

23RSS

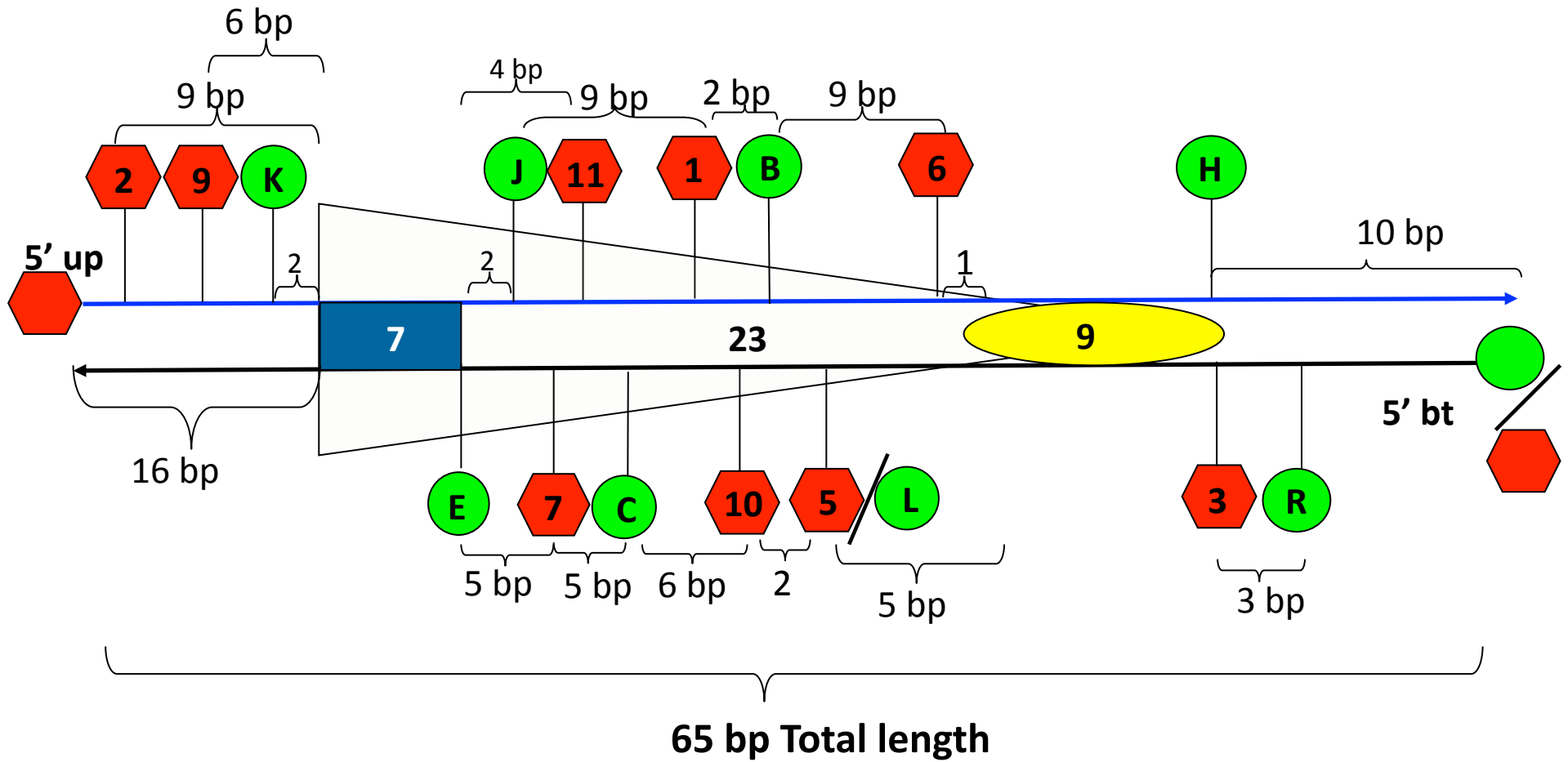


Fig. S1

● = FAM

⬡ = TAMRA

Figure S1. Location of the fluorophore-labeling positions used in this study. Donor labeling positions (green) are designated with a letter, and acceptor labeling positions (red) are designated with a number, while labels at the 5' ends of the two DNA strands are indicated as 5' up for the top strand and 5' bt for the bottom strand.

Fig. S2

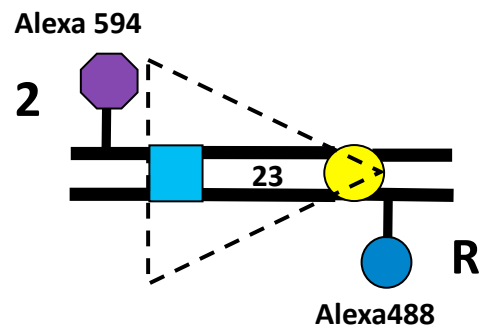
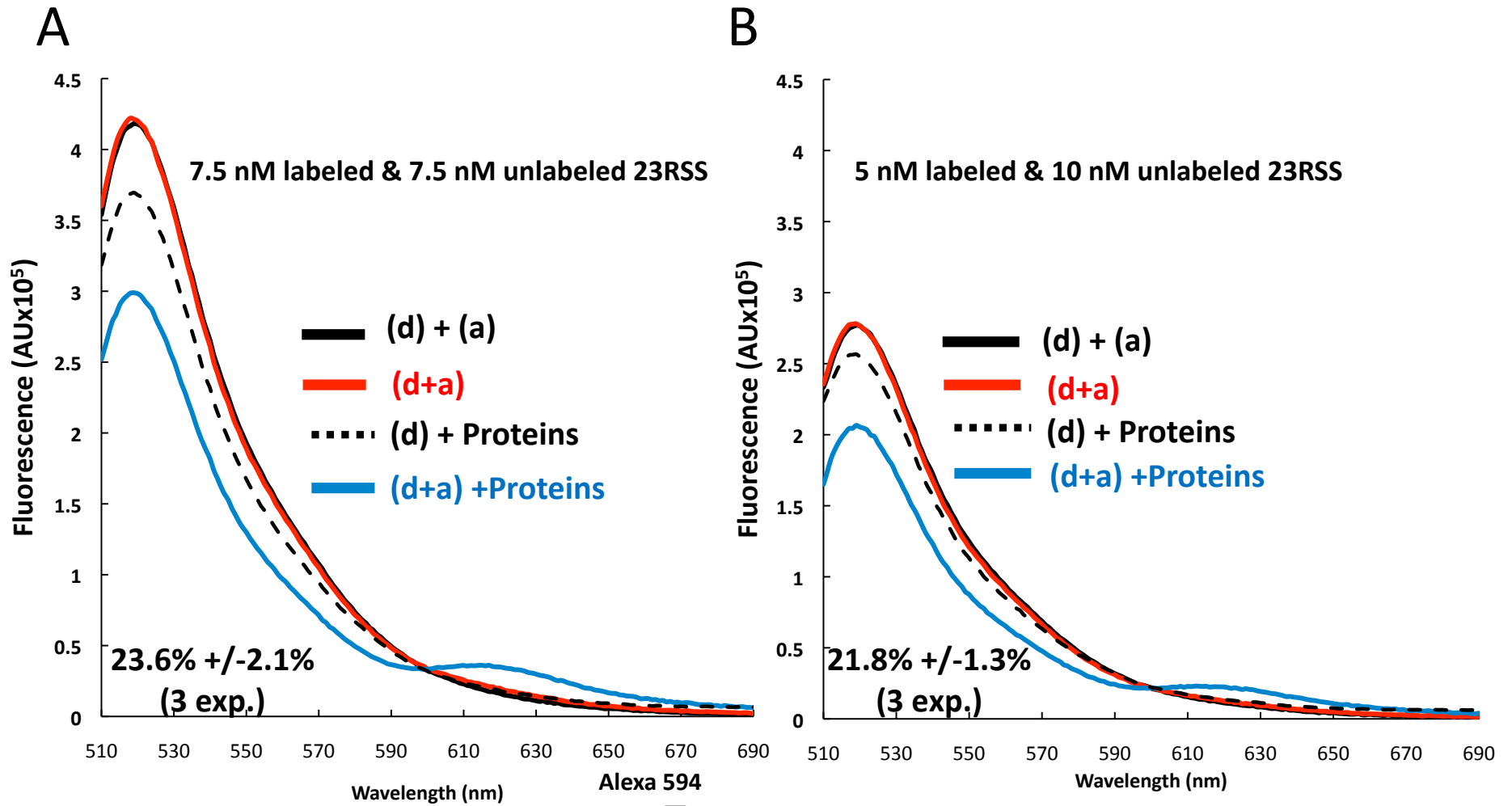


Figure S2. Analysis of energy transfer when the 23RSS_dR2_a substrate is diluted with unlabeled 23RSS. FRET experiments were performed as in Fig. 2E (co-expressed MBP-RAG1c and MBP-RAG2c proteins) but instead of 15 nM 23RSS_dR2_a substrate as in Fig. 2E, 7.5 nM 23RSS_dR2_a substrate and 7.5 nM unlabeled 23RSS (A), or 5 nM 23RSS_dR2_a substrate and 10 nM unlabeled 23RSS (B) were used. Data are displayed as in Fig. 2E. Robust energy transfer is detected even after dilution of the labeled substrate, strongly arguing against a major contribution of energy transfer between different 23RSS molecules (in *trans*).

Nonamer Intrinsic Bend

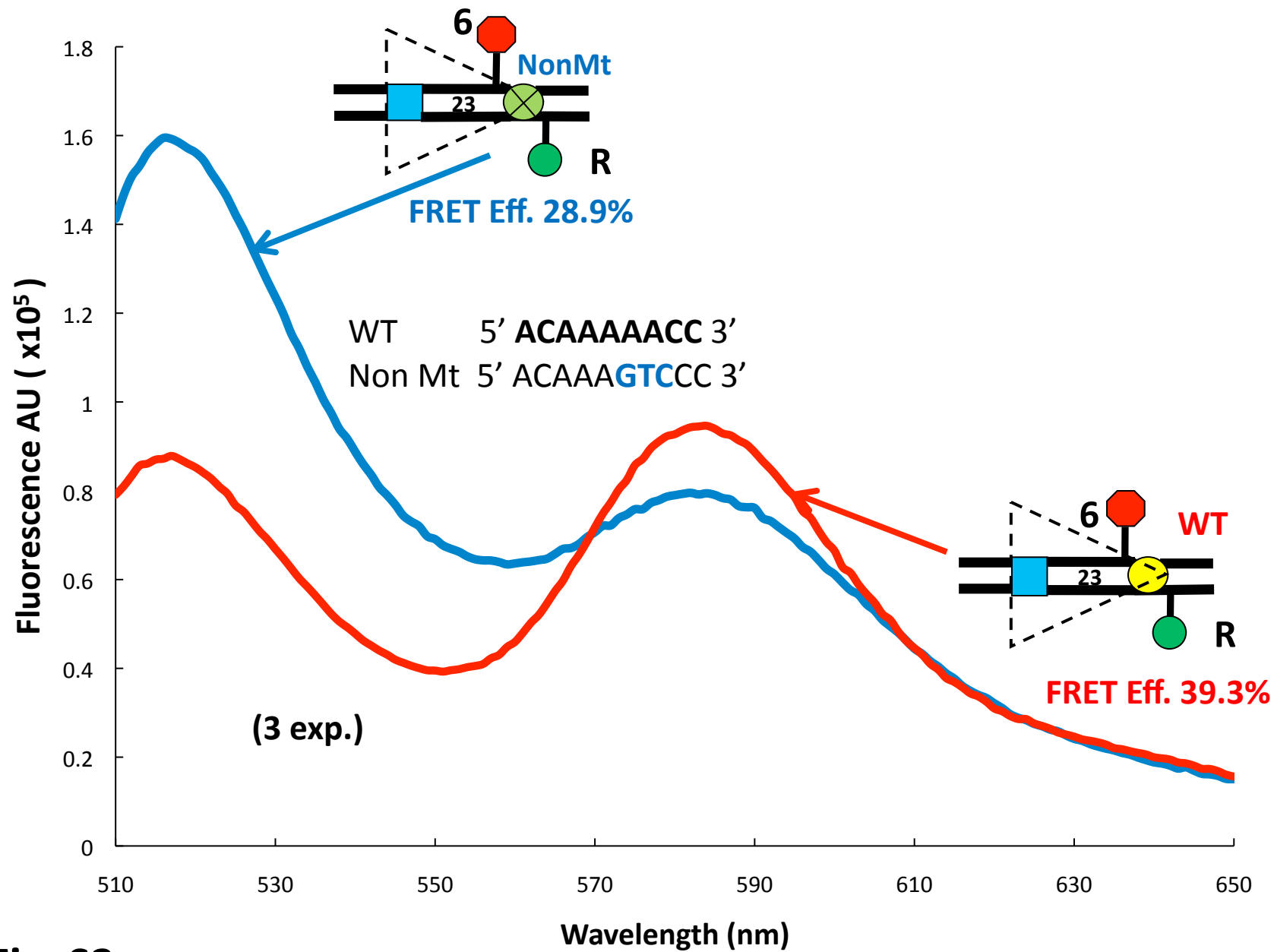


Fig. S3

Figure S3. Evidence for an intrinsic bend in the 23RSS nonamer. Emission spectra are shown for the 23RSS_dR6_a substrate (red) and the 23RSS_dR6_a NonMt substrate (blue) in which three bp in the A₅ tract of the nonamer have been mutated, as indicated. The fluorophores in this substrate are positioned so that the predicted intrinsic bend associated with an A₅ tract (36) would bring them closer together. The decrease in E-FRET observed in the 23RSS_dR6_a NonMt substrate as compared to the 23RSS_dR6_a substrate, while not statistically significant (P=0.25), is consistent with a loss of this intrinsic bend as a consequence of disrupting the A₅ tract.