THE CHARACTERISTICS OF SOME RUMEN LACTOBACILLI1

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MATERIALS AND METHODS

During the past few years several investigators have reported the isolation of lactobacilli or lactobacilli-like organisms from the rumen liquor of cattle and sheep. Gall and Huhtanen (1951) isolated several gram positive, nonsporeforming rods from rumen fluid which produced lactic acid from glucose, but they were not further identified. Some lactobacilli found by Rodwell (1953) in a sheep's rumen were described resembling Lactobacillus leichmannii **as** or Lactobacillus delbrueckii. Huhtanen and Gall (1953) isolated two types of rods from rumen fluid which they stated were closely related to the genus Lactobacillus. Wasserman et al. (1953) characterized some isolates from a bovine rumen as variants of Lactobacillus bifidus. A rumen lactobacillus was classified into a group containing Lactobacillus brevis, Lactobacillus buchneri, and other heterofermenters by Briggs (1953b). L. brevis, an anaerobic variety of Lactobacillus lactis and an organism similar to the motile lactobacillus of Harrison and Hansen (1950) were discovered by Mann and Oxford (1954) in the rumens of young calves. Mann and Oxford (1955) also found Lactobacillus fermenti and Lactobacillus acidophilus in the rumens and abomasums of calves and kids. The number of lactobacilli in the rumen appears to vary with the diet, according to Briggs (1955), who reported that lactobacilli predominated in the rumen of animals on concentrate diets, but were less numerous in grass-fed animals.

This report deals with the isolation and identification of lactobacilli mainly from bovine rumen fluid. A sample of ovine rumen fluid yielded two cultures in some preliminary work which stimulated the investigation leading to this report.

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² Present Address Department of Animal Industries, College of Agriculture, University of Connecticut, Storrs, Connecticut. One sample was taken from a fistulated sheep and eight from a fistulated cow, both of which received a high roughage diet for the duration of the study. Rumen fluid was drawn through a tube into a flask by a vacuum pump, shaken and strained through cheese cloth and used for inoculation.

Preliminary studies indicated that enrichments with plain skim milk and rumen fluid plus glucose or plating with SL agar (Rogosa et al., 1951), incubated anaerobically, would not suffice as isolation media. Consequently, serial dilutions were made with rumen fluid to 10^{-12} in SL medium, Briggs medium (1953a), and fortified skim milk. The fortified skim milk contained yeast extract, 5 g; glucose, 10 g; "V-8 juice," 100 ml; and skim milk, 1,000 ml; final pH, 6.8. These media were dispensed into screw top tubes in 9 ml amounts, one ml of rumen fluid placed in the first tube and the series carried to 10^{-12} . All routine incubations were at 37 C and extended from 2 to 7 days. V-8 juice strained through 2 layers of cheese cloth was substituted for tomato juice in all media. Several unsuccessful attempts were made to isolate obligate anaerobes by adding 0.05 per cent cysteine to the Briggs' and SL media and flushing with nitrogen.

After incubation V-8 juice agar was used for plating and isolation. This medium contained strained V-8 juice, 200 ml; tryptone, 5 g; tryptose, 5 g; yeast extract, 5 g; agar, 15 g; and distilled water, 800 ml, with a final pH of 6.8. The composition was obtained from Dr. R. P. Tittsler in a personal communication. In many cases duplicate plates were prepared and one set incubated anaerobically. The plates were incubated for 48 hr and colonies were picked into tubes of fortified litmus milk. These cultures were replated for purification and to determine the colony type.

Preliminary procedures were performed as follows: gram stain, examination for spores and

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Arabinose			1	+	+	I	I	+	+	or pr	
Raffinose			4	4	+	I	I	ũ	+	erol	
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		Coagi	2nd day	5†	1	1	+	36	1	43	lucos
	Plain		%	0.5	0.1	0.2	1.0	0.1	0.5	0.6	lk. G
		°.	agu- lated	+	I	1	+	I	4	+	us mi
No. of Strains			12	5	30	4	46	9	52	e-litm	
Identified as			Lactobacillus acidophilus	Lactooactitus brevis	Lactobacillus buchneri	Lactobacillus casei	L. casei (non- lactose fer- menting)	Lactobacillus fermenti	Lactobacillus plantarum	* Yeast-glucos	

Characteristics of identified rumen lactobacilli TABLE 1

5 ~ 2 Yeast-glucose-litmus milk. Glucose, maltose, galactose, and sucrose v Proskauer reaction.
† The figures indicate the number of strains giving a positive result.
‡ Growth slight.

motility, and tests for catalase. Tests for nitrate reduction were made with both Briggs' medium and indole-nitrate broth (BBL), since Costilow and Humphreys (1955) noted that several strains of *Lactobacillus plantarum* reduced nitrate in the latter medium. Gas production was determined in Briggs' medium by the method of Rogosa *et al.* (1953).

Tests for the amount of acidity produced in skim milk, growth at 16, 45, and 48 C, type of lactic acid, carbohydrate fermentation, and hydrolysis of hippurate were performed as described by Rogosa *et al.* (1953) and Tittsler in a personal communication. The methods used to determine NaCl tolerance, bile salt tolerance, acetylmethylcarbinol production, action in yeastglucose-litmus milk, and starch hydrolysis were described by Wheater (1955*a*, *b*). Briggs' medium was used for the growth temperature tests and for the preparation of inocula.

The descriptions of various lactobacilli in the papers of Tittsler *et al.* (1947), Rogosa *et al.* (1953), Wheater (1955*a*, *b*) and Sharpe (1955) were employed as guides for identification.

RESULTS

One hundred sixty-eight cultures were studied and 155 were identified. All of the strains were gram positive, catalase negative, nonsporeforming, nonmotile rods. All of the isolates except four strains of L. plantarum failed to reduce nitrate. These four strains reduced nitrate to nitrite in indole-nitrate medium (BBL). In every other respect they resembled L. plantarum. No bifid forms were seen. The isolates grew on the surface of complex solid media containing reducing substances.

The characteristics of the identified isolates are summarized in table 1. In addition, volatile acid determinations helped to confirm the identifications. Cultures of L. brevis, L. buchneri, and L. fermenti all produced relatively large amounts of volatile acidity. Determinations were made of the optical rotation of the lactic acid produced by two strains of L. acidophilus and one strain of Lactobacillus casei (nonlactose fermenting). The former was inactive and with the latter, the D form was slightly predominant.

A group of 13 lactobacilli could not be identified on the species level. All were homofermenters that did not ferment lactose, but the fermentation patterns of the other carbohydrates were unique. At first growth failures were believed to be the cause, but the same results were obtained on repeated testing.

Because the isolation of lactobacilli resembling

	Plain Skim Enrich.	SL Agar	SL Broth	Briggs' Broth	Fortified Skim	Rumen Fluid Plus
						Glucose
Lactobacillus acidophilus	2*		4	1	3	2
	(2)†		(3)	(2)	(2)	(2)
Lactobacillus brevis			4	1		
			(5)	(6)		
Lactobacillus buchneri	2	1	15	4	8	
	(1)	(3)	(7)	(4)	(4)	
Lactobacillus casei			4			
			(4)			
L. casei (non lactose fermenting)	2		30	8	6	
	(3)		(6)	(5)	(4)	
Lactobacillus fermenti			3	1		2
			(4)	(7)		(2)
Lactobacillus plantarum	1	18	21	4	8	
	(3)	(3)	(6)	(6)	(6)	
Unidentified	. /	. /	7	6		
			(7)	(7)		

TABLE 2Cultures isolated from various media

* These two cultures were isolated from ovine rumen fluid.

† Figures in parentheses are the logarithms of the highest dilution from which the cultures were isolated.

L. casei var. alactosus (Rogosa et al., 1953) from rumen fluid was surprising, further testing was deemed necessary. A known culture (ATCC 11582) was obtained and studied. The ability of the isolates and ATCC 11582 to ferment inositol. dulcitol, adonitol, and sorbitol was determined. Dr. M. E. Sharpe kindly investigated the biochemical and serological characteristics of ATCC 11582 and three of the isolates. In the main, ATCC 11582 had the same characteristics as the isolated strains listed in table 1. All fermented adonitol, dulcitol, and sorbitol, but not inositol. Dr. Sharpe stated, in a personal communication, that she obtained almost the same results biochemically. She found that ATCC 11582 differed from the three isolates in fermenting inositol and dextrin but not dulcitol and all four hydrolyzed esculin. However, serologically the rumen strains differed from ATCC 11582. The three isolates reacted specifically, but weakly with a serum for the L. casei group. ATCC 11582 reacted specifically and strongly with a serum for the L. casei-helveticus group (Sharpe, 1955). The rumen isolates can apparently be classified as a nonlactose fermenting variety of L. casei not closely related serologically to ATCC 11582. The growth of these strains in a 45 C water bath was slight and in some cases questionable, but none of the strains fermented lactose or coagulated skim milk.

Table 2 presents data concerning the media with which the cultures were isolated. Lactobacilli were not isolated from SL agar at rumen fluid dilutions greater than 10^{-3} even when cysteine was added and the plates were incubated anaerobically.

The first 2 of 9 samples, including the ovine, were used for preliminary tests to develop the method of isolation. Strains of *L. casei*, *L. plantarum*, and *L. buchneri* were found in all of the next 7 samples; *L. acidophilus* in 4, *L. brevis* in 2, and *L. fermenti* in 2. The 7 samples were taken over a period of 2 months. Two cultures of *L. acidophilus* were isolated by an enrichment in skim milk from the only sample of ovine rumen fluid studied.

DISCUSSION

It has been demonstrated in this study that several species of lactobacilli were regularly found in the rumen fluid of 1 cow. The identified species do not appear to be indigenous to the rumen as all of them have been isolated from other sources. Although no total cultural counts were made, it is evident that the lactobacilli represented a numerically important part of the total rumen population, since some of the isolated species were present in numbers greater than 10^7 (table 2). In this connection, Wilson and Briggs (1954) obtained total counts ranging from 10^8 to 10^{10} per g. Briggs (1955) noted that lactobacilli predominated in the rumens of animals fed high concentrate diets, whereas the cow used in the present study was fed a high roughage diet.

Table 2 indicates that L. acidophilus and L. casei were probably passengers because they were not found in the higher dilutions nor in all of the samples. Only one culture, a strain of L. acidophilus, had amylolytic powers, indicating that the lactobacilli isolated from the roughage-fed animal used in this study were not important in the fermentation of starch. However, Mann and Oxford (1955) isolated several amylolytic strains of L. acidophilus from kids.

L. brevis and L. fermenti have both been found before in calves and kids. (Mann and Oxford, 1954, 1955).

Although the idea of a nonlactose fermenting variety of L. casei seems somewhat surprising, a number of such cultures have been isolated. Orla-Jensen (1942) listed two nonlactose fermenters. Rogosa et al., (1953) isolated 30 oral strains which they classified as L. casei var. alactosus. Rodwell (1953) suggested that one of the strains he isolated from a sheep's rumen was similar to L. delbrueckii or L. leichmannii because it did not ferment lactose. It was a homofermenter which grew at 40 C and fermented glucose, fructose, mannose, sucrose, and trehalose. It is certainly possible that Rodwell's isolate may have been similiar to the nonlactose fermenting strains of L. casei isolated in the present study. Sharpe, in a personal communication, stated that her group had come across several strains of L. casei which did not ferment lactose. Hunt and Rettger (1930) found a number of nonlactose fermenting lactobacilli in soil and grain. The descriptions of some of their strains indicate that they were similar in many ways to the rumen isolates.

The presence of L. plantarum in the rumen was expected because this organism has been isolated many times from many types of vegetable material. Several attempts were made to isolate obligately anaerobic lactobacilli, but none were found. This, of course, does not mean that they were not present in the rumen. Mann and Oxford (1954) found an anaerobic variety of L. lactis in a calf's rumen.

Several characteristics of the lactobacilli seem to be important in the over-all rumen fermentation. They are active fermenters of many carbohydrates, producing lactic acid, or lactic acid, CO₂, and acetic acid. Their requirements for vitamins and amino acids are well known. Certain strains decarboxylate histidine to histamine which can cause rumen paralysis (Rodwell, 1953). Heavy grain feeding caused the lactobacilli to predominate in the rumen (Briggs, 1955) and increased the amount of lactic acid (Balch et al., 1955). With these characteristics in mind the need for further research upon the numbers and species of lactobacilli in the rumen with different conditions of feeding and in different geographic locations certainly seems evident.

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The authors are indebted to Dr. M. E. Sharpe, National Institute for Research in Dairying, University of Reading, England, who clarified the relationship of *Lactobacillus casei* (nonlactose fermenting) to *L. casei* var. *alactosus* by serological methods.

SUMMARY

One hundred sixty-eight cultures of lactobacilli were isolated, 2 from a sample of ovine rumen fluid in a preliminary trial and the rest from 8 samples of bovine rumen fluid. The methods of isolation are described. One hundred fifty-five were identified as follows: 12 were Lactobacillus acidophilus, 5 were Lactobacillus brevis, 30 were Lactobacillus buchneri, 4 were Lactobacillus casei, 46 were Lactobacillus casei (nonlactose fermenting), 6 were Lactobacillus fermenti, and 52 were Lactobacillus plantarum. Two cultures of L. acidophilus were found in the ovine sample. All, except L. acidophilus and L. casei, were found in rumen fluid dilutions of 10^{-6} or 10^{-7} . The classification of nonlactose fermenting variety of L. casei is discussed.

REFERENCES

BALCH, C. C., BALCH, D. A., BARTLETT, S., BARTRUM, M. P., JOHNSON, V. W., ROW- LAND, S. J., AND TURNER, J. 1955 Studies of the secretion of milk of low fat content by cows on diets low in hay and high in concentrates. VI. The effect on the physical and biochemical processes of the reticulo-rumen. J. Dairy Research, 22, 270-289.

- BRIGGS, C. A. E. 1955 Viable bacteria in the rumen. Dairy Sci. Abstr., 17, 714-722.
- BRIGGS, M. 1953a An improved medium for lactobacilli. J. Dairy Research, 20, 36-39.
- BRIGGS, M. 1953b The classification of lactobacilli by means of physiological tests. J. Gen. Microbiol., 9, 234-248.
- COSTILOW, R. N. AND HUMPHREYS, T. W. 1955 Nitrate reduction by certain strains of *Lactobacillus plantarum*. Science, **121**, 168.
- GALL, L. S. AND HUHTANEN, C. N. 1951 Criteria for judging a true rumen organism and a description of five rumen bacteria. J. Dairy Sci., 34, 353-362.
- HARRISON, A. P. AND HANSEN, P. A. 1950 A motile Lactobacillus from the cecal feces of turkeys. J. Bacteriol., 59, 444-446.
- HUHTANEN, C. N. AND GALL, L. S. 1953 Rumen organisms. II. Two lactate utilizers and six miscellaneous types. J. Bacteriol., 65, 554-559.
- HUNT, G. A. AND RETTGER, L. F. 1930 A comparative study of members of the Lactobacillus genus with special emphasis on the lactobacilli of soil and grain. J. Bacteriol., 20, 61-83.
- MANN, S. O. AND OXFORD, A. E. 1954 Studies of some presumptive lactobacilli isolated from the rumens of young calves. J. Gen. Microbiol., **11**, 83-90.
- MANN, S. O. AND OXFORD, A. E. 1955 Relationships between viable saccharolytic bacteria in rumen and abomasum of the young calf and kid. J. Gen. Microbiol., 12, 140-146.
- ORLA-JENSEN, S. 1942 The lactic acid bacteria, 2nd ed. Ejnar Munksgaard, Copenhagen.
- RODWELL, A. W. 1953 The occurrence and distribution of amino acid decarboxylases within the genus Lactobacillus. J. Gen. Microbiol., 8, 224-232.
- ROGOSA, M., MITCHELL, J. A., AND WISEMAN, R. F. 1951 A selective medium for the isolation and enumeration of oral lactobacilli. J. Dental Research, **30**, 682-689.
- ROGOSA, M., WISEMAN, R. F., MITCHELL, J. A., DISREALY, M. N., AND BEAMAN, A. J. 1953 Species differentiation of oral lactobacilli from man including descriptions of *Lactobacillus salivarius* n. sp. and *Lactobacillus cellobiosus* n. sp. J. Bacteriol., 65, 681-699.

- SHARPE, M. E. 1955 A serological classification of lactobacilli. J. Gen. Microbiol., 12, 107-122.
- TITTSLER, R. P., GEIB, D. S., AND ROGOSA, M. 1947 Taxonomy of the genus Lactobacillus, with special reference to correlations of differential characteristics. J. Bacteriol., 54, 12-13.
- WASSERMAN, R. H., SEELEY, H. W., AND LOOSLI, J. K. 1953 The physiology and nutrition of a rumen Lactobacillus. J. Animal Sci., 12, 935-936.
- WHEATER, D. M. 1955a The characteristics of Lactobacillus acidophilus and Lactobacillus bulgaricus. J. Gen. Microbiol., 12, 123-132.
- WHEATER, D. M. 1955b The characteristics of Lactobacillus plantarum, L. helveticus and L. casei. J. Gen. Microbiol., 12, 133-139.
- WILSON, M. K. AND BRIGGS, C. A. E. 1954 The normal flora of the bovine rumen. I. Bacteriological methods for quantitative studies. Vet. Record., 66, 187-188.