

Figure S1 Chromosomes carrying *zuc*^{HM} and *squ*^{PP} suppress distortion by SD-LA in both *aub*^{HR-wt} and *aub*^{HR-HN} backgrounds. Females of the genotype SD-LA *aub*^{HR-wt} and *aub*^{HR-HN} were crossed with males carrying either a mutation in either *zuc*, *cuff* or *squ*. The 15-30 individual male F1 were then backcrossed to *cn bw* females to test for distortion. The *squ*^{PP} and *zuc*^{HM} chromosomes significantly suppressed distortion when paired with either the *aub*^{HR-HN} or *aub*^{HR-wt} chromosome. The *cuff*^{MM} chromosome gave a small but significant enhancement of distortion ($p < 0.05$) with the wild type allele and there was no significant change with the *aub*^{HR-HN} mutant (** $p < 0.0005$, * $p < 0.05$, 2-tailed Z-test).

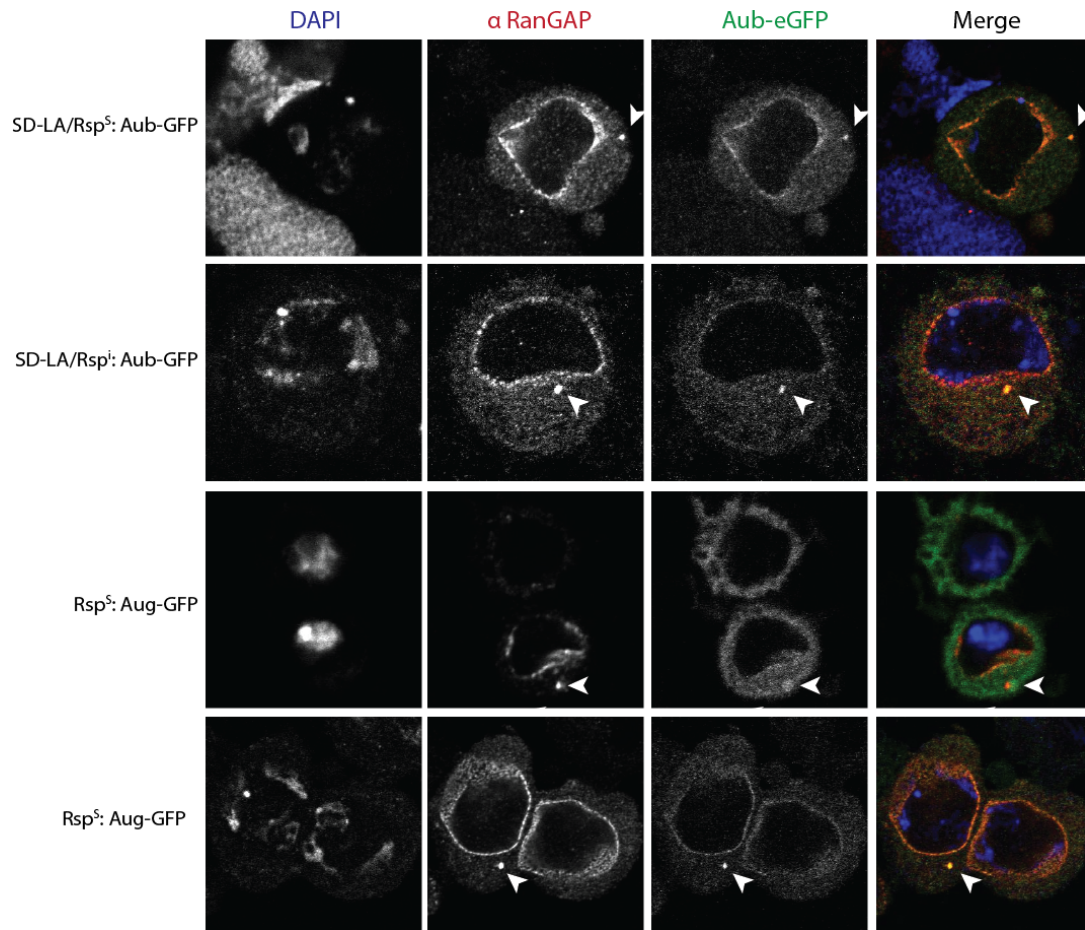


Figure S2 Aub-GFP and RanGAP colocalized in a single large body of primary spermatocytes. Whole mount testis of males expressing Aub-GFP were fixed with paraformaldehyde and stained with anti-RanGAP (1:1000) primary and Alexa Fluor (1:800) secondary antibodies. Confocal imaging was done on a Zeiss LSM 510. Spermatocytes in the testis of both distorting (*SD-LA/Rsp⁵*) and non-distorting (*SD⁺ Rsp⁵*) males show a single large point of colocalization between Aub-GFP and Ran-GAP (arrowhead)

Table S1 Genetic stocks used in this study

Line	Notes on genotype	Source	Reference
HR stocks			
<i>y w; FLP-I-Scel/TM6</i>	$y^1 w^*$; $P\{ry^{+17.2}=70FLP\}11 P\{v^{+11.8}=70I - Scel\}2B noc^{ScO}/CyO, S^2$	BSC 6930	Staber <i>et al.</i> 2011
<i>y w ey-FLP</i>	$y^{02} w^{1118} P\{ry[+t7.2]=ey-FLP.N\}2$	BSC 5580	Staber <i>et al.</i> 2011
<i>y w Cre; noc^{ScO}/CyO</i>	$y^1 w^{67c23} P\{y^{+mbDint2}=Crey\}1b;$ noc^{ScO}/CyO	BSC 766	Staber <i>et al.</i> 2011
w^1		BSC 145	Staber <i>et al.</i> 2011
w^{1118}		BSC 3605	Staber <i>et al.</i> 2011
Balancer Stocks			
<i>w; TM3 sb /TM6 tb</i>		B. Ganetzky	
<i>+</i> ; $CyO^{It\ ap\ pr\ cn}/Sco$		B. Ganetzky	
<i>w</i> ; $CyO^{cn\ bw\ Roi}/Sco$		B. Ganetzky	
SD Stocks			
<i>SD-5</i>	Strong distortion phenotype and contains <i>Sd</i> , <i>E(SD)</i> , <i>Rspⁱ</i> , <i>M(SD)</i> and <i>St(SD)</i> as well as two paracentric inversions on 2R.	B. Ganetzky	<i>Sandler and Hiraizumi 1959;</i> <i>Sandler et al. 1959</i>
<i>SD-5*</i>	Uncharacterized derivative of <i>SD-5*</i> which exhibits an intermediate level of distortion	B. Ganetzky	
<i>SD-72</i>	Strong distorter with a both pericentric and paracentric inversions	B. Ganetzky	Sandler and Hiraizumi 1959)
<i>SD-Mad It cn</i>	Derived from the original <i>SD-Mad</i> by recombination with chromosomes carrying the recessive markers <i>cn</i> , <i>It</i>	B. Ganetzky	R.G. Temin 1979
<i>SD-Mad bw³</i>	Derived from the original <i>SD-Mad</i> by recombination with chromosomes carrying the recessive markers <i>bw³</i>	B. Ganetzky	R.G. Temin 1979
<i>SD-Roma</i>	Inversion free moderately distorting SD chromosome isolated in Italy	B. Ganetzky	Nicoletti and Trippa 1967)
<i>SD-Los Arrenos</i>	Weak distorter	B. Ganetzky	
<i>Rsp^S cn bw</i>	$[Sd^+, E(SD)^+, Rsp^S]$ standard Rsp sensitive chromosome	B. Ganetzky	Lyttle 1991
<i>Rspⁱ¹⁶ cn bw</i>	$[Sd^+ E(SD)^+ Rsp^i]$ Radiation-induced derivative of the Rsp ^S cn bw chromosome where the Rsp locus has	B. Ganetzky	Ganetzky 1977

	been completely deleted		
<i>Rsp^{SS} It pk cn</i>	[<i>Sd^r E(SD)^r Rsp^{SS}</i>] the canonical RspSS chromosome	B. Ganetzky	Lyttle 1991
RNAi Mutants			
<i>aub^{CC42} cn bw/CyO</i>	EMS; Strong allele	T. Schüpbach	Schupbach and Wieschaus 1991
<i>aub^{HN2} cn bw/CyO</i>	EMS; Strong allele	T. Schüpbach	Schupbach and Wieschaus 1991
<i>aub^{HM23} cn bw/CyO</i>	EMS; Strong allele	T. Schüpbach	Schupbach and Wieschaus 1991
<i>aub^{AHN56} cn bw/CyO</i>	EMS: Strong allele, has secondary mutation	T. Schüpbach	Schupbach and Wieschaus 1991
<i>aub^{AHE13} cn bw/CyO</i>	EMS: Strong allele, has secondary mutation	T. Schüpbach	Schupbach and Wieschaus 1991
<i>zuc^{HM27} cn bw/CyO</i>	EMS: Strong allele	T. Schüpbach	Schupbach and Wieschaus 1991
<i>zuc^{SG63,rec2} pr c px sp/CyO</i>	EMS: homozygotes poorly viable	T. Schüpbach	Schupbach and Wieschaus 1991
<i>squ^{PP32} cn bw/CyO</i>	EMS; Strong allele	T. Schüpbach	Schupbach and Wieschaus 1991
<i>squ^{HE47} cn bw/CyO</i>	EMS; Strong allele	T. Schüpbach	Schupbach and Wieschaus 1991
<i>cuff^{WM25} cn bw/CyO</i>	EMS; Strong allele	T. Schüpbach	Schupbach and Wieschaus 1991
<i>cuff^{Q037} cn bw/CyO</i>	EMS; Strong allele	T. Schüpbach	Schupbach and Wieschaus 1991
<i>w¹; aub^{N11}/CyO</i>	110bp deletion	P. Macdonald	Harris and Macdonald 2001
<i>piwi⁰⁶⁸⁴³ cn /CyO</i>	P{PZ} in first exon	BSC 12225	Cox et al 1998
nosGAL4, Aub-GFP	UAS-Aubergine-GFP recombined on to the same chromosome as nos-GAL4 VP16	P. Macdonald	Harris and Macdonald 2001

Table S2 Primer sequences used in this study

Name	Sequence
Cloning Primers	
AUBarm1-F <i>Bs</i> WI	TCGTACGCTCTCCATGTAGCGAAAAGCGTATAG
AUBarm1-R <i>As</i> cl	TGGCGCGCCAATGCATATATTCGTATGAAATGAGC
AUBarm2-F <i>Acc</i> 65I	TGGTACCTTCCAGCAACGGAACCTTTATAGAC
AUBarm2-R <i>Not</i> I	TGCGGCCGCTGGTTACGAAGAGTCGTGCTGGCG
Sequencing Primers	
AUBarm1-s1	GTGACAGTCGCTGCCATCGTATTATTGG
AUBarm1-s2	CTGACAAGTGGACTGTCTGCAGTGGGTG
AUBarm1-s3	CTTTATATCGAAGGGTAAGTACATG
AUBarm1-s4	GA CT CGA ACG AT GA A G A A C A A C A C C A G C G
AUBarm2-s1	GTAGCGAGAATTAATGCGCTTGGATAC
AUBarm2-s2	GTGTCACCACCTGCGATGGCACC GGTC
AUBarm2-s3	CCGGTGGCAACACGCGTGCTGGAATCTC
AUBarm2-s4	CGATTGAAAGTCGACATCGTCAATGCG
aub arm1-RS1	GGGCAACTCAAAAAGTGGTAACAAG
aub arm2-RS1	GATATATGTATGTAGATATGTACATC
AUBarm1-R3	CTTGTTTATTTACATGTAAGTACTGTCC
AUBarm1-R4	CCTGGTCAACCCCTTAAATGCAGATGC
AUBarm1-R5	GCTTAGTAAATAAGTTAACACAATTTAC
AUB HN1 VR	CCGCAACGGCACTTACTCCCAAGCG
Mutagenic Primers	
AUB HN m1 F	GATGGGAGCTCCCTGGTAGGTAGTCATCCCCCTCCACGGTCTGA
AUB HN m1 R	TCAGACCGTGGAGGGGGATGACTACCTACCAGGGAGCTCCCATC
aub E721A R	GGTCAGCTCTACCAGGTGGTAAACAGCGCGGTGAACACCCTAAAGGACAGG
aub E721A F	CCTGTCCTTTAGGGTGTTCACCGCGCTGTTTACCACCTGGTAGAGCTGACC
Arm Specific Validation Primers	
AUBarm2-V1	CTTGATGAACATAAAGGGATCC
AUBarm2-V2	CGTCGATTTTACGCCTGATGTGG
AUBarm2-V3	GGCACTCATATTACCGTGCAGGC
AUBarm1-V1	GGTTTATAATTCCACGGACTTC
AUBarm1-V2	CCATAGCGAATGTAGTGTG
AUBarm1-V3	CCCGGACCATCGAAAGTAACTGC
AUBarm2-VR2	CGGTGAACATGGCGTGCGCCCTGAGG
AUBarm1-R3	CTTGTTTATTTACATGTAAGTACTGTCC
AUBarm1-R4	CCTGGTCAACCCCTTAAATGCAGATGC
AUBarm1-R5	GCTTAGTAAATAAGTTAACACAATTTAC

p[W25.2] Specific Validation Primers	
pW-Not 1	CACTGTTACGTCGCACTCGAGGGTAC
pW-Not 2	GCACTCGAGAGCTCGTTACAGTCCG
pW-Bsi 1	CGCACCGGACTGTAACGAGCTAC
pW-Bsi 2	GGCGACTCAACGCAGATGCCGTACC
pW-Asc 1	GTATGCTATACGAAGTTATCTAGACTAGTCTAGGGCG
pW-Asc 2	GCTTGGCTGCAGGTCGACTCTAGAGG
pW-Asc 3	CGATCATTATTTCGCTGCATGAATTAGC
pW-Acc 1	CATTATACGAAGTTATCTAGACTAGTCTAGGGTAC
pW-Acc 2	GACGCTCCGTCGACGAAGCGCTC
pW-Acc 3	GCTCAGCTTGCTTCGCGATGTGTTAC
WTPR	CGCGAACATTTCGAGGCGCGCTCTCTCG
Wt SQ10F	GTGACCTGTTCCGGAGTGATTAGCG
Southern Blotting	
SP6	GCCAAGCTATTTAGGTGACACTATAG
T7	GAATTGTAATACGACTCACTATAGGG
Rsp13 clone sequence	GGAGCTCCACCGCGGTGGCGGCCGGCCCTGCAGATCTGCGGCCGCTCTAGAAGGTGTCTTCTGTTGCGCT GGTACTTGAAATCGAAAAATCACTCATTGACCGCTAAAATGACATAACTTAGTCAATTTATTGTTTTGTG TACCAGTTTAAATAATCTGTAGAAGGTATCTTCTGTTTGTCTGGTACTTGAAATCGGAAAATCACTCATT TGACCGCTTAAAATGTA AAACTTAGGCAATTTACTGTTTTTCTTACCAGTTGAACAGAATCTCTAGAAGG TGTTCTTCTGTTGCGCTGGTACTTGAAATCGAAAAATCACTCATTGACCGCTAAAATGACATAACTTAGTCA ATTTATTGTTTTGTGTACCAGTTTAAATAATCTGTAGAAGGTATCTTCTGTTTGTCTGGTACTTGAAATC GGAAAATCACTCATTGACCGCTTAAAATGTA AAACTTAGGCAATTTACTGTTTTTCTTACCAGTTGAAC AGAATCTCTAGAACTAGTGGATCCCCGGGCTGCAGGAATTCGATATCAAGCTTATCGATACCGTCGACC TCGAGGGGGGGCCCGGTACCAGCTTTTGTCCCTTTAGTGAGGGTTAATTCGAGCTTGGCGTAATCATG GTCATAGCTGTTTCTGTGTGAAATTGTTATCCGCTCAC