

Table S1. List of primers and conditions used for amplifying macropodid DNA

| Genome | Gene amplified ¹ | Primer name | Sequence (5' – 3') | ~ Amplicon Length (bp) | Annealing temp. (°C) | Primer reference | |
|---------------|-----------------------------|--|---|------------------------|----------------------|--|----------------------|
| Nuclear | <i>IRBP</i> | IR_A_F | ATGGCCAAGGTCCTCTTGGATAACTACTGCTT | 1300 | 58 ² | Meredith et al. 2008 | |
| | | IR_B_R | AGGTTCCATGATGAGGTGCTCCGTGTCCTG | | | | |
| | <i>IRBP</i> | IR_A_F | ATGGCCAAGGTCCTCTTGGATAACTACTGCTT | 600 | 62 ² | | Meredith et al. 2008 |
| | | IR_N_R | CGCATCTTCTTGAGGATGTAG | | | | |
| | <i>IRBP</i> | IR_M_F | CCCSTTTGTCATTTCTACC | 700 | 58 ² | | Meredith et al. 2008 |
| | | IR_B_R | AGGTTCCATGATGAGGTGCTCCGTGTCCTG | | | | |
| <i>ApoB</i> | F60 MARS_R820 | GGAGAAGCCAAYCTGGATTTTCYT GTYGTCCCATCTAACTTATACTGC | 750 | 60 ² | Meredith et al. 2008 | | |
| <i>ApoB</i> | F60 MARS_R474 | GGAGAAGCCAAYCTGGATTTTCYT CATGTAGCCAAWTGRTGGCATCTC | 500 | 60 ² | Meredith et al. 2008 | | |
| <i>ApoB</i> | MARS_F420 MARS_R820 | CAATTCCTGAAATGACTCTGCC GTYGTCCCATCTAACTTATACTGC | 450 | 60 ² | Meredith et al. 2008 | | |
| Mitochondrial | NADH1 | ND1_1F ND1_1R | GGCAGAGCTGGYAATTGCAT GAAATAAGGGGGCTTDCAC | 960 | 55 ^{3,4} | Cao et al. 1998 | |
| | NADH2 | tMetFor tAlaRev | ATACCCCGAAAATGTTGGTTC GATTTGCGTTCGAKTGATGCAAG | 1050 | 55 ^{3,4} | Osborne & Christidis, 2001 | |
| | <i>Cytb</i> | <i>Cytb</i> _F14195 <i>Cytb</i> _R15472 | CATTTTAGTATGGACTCTAACCATAACC AGGGTGTTATACCTTCATTTTGG | 1150 | 54 ⁴ | This study | |
| | 12S rRNA | L2180_F M_12S6_R | AAAGCACAGCACTGAAGATGC GCTCAAAATGGTCAGGGTTAGCTG | 950 | 58 ⁴ | mod. from Titus & Frost 1996 This study | |
| | 16S rRNA | M_12S3_F M_12S5_R | AAACGTTAGGTCAAGGTGTAGC GCAATTGCCAGCYCTGCCAC | 1950 | 58 ⁴ | This study | |
| | 12/16S rRNA | L2180 H3628 | AAAGCACAGCACTGAAGATGCTG GCTGTCTTTACAGGTGGCTGCT | 1560 | 55 ³ | mod. from Titus & Frost 1996 Reed et al. 2001 | |

| | | | | | |
|----------|--------------------|---|------|-----------------|--|
| 16S rRNA | 16Sar 16Sbr | CGCCTGTTTATCAAAAACAT CCGGTYTGAACCTCAGATCAYGT | 550 | 55 ³ | Palumbi et al. 1996 Palumbi et al. 1996 |
| NADH2 | ND2a_3F tAlaRev | GGGGTGMAAGCCCCCTTATT GATTTGCGTTCGAKTGATGCAAG | 1205 | 55 ³ | This study |
| NADH1 | ND1_1F ND1_1R | GGCAGAGCTGGYAATTGCAT GAAATAAGGGGGCTTDCAC | 960 | 55 ³ | Cao et al. 1998 |

¹ Post-PCR purifications were performed either using Shrimp Alkaline Phosphatase and Exonuclease I (GE Healthcare/Crown scientific) or alternatively, using a Qiaquick purification kit (Qiagen Sciences, MD, USA). All amplicons were sequenced by Macrogen (Seoul, South Korea) on an Applied Biosystems 3730XL capillary sequencer. Geneious 5.4.3 (Biomatters, NZ) was used to examine the chromatograms and assemble the forward and reverse contigs for each gene.

² Murdoch University, nuclear genes: initial denaturation (5 min at 94°C); 35 cycles of denaturation (45 sec at 94°C); variable annealing temperatures (see Table above, for 45 sec); extension (68°C for 1-2 min, depending on fragment length); final extension (10 min at 72°C).

³ Australian National University, mitochondrial genes: initial denaturation (2 min at 94 C); 30 cycles of denaturation (15 sec at 94 C), annealing (30 sec at 50 – 55 C), extension (90 sec at 68 C); final extension (4 min at 68C).

⁴ Murdoch University, mitochondrial genes: initial denaturation (3 min at 94°C); 35 cycles of denaturation (30 sec at 94°C); variable annealing temperatures (see Table above, for 30 sec); extension (90 sec at 68°C); final extension (10 min at 72°C).

References

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