathology changes in the liver (9), the blood chemistry of this one particular animal may suggest a pathological change in the liver. Although, slight changes in clinical chemistry parameters of other individual mice were observed they were most likely attributable to the increased variability often seen in laboratory animals of advanced age.

References

1. Chen, H., McCaffery, J.M. and Chan, D.C. (2007) Mitochondrial fusion protects against neurodegeneration in the cerebellum. *Cell*, **130**, 548-562.
2. Bibb, M.J., Van Etten, R.A., Wright, C.T., Walberg, M.W. and Clayton, D.A. (1981) Sequence and gene organization of mouse mitochondrial DNA. *Cell*, **26**, 167-180.
3. Zeng, Z., Zhang, Z., Yu, H., Corbley, M.J., Tang, Z. and Tong, T. (1999) Mitochondrial DNA deletions are associated with ischemia and aging in Balb/c mouse brain. *J. Cell. Biochem.*, **73**, 545-553.
4. Tanhauser, S.M. and Laipis, P.J. (1995) Multiple deletions are detectable in mitochondrial DNA of aging mice. *J. Biol. Chem.*, **270**, 24769-24775.
5. He, L., Chinnery, P.F., Durham, S.E., Blakely, E.L., Wardell, T.M., Borthwick, G.M., Taylor, R.W. and Turnbull, D.M. (2002) Detection and quantification of mitochondrial DNA deletions in individual cells by real-time PCR. *Nucleic Acids Res.*, **30**, e68.
6. Grattagliano, I., de Bari, O., Bernardo, T.C., Oliveira, P.J., Wang, D.Q. and Portincasa, P. (2012) Role of mitochondria in nonalcoholic fatty liver disease-from origin to propagation. *Clinical biochemistry*, **45**, 610-618.
7. Dimauro, S. and Davidzon, G. (2005) Mitochondrial DNA and disease. *Ann. Med.*, **37**, 222-232.

8. Cortopassi, G.A., Shibata, D., Soong, N.W. and Arnheim, N. (1992) A pattern of accumulation of a somatic deletion of mitochondrial DNA in aging human tissues. *Proc. Natl. Acad. Sci. U. S. A.*, **89**, 7370-7374.
9. Travlos, G.S., Morris, R.W., Elwell, M.R., Duke, A., Rosenblum, S. and Thompson, M.B. (1996) Frequency and relationships of clinical chemistry and liver and kidney histopathology findings in 13-week toxicity studies in rats. *Toxicology*, **107**, 17-29.

SUPPLEMENTAL FIGURE LEGENDS

Supplemental Figure 1. The protein alignment of human and mouse POLG2 shows a highly conserved C-terminus. Amino acids that are identical are highlighted in yellow, and the green highlighted amino acids indicated a change in the amino acid with a conservation of charge. Exons 5, 6, and 7 are deleted in our Polg2 mouse model. Exon 5 (red) encodes for the dimerization of POLG2, while exons 6 (blue) and 7 (purple) both code for the POLG and DNA binding domains.

Supplemental Figure 2. Electron microscopy of heart tissue from 1 y.o. wild type (+/+) and Polg2 heterozygous (+/-) mice. At 1700X magnification the normal pattern of clustered mitochondria (darker grey) that are perpendicular to myofibrils of cardiac tissue, and are similar in both the wild type and heterozygous mouse. At 20,500X magnification the mitochondria of the Polg2^{+/-} are similar in size, shape, and cristae pattern compared to the wild type.

Supplemental Figure 3. Measuring expression levels of genes encoding proteins important for mtDNA replication. qPCR was used to determine transcript levels of Polg2, Polg, Tfam, mtRnap, Tfb1, Tfb2, Twinkle, and mtSsb. The standard $\Delta\Delta C_T$ method of analysis was used to report data as fold change from the mean levels of wild-type tissues. The transcripts levels were measured in 1-year Polg2^{+/-} tissues (n=10). Tissues included: liver (blue), muscle (red), brain (green), heart (orange), and kidney (light blue). The Mann-Whitney U-test showed no statistical difference ($p>0.05$) in expression from the wild-type. Error bars indicate \pm standard deviation.

Supplemental Figure 4. Detection of mtDNA deletions in mouse tissue. Mouse mtDNA deletions were detected using Long Range PCR in Polg2^{+/-} (n=3) and age-matched WT (n=3). L1/H primer pair resulted in three fragments: 1) 262 bp corresponding to a 4236 bp deletion, 2) 630 bp indicative of a 3867 bp deletion, and 3) 772 bp representing a 3726 bp deletion. Tissue samples were amplified with an internal control primer set L2/H resulting in a 470 bp fragment. A) mtDNA deletions in 6 month brain (B), liver (L), muscle (M), and heart (H) tissues. Deletions were observed in brain and heart of all 3 WT and 2 Polg2^{+/-} animals. B) mtDNA deletions in 10 month heart samples. The three most common mouse mtDNA deletions were observed in both the wild-type and heterozygous mice at 10 month of age.

Supplemental Figure 5. Determining mtDNA heteroplasmy in *Polg2*^{+/-} tissues. mtDNA heteroplasmy was evaluated using qPCR in muscle, brain, heart, and liver tissue of mice 1 year of age. The relative copy number was determined using the following calculation: $R = 2^{-\Delta Ct}$, where ΔCt is the $Ct_{Nd1} - Ct_{Nd4}$. Wild type mice tissues are shown in blue and heterozygous mice tissues in red. There was no significant difference in mtDNA heteroplasmy between the wild type and *Polg2* heterozygous mice.

Supplemental Figure 6. The morphology of littermates was analyzed at E8.0-8.5 post coitum. **A)** A sagittal view of a WT embryo enveloped in the amniotic sac with the allantois (a) extending from the embryo, a well formed cardiac sac (c), distinct somites (s), and normal head (h) region. **B)** A sagittal view of a *Polg2*^{-/-} embryo littermate within an intact amniotic sac showing the complete loss of a cardiac sac, abnormally developed head region, and undefined somite architecture.

Supplemental Table 1. A. The body weights of *Polg2*^{+/-} mice at 40 weeks and 2 years of age. No statistical differences in the total body weight of the wild-type and *Polg2*^{+/-} mice were observed at 40 weeks and 2 years of age. B. The relative liver weights were measured in a population of 2 year old mice. The relative liver weight is calculated based on the ratio of liver weight to the body weight of each mouse. Again, no significant difference was found in the relative liver weights of the wild type in comparison to the *Polg2*^{+/-} mice.

Supplemental Table 2. Serum clinical chemistry of 2 y.o. male mice. Serum chemistries included: alanine aminotransferase (ALT), sorbitol dehydrogenase (SDH), aspartate aminotransferase (AST), alkaline phosphatase (ALP), total bile acids (TBA), cholesterol (Chol), triglycerides (Trig), high-density lipoprotein (HDL), low-density lipoprotein (LDL), calcium (Ca), creatinine (Creat) and creatine kinase (CK). Changes in individual mice were observed, but they are most likely attributed to the age of the mice, and not the loss of *Polg2* on one allele. The Mann-Whitney test was used to determine the significance at $p < 0.05$. No sample received (nsr). Quantity not sufficient for assay (qns). Values determined by dilution (*).

Supplemental Table 3. Serum clinical chemistry of 2 y.o. female mice. Serum chemistries included: alanine aminotransferase (ALT), sorbitol dehydrogenase (SDH), aspartate aminotransferase (AST), alkaline phosphatase (ALP), total bile acids (TBA), cholesterol (Chol), triglycerides (Trig), high-density lipoprotein (HDL), low-density lipoprotein (LDL), calcium (Ca), creatinine (Creat) and creatine kinase (CK). Changes in individual mice were observed, but they are most likely attributed to the age of the mice, and not the loss of *Polg2* on one allele. The Mann-Whitney test was used to determine the significance at $p < 0.05$. No sample received (nsr). Quantity not sufficient for assay (qns). Values determined by dilution (*).

Supplemental Table 4. Cell blood counts (CBC) were determined on the male mouse aging colony. Parameters measured included: white blood cells (WBC), red blood cells (RBC), neutrophils (Neut), lymphocytes (Lym), monocytes (Mono), eosinophils (Eos), basophils (Baso), hemoglobin (Hgh), hematocrit (Hct), platelet count (Plts), reticulocyte

count (Retic), mean corpuscular volume (MCV), mean corpuscular hemoglobin (MCH), mean corpuscular hemoglobin concentration (MCHC), percentage of hematocrit (Spun), and large unstained cells (LUC). There were no significant changes observed between the wild type and the heterozygous mice that can be attributed to the loss of one copy of Polg2 at 2 years of age. The Mann-Whitney U-test was used to determine the significance at $p < 0.05$.

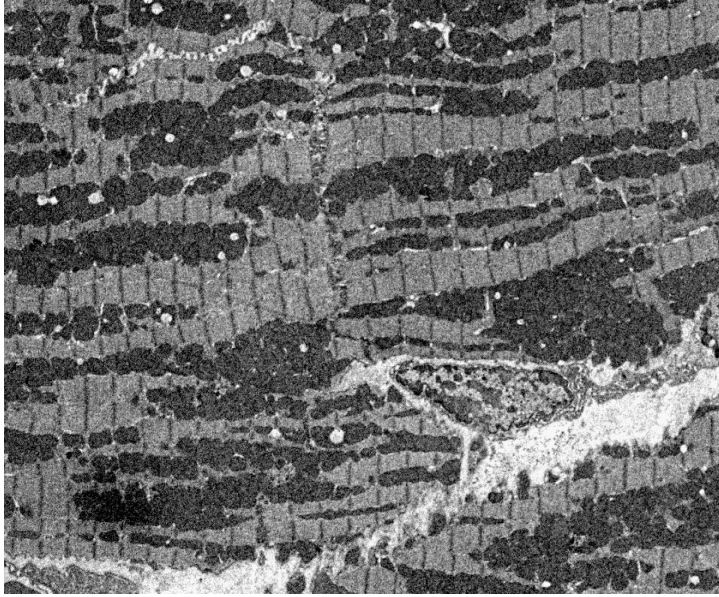
Supplemental Table 5. Cell blood counts (CBC) were determined on the female mouse aging colony. Parameters measured included: white blood cells (WBC), red blood cells (RBC), neutrophils (Neut), lymphocytes (Lym), monocytes (Mono), eosinophils (Eos), basophils (Baso), hemoglobin (Hgh), hematocrit (Hct), platelet count (Plts), reticulocyte count (Retic), mean corpuscular volume (MCV), mean corpuscular hemoglobin (MCH), mean corpuscular hemoglobin concentration (MCHC), percentage of hematocrit (Spun), and large unstained cells (LUC). There were no significant changes observed between the wild type and the heterozygous mice that can be attributed to the loss of one copy of Polg2 at 2 years of age. The Mann-Whitney U-test was used to determine the significance at $p < 0.05$.

		1		60
Homo sapien POLG2	(1)	MRSRVAVRACHKVCRCILSGFGGRVLAGQPELLTERS	SSPKGGHVKSHAELEGNGEHEEAP	
Mus musculus POLG2	(1)	MRCGGARACRRACRCWLSGYAGPADG	-----TQQPDAP	
Consensus	(1)	MR A RAC K CRC LSGFAG DA		PDAP
		61		120
Homo sapien POLG2	(61)	GSGEGSEALELTCQRRHFLSGSKQQLSRD	LLSGCHPGFGPLGVELRKNLAAEWWT	SVVV
Mus musculus POLG2	(35)	EHAVAREALVDLCRRHFFSCTPQQLSTA	ALLSGCHARFGPLGVELRKNLASQWSS	SMVY
Consensus	(61)	A A EALLDIC RRHF SGS QQLS	ALLSGCH FGPLGVELRKNLAA	WWSSMVY
		121		180
Homo sapien POLG2	(121)	FREQVFPVDALHHKPGPLLPGDSAFRLV	SAETLREILQDKELSKEQLVAFLEN	VLKTS
Mus musculus POLG2	(95)	FREQVFAVDSLHQEPGSSQPRDSAFRLV	PEISIREILQDREPSKEQLVAFLEN	LLKTS
Consensus	(121)	FREQVF VDALH PG P DSAFRLVS	ESIREILQDKE SKEQLVAFLEN	LLKTS
		181		240
Homo sapien POLG2	(181)	LRLENLLHGALEHYVNCLDLVNRKLFY	GLAQIGVCFHPVFDTKQIRNGVKS	IGEKTEASLV
Mus musculus POLG2	(155)	LRATLLHGALEHYVNCLDLVNRKLFY	GLAQIGVCFHPVSNSTPSSVTRV	IGEKTEASLV
Consensus	(181)	LR LLHGALEHYVNCLDLVNRKLLP	FGLAQIGVCFHPV S Q V	IGEKTEASLV
		241		300
Homo sapien POLG2	(241)	WFTPRTSNQWLDFWLRHRLQWWRKFAM	SPSNFSSDCQDEEGRKGNKLYYN	FPWGKELI
Mus musculus POLG2	(215)	WFTPRTSSQWLDFWLRHRLLWWRKFAM	SPSNFSSADCQDELGRKGSKLYYS	FPWGKEPI
Consensus	(241)	WFTP RTS QWLDFWLRHRL WWRKFAM	SPSNFSSADCQDE GRKG KLYY	FPWGKE I
		301		360
Homo sapien POLG2	(301)	ETLWNLGDHELLHMYPGNVSKLHG	GRDGRKNVVPVLSVNGLDRCMLAYLY	DSFQLTENS
Mus musculus POLG2	(275)	ETLWNLGDQELLHTYPGNVSTIQ	GRDGRKNVVPVLSVSGIVDLGTLAYLY	DSFQLAENS
Consensus	(301)	ETLWNLGD ELLH YPGNVS I	GRDGRKNVVPVLSV GDLD G	LAYLYDSFQL ENS
		361		420
			EXON 5	
			EXON 6	EXON 7
Homo sapien POLG2	(361)	FTRKKNLHRRKVLKLLHPCLAPIKVALD	VGKPTLELRQVCQGLFNELLENGIS	VWPGYLET
Mus musculus POLG2	(335)	FARKKSLQRKVLKLLHPCLAPIKVALD	VGKPTVELRQVCQGLLNELLENGIS	VWPGYSET
Consensus	(361)	F RKK L R KVLKLLHPCLAPIKVALD	VGKPTLELRQVCQGL NELLENGIS	VWPGY ET
		421		480
Homo sapien POLG2	(421)	MQSSLEQLIYSKYDEMSEILFTVLVTET	TLENGLIHLRSRDTTMKEMMHISKIK	DFLIKYIS
Mus musculus POLG2	(395)	VHSSLEQLHSKYDEMSEILFVSVLVTET	TLENGLIQLRSRDTTMKEMMHISKIR	DFIVKYLA
Consensus	(421)	M SSLEQLHSKYDEMSEILFVSVLVTET	TLENGLI LRSRDTTMKEMMHISKIK	DFLIKYIA
		481		
Homo sapien POLG2	(481)	SAKNV		
Mus musculus POLG2	(455)	SASNV		
Consensus	(481)	SA NV		

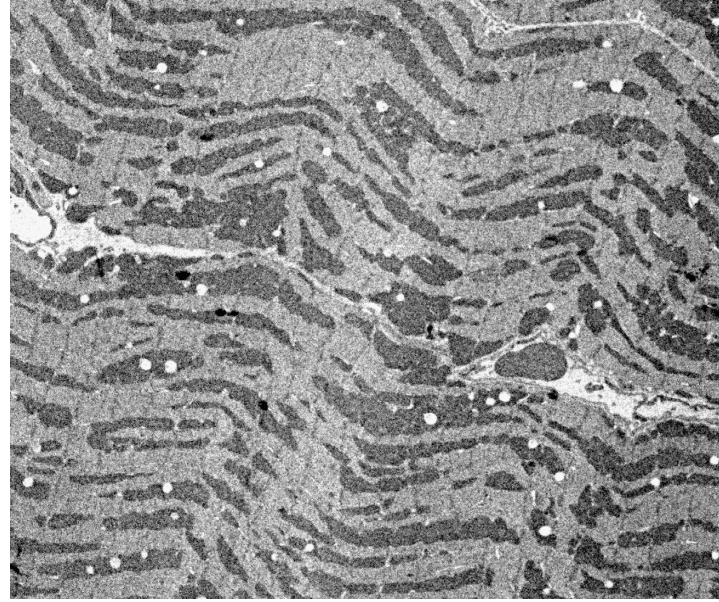
Supplemental figure 1

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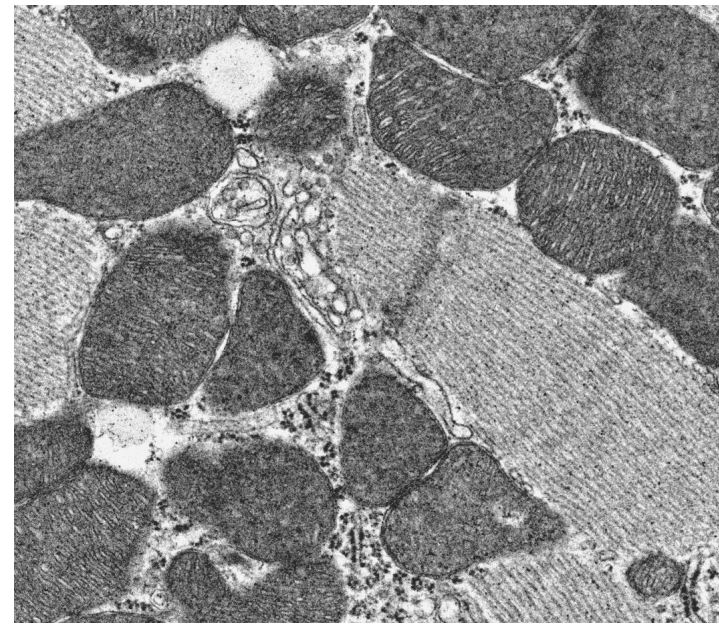
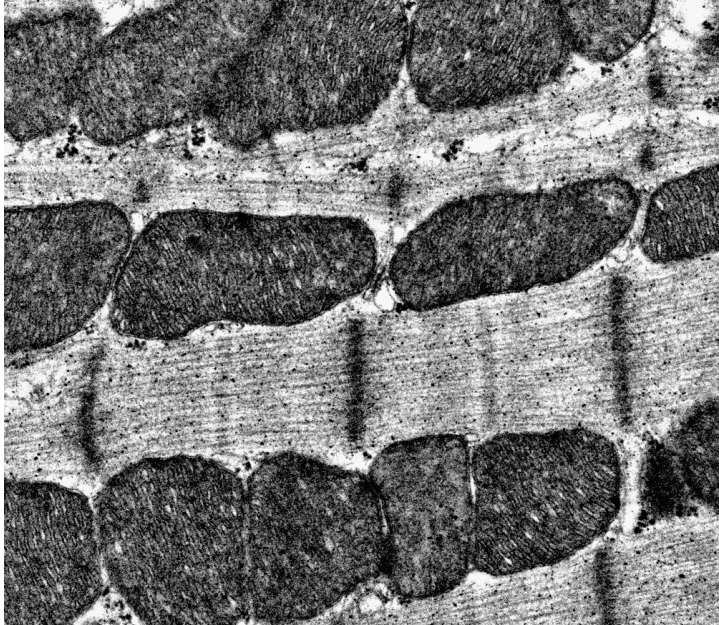
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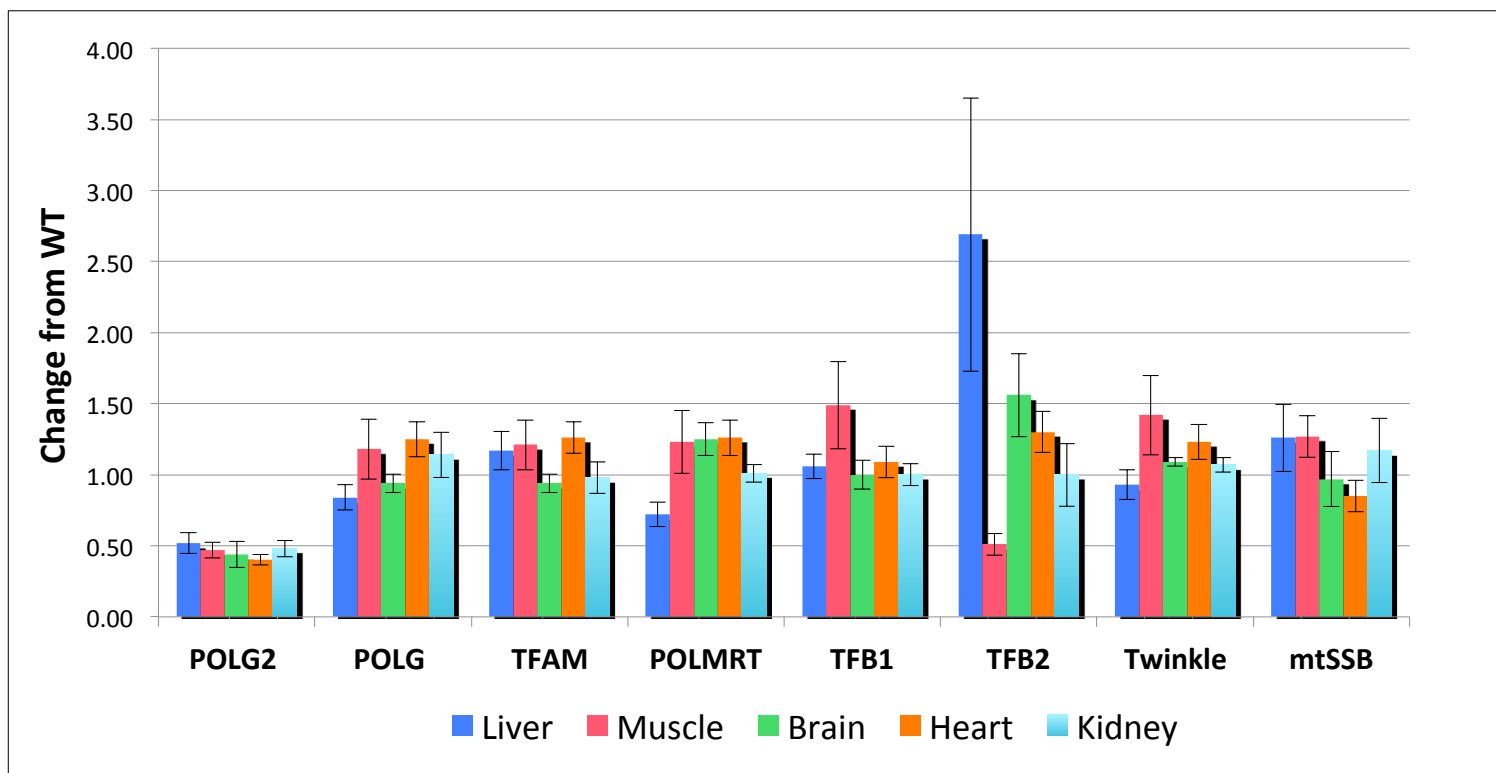
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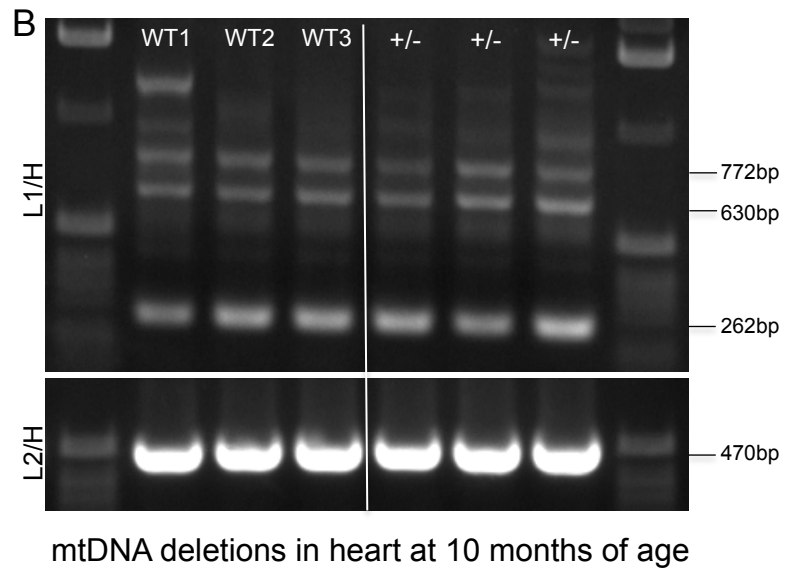
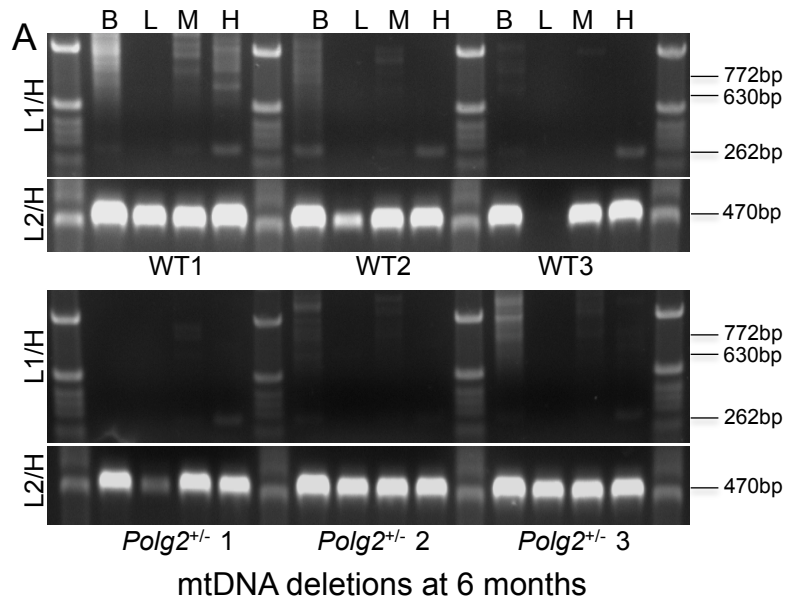
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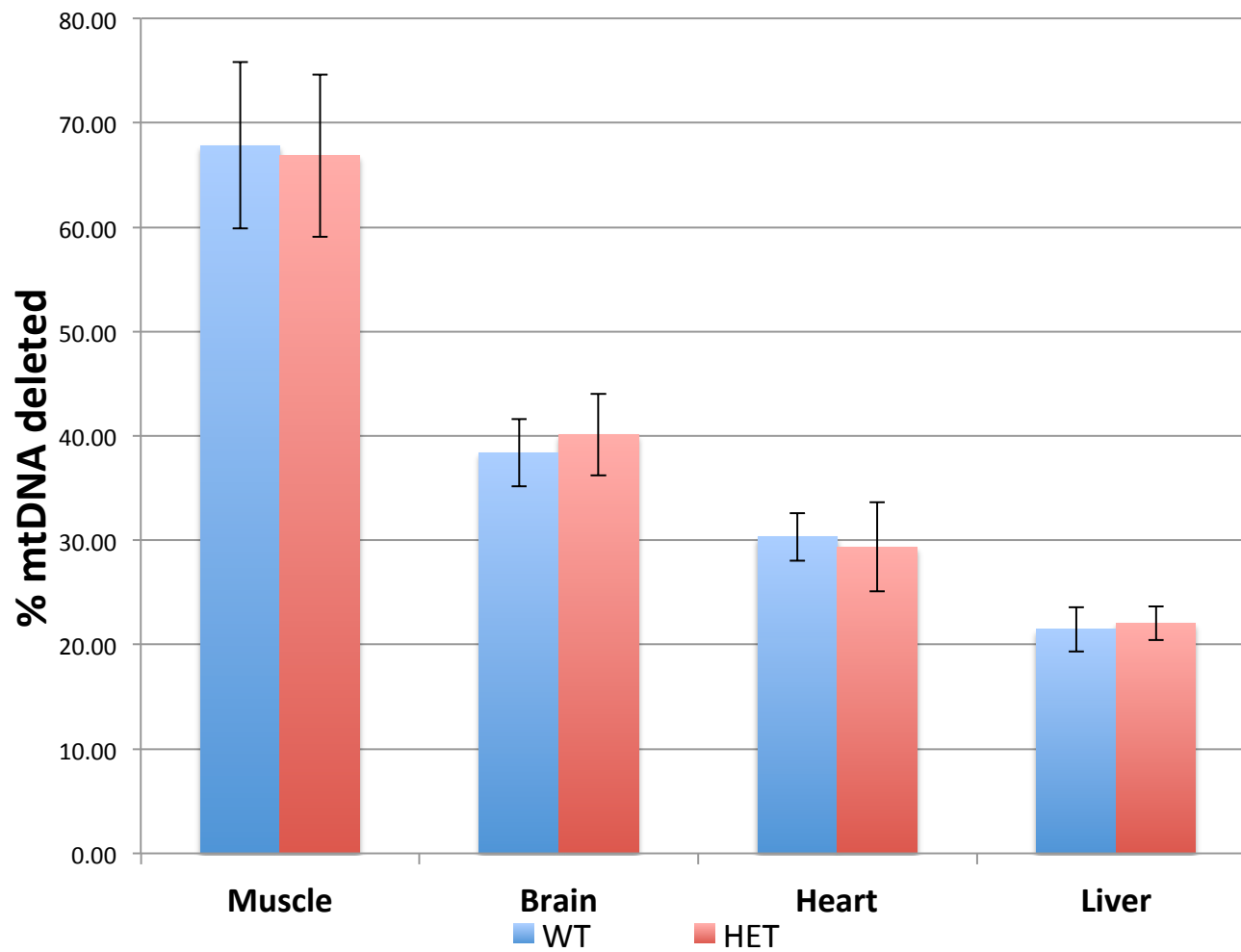


Supplemental figure 2



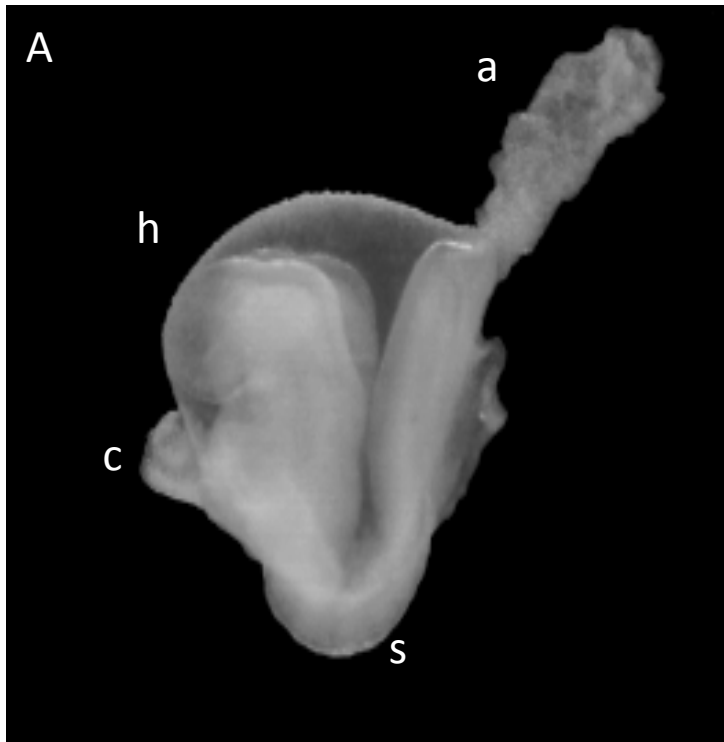
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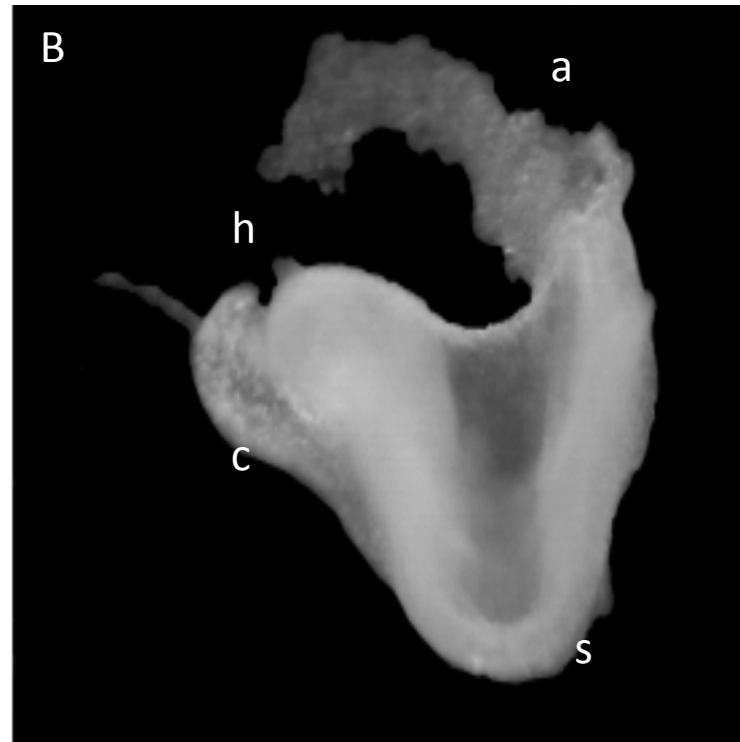


Supplemental figure 5

POLG2 +/+



POLG2 -/-



A.

<i>Polg2 +/-</i>	Body Weight (gm)	STD
40 wk WT Male	36.88	5.73
40 wk HET Male	39.48	6.34
40 wk WT Female	36.38	7.88
40 wk HET Female	26.43	3.71
2 yr WT Male	41.55	7.88
2 yr HET Male	38.60	4.73
2 yr WT Female	34.50	10.9
2 yr HET Female	36.27	7.03

B.

2 yr <i>Polg2 +/-</i>	Liver Weight (gm)	STD	Relative Liver Weight Ratio
WT Male	1.70	0.28	0.04 – 0.05
HET Male	2.23	0.70	0.04 – 0.11
WT Female	1.62	0.46	0.04 – 0.05
HET Female	1.60	0.21	0.04 – 0.05

Sample#	Group ID	Sex	ALT	SDH	AST	ALP	TBA	Chol	Trig	HDL	LDL	Ca	Creat	CK
			U/L	U/L	U/L	U/L	uMol/L	mg/dl	mg/dl	mg/dl	mg/dl	mg/dl	mg/dl	U/L
38-1	MPOLG2	M	483	351.2*	386	31	3.8	69	66	35	6	5.8	0.1	481
38-2	MPOLG2	M	79	65.8	75	57	39.2	108	68	52	8	7.4	0.1	83
38-3	MPOLG2	M	19	37.2	128	22	20.8	75	70	30	10	8.1	0.1	59
51-2	MPOLG2	M	133	93.2	102	41	6.3	99	67	44	8	7.4	0.1	178
53-1	MPOLG2	M	38	57.6	53	47	6.5	97	57	48	6	7.4	0.1	54
54-1	MPOLG2	M	83	146*	77	37	9.8	139	47	63	8	6.7	0.1	33
54-2	MPOLG2	M	29	48.5	41	42	6.7	114	51	60	6	7.0	0.1	114
54-3	MPOLG2	M	27	29.2	50	37	6.6	110	68	57	5	7.3	0.1	58
54-4	MPOLG2	M	132	285.1*	127	110	13.4	121	51	53	7	8.4	0.2	79
57-1	MPOLG2	M	218	qns	193	45	4.3	qns	33	21	6	5.7	qns	qns
62-1	MPOLG2	M	nsr	nsr	nsr	nsr	nsr	nsr	nsr	nsr	nsr	nsr	nsr	nsr
62-2	MPOLG2	M	nsr	nsr	nsr	nsr	nsr	nsr	nsr	nsr	nsr	nsr	nsr	nsr
67-2	MPOLG2	M	37	37.6	65	37	7.2	80	66	38	5	6.9	0.1	327
67-3	MPOLG2	M	nsr	nsr	nsr	nsr	nsr	nsr	nsr	nsr	nsr	nsr	nsr	nsr
n =		14	11	10	11	11	11	10	11	11	11	11	10	10
52609M1	WT	M	29	38.9	43	57	4.1	73	45	37	6	7.4	0.9	136
52609M2	WT	M	26	41.1	43	53	4.2	53	47	25	4	7.4	0.2	93
080309M1	WT	M	43	64.5	63	55	4.0	71	61	36	5	7.4	0.2	71
080309M2	WT	M	34	58.6	57	51	4.9	109	45	50	5	7.2	0.1	43
n =		4	4	4	4	4	4	4	4	4	4	4	4	4
MEANS														
Group ID	Sex	ALT	SDH	AST	ALP	TBA	Chol	Trig	HDL	LDL	Ca	Creat	CK	
MPOLG2	M	116	115.1	118	46	11.3	101	59	46	7	7.1	0.1	147	
WT	M	33	50.8	52	54	4.3	77	50	37	5	7.4	0.4	86	
STANDARD ERRORS														
MPOLG2	M	41	35.9	30	7	3.1	7	4	4	0	0.2	0.0	46	
WT	M	4	6.4	5	1	0.2	12	4	5	0	0.1	0.2	20	
Mann-Whitney p-value		0.186	0.635	0.074	0.057	0.026	0.106	0.087	0.280	0.040	0.492	0.031	0.839	
★Significant at p < 0.05						★				★		★		

Supplemental table 2

Sample#	Group ID	Sex	ALT	SDH	AST	ALP	TBA	Chol	Trig	HDL	LDL	Ca	Creat	CK
			U/L	U/L	U/L	U/L	uMol/L	mg/dl	mg/dl	mg/dl	mg/dl	mg/dl	mg/dl	U/L
23-5	MPOLG2	F	33	qns	87	292	17.1	107	50	49	9	9.0	0.1	956
23-6	MPOLG2	F	40	42.1	68	163	9.8	79	56	40	8	8.3	0.1	62
38-5	MPOLG2	F	48	40.5	95	178	12.8	65	57	34	5	6.9	0.04	67
38-6	MPOLG2	F	271	191.5*	273	56	17.8	66	60	33	5	6.4	0.1	88
38-8	MPOLG2	F	322	218.6*	346	32	9.5	44	53	22	5	3.9	<0.01	1366
53-5	MPOLG2	F	36	49.4	67	54	10.4	64	62	32	4	7.2	0.1	42
54-5	MPOLG2	F	39	47.1	70	95	5.4	82	51	42	5	7.6	0.1	49
57-2	MPOLG2	F	36	48.4	79	116	8.1	75	55	35	4	7.6	0.1	46
62-5	MPOLG2	F	nsr	nsr	nsr	nsr	nsr	nsr	nsr	nsr	nsr	nsr	nsr	nsr
67-8	MPOLG2	F	34	34.9	61	47	21.0	70	60	32	4	7.4	0.1	107
	n =	10	9	9	9	9	9	9	9	9	9	9	9	9
52609F1	WT	F	80	64.2	100	38	8.0	66	51	34	5	7.3	0.1	146
52609F2	WT	F	32	26.8	67	67	17.0	51	41	26	4	7.0	0.1	110
52609F3	WT	F	70	54.4	113	32	8.3	92	49	42	6	8.1	0.1	405
	n =	3	3	3	3	3	3	3	3	3	3	3	3	3

MEAN VALUES

Group ID	Sex	ALT	SDH	AST	ALP	TBA	Chol	Trig	HDL	LDL	Ca	Creat	CK
MPOLG2	F	95	84.1	127	115	12.4	72	56	35	5	7.1	0.1	309
WT	F	61	48.5	93	46	11.1	70	47	34	5	7.5	0.1	220

STANDARD ERRORS

MPOLG2	F	38	26.8	35	28	1.7	6	1	3	1	0.5	0.0	165
WT	F	15	11.2	14	11	3.0	12	3	5	1	0.3	0.0	93

Mann-Whitney p-value 0.973 1.000 0.759 0.155 0.482 0.896 0.027 0.973 1.000 0.973 0.796 0.209

★ Significant at p < 0.05

★

Lab Acc	ID #	Sex	WBC 10 ³ /ul	RBC 10 ⁶ /ml	Hgb g/dl	Hct %	Spun Hct %	MCV fL	MCH pg	MCHC g/dl	Plts 10 ³ /ul	Neut %	Lym %	Mono %	Eos %	Baso %	LUC %	Neut 10 ³ /ul	Lymph 10 ³ /ul	Mono 10 ³ /ul	Eos 10 ³ /ul	Baso 10 ³ /ul	LUC 10 ³ /ul	Retic %	Retic 10 ⁶ /ul		
MPOLG2																											
1687	38-1	M	3.8	8.43	12.6	37.9	37.0	44.9	14.9	33.2	959	25	65	7	0	0	3	1.0	2.5	0.3	0.0	0.0	0.1	2.5	0.2		
1689	38-2	M	9.4	9.71	14.7	45.7	44.0	47.1	15.1	32.2	1738	11	85	3	0	0	1	1.0	8.0	0.3	0.0	0.0	0.1	1.9	0.2		
1688	38-3	M	3.7	5.82	8.8	27.1	27.0	46.6	15.1	32.5	1767	45	50	4	0	0	1	1.7	1.9	0.1	0.0	0.0	0.0	9.0	0.5		
1692	51-2	M	4.5	6.28	11.1	34.4	33.5	54.7	17.7	32.3	1841	21	70	8	1	0	0	0.9	3.2	0.4	0.0	0.0	0.0	4.1	0.3		
1696	53-1	M	5.9	9.30	14.5	44.9	43.5	48.3	15.6	32.3	1558	17	75	6	0	0	2	1.0	4.5	0.4	0.0	0.0	0.1	2.1	0.2		
1698	54-1	M	4.5	9.48	14.8	43.6	43.5	46.0	15.6	33.9	1377	25	66	7	0	0	2	1.1	3.0	0.3	0.0	0.0	0.1	3.0	0.3		
1703	54-2	M	2.7	7.60	12.8	40.4	38.0	53.2	16.8	31.7	1468	46	53	1	0	0	0	1.3	1.4	0.0	0.0	0.0	0.0	1.2	0.1		
1707	54-3	M	4.1	9.27	13.4	41.6	41.5	44.9	14.5	32.2	2065	37	58	5	0	0	0	1.5	2.4	0.2	0.0	0.0	0.0	2.0	0.2		
1702	54-4	M	3.3	7.40	12.7	35.8	37.0	48.4	17.2	35.5	1189	27	68	2	2	0	1	0.9	2.2	0.1	0.1	0.0	0.0	2.9	0.2		
1706	57-1	M	3.8	9.69	14.6	44.5	44.5	45.9	15.1	32.8	1382	35	61	2	1	0	1	1.3	2.3	0.1	0.0	0.0	0.0	2.2	0.2		
1709	67-2	M	3.8	9.63	14.5	44.6	44.0	46.3	15.1	32.5	1910	33	61	4	0	0	2	1.2	2.3	0.2	0.0	0.0	0.1	2.4	0.2		
Mean			4.5	8.4	13.1	40.0	39.4	47.8	15.7	32.8	1568.5 45455	29.3	64.7	4.5	0.4	0.0	1.2	1.2	3.1	0.2	0.0	0.0	0.1	3.0	0.2		
SD			1.8	1.4	1.9	5.8	5.6	3.2	1.0	1.1	332	11.1	9.9	2.3	0.7	0.0	1.0	0.3	1.8	0.1	0.0	0.0	0.0	2.1	0.1		
n			11	11	11	11	11	11	11	11	11	11	11	11	11	11	11	11	11	11	11	11	11	11	11	11	
WT																											
1690	52609M1	M	7.2	9.33	14.2	43.6	43.0	46.7	15.2	32.6	1929	24	71	5	0	0	0	1.7	5.1	0.4	0.0	0.0	0.0	3.0	0.3		
1697	52609M2	M	4.0	9.14	13.6	41.1	41.0	45.0	14.9	33.1	1472	25	67	5	2	0	1	1.0	2.7	0.2	0.1	0.0	0.0	3.4	0.3		
1701	080309M1	M	4.5	9.26	14.0	41.1	42.0	44.4	15.1	34.1	1246	24	70	4	0	0	2	1.1	3.2	0.2	0.0	0.0	0.1	2.9	0.3		
1704	080309M2	M	3.8	9.01	13.3	40.8	40.0	45.3	14.8	32.6	1194	50	48	0	2	0	0	1.9	1.8	0.0	0.1	0.0	0.0	2.8	0.3		
Mean			4.9	9.2	13.8	41.7	41.5	45.4	15.0	33.1	1460	30.8	64.0	3.5	1.0	0.0	0.8	1.4	3.2	0.2	0.0	0.0	0.0	3.0	0.3		
SD			1.6	0.1	0.4	1.3	1.3	1.0	0.2	0.7	335	12.8	10.8	2.4	1.2	0.0	1.0	0.5	1.4	0.1	0.0	0.0	0.0	0.3	0.0		
n			4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	4	
MEAN																											
MPOLG2			4.5	8.42	13.1	40.0	39.4	47.8	15.7	32.8	1569	29	65	4	0	0	1	1.2	3.1	0.2	0.0	0.0	0.1	3.0	0.2		
WT			4.9	9.19	13.8	41.7	41.5	45.4	15.0	33.1	1460	31	64	4	1	0	1	1.4	3.2	0.2	0.0	0.0	0.0	3.0	0.3		
STANDARD ERROR																											
MPOLG2			0.5	0.43	0.6	1.7	1.7	1.0	0.3	0.3	100	3	3	1	0	0	0	0.1	0.5	0.0	0.0	0.0	0.0	0.6	0.0		
WT			0.8	0.07	0.2	0.7	0.6	0.5	0.1	0.4	167	6	5	1	1	0	0	0.2	0.7	0.1	0.0	0.0	0.0	0.1	0.0		
Mann-Whitney p-value			0.431	1.000	0.928	0.974	1.000	0.131	0.223	0.218	0.753	0.929	0.776	0.634	0.330	1.000	0.528	0.343	0.641	0.851	0.264	1.000	0.498	0.213	0.078		

★ Significant at p < 0.05

Lab	ID	Sex	WBC	RBC	Hgb	Hct	Spun	MCV	MCH	MCHC	Plts	Neut	Lym	Mono	Eos	Baso	LUC	Neut	Lymph	Mono	Eos	Baso	LUC	Retic	Retic		
Acc	#		10 ³ /ul	10 ⁶ /ml	g/dl	%	Hct %	fL	pg	g/dl	10 ³ /ul	%	%	%	%	%	%	10 ³ /ul	10 ³ /ul	10 ³ /ul	10 ³ /ul	10 ³ /ul	10 ³ /ul	%	10 ⁶ /ul		
MPOLG2																											
1685	23-6	F	3.2	8.37	13.4	40.4	40.0	48.3	16.0	33.2	1078	15	80	1	0	0	4	0.5	2.5	0.0	0.0	0.0	0.1	4.8	0.4		
1686	23-5	F	2.2	6.03	11.0	31.8	32.0	52.7	18.2	34.6	854	38	58	4	0	0	0	0.8	1.3	0.1	0.0	0.0	0.0	2.7	0.2		
1691	38-5	F	2.4	9.28	14.8	46.1	45.0	49.7	15.9	32.1	1345	32	66	2	0	0	0	0.8	1.6	0.0	0.0	0.0	0.0	1.9	0.2		
1693	38-6	F	3.5	7.60	12.6	37.3	37.5	49.1	16.6	33.8	776	21	75	3	0	0	1	0.7	2.7	0.1	0.0	0.0	0.0	1.9	0.1		
1694	38-8	F	3.7	7.70	12.7	38.2	38.0	49.6	16.5	33.2	758	15	82	1	1	0	1	0.6	3.0	0.0	0.0	0.0	0.0	3.1	0.2		
1699	53-5	F	4.3	9.28	15.1	45.2	45.0	48.7	16.3	33.4	1110	33	50	13	0	0	4	1.4	2.1	0.6	0.0	0.0	0.2	3.6	0.3		
1700	54-5	F	2.4	8.38	14.6	44.7	43.5	53.4	17.4	32.7	884	21	74	3	1	0	1	0.5	1.8	0.1	0.0	0.0	0.0	4.0	0.3		
1705	57-2	F	1.9	7.93	12.9	38.7	39.0	48.8	16.3	33.3	966	41	55	2	0	0	2	0.8	1.0	0.0	0.0	0.0	0.0	2.9	0.2		
1710	67-8	F	4.2	9.49	14.6	44.2	44.0	46.6	15.4	33.0	2035	33	61	3	0	0	3	1.4	2.6	0.1	0.0	0.0	0.1	2.8	0.3		
Mean			3.1	8.2	13.5	40.7	40.4	49.7	16.5	33.3	1090	27.7	66.8	3.6	0.2	0.0	1.8	0.8	2.1	0.1	0.0	0.0	0.1	3.1	0.3		
SD			0.9	1.1	1.4	4.7	4.4	2.1	0.8	0.7	400	9.8	11.5	3.7	0.4	0.0	1.6	0.3	0.7	0.2	0.0	0.0	0.1	0.9	0.1		
n			9	9	9	9	9	9	9	9	9	9	9	9	9	9	9	9	9	9	9	9	9	9	9	9	
WT																											
1684	52609F1	F	5.6	9.78	15.4	46.8	45.0	47.9	15.7	32.9	948	25	68	3	1	0	3	1.4	3.8	0.2	0.1	0.0	0.2	2.8	0.3		
1695	52609F2	F	1.8	8.23	12.9	38.2	38.0	46.4	15.7	33.8	1361	51	48	1	0	0	0	0.9	0.8	0.0	0.0	0.0	0.0	4.2	0.3		
1708	52609F3	F	8.2	8.65	13.5	40.6	41.0	46.9	15.6	33.3	1379	22	69	5	0	0	4	1.8	5.7	0.4	0.0	0.0	0.3	2.6	0.2		
Mean			5.2	8.9	13.9	41.9	41.3	47.1	15.7	33.3	1229	32.7	61.7	3.0	0.3	0.0	2.3	1.4	3.4	0.2	0.0	0.0	0.2	3.2	0.3		
SD			3.2	0.8	1.3	4.4	3.5	0.8	0.1	0.5	244	15.9	11.8	2.0	0.6	0.0	2.1	0.5	2.4	0.2	0.0	0.0	0.2	0.9	0.1		
n			3	3	3	3	3	3	3	3	3	3	3	3	3	3	3	3	3	3	3	3	3	3	3	3	
MEAN																											
	MPOLG2		3.1	8.23	13.5	40.7	40.4	49.7	16.5	33.3	1090	28	67	4	0	0	2	0.8	2.1	0.1	0.0	0.0	0.1	3.1	0.3		
	WT		5.2	8.89	13.9	41.9	41.3	47.1	15.7	33.3	1229	33	62	3	0	0	2	1.4	3.4	0.2	0.0	0.0	0.2	3.2	0.3		
STANDARD ERROR																											
	MPOLG2		0.3	0.36	0.5	1.6	1.5	0.7	0.3	0.2	133	3	4	1	0	0	1	0.1	0.2	0.1	0.0	0.0	0.0	0.3	0.0		
	WT		1.9	0.46	0.8	2.6	2.0	0.4	0.0	0.3	141	9	7	1	0	0	1	0.3	1.4	0.1	0.0	0.0	0.1	0.5	0.0		
Mann-Whitney p-value			0.482	0.350	0.500	0.632	0.746	0.036	0.055	0.814	0.282	0.573	0.600	0.882	1.000	1.000	0.727	0.100	0.482	0.727	0.673	1.000	0.477	1.000	0.600		

★Significant at p < 0.05

★