

Osmotic Challenge Drives Rapid and Reversible Chromatin Condensation in Chondrocytes

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Supporting Material

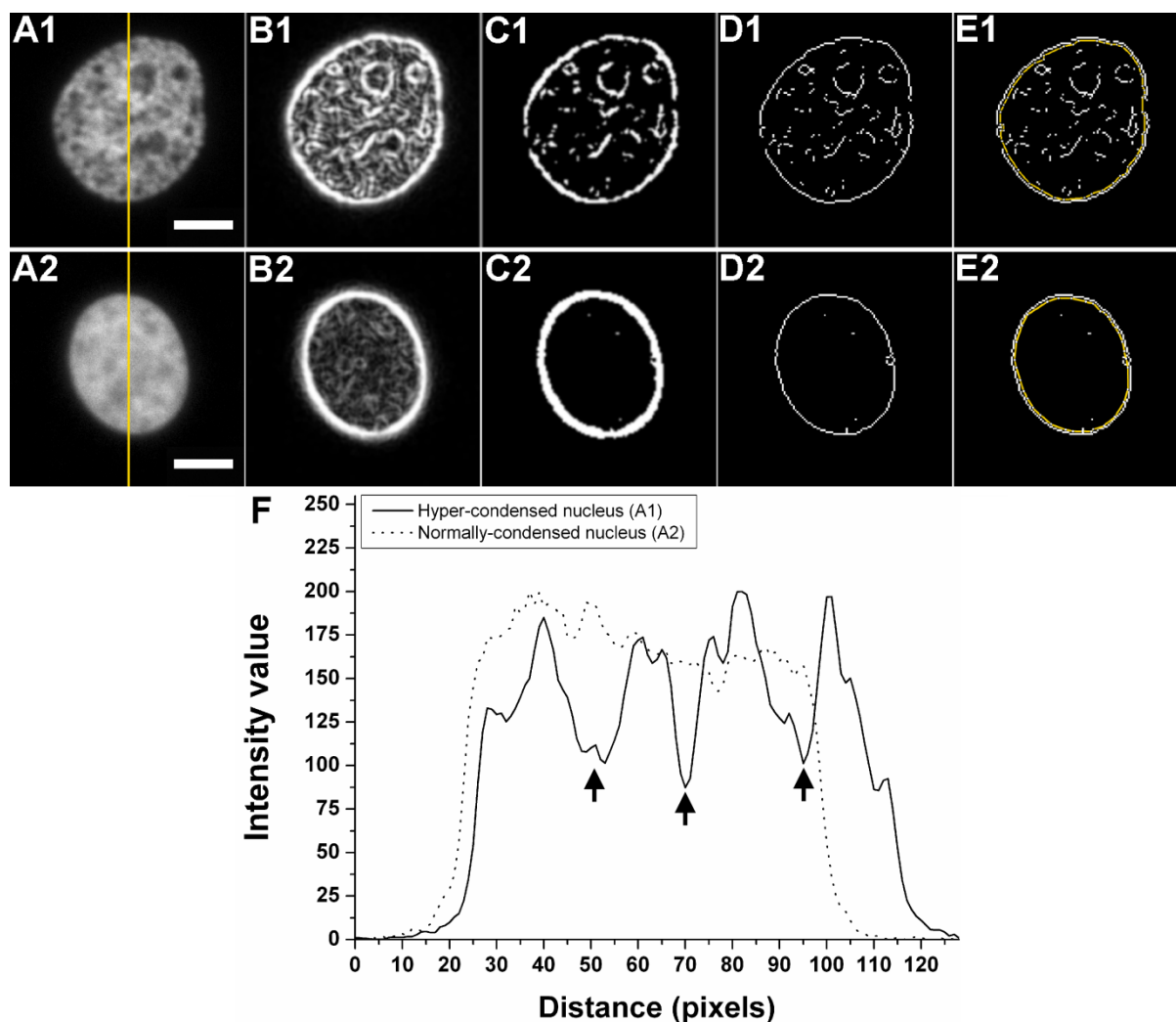


FIGURE S1 Monolayer cultured chondrocyte nuclei with hyper- (A1) and normally- (A2) condensed chromatin as the source images, both with a region of interest drawn across the nuclei (yellow). Image after Sobel algorithm (B), Sobel image post-thresholding (C), thresholded image post-thinning morphological operation (D), region of interest (yellow) showing the inner part of the nucleus (E). From this quantification method, the nucleus with hyper-condensed chromatin has an chromatin condensation parameter of 4.86%, whereas the nucleus without hyper-condensed chromatin has an chromatin condensation parameter of 0.16%. (F) The intensity profile across the nucleus with hyper- (A1, —) and normally- (A2,) condensed chromatin, arrows showing the intensity dips (Bar: 5 μ m).

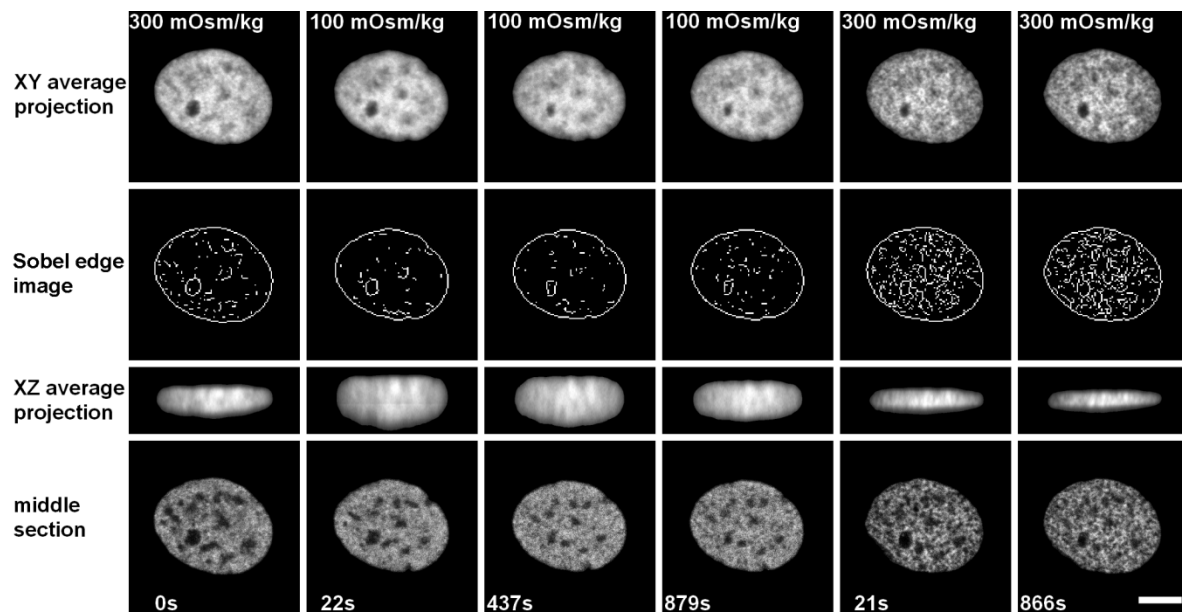


FIGURE S2 Images of monolayer cultured chondrocytes expressing H2B-GFP fusion-protein in an isoosmotic environment (300 mOsm/kg), followed by hypoosmotic challenge (100 mOsm/kg), which was then brought back to the isoosmotic conditions. Images were thresholded using the Iterative Self Organizing Data algorithm (Bar: 5 μ m).

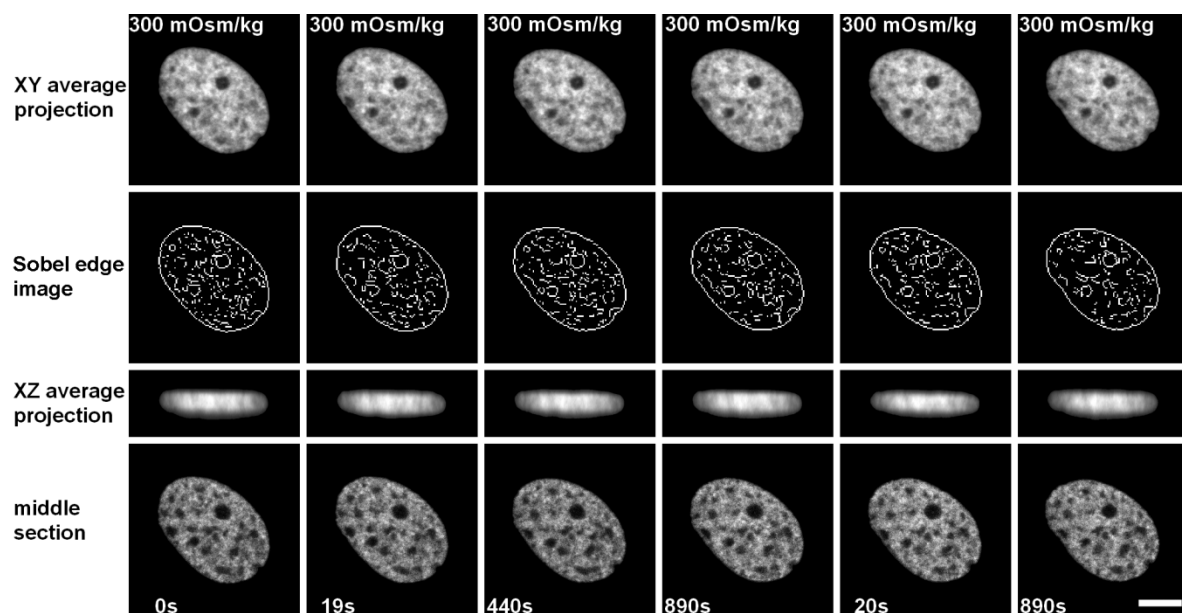


FIGURE S3 Images of monolayer cultured chondrocytes expressing H2B-GFP fusion-protein in an isoosmotic environment (300 mOsm/kg), followed by isoosmotic challenge (300 mOsm/kg), which was then brought back to the isoosmotic conditions. Images were thresholded using the Iterative Self Organizing Data algorithm (Bar: 5 μ m).

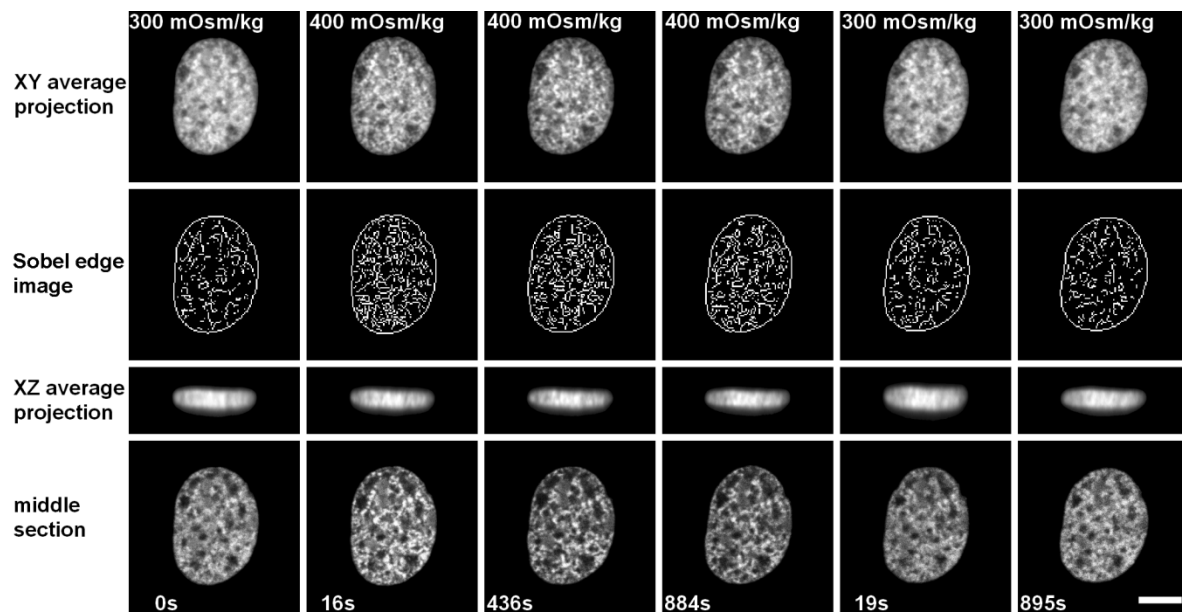


FIGURE S4 Images of monolayer cultured chondrocytes expressing H2B-GFP fusion-protein in an isoosmotic environment (300 mOsm/kg), followed by hyperosmotic challenge (400 mOsm/kg), which was then brought back to the isoosmotic conditions. Images were thresholded using the Iterative Self Organizing Data algorithm (Bar: 5 μ m).

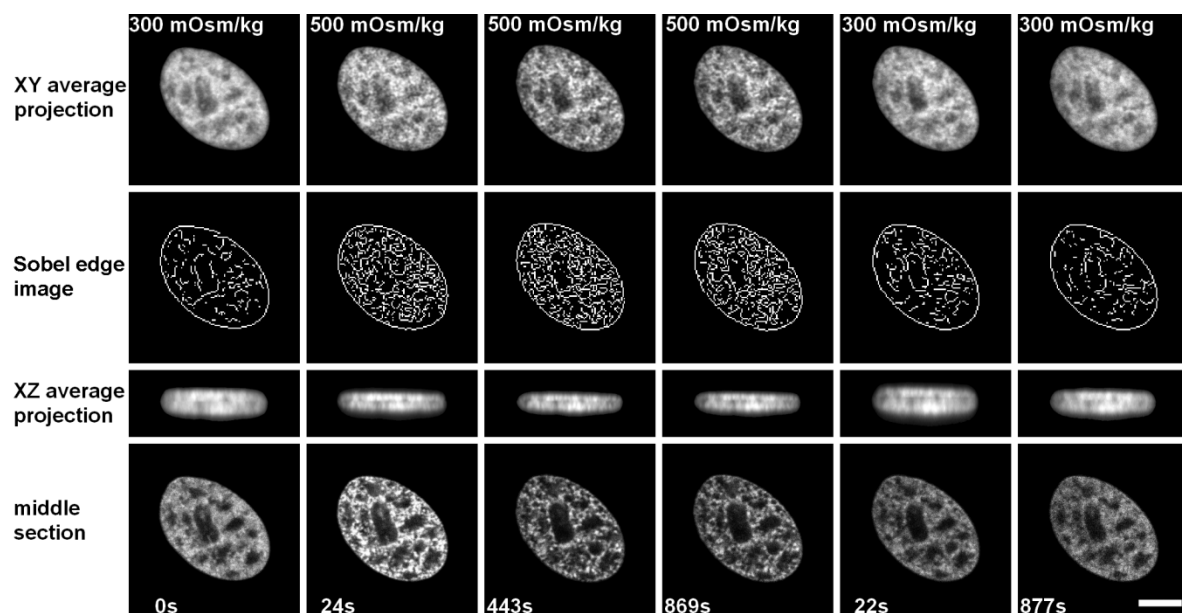


FIGURE S5 Images of monolayer cultured chondrocytes expressing H2B-GFP fusion-protein in an isoosmotic environment (300 mOsm/kg), followed by hyperosmotic challenge (500 mOsm/kg), which was then brought back to the isoosmotic conditions. Images were thresholded using the Iterative Self Organizing Data algorithm (Bar: 5 μ m).

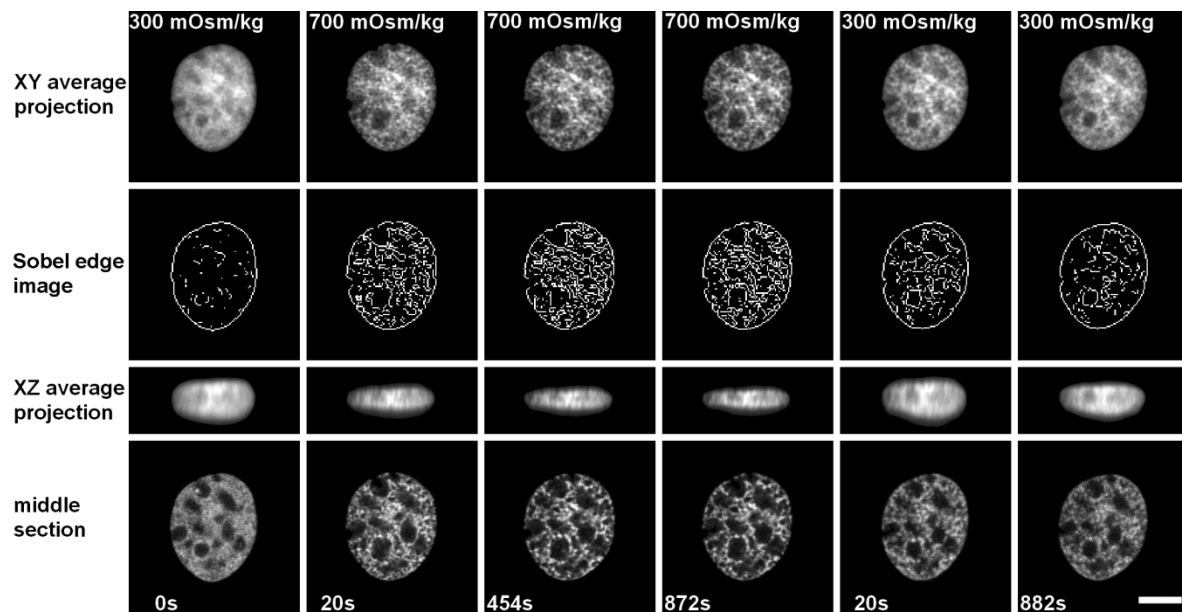


FIGURE S6 Images of monolayer cultured chondrocytes expressing H2B-GFP fusion-protein in an isoosmotic environment (300 mOsm/kg), followed by hyperosmotic challenge (700 mOsm/kg), which was then brought back to the isoosmotic conditions. Images were thresholded using the Iterative Self Organizing Data algorithm (Bar: 5 μ m).

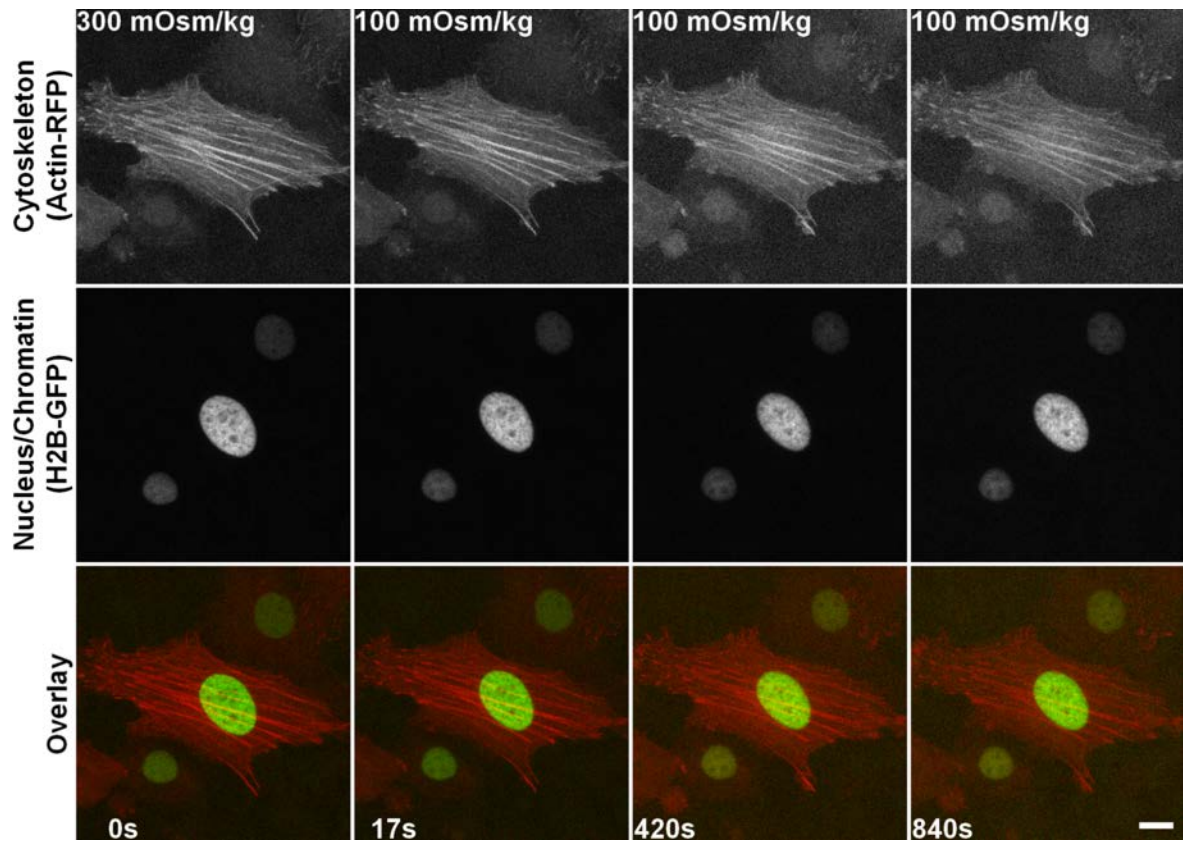


FIGURE S7 Average projection images of monolayer cultured chondrocytes expressing actin-RFP and H2B-GFP fusion-protein in an isoosmotic environment (300 mOsm/kg), followed with hypoosmotic challenge (100 mOsm/kg) (Bar: 10 μ m).

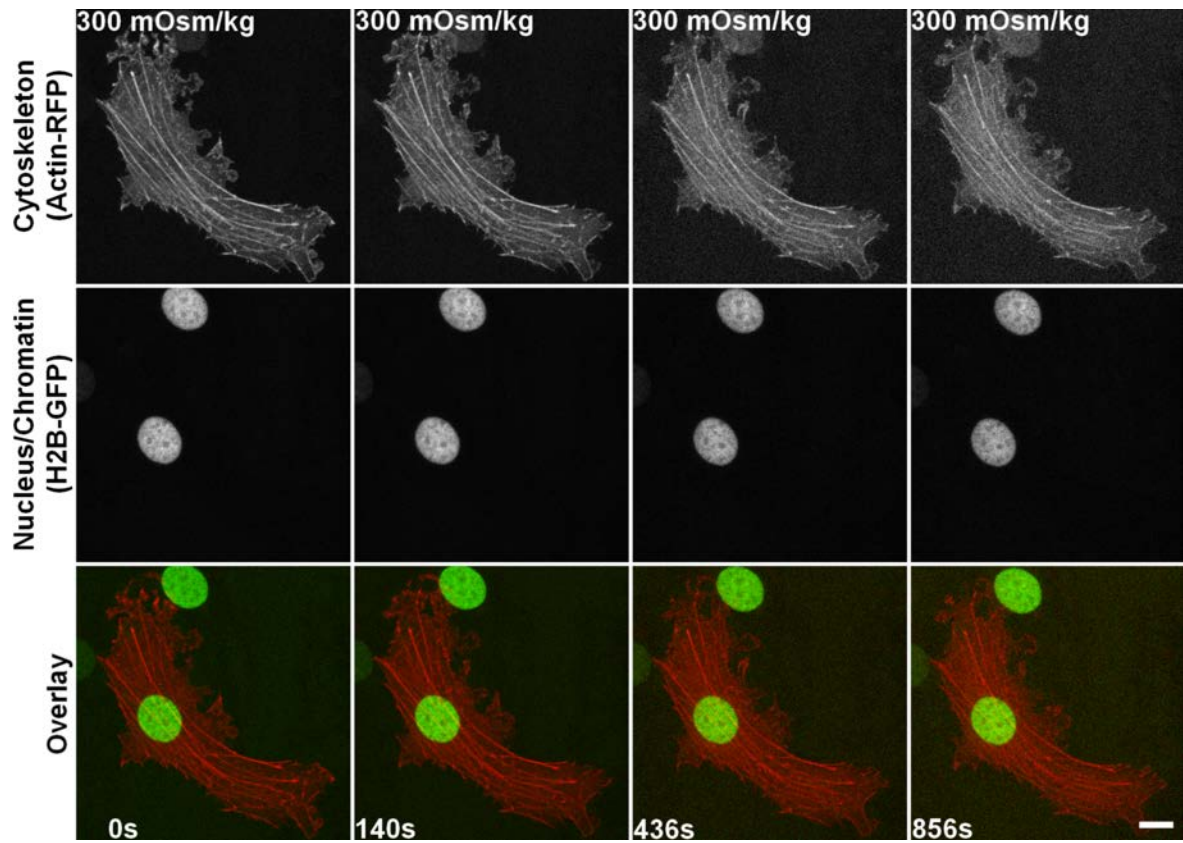


FIGURE S8 Average projection images of monolayer cultured chondrocytes expressing actin-RFP and H2B-GFP fusion-protein in an isoosmotic environment (300 mOsm/kg), followed with isoosmotic challenge (300 mOsm/kg) (Bar: 10 μ m).

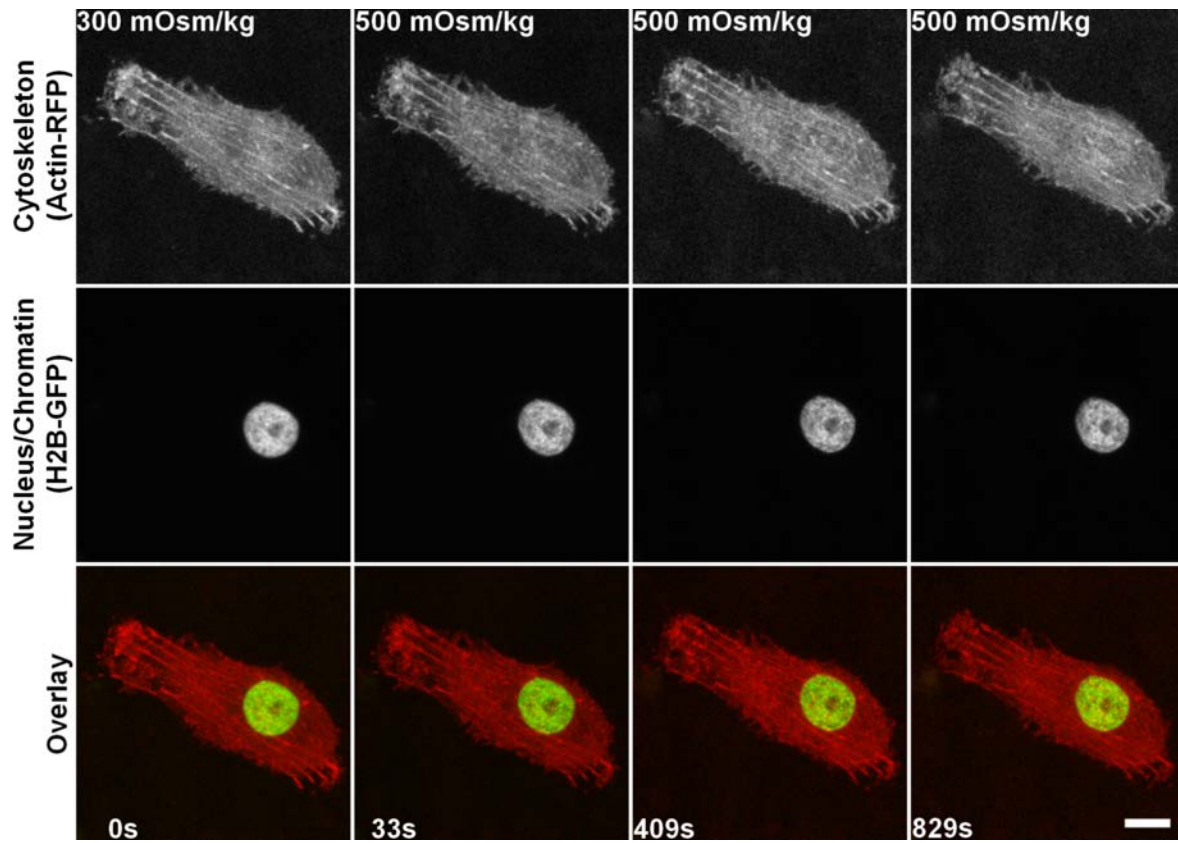


FIGURE S9 Average projection images of monolayer cultured chondrocytes expressing actin-RFP and H2B-GFP fusion-protein in an isoosmotic environment (300 mOsm/kg), followed with hyperosmotic challenge (500 mOsm/kg) (Bar: 10 μ m).