

Supplementary Figure 1. Cryo-electron tomography and GRAFIX optimization of COPII cages. a) Tomogram of Sec13-31 cages viewed along the air/water interface. The tomogram shows a large amount of disordered protein at the air/water interface. The cages are obscured by the large mass of disordered protein. b) CryoEM image of cages before fixation. c) Cages after GRAFIX preparation. The cages are more distinct and featured after gradient fixation.

Supplementary Figure 2. Sec13-31 homology model before and after flexible fitting. a) Human homology of Sec13-31. Loops colored in red were unresolved in the yeast crystal structure and were modeled in. b) The per-atom average RMSD after symmetrically realigning the 24 flexibly fitted Sec13-31 edges mapped onto an averaged crystal structure of the edges. The mean RMSD is 1.173 Å. c) The 24 symmetrically realigned flexibly fitted Sec13-31 edges superimposed.

Supplementary Figure 3 12 Å reconstruction of the Sec13-31 COPII cage fitted with the pseudo-atomic structure. The cryoEM density map is radially colored. The entire flexibly fitted pseudo-atomic model of human Sec13-31 is colored as in Figure 6.

Supplementary Figure 4. See Supplemental Data file. Deuterium uptake kinetics. a)

Deuterium incorporation vs. H/D exchange period (in hours) for segments of edge and cage forms of Sec31. Blue indicates edge and magenta indicates cage. b) Deuterium incorporation vs. H/D exchange period (in hours) for segments of edge and cage forms of Sec13. Blue indicates edge and magenta indicates cage.

Supplementary Figure 5. Difference in deuterium uptake between Sec13-31 edge and cage, for all peptides. The deuterium uptake difference for each peptide is calculated by the ARDD method as in Fig. 6.

Supplementary Figure 6. Isotopic distributions (normalized) of peptide 439-456, showing a bimodal distribution for cage (right) but not edge (left) forms. This peptide is located in the Sec13-31 hinge region.

Supplementary Movie 1. 12 Å reconstruction of the Sec13-31 COPII cage, radially colored. The human Sec13-31 homology model, colored by chain, is then docked into the density map. An asymmetric unit of the density map is segmented out and MDFF is applied to the homology model. Octahedral symmetry is imposed when introducing 23 additional copies of the Sec13-31 atomic model. The 24 Sec13-31 atomic models are again subjected to MDFF. Multiple views of the resulting atomic model are shown at a vertex, colored as in Figure 6. At the end of the movie, an individual flexibly fitted Sec13- 31 unit is shown. The two Sec13 chains are aligned, superimposed, and colored as in Figure 2. The resulting hinge with a range of 17° in the plane normal to the 4-fold axis is shown.

Supplementary Note

Flexible Fitting

The flexible fitting processes were performed by use of Molecular Dynamics Flexible Fitting (MDFF)¹. The NAMD configuration files were created from within Visual Molecular Dynamics $(VMD)^2$.

MDFF uses a user-defined 3D density map to generate a grid based set of force field vectors that are proportional to the gradient of the density map. Given an atomic model that has been rigidly fit to said density map, MDFF imposes molecular dynamics simulations with the additional grid potential:

$$
U_{EM}(\overline{R}) = \xi \sum_{j} \omega_j \begin{cases} \frac{1 - \phi(\overline{r_j}) - \phi_{thr}}{\phi_{max} - \phi_{thr}} & \text{if } \phi(\overline{r_j}) \geq \phi_{thr}, \\ 1 & \text{if } \phi(\overline{r_j}) < \phi_{thr}. \end{cases}
$$

ξ is a global scaling factor, $ω_j$ is a per-atom scaling factor, $φ(r_j)$ is the potential at atom *j*, ϕ _{max} is the maximum value of $\phi(r)$, and ϕ_{thr} is the user-defined minimum potential threshold.

In addition, secondary structures are identified and locally restrained during the molecular dynamics simulations. The following MDFF simulations were carried out *in vacuo* at a temperature of 300 K with the scaling factor ξ = 0.3 kcal/mol and with 1 fs time steps.

The flexible fitting was done in two steps. The first step was to flexibly fit the Sec13-31 homology model to an asymmetric unit of density map. UCSF Chimera 3 was used to obtain this density map. An initial rigidly fit model of the COPII cage was made by use of the fit command in Chimera. The Color Zone feature in Chimera was used in conjunction with the initial model and the COPII cage 3D reconstruction to extract a density map consisting of one asymmetric unit. Using the fit command in Chimera, the Sec13-31 homology model was rigidly fitted to the asymmetric density map. We used MDFF to flexibly fit the rigidly fit Sec13-31 homology model to the asymmetric density map. During this process, the energy of the model was first minimized for 200 fs. The model was then subjected to the molecular dynamics simulation for 70.2 ps by use of 20 processors at FSU's High Performance Computing (HPC) cluster.

For the second step in the flexible fitting process, we used the fit and symmetry commands in Chimera to create an initial model of the COPII cage by fitting 23 additional flexibly fit asymmetric models into the COPII cage density map. To relieve potential conflicting areas at and near the 12 vertices of the COPII cage, MDFF was again used to flexibly fit this model into the COPII cage density map. The energy was first minimized for 1 ps, followed by 134.5 ps of molecular dynamics simulation by use of 72 processors of FSU's HPC cluster.

Michel Sanner's Molecular Surface (MSMS) was used to create a computational model analogous to H/D exchange ⁴. The solvent-accessible surface area (SASA) was computed for a single, flexibly fit COPII vertex and for each individual Sec13-31 asymmetric unit for which the vertex is comprised. The SASA of the vertex was subtracted from the combined SASAs of the constituent asymmetric units of the vertex.

References

- 1. Trabuco, L. G., Villa, E., Mitra, K., Frank, J. & Schulten, K. Flexible fitting of atomic structures into electron microscopy maps using molecular dynamics. *Structure* **16,** 673-683 (2008).
- 2. Humphrey, W., Dalke, A. & Schulten, K. VMD: visual molecular dynamics. *J Mol Graph* **14,** 33-8, 27-8 (1996).
- 3. Pettersen, E. F., Goddard, T. D., Huang, C. C., Couch, G. S., *et al.* UCSF Chimera A Visualization System for Exploratory Research and Analysis. *J. Comput. Chem.* **25,** 1605-1612 (2004).
- 4. Sanner, M. F., Olson, A. J. & Spehner, J. C. Reduced surface: an efficient way to compute molecular surfaces. *Biopolymers* **38,** 305-320 (1996).

Supplementary Table 1. Relative difference in HDX between the unassembled

Sec13/31 edge and the assembled COPII cage for all HDX fragments. Positive percent relative difference values indicate a fragment that underwent greater HDX in the edge than in the cage. CC, Conformational Change; FX, Flexibility; B, Buried; UB, Unburied; ME, Minus End; PE, Positive End.