

Supporting Information

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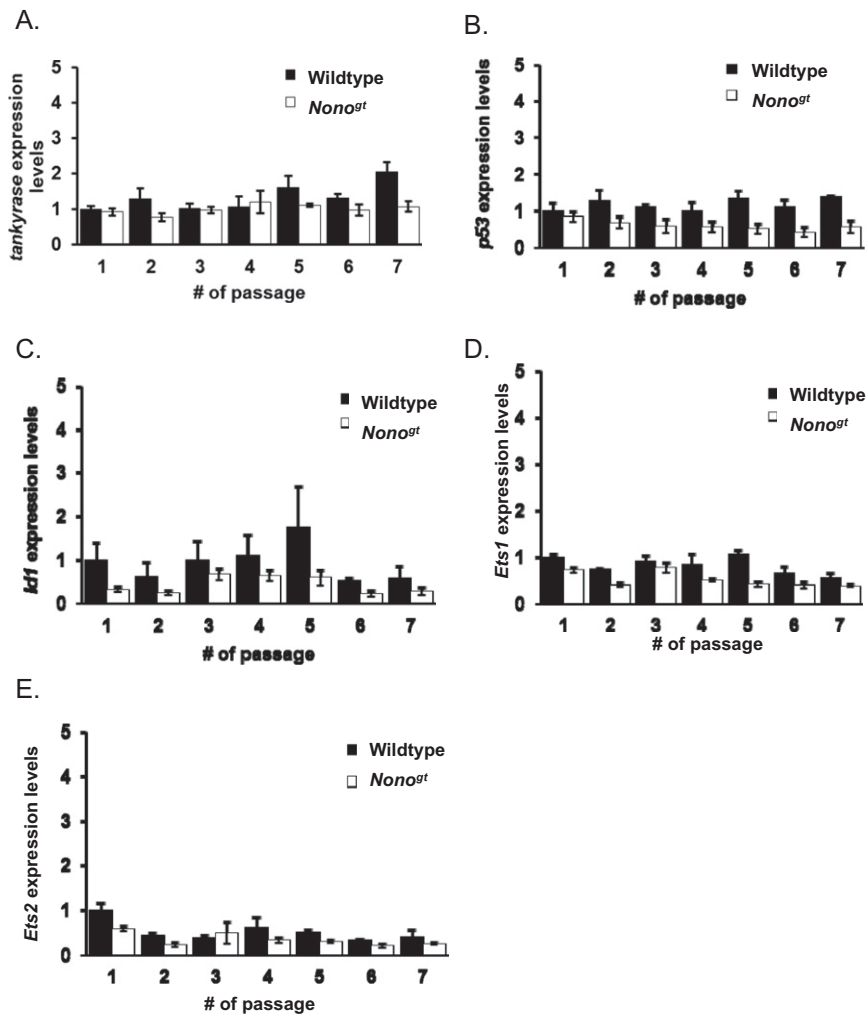


Fig. S1. Transcription of senescence-implicated genes in serially passaged WT and *Nono^{gt}* primary fibroblasts. WT and *Nono^{gt}* primary fibroblasts were counted and passaged every 2 d and a constant number of cells plated to a new dish. Total RNA was harvested from cells in each passage, and quantitative PCR (qPCR) was used to quantify the transcript levels of senescence-implicated genes *tankyrase* (A) and *p53* (B), as well as upstream regulators of *p16-Ink4A*, the genes *Id1* (C), *Ets1* (D), and *Ets2* (E). Values are plotted in arbitrary units relative to WT levels at passage 1.

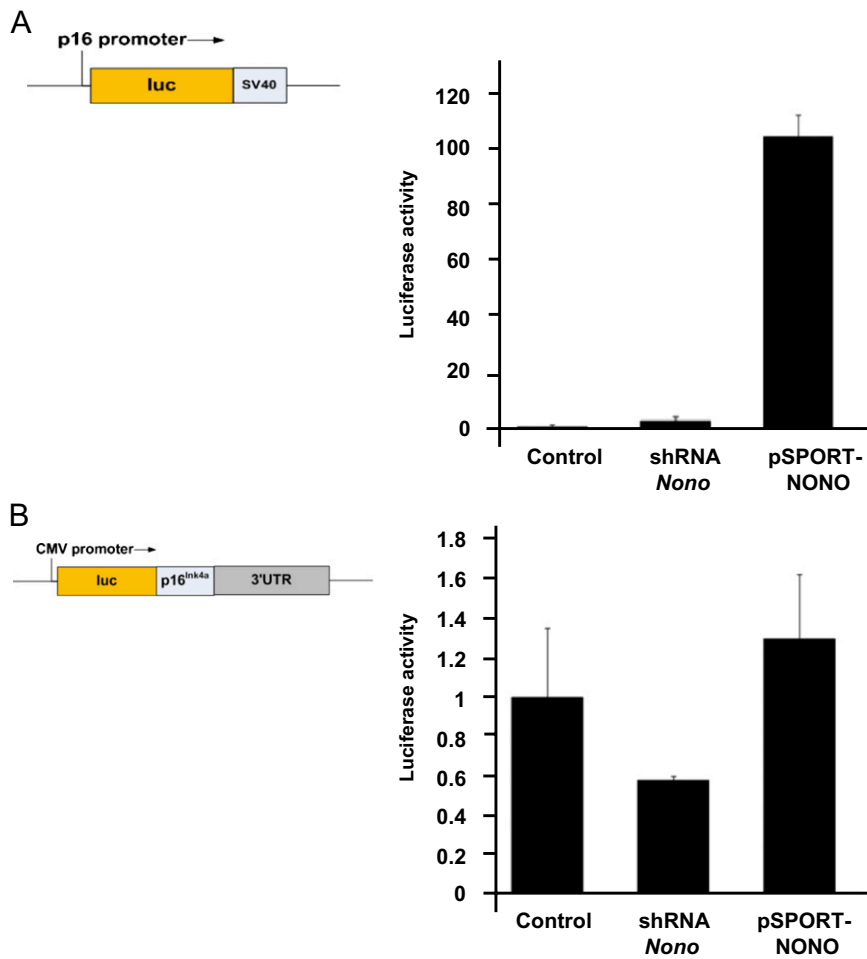


Fig. 52. NONO activates transcription of *p16-Ink4A* promoter reporters. (A) Relative expression levels of the diagrammed *p16-luc* promoter construct alone, in the presence of a NONO-targeting RNAi hairpin that reduces NONO levels 10-fold, and in the presence of a NONO-overexpressing vector. (B) Similar experiments using the diagrammed construct containing the *p16-Ink4A* 3' UTR.

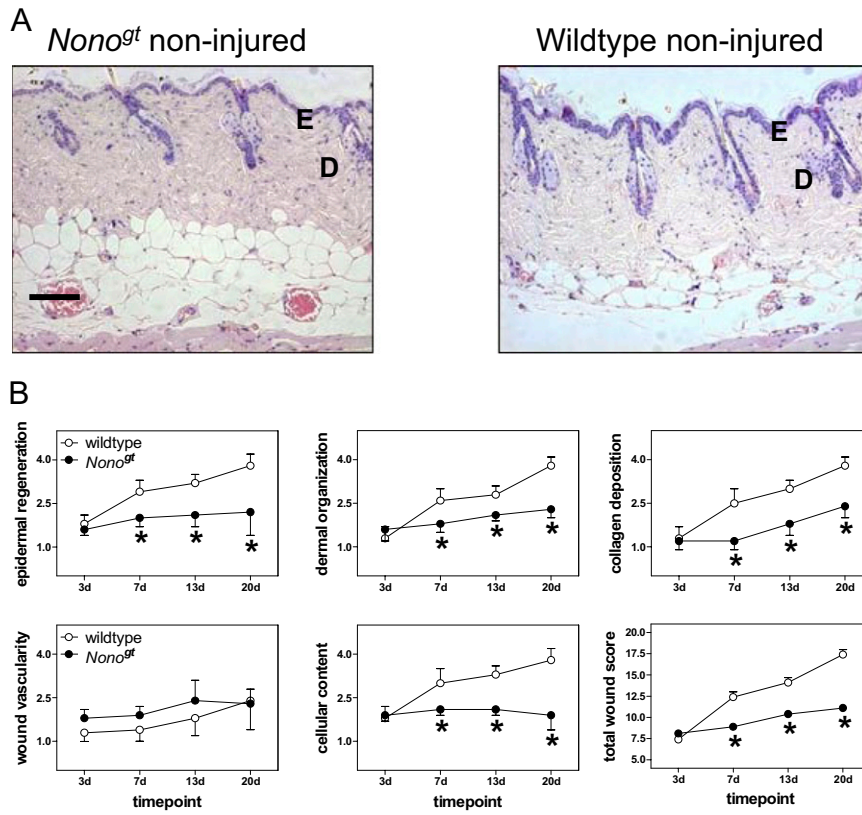


Fig. S3. Dermal structure and incisional wound healing in WT and *Nono^{gt}* mice. (A) H&E-stained paraffin sections from uninjured dorsal skin. Normal skin morphogenesis is not affected in *Nono^{gt}* mice compared with WT littermates (10–11 wk of age, 24 g body weight). (B) Full wound healing subscores of normal healing WT mice compared with *Nono^{gt}* mice on day 3, 7, 13, and 20 after incisional wounding in the dorsal skinfold. Data represent means ± SD (*n* = 5).

Table S1. Regulation of cell cycle genes in WT and *Nono*^{gt} fibroblasts

| Gene | Fold difference, <i>gt</i> /WT |
|-----------------|--------------------------------|
| <i>Abl1</i> | 2.11 |
| <i>Ak1</i> | 3.90 |
| <i>Apbb1</i> | 1.91 |
| <i>Atm</i> | 2.14 |
| <i>Brca1</i> | 3.09 |
| <i>Brca2</i> | 3.56 |
| <i>Camk2a</i> | 2.18 |
| <i>Camk2b</i> | 0.37 |
| <i>Casp3</i> | 2.67 |
| <i>Ccna1</i> | 0.37 |
| <i>Ccna2</i> | 19.68 |
| <i>Ccnb1</i> | 27.08 |
| <i>Ccnb2</i> | 10.26 |
| <i>Ccnc</i> | 5.27 |
| <i>Ccnd1</i> | 6.05 |
| <i>Ccne1</i> | 2.83 |
| <i>Ccnf</i> | 5.21 |
| <i>Cdc25a</i> | 2.58 |
| <i>Cdk2</i> | 12.98 |
| <i>Cdk4</i> | 4.51 |
| <i>Cdk5rap1</i> | 2.78 |
| <i>Cdkn1a</i> | 1.57 |
| <i>Cdkn1b</i> | 3.44 |
| <i>Cdkn2arf</i> | 1.85 |
| <i>Chek1</i> | 3.31 |
| <i>Cks1b</i> | 2.37 |
| <i>Ddit3</i> | 3.07 |
| <i>Dnajc2</i> | 1.73 |
| <i>Dst</i> | 1.12 |
| <i>E2f1</i> | 1.64 |
| <i>E2f2</i> | 0.37 |
| <i>E2f3</i> | 6.78 |
| <i>E2f4</i> | 1.70 |
| <i>Gadd45a</i> | 3.13 |
| <i>Gpr132</i> | 0.37 |
| <i>Hus1</i> | 3.04 |
| <i>Inha</i> | 0.82 |
| <i>Itgb1</i> | 1.81 |
| <i>Macf1</i> | 2.72 |
| <i>Mad2l1</i> | 2.93 |
| <i>Mcm2</i> | 7.62 |
| <i>Mcm3</i> | 9.82 |
| <i>Mcm4</i> | 5.67 |
| <i>Mdm2</i> | 2.94 |
| <i>Mki67</i> | 5.58 |
| <i>Mre11a</i> | 1.16 |
| <i>Msh2</i> | 1.57 |
| <i>Mtbp</i> | 4.46 |
| <i>Myb</i> | 0.17 |
| <i>Nek2</i> | 3.11 |
| <i>Nfatc1</i> | 2.82 |
| <i>Notch2</i> | 4.04 |
| <i>Npm2</i> | 1.06 |
| <i>Pcna</i> | 3.95 |
| <i>Pes1</i> | 2.48 |
| <i>Pkd1</i> | 1.93 |
| <i>Pmp22</i> | 1.74 |
| <i>Ppm1d</i> | 2.03 |
| <i>Ppp2r3a</i> | 0.90 |
| <i>Ppp3ca</i> | 1.86 |
| <i>Prm1</i> | 0.37 |
| <i>Rad17</i> | 3.17 |

Table S1. Cont.

| Gene | Fold difference, <i>gt</i> /WT |
|-----------------|--------------------------------|
| <i>Rad21</i> | 2.38 |
| <i>Rad51</i> | 0.43 |
| <i>Rad9</i> | 2.66 |
| <i>Ran</i> | 3.18 |
| <i>Rbl1</i> | 0.94 |
| <i>Rbl2</i> | 1.04 |
| <i>Sesn2</i> | 1.53 |
| <i>Sfn</i> | 2.05 |
| <i>Shc1</i> | 5.39 |
| <i>Skp2</i> | 3.24 |
| <i>Slfn1</i> | 0.37 |
| <i>Smc1a</i> | 5.86 |
| <i>Stag1</i> | 3.45 |
| <i>Sumo1</i> | 3.45 |
| <i>Taf10</i> | 3.08 |
| <i>Terf1</i> | 2.62 |
| <i>Tfdp1</i> | 3.53 |
| <i>Psmg2</i> | 2.74 |
| <i>Trp53</i> | 3.18 |
| <i>Trp63</i> | 0.15 |
| <i>Tsg101</i> | 1.25 |
| <i>Wee1</i> | 1.23 |
| <i>Gusb</i> | 5.23 |
| <i>Hprt1</i> | 0.01 |
| <i>Hsp90ab1</i> | 3.04 |
| <i>Gapdh</i> | 2.53 |
| <i>Actb</i> | 2.77 |

Total RNA was harvested from dividing cultures of WT and *Nono^{gt}* primary fibroblasts and subjected to qPCR array analysis. All array targets are shown, with fold-regulation of *Nono^{gt}* vs. WT fibroblasts. Note that the p16-*Ink4A* transcript itself was not probed on this commercial array. Independent qPCR analysis using the primers in Table S2 confirmed fivefold reduction in expression of p16-*Ink4A* in *Nono^{gt}* vs. WT fibroblasts in both replicates of the cellular RNA used for this array.

Table S2. Primer sequences

| Gene | Orientation | Sequence (5'–3') |
|------------------|-------------|---|
| <i>Ets1</i> | Sense | CGG CAT CAT AGC ACA GTT CAA G |
| <i>Ets1</i> | Antisense | CCC ATG CAA ACG GCT TTT AT |
| <i>Ets1</i> | Probe | FAM-AAC CGC TAC CCG AAA CAT GGA AGA CTC AG-TAMRA |
| <i>Id1</i> | Primer Set | 1) Assay ID: Mm00775963_g1 |
| <i>Ets2</i> | Primer Set | 1) Assay ID: Mm00468972_m1 |
| <i>NONO</i> | Sense | TGC GCT TCG CCT GTC A |
| <i>NONO</i> | Antisense | GCA GTT CGT TCG ACA GTA CTG |
| <i>NONO</i> | Probe | FAM-AGT GCA CCC TTA CAG TCC GCA ACC TT-TAMRA |
| <i>qPCR</i> | | |
| <i>p16-Ink4A</i> | Sense | CCC AAC GCC CCG AAC T |
| <i>p16-Ink4A</i> | Antisense | GTG AAC GTT GCC CAT CAT CA |
| <i>p16-Ink4A</i> | Probe | FAM-TTT CGG TCG TAC CCC GAT TCA GG-TAMRA |
| <i>ChIP</i> | | |
| <i>p16-Ink4A</i> | Sense | TTT CGC CCA ACG CCC CGA A |
| <i>p16-Ink4A</i> | Antisense | ACC CGA CTG CAG ATG GGA CAC |
| <i>p16-Ink4A</i> | Probe | FAM- CGA ACT CTT TCG GTC GTA CCC CGA TTC-TAMRA |
| <i>p53</i> | Sense | GCA TCC CGT CCC CAT CA |
| <i>p53</i> | Antisense | GGA TTG TGT CTC AGC CCT GAA G |
| <i>p53</i> | Probe | FAM-CAG CCT CCC CCT CTC CTT GCT GTC TTA-TAMRA |
| <i>Tankyrase</i> | Sense | CGG CAG CAG AGC AGA AGA C |
| <i>Tankyrase</i> | Antisense | TGT ACT CCA GTT GCA GGT TTG AAT |
| <i>Tankyrase</i> | Probe | TAG TGA CCA CCC CTG GTA AAG GCC AGA-TAMRA |
| <i>GAPDH</i> | Sense | CAT GGC CTT CCG TGT TCC TA |
| <i>GAPDH</i> | Antisense | CCT GCT TCA CCA CCT TCT TGA |
| <i>GAPDH</i> | Probe | YAK-CCG CCT GGA GAA ACC TGC CAA GTA TG-TAMRA |

TaqMan Gene Expression Assays from Applied Biosystems are tested, but sequences are not provided. Assay consists of primer forward, primer reverse, and probe, as usual.

Table S3. Histological wound healing scores

| Score | Dermal organization | Epidermal regeneration | Collagen deposition | Cellular content | Wound vascularity |
|-------|--|--|--|--|----------------------------------|
| 1 | 25% thickness of granulation tissue compared with healthy tissue | No epithelial closure | Gap without ingrowing collagen fibrils | Low cell proliferation, mainly inflammatory cells | 1–3 capillaries per visual field |
| 2 | 50% thickness of granulation tissue compared with healthy tissue | Strong hyperproliferative epithelium | Gap with ingrowing collagen fibrils | Predominantly inflammatory cells or dysfunctional fibroblasts, hyper-proliferation | 4–6 capillaries per visual field |
| 3 | 75% thickness of granulation tissue compared with healthy tissue | Moderate hyperproliferative epithelium | No gap, but unstable adhesion | Predominantly normal fibroblasts | 7–9 capillaries per visual field |
| 4 | Thickness of granulation tissue equal to healthy tissue | Thickness and structure equal to normal epithelium | No gap, stable adhesion | Low cell proliferation, mainly fibroblasts | >9 capillaries per visual field |