

Supplementary Table III - Overall numbers of genes upregulated or downregulated more than 2 fold in *teb1-1* compared to wt.

	25 °C	36 °C	Both temperatures
Genes upregulated more than 2 fold	244	224	152
Genes downregulated more than 2 fold	84	41	29

Supplementary Table IV - Iron metabolism genes downregulated in *teb1-1* strain at 25°C and 36°C

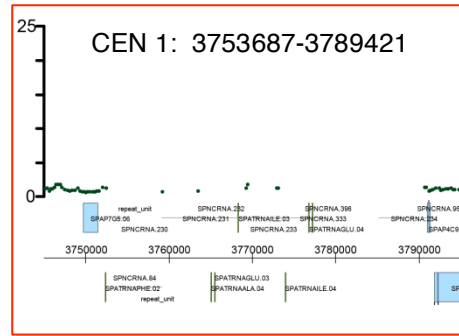
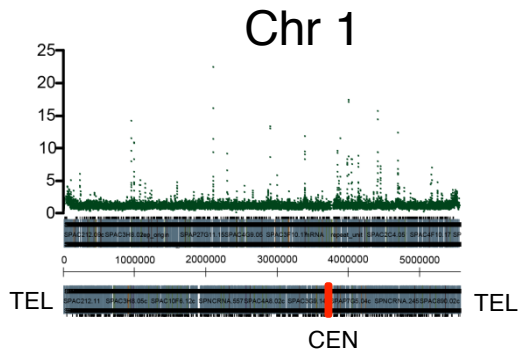
<i>Iron metabolism gene</i>	Gene expression compared to wt (microarray cells 25°C)	Gene expression compared to wt (microarray cells 36°C)
<i>str1</i>	0.293	0.177
<i>str3</i>	0.214	0.1395
<i>fip1</i>	0.4485	0.4705
<i>frp1</i>	0.212	0.2215

Supplementary Table V – Histone average expression levels in *teb1-1* strain at 25°C and 36°C

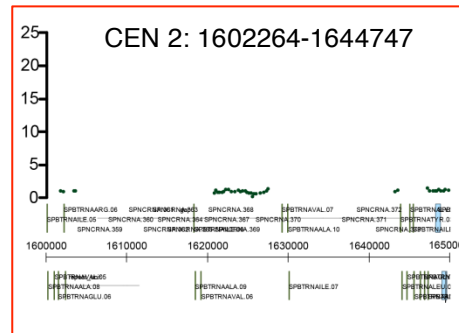
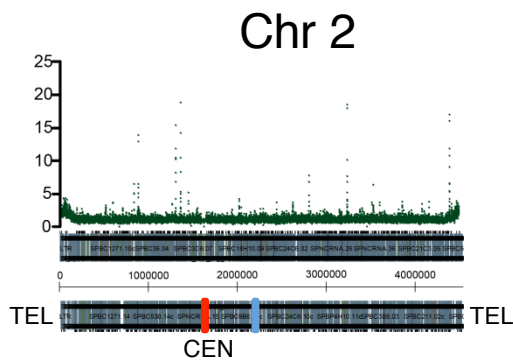
Histone genes	Gene expression compared to wt (microarray – cells at 25°C)	Gene expression compared to wt (microarray – cells at 36°C)
<i>hta1</i>	0.77	1.022
<i>htb1</i>	0.6025	0.697
<i>hta2</i>	0.286	0.3395
h3.1: <i>hht1</i>	0.3915	0.4505
h3.2: <i>hht2</i>	0.4215	0.5745
h3.3: <i>hht3</i>	0.3135	0.37
h4.1: <i>hhf1</i>	0.321	0.3465
h4.2: <i>hhf2</i>	0.409	0.5155
h4.3: <i>hhf3</i>	0.4115	0.429

Figure S1

A



B



C

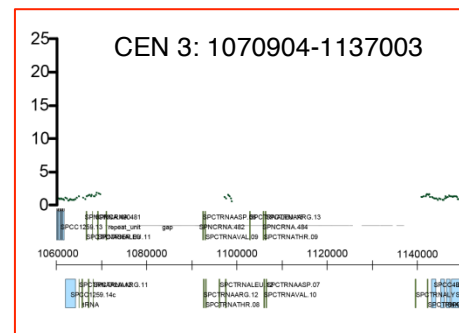
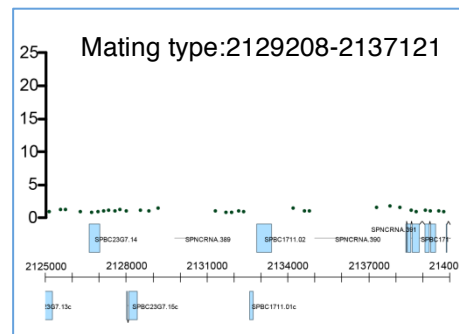
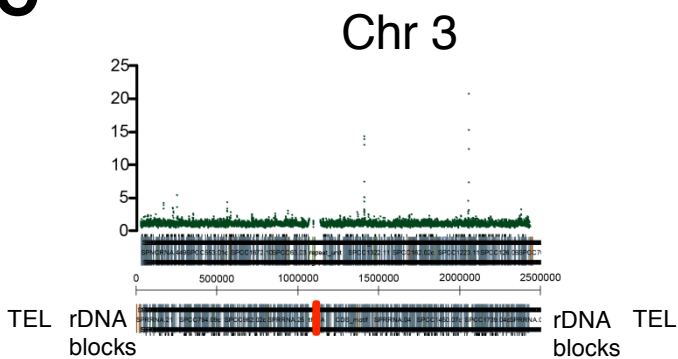
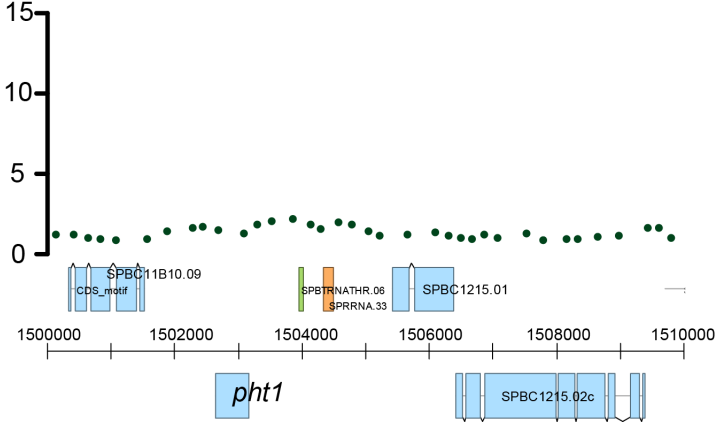
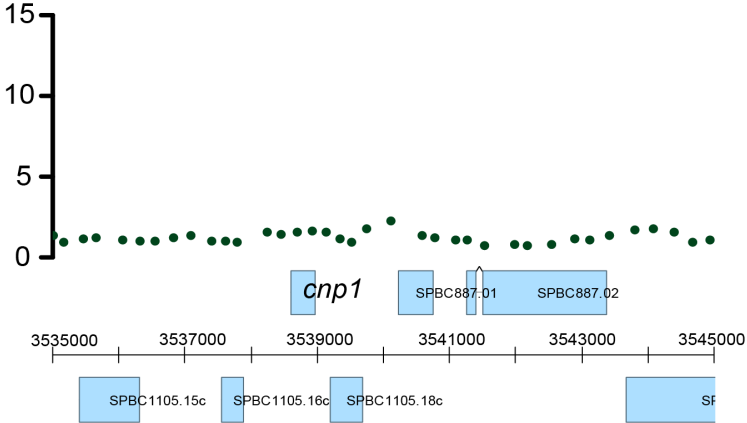


Figure S2

A



B



C

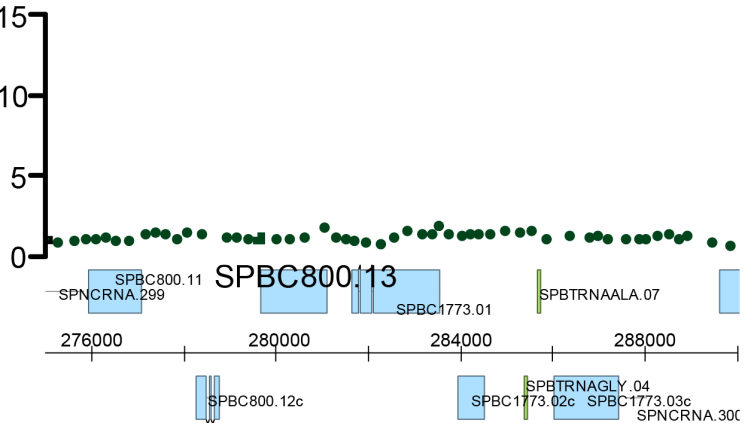


Figure S3

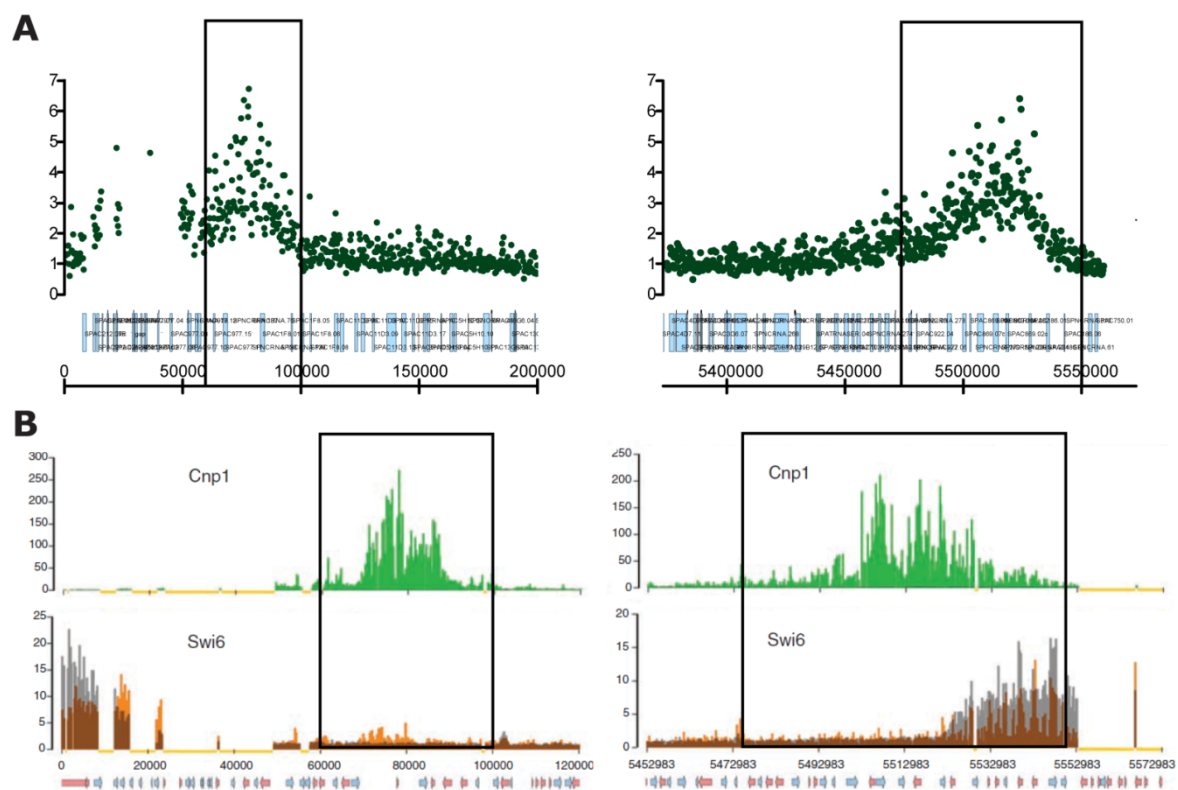
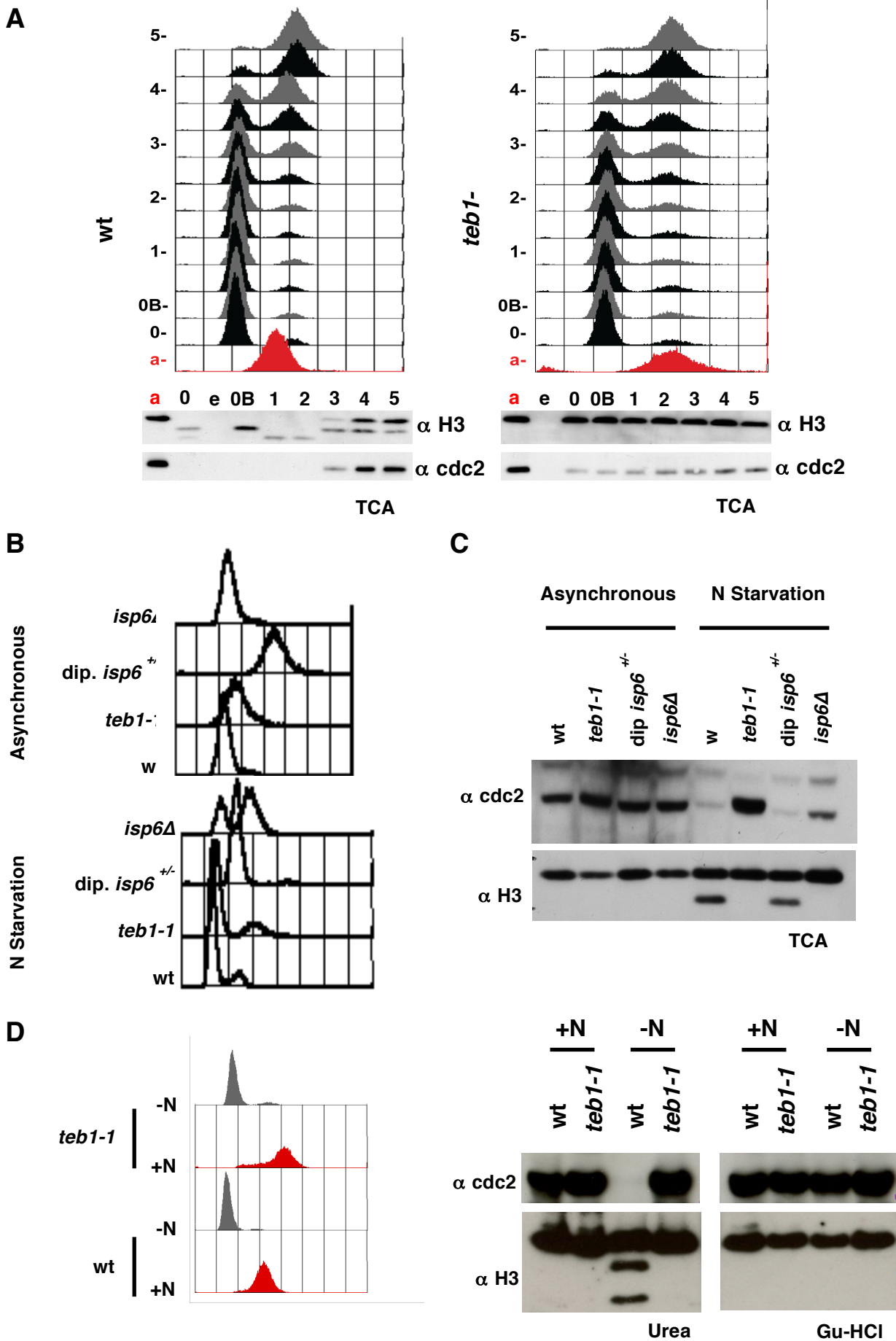


Figure S4



Supplementary Figure Legends

Figure S1. *Teb1* binds specific genomic loci. Representation of ChIP-chip data demonstrating *Teb1* binding to specific regions across the genome, and the absence of binding at the non-repetitive regions of the centromere. Repetitive regions were not included in the analysis as these sequences are absent from the microarray data. (A) *Teb1* binding at Chr 1 and its centromere (B) *Teb1* binding on Chr 2, which contains also the mating type locus. (C) *Teb1* binding on Chr 3.

Figure S2. *Teb1* does not bind promoters of the histone variants (A) *pht1* (H2A variant); (B) *cnp1* and (C) SPBC800.13 (H4 variant).

Figure S3. *Teb1* binds the same regions near the ends of Chr I that are bound by CENP-A upon neocentromere formation. (A) Representation of ChIP-chip data highlighting *Teb1* binding near the left and right termini of Chr 1. (B) Sites of *Cnp1* or *Swi6* binding after centromere disruption and neo-centromere formation on Chr 1 (from *Ishii et al*, 2008). Boxes highlighting regions of the left and right arms depict the same positions shown in the boxes in (A), allowing direct comparison.

Figure S4. Histone H3 is degraded in a *Teb1* and *Isp6*-dependent manner during extraction of G1-arrested cells, but not extracted with Gu-HCl. (A) Top: FACS analysis of wt and *teb1-1* cells arrested in G1 by nitrogen starvation for 16h at 25°C, shifted to 36°C for 1.5h, re-fed to allow resumption of growth and sampled every 30 min for 5 hours. Bottom: Western blot of TCA extracts from the following: (a) asynchronously growing cells; (0) cells starved for nitrogen for 16h; (OB) the same cells 1.5h at 36 °C, before re-feeding; (1 through 5) following re-feeding for 1 to 5 hours as indicated. Both wt and *teb1-1* cells recover efficiently from G1 arrest. Lane (e) was used for the protein marker. Both *Cdc2* proteolysis and histone

H3 clipping occur in extracts of wt G1 arrested cells (left panel). However, neither histone H3 clipping nor Cdc2 proteolysis occur in extracts of *teb1-1* cells (right panel). **(B)** Isp6 is required for histone clipping in extracts of G1-arrested cells. Asynchronous or nitrogen starved (16 hours) cultures of cells of the indicated genotypes were analysed by FACS. **(C)** Western blot analysis reveals that impairment of Isp6 function abolishes histone clipping in these TCA extracts. **(D)** Histone H3 clipping is detected in extracts of nitrogen starved cells prepared in the presence of 8M Urea but not in the presence of Gu-HCl.