## **Supplementary Information**

# Synthesis and activity of biomimetic biofilm disruptors

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## Materials

Chemicals and solvents were purchased from Sigma Aldrich or EMD chemicals. Solvents were anhydrous or HPLC grade and chemicals were of reagent grade or better and used without further purification. Temperatures were measured externally.

<sup>1</sup>H NMR spectra and <sup>13</sup>C NMR spectra were recorded on a Varian 400 MHz NMR-System and referenced to the residual proton and carbon signals of the deuterated solvent, respectively. Spectra in D<sub>2</sub>O were referenced to 3-(trimethylsilyl)propionic-2,2,3,3- $d_4$  acid sodium salt (0.05 wt %). Splitting patterns that appear different from their actual spin systems are denoted as peseudo-plets ( $\Psi$ ).

Low resolution mass spectra were obtained by LC/MS (ESI) with an Agilent 6130 Quadrupole LC/MS coupled to an Agilent 1200 series HPLC system. High resolution mass spectrometry was carried out by the mass spectrometry laboratory of the University of Illinois by Q-TOF MS in positive mode or by the WM Keck Foundation Biotechnology Resource Laboratory of Yale University using a 9.4T Bruker Qe FT-ICR MS in negative mode. Infrared (IR) spectra were measured using an ALPHA FT-IR Spectrometer (Bruker).

#### 1) Synthesis

Guanidines were prepared via different routes.

#### Preparation of 1,1'-(ethane-1,2-diyl)diguanidine sulfate (1)

$$H_2N \xrightarrow{NH}_{H_2}N \xrightarrow{H}_{NH} \xrightarrow{NH}_{NH_2}$$

Ethylenediamine (1.67 mL, 25 mmol) was added to a solution of S-methylisothiourea  $\cdot \frac{1}{2}$  H<sub>2</sub>SO<sub>4</sub> (1.39 g, 10 mmol) in 8 mL of 1:3 H<sub>2</sub>O/EtOH and stirred for 24 h at room

temperature. A white viscous precipitate had formed. The supernatant was decanted and the residue washed with ethanol. Crystallization from ethanol/H<sub>2</sub>O afforded transparent crystals which were sucked off and washed with 50% ethanol in water and ethanol (yield: 214 mg, 18%).

<sup>1</sup>H NMR (400 MHz, D<sub>2</sub>O): δ 3.44 (s, 4H, C<u>H</u><sub>2</sub>-C<u>H</u><sub>2</sub>).

<sup>13</sup>C NMR (400 MHz, D<sub>2</sub>O): δ 159.95 (2x <u>C</u>=NH), 42.95 (<u>C</u>H<sub>2</sub>-<u>C</u>H<sub>2</sub>).

The structure was confirmed via gCOSY, HSQCAD and gHMBCAD.

IR (solid): v (cm<sup>-1</sup>) 3319, 3160, 3111, 3034, 1686, 1627, 1153, 1078, 1029, 970, 596, 529, 472, 437.

LC-MS (LR) m/z: 145.2  $[M+H]^+$  (calc.: 145.1).

HR ESI-MS, m/z: 145.1200 [M+H]<sup>+</sup> (calc.: 145.1202).

Anal. calcd for C<sub>4</sub>H<sub>14</sub>N<sub>6</sub>SO<sub>4</sub> (C<sub>4</sub>H<sub>12</sub>N<sub>6</sub> · H<sub>2</sub>SO<sub>4</sub>): C, 19.83; H, 5.82; N, 34.69; S, 13.24. Found: C, 19.81; H, 5.74; N, 33.96; S, 13.23.

## Preparation of N,N-di(2-guanidinylethyl)amine sulfate hydrate (2a)



Diethylenetriamine (540  $\mu$ L, 5 mmol) was added to a solution of *S*-methylisothiourea  $\cdot \frac{1}{2}$  H<sub>2</sub>SO<sub>4</sub> (1392 mg, 10 mmol) in 5 mL of 1:1 H<sub>2</sub>O/EtOH and stirred for 16 h at room temperature. A white crystalline precipitate formed, which was sucked off and washed with ethanol. The raw crystals were dissolved in the minimum volume H<sub>2</sub>O at 30°C and isopropanol was carefully layered on top of the solution. The pure product crystallized over night in at 4°C as fine white needles (910 mg, 60%).

<sup>1</sup>H NMR (400 MHz, D<sub>2</sub>O):  $\delta$  3.34 (t, J = 6.0 Hz, 4H, 2x <sup>Gua</sup>NH-C<u>H</u><sub>2</sub>), 2.83 (t, J = 6.0 Hz, 4H, C<u>H</u><sub>2</sub>-NH-C<u>H</u><sub>2</sub>).

<sup>13</sup>C NMR (400 MHz, D<sub>2</sub>O): δ 159.92 (2x <u>C</u>=NH), 49.43 (<u>C</u>H<sub>2</sub>-NH-<u>C</u>H<sub>2</sub>), 43.47 (2x <sup>Gua</sup>NH-<u>C</u>H<sub>2</sub>).

The structure was confirmed via gCOSY, HSQCAD and gHMBCAD.

IR (solid): v (cm<sup>-1</sup>) 3334, 3282, 3151, 2852, 1626, 1459, 1345, 1046, 749, 601, 508.

HR ESI-MS, m/z: 188.1626 [M+H]<sup>+</sup> (calc.: 188.1624).

Anal. calcd for C<sub>6</sub>H<sub>21</sub>N<sub>7</sub>SO<sub>5</sub> (C<sub>6</sub>H<sub>17</sub>N<sub>7</sub> · H<sub>2</sub>SO<sub>4</sub> · H<sub>2</sub>O): C, 23.76; H, 6.98; N, 32.32; S, 10.57. Found: C, 23.75; H, 6.87; N, 32.12; S, 10.59. Free base (<u>2b</u>):

The sulfate salt (200 mg, 0.659 mmol) was dissolved in 10 mL H<sub>2</sub>O and a solution of  $Ba(OH)_2$  (113 mg, 0.659 mmol) in 6 mL H<sub>2</sub>O was added. Precipitation was completed at 4°C, the suspension was filtered through celite and lyophilized. The residue was extracted with hot methanol and filtered through celite giving the free base in quantitative yield. The free base was well soluble in hot methanol, the sulfate salt and barium salts were insoluble.

Preparation of  $1-[2-N^2, N^3$ -diisopropylguanidinylethane-1-yl]-1,1'-(ethane-1,2-diyl)-bis-( $N^2, N^3$ -diisopropylguanidine) trihydrochloride dihydrate (<u>3</u>)



Diethylenetriamine (216  $\mu$ L, 2.0 mmol) was dissolved in 4 mL acetonitril, *N,N'*diisopropylcarbodiimide (1174  $\mu$ L, 7.5 mmol) was added and the solution was stirred at 70°C for 24 h. The solvent was removed, the oily residue was taken up in 6 mL ethanol and 4 mL of a 1:1 mixture of ethanol and 37% HCl aq. were added. After evaporation of the solvent, the solid residue was taken up in a small volume of ethanol and acetone was added until a precipitate appeared. The liquid phase was decanted and the solid material taken up in ethanol. After filtration the solution was evaporated until a highly viscous oil was obtained. Isopropanol was layered atop the oil and after standing for three days at room temperature, crystalline material had formed. The crystals were collected, washed with ice cold ethanol and isopropanol and re-crystallized twice from isopropanol/ethanol and isopropanol/H<sub>2</sub>O yielding 58.6 mg (4.7 %) of the product as the trihydrochloride dihydrate.

<sup>1</sup>H NMR (400 MHz, D<sub>2</sub>O):  $\delta$  3.82-3.72 (m, 6H, 2x C<u>H</u>(CH<sub>3</sub>)<sub>2</sub>, 4x C<u>H</u>(CH<sub>3</sub>)<sub>2</sub>), 3.62 (t, *J* = 4.8 Hz, 4H, C<u>H</u><sub>2</sub>-N-C<u>H</u><sub>2</sub>), 3.53 (t, *J* = 4.7 Hz, 4H, 2x C<u>H</u><sub>2</sub>-NH), 1.29 (d, *J* = 6.5 Hz, 12H, 2x CH(C<u>H</u><sub>3</sub>)<sub>2</sub>), 1.25 (d, *J* = 6.4 Hz, 24H, 4x CH(C<u>H</u><sub>3</sub>)<sub>2</sub>).

<sup>13</sup>C NMR (400 MHz, D<sub>2</sub>O): δ 161.95 (<u>C</u>=NH), 155.60 (2x <u>C</u>=NH), 50.56 (2x <u>C</u>H(CH<sub>3</sub>)<sub>2</sub>), 49.71 (<u>C</u>H<sub>2</sub>-N-<u>C</u>H<sub>2</sub>), 47.07 (4x <u>C</u>H(CH<sub>3</sub>)<sub>2</sub>), 40.78 (2x <u>C</u>H<sub>2</sub>-NH), 25.03 (2x CH(<u>C</u>H<sub>3</sub>)<sub>2</sub>), 24.45 (4x CH(<u>C</u>H<sub>3</sub>)<sub>2</sub>).

The structure was confirmed via gCOSY, HSQCAD and gHMBCAD.

IR (solid): v (cm<sup>-1</sup>) 3199, 2973, 1615, 1430, 1388, 1368, 1335, 1268, 1167, 1147, 1133, 1073, 734, 591. LC-MS (LR) m/z: 161.6 [M+3H]<sup>3+</sup> (calc.: 161.5), 241.8 [M+2H]<sup>2+</sup> (calc.: 241.7), 482.4 [M+H]<sup>+</sup> (calc.: 482.5) HR ESI-MS, m/z: 482.4653 [M+H]<sup>+</sup> (calc.: 482.4659) Anal. calcd for  $C_{25}H_{62}O_2N_9Cl_3$  ( $C_{25}H_{55}N_9 \cdot 3HCl \cdot 2H_2O$ ): C, 47.88; H, 9.96; N, 20.10; Cl, 16.96. Found: C, 48.01; H, 10.35; N, 19.73; Cl, 17.44.

## Preparation of 1,1'-(propane-1,3-diyl)diguanidine sulfate (4)



Compound <u>9</u> (740 mg, 3.19 mmol) and S-methylisothiourea  $\cdot \frac{1}{2}$  H<sub>2</sub>SO<sub>4</sub> (481 mg, 3.45 mmol) were dissolved in 5 mL H<sub>2</sub>O and the solution was stirred under reflux. 5 M sodium hydroxide solution (690 µL, 3.45 mmol) was added dropwise and stirred for 4 h. Upon cooling to 4°C a mixture of <u>4</u> and <u>9</u> crystallized which was collected, dissolved in 5 mL H<sub>2</sub>O, and again treated with S-methylisothiourea  $\cdot \frac{1}{2}$  H<sub>2</sub>SO<sub>4</sub> (481 mg, 3.45 mmol) and sodium hydroxide solution (690 µL, 3.45 mmol) under reflux for 4 h. Large white crystals formed at 4°C which were sucked off and re-crystallized from H<sub>2</sub>O, yielding 206.5 mg (25%) of the product as sulfate salt.

<sup>1</sup>H NMR (400 MHz, D<sub>2</sub>O):  $\delta$  3.29 (t, *J* = 6.8 Hz, 4H, 2x C<u>H</u><sub>2</sub>-NH<sup>Gua</sup>), 1.9 (p, *J* = 6.8 Hz, 2H, C<u>H</u><sub>2</sub>).

<sup>13</sup>C NMR (400 MHz, D<sub>2</sub>O): δ 159.71 (2x <u>C</u>=NH), 41.22 (2x <u>C</u>H<sub>2</sub>-NH<sup>Gua</sup>), 29.85 (<u>C</u>H<sub>2</sub>).

The structure was confirmed via gCOSY, HSQCAD and gHMBCAD.

IR (solid): v (cm<sup>-1</sup>) 3304, 3131, 3026, 1690, 1667, 1636, 1465, 1223, 1085, 1060, 729, 628, 603, 556, 452.

HR ESI-MS, m/z: 159.1362 [M+H]<sup>+</sup> (calc.: 159.1358).

Anal. calcd for C<sub>5</sub>H<sub>16</sub>N<sub>6</sub>SO<sub>4</sub> (C<sub>5</sub>H<sub>14</sub>N<sub>6</sub> · H<sub>2</sub>SO<sub>4</sub>): C, 23.43; H, 6.29; N, 32.79; S, 12.51. Found: C, 23.47; H, 6.58; N, 32.36; S, 12.53.

#### Preparation of N,N-di-(3-guanidinylpropyl)amine sesquisulfate hydrate (5a)



Norspermidine (700  $\mu$ L, 5 mmol) was added to a solution of *S*-Methylisothiourea · ½ H<sub>2</sub>SO<sub>4</sub> (2505 mg, 18 mmol) in 7 mL H<sub>2</sub>O and stirred for 3 days at 105°C. A white precipitate formed after standing overnight at 4°C. The precipitate was sucked off and washed with ice cold H<sub>2</sub>O followed by ethanol. Recrystallization from H<sub>2</sub>O afforded the sesquisulfate hydrate of the product (1275 mg, 67%) as white solid.

<sup>1</sup>H NMR (400 MHz, D<sub>2</sub>O):  $\delta$  3.33 (t, *J* = 6.8 Hz, 4H, 2x HN=C-NH-C<u>H</u><sub>2</sub>), 3.15 ( $\Psi$ t, *J* = 7.9 Hz, 4H, C<u>H</u><sub>2</sub>-NH-C<u>H</u><sub>2</sub>), 2.02 ( $\Psi$ q, *J* = 7.4 Hz, 4H, 2x CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>).

<sup>13</sup>C NMR (400 MHz, D<sub>2</sub>O): δ 159.73 (2x <u>C</u>=NH), 47.85 (2x <u>C</u>H<sub>2</sub>-NH-<u>C</u>H<sub>2</sub>), 41.04 (2x HN=C-NH-<u>C</u>H<sub>2</sub>), 27.83 (2x CH<sub>2</sub>-<u>C</u>H<sub>2</sub>-CH<sub>2</sub>).

The structure was confirmed via gCOSY, HSQCAD and gHMBCAD

IR (solid): v (cm<sup>-1</sup>) 1682, 1638, 1479, 1392, 1049, 969, 770, 617, 595, 555, 462.

LC-MS (LR) m/z: 108.8 [M+2H]<sup>2+</sup> (calc.: 108.6), 216.2 [M+H]<sup>+</sup> (calc.: 216.2).

HR ESI-MS, m/z: 216.1930 [M+H]<sup>+</sup> (calc.: 216.1937).

Anal. calcd for  $C_8H_{26}N_7S_{1.5}O_7$  ( $C_8H_{21}N_7 \cdot 1.5H_2SO_4 \cdot H_2O$ ): C, 25.26; H, 6.89; N, 25.77; S, 12.64. Found: C, 25.65; H, 7.06; N, 25.42; S, 12.72.

Free base (<u>5b</u>):

The sulfate salt (200 mg, 0.526 mmol) was dissolved in 10 mL H<sub>2</sub>O and a solution of  $Ba(OH)_2$  (135 mg, 0.789 mmol) in 6 mL H<sub>2</sub>O was added. Precipitation was completed at 4°C, the suspension was filtered through celite and lyophilized. The residue was extracted with hot methanol giving the free base in quantitative yield. The free base was well soluble in hot methanol, the sulfate salt and barium salts were insoluble.

Preparation of 1-(3-guanidinylpropane-1-yl)-1,1'-(propane-1,3-diyl)-diguanidine trihydrochloride hydrate (<u>6a</u>)



Norspermidine (1399  $\mu$ L, 10 mmol) was dissolved in 7 mL ethanol, 1.5 mL of concentrated hydrochloric acid were added whereupon a white precipitate formed. Cyanamide (1640 mg, 39 mmol) was added and the reaction mixture was refluxed for 36.5 h and let to cool. The solvent was removed resulting in a highly viscous mass which crystallized after 8 days at room temperature. The crystalline material was washed with isopropanol and ice cold ethanol. It was recrystallized from hot methanol/isopropanol, yielding the trihydrochloride hydrate of the product (910 mg, 23.7 %) as white crystals.

<sup>1</sup>H NMR (400 MHz, D<sub>2</sub>O):  $\delta$  3.46 ( $\Psi$ t, J = 7.6 Hz, 4H, C<u>H</u><sub>2</sub>-N-C<u>H</u><sub>2</sub>), 3.27 (t, J = 6.9 Hz, 4H, 2x C<u>H</u><sub>2</sub>NH), 1.98 ( $\Psi$ q, J = 7.2 Hz, 4H, 2x CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>).

<sup>13</sup>C NMR (400 MHz, D<sub>2</sub>O): δ 159.62 (2x <u>C</u>=NH), 158.91 (<u>C</u>=NH), 48.79 (<u>C</u>H<sub>2</sub>-N-<u>C</u>H<sub>2</sub>), 41.18 (2x <u>C</u>H<sub>2</sub>NH), 28.35 (2x CH<sub>2</sub>-<u>C</u>H<sub>2</sub>-CH<sub>2</sub>).

The structure was confirmed via gCOSY, HSQCAD and gHMBCAD

IR (solid): v (cm<sup>-1</sup>) 3305, 3128, 1640, 1610, 1534, 1464, 1391, 1165, 1095, 1064, 464.

LC-MS (LR) m/z: 129.7 [M+2H]<sup>2+</sup> (calc.: 129.6), 258.2 [M+H]<sup>+</sup> (calc.: 258.2).

HR ESI-MS, m/z: 258.2147 [M+H]<sup>+</sup> (calc.: 258.2155).

Anal. calcd for  $C_9H_{28}N_9Cl_3O$  ( $C_9H_{23}N_9 \cdot 3HCl \cdot H_2O$ ): C, 28.10; H, 7.34; N, 32.77; Cl, 27.64.21. Found: C, 28.34; H, 7.54; N, 32.02; Cl, 27.14.

Free base (<u>6b</u>):

The chloride salt (350 mg, 0.91 mmol) was dissolved in 10 mL H<sub>2</sub>O and a suspension of  $Ag_2SO_4$  (423 mg, 1.36 mmol) was added in 6 mL H<sub>2</sub>O. After sonication, precipitation was completed at 4°C, the suspension was filtered through celite and a solution of Ba(OH)<sub>2</sub> (234 mg, 1.36 mmol) in 5 mL H<sub>2</sub>O was added. Precipitation was completed at 4°C and the suspension was filtered through celite and lyophilized. The residue was extracted with hot methanol and filtered through celite giving the free base in quantitative yield. The free base was well soluble in hot methanol, the sulfate salt, the silver and barium salts were insoluble.

## Preparation of N,N'-di-(3-guanidinylpropyl)-1,3-propanediamine disulfate hydrate (7a)



Norspermine (512  $\mu$ L, 2.5 mmol) was added to a solution of *S*-methylisothiourea ·  $\frac{1}{2}$  H<sub>2</sub>SO<sub>4</sub> (1670 mg, 12 mmol) in 4 mL H<sub>2</sub>O and stirred for 3 days at 105°C. The solvent was largely removed under reduced pressure and the residue dissolved by refluxing in ethanol/H<sub>2</sub>O. A

white precipitate formed upon cooling which was sucked off, washed with ethanol and recrystallized from ethanol/ $H_2O$ . The disulfate hydrate salt of the product (560 mg, 46%) was obtained as white solid.

<sup>1</sup>H NMR (400 MHz, D<sub>2</sub>O):  $\delta$  3.33 (t, *J* = 6.8 Hz, 4H, 2x HN=C-NH-C<u>H</u><sub>2</sub>), 3.21-3.17 (m, 4H, 2x HN=C-NH-(CH<sub>2</sub>)<sub>3</sub>-NH-C<u>H</u><sub>2</sub>), 3.19-3.15 (m, 4H, 2x HN=C-NH-(CH<sub>2</sub>)<sub>2</sub>-C<u>H</u><sub>2</sub>), 2.22-2.14 (m, 2H, HN-CH<sub>2</sub>-C<u>H</u><sub>2</sub>-CH<sub>2</sub>-NH), 2.07-2.00 (m, 4H, 2x HN=C-NH-CH<sub>2</sub>-<u>C</u>H<sub>2</sub>) <sup>13</sup>C NMR (400 MHz, D<sub>2</sub>O):  $\delta$  159.72 (2x C=NH), 47.88 (2x HN=C-NH-(CH<sub>2</sub>)<sub>2</sub>-CH<sub>2</sub>), 47.42

 $(2x \text{ HN}=\text{C-NH-(CH}_2)_3\text{-NH-}\underline{CH}_2), 41.02 (2x \text{ HN}=\text{C-}\underline{CH}_2), 27.80 (2x \text{ HN}=\text{C-NH-CH}_2-\underline{CH}_2), 25.52 (\text{HN-}CH_2-\underline{CH}_2-\text{CH}_2-\text{NH}).$ 

The structure was confirmed via gCOSY, HSQCAD and gHMBCAD.

IR (solid): v (cm<sup>-1</sup>) 3118, 1685, 1638, 1478, 1060, 601, 557.

LC-MS (LR) m/z: 137.2 [M+2H]<sup>2+</sup> (calc.: 137.1), 273.3 [M+H]<sup>+</sup> (calc.: 273.3).

HR ESI-MS, m/z: 273.2514 [M+H]<sup>+</sup> (calc.: 273.2515).

Anal. calcd for  $C_{11}H_{34}N_8S_2O_9$  ( $C_{11}H_{28}N_8 \cdot 2H_2SO_4 \cdot H_2O$ ): C, 27.15; H, 7.04; N, 23.03; S, 13.18. Found: C, 27.27; H, 7.13; N, 23.23; S, 12.98.

Free base (<u>7b</u>):

The sulfate salt (200 mg, 0.411 mmol) was dissolved in 10 mL H<sub>2</sub>O and a solution of  $Ba(OH)_2$  (141 mg, 0.822 mmol) in 6 mL H<sub>2</sub>O was added. Precipitation was completed at 4°C, the suspension was filtered through celite and lyophilized. The residue was extracted with hot methanol and filtered through celite giving the free base in quantitative yield. The free base was well soluble in hot methanol, the sulfate salt and barium salts were insoluble.

# Preparation of *N*-(4-guanidinylbutyl)-*N*-(3-guanidinylpropyl)amine sesquisulfate dihydrate (<u>8a</u>)



Spermidine (393  $\mu$ L, 2.5 mmol) was added to a solution of *S*-Methylisothiourea ·  $\frac{1}{2}$  H<sub>2</sub>SO<sub>4</sub> (1253 mg, 9.0 mmol) in 3.5 mL H<sub>2</sub>O and stirred for 3 days at 105°C. After cooling to room temperature a white precipitate was sucked off and washed with ice cold ethanol followed by ethanol. Crystallization from ethanol/H<sub>2</sub>O afforded the sesquisulfate dihydrate of the product (557 mg, 54%) as white solid.

<sup>1</sup>H NMR (400 MHz, D<sub>2</sub>O):  $\delta$  3.33 (t, *J* = 6.8 Hz, 2H, NH-C<u>H</u><sub>2</sub>-(CH<sub>2</sub>)<sub>2</sub>-NH-CH<sub>2</sub>), 3.25 (t, *J* = 6.7 Hz, 2H, NH-C<u>H</u><sub>2</sub>-(CH<sub>2</sub>)<sub>3</sub>-NH-CH<sub>2</sub>), 3.15-3.12 (m, 2H, NH-(CH<sub>2</sub>)<sub>2</sub>-C<u>H</u><sub>2</sub>-NH-CH<sub>2</sub>), 3.12-3.08 (m, 2H, NH-(CH<sub>2</sub>)<sub>3</sub>-C<u>H</u><sub>2</sub>-NH-CH<sub>2</sub>), 2.02 (tt, *J* = 7.9, 6.8 Hz, 2H, NH-CH<sub>2</sub>-C<u>H</u><sub>2</sub>-CH<sub>2</sub>-NH), 1.82-1.75 (m, 2H, NH-(CH<sub>2</sub>)<sub>2</sub>-C<u>H</u><sub>2</sub>-CH<sub>2</sub>-NH-CH<sub>2</sub>), 1.72-1.65 (m, 2H, NH-CH<sub>2</sub>-C<u>H</u><sub>2</sub>-(CH<sub>2</sub>)<sub>2</sub>-NH-CH<sub>2</sub>),

<sup>13</sup>C NMR (400 MHz, D<sub>2</sub>O): δ 159.72 (<u>C</u>=NH), 159.67 (<u>C</u>=NH), 50.05 (NH-(CH<sub>2</sub>)<sub>3</sub>-<u>C</u>H<sub>2</sub>-NH-CH<sub>2</sub>), 47.67 (NH-(CH<sub>2</sub>)<sub>2</sub>-<u>C</u>H<sub>2</sub>-NH-CH<sub>2</sub>), 43.26 (NH-<u>C</u>H<sub>2</sub>-(CH<sub>2</sub>)<sub>3</sub>-NH-CH<sub>2</sub>), 41.07 (NH-<u>C</u>H<sub>2</sub>-(CH<sub>2</sub>)<sub>2</sub>-NH-CH<sub>2</sub>), 27.87 (NH-CH<sub>2</sub>-<u>C</u>H<sub>2</sub>-(CH<sub>2</sub>)<sub>2</sub>-NH-CH<sub>2</sub>), 27.81 (NH-CH<sub>2</sub>-<u>C</u>H<sub>2</sub>-CH<sub>2</sub>-NH), 25.66 (NH-(CH<sub>2</sub>)<sub>2</sub>-<u>C</u>H<sub>2</sub>-CH<sub>2</sub>-NH-CH<sub>2</sub>).

The structure was confirmed via gCOSY, HSQCAD and gHMBCAD.

IR (solid): v (cm<sup>-1</sup>) 3118, 1629, 1553, 1478, 1044, 971, 748, 607.

LC-MS (LR) m/z: 115.7 [M+2H]<sup>2+</sup> (calc.: 115.6), 230.2 [M+H]<sup>+</sup> (calc.: 230.2).

HR ESI-MS, m/z: 230.2094 [M+H]<sup>+</sup> (calc.: 230.2093).

Anal. calcd for  $C_9H_{30}N_7S_{1.5}O_8$  ( $C_9H_{23}N_7 \cdot 1.5H_2SO_4 \cdot 2H_2O$ ): C, 26.21; H, 7.33; N, 23.77; S, 11.66. Found: C, 26.22; H, 7.23; N, 23.45; S, 10.93.

Free base (<u>8b</u>):

The sulfate salt (200 mg, 0.485 mmol) was dissolved in 10 mL H<sub>2</sub>O and a solution of  $Ba(OH)_2$  (125 mg, 0.727 mmol) in 6 mL H<sub>2</sub>O was added. Precipitation was completed at 4°C, the suspension was filtered through celite and lyophilized. The residue was extracted with hot methanol and filtered through celite giving the free base in quantitative yield. The free base was well soluble in hot methanol, the sulfate salt and barium salts were insoluble.

#### Preparation of (3-Aminopropyl)guanidine sulfate hydrate (9)

S-Methylisothiourea  $\cdot \frac{1}{2}$  H<sub>2</sub>SO<sub>4</sub> (696 mg, 5 mmol) was added to a solution of 1,3diaminopropane (1043 µL, 12.5 mmol) in 5 mL of 1:3 H<sub>2</sub>O/EtOH and stirred for a few minutes at room temperature until precipitation occurred. Another 5 mL of 1:3 H<sub>2</sub>O/EtOH were added and stirring continued for 30 min. The precipitate was sucked off and crystallized from hot H<sub>2</sub>O (yield: 416 mg, 36%).

<sup>1</sup>H NMR (400 MHz, D<sub>2</sub>O):  $\delta$  3.32 (t, *J* = 6.9 Hz, 2H, <sup>Gua</sup>NH-C<u>H</u><sub>2</sub>), 3.09 ( $\Psi$ t, J = 7.8 Hz, 2H, C<u>H</u><sub>2</sub>-NH<sub>2</sub>, 2.03-1.96 (m, 2H, <sup>Gua</sup>NH-CH<sub>2</sub>-C<u>H</u><sub>2</sub>-CH<sub>2</sub>-NH<sub>2</sub>)

<sup>13</sup>C NMR (400 MHz, D<sub>2</sub>O): δ 159.74 (<u>C</u>=NH), 41.05 (<sup>Gua</sup>NH-<u>C</u>H<sub>2</sub>), 39.73 (<u>C</u>H<sub>2</sub>-NH<sub>2</sub>), 28.95 (<u>C</u>H<sub>2</sub>).

The structure was confirmed via gCOSY, HSQCAD and gHMBCAD. IR (solid): v (cm<sup>-1</sup>) 2884, 2805, 1684, 1638, 1079, 863, 738, 618, 601, 552, 469. LC-MS (LR) m/z: 117.2 [M+H]<sup>+</sup> (calc.: 117.1). HR ESI-MS, m/z: 117.1140 [M+H]<sup>+</sup> (calc.: 117.1149). Anal. calcd for  $C_4H_{16}N_4SO_5$  ( $C_4H_{12}N_4 \cdot H_2SO_4 \cdot H_2O$ ): C, 20.69; H, 6.94; N, 24.12; S, 13.81. Found: C, 21.05; H, 6.95; N, 23.87; S, 13.89.

Preparation of *N*-(3-aminopropyl)-*N*-(3- $N^2$ , $N^3$ -diisopropylguanidinylpropyl)amine triformate (10)



*N,N*-Diisopropylcarbodiimide (313  $\mu$ L, 2.0 mmol) was dissolved in 1 mL dichloromethane and norspermidine (140  $\mu$ L, 1.0 mmol) was added under vigorous stirring. After stirring over night, the solvent was evaporated and the residue dissolved in a mixture of 1.1 mL water and 0.4 mL methanol. 50  $\mu$ L of the solution were purified by HPLC with on a Hypercarb column (250 x 10 mm, 5  $\mu$ m particle size, Thermo Scientific). Mobile phase A = water, B = acetonitril, gradient (T<sub>min</sub>) T<sub>0</sub>: B = 0%, T<sub>30</sub>: B = 100%. The product eluted with a retention time of 8 min (yield 1.78 mg, 14%).

<sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>OD):  $\delta$  3.82 (sept, J = 6.6 Hz, 2H, 2x C<u>H</u>(CH<sub>3</sub>)<sub>2</sub>), 3.36 (t, J = 7.0 Hz, 2 H, NH<sup>Gua</sup>-C<u>H</u><sub>2</sub>), 3.03 (t, J = 7.6, 2H, C<u>H</u><sub>2</sub>-NH<sub>2</sub>), 2.99 (Ψt, J = 7.4 Hz, 2H, C<u>H</u><sub>2</sub>-(CH<sub>2</sub>)<sub>2</sub>-NH<sub>2</sub>), 2.95 (Ψt, J = 7.4 Hz, 2H, NH<sup>Gua</sup>-(CH<sub>2</sub>)<sub>2</sub>-C<u>H</u><sub>2</sub>-NH), 2.01 (Ψq, J = 7.5 Hz, 2H, C<u>H</u><sub>2</sub>-CH<sub>2</sub>-NH<sub>2</sub>), 1.94 (Ψq, J = 7.1 Hz, 2H, C<u>H</u><sub>2</sub>-CH<sub>2</sub>-NH), 1.27 (d, J = 6.4, 12H, 4x CH<sub>3</sub>).

<sup>13</sup>C NMR (400 MHz, CD<sub>3</sub>OD): δ 154.50 (<u>C</u>=N), 46.61 (NH<sup>Gua</sup>-(CH<sub>2</sub>)<sub>2</sub>-<u>C</u>H<sub>2</sub>-NH), 46.50 (<u>C</u>H<sub>2</sub>-(CH<sub>2</sub>)<sub>2</sub>-NH<sub>2</sub>), 45.52 (<u>C</u>H(CH<sub>3</sub>)<sub>2</sub>), 40.09 (NH<sup>Gua</sup>-<u>C</u>H<sub>2</sub>), 38.37 (<u>C</u>H<sub>2</sub>-NH<sub>2</sub>), 27.86 (<u>C</u>H<sub>2</sub>-CH<sub>2</sub>-NH), 26.53 (<u>C</u>H<sub>2</sub>-CH<sub>2</sub>-NH<sub>2</sub>), 22.76 (<u>C</u>H<sub>3</sub>).

The structure was confirmed via gCOSY, HSQCAD and gHMBCAD.

HR ESI-MS, m/z: 258.2662 [M+H]<sup>+</sup> (calc.: 258.2658).

#### Preparation of *m*-Phenylenedibiguanide dihydrochloride (11a)



1,3-Diaminobenzene (1081 mg, 10 mmol) was dissolved in 7 mL H<sub>2</sub>O, concentrated hydrochloric acid was added until pH = 1 and dicyandiamide (2186 mg, 26 mmol) was added. The reaction was refluxed for 4 h, let to cool and acetone was added until a white precipitate formed. After standing over night the fine suspended material had settled and was washed with two times 20 mL acetone. After drying, the material was crystallized from hot ethanol/H<sub>2</sub>O. The white crystalline material was sucked off and washed with cold ethanol (yield: 1.956 g, 56%).

<sup>1</sup>H NMR (400 MHz, D<sub>2</sub>O):  $\delta$  7.45 (t, J = 8.1 Hz, 1H, aromate: CH=C<u>H</u>-CH), 7.33 (t, J = 2.0

Hz, 1H, aromate: N-C=C<u>H</u>-C-N), 7.15 (dd, *J* = 8.1, 2.1 Hz, 2H, aromate: C<u>H</u>=CH-C<u>H</u>).

<sup>13</sup>C NMR (400 MHz, D<sub>2</sub>O): δ 163.83 (2x <u>C</u>=NH), 160.34 (2x <u>C</u>=NH), 140.16 (N-<u>C</u>=CH-<u>C</u>-

N), 133.12 (CH=<u>C</u>H-CH), 123.68 (<u>C</u>H=CH-<u>C</u>H), 121.68 (N-C=<u>C</u>H-C-N).

The structure was confirmed via gCOSY, HSQCAD and gHMBCAD

IR (solid): v (cm<sup>-1</sup>) 3292, 3136, 1628, 1575, 1541, 1525, 1448, 1394, 1283, 781, 600, 468.

HR ESI-MS, m/z: 277.1632 [M+H]<sup>+</sup> (calc.: 277.1638).

Anal. calcd for  $C_{10}H_{18}N_{10}Cl_2$  ( $C_{10}H_{16}N_{10} \cdot 2HCl$ ): C, 34.39; H, 5.20; N, 40.11; Cl, 20.30. Found: C, 34.52; H, 5.05; N, 38.94; Cl, 20.85.

Free base (<u>11b</u>):

The chloride salt (350 mg, 1.00 mmol) was dissolved in 10 mL H<sub>2</sub>O and a suspension of  $Ag_2SO_4$  (313 mg, 1.00 mmol) was added in 10 mL H<sub>2</sub>O. After sonication, precipitation was completed at 4°C and a solution of Ba(OH)<sub>2</sub> (172 mg, 1.00 mmol) in 5 mL H<sub>2</sub>O was added. Precipitation was completed at 4°C, 5 mL ethanol were added and the suspension was filtered through celite and lyophilized. The residue was extracted with hot ethanol and filtered through celite giving the free base in 33% yield. The free base was well soluble in hot ethanol, the sulfate salt, the silver and barium salts were insoluble.

#### Preparation of 1-(2-(2-Iminoimidazolidin-1-yl)ethyl)guanidine sulfate (12)



Diethylenetriamine (540  $\mu$ L, 5.0 mmol) was added to a solution of *S*-methylisothiourea (1392 mg, 10 mmol) in 2.5 mL H<sub>2</sub>O and refluxed for 3 days. After cooling to room temperature, 10 mL isopropanol were added followed by 1.3 mL of a 10% solution of sulfuric acid in isopropanol. The solvent was removed and the residue crystallized from hot ethanol/H<sub>2</sub>O. The crystals were sucked off, washed with ice cold ethanol and re-crystallized from hot ethanol/H<sub>2</sub>O affording 794 mg (59 %) of white crystalline material.

<sup>1</sup>H NMR (400 MHz, D<sub>2</sub>O): δ 3.81-3.76 (m, 2H, cyclic NH-CH<sub>2</sub>-C<u>H</u><sub>2</sub>-N), 3.68-3.64 (m, 2H, cyclic NH-C<u>H</u><sub>2</sub>-CH<sub>2</sub>-N), 3.55-3.49 (m, 4H, N-C<u>H</u><sub>2</sub>-CH<sub>2</sub>-NH<sup>Gua</sup> and N-CH<sub>2</sub>-C<u>H</u><sub>2</sub>-NH<sup>Gua</sup>). <sup>13</sup>C NMR (400 MHz, D<sub>2</sub>O): δ 161.93 (ring <u>C</u>=NH), 159.91 (<u>C</u>=NH), 50.70 (cyclic NH-CH<sub>2</sub>-<u>C</u>H<sub>2</sub>-N), 46.49 (N-<u>C</u>H<sub>2</sub>-CH<sub>2</sub>-NH<sup>Gua</sup>), 43.59 (cyclic NH-<u>C</u>H<sub>2</sub>-CH<sub>2</sub>-N), 41.38 (N-CH<sub>2</sub>-<u>C</u>H<sub>2</sub>-NH<sup>Gua</sup>).

The structure was confirmed via gCOSY, HSQCAD and gHMBCAD.

IR (solid): v (cm<sup>-1</sup>) 3097, 2996, 2899, 1674, 1646, 1631, 1587, 1462, 1312, 1074, 797, 647, 614, 593, 549.

HR ESI-MS, m/z: 171.1367 [M+H]<sup>+</sup> (calc.: 171.1358)

Anal. calcd for C<sub>6</sub>H<sub>16</sub>N<sub>6</sub>SO<sub>4</sub> (C<sub>6</sub>H<sub>14</sub>N<sub>6</sub> · H<sub>2</sub>SO<sub>4</sub>): C, 26.86; H, 6.01; N, 31.32; S, 11.95. Found: C, 26.85; H, 6.27; N, 31.06; S, 11.92.

## Preparation of Norspermidine trihydrochloride

$$H_2N$$
  $N$   $H_2$   $NH_2$ 

Norspermidine (2.80 mL, 20 mmol) was dissolved in 40 mL ethanol and 12 mL of a 1:1 (vol.) mixture of 37% aqueous HCl and ethanol was added. The precipitate was sucked off, washed with ethanol and the product crystallized from hot ethanol/ H<sub>2</sub>O (yield 3.73 g, 78%). <sup>1</sup>H NMR (400 MHz, D<sub>2</sub>O):  $\delta$  3.20 ( $\Psi$ t, *J* = 8.0 Hz, 4H, C<u>H</u><sub>2</sub>-N-C<u>H</u><sub>2</sub>), 3.13 ( $\Psi$ t, *J* = 7.9 Hz, 4H, 2x CH<sub>2</sub>NH<sub>2</sub>), 2.16-2.08 (m, 4H, 2x CH<sub>2</sub>-CH<sub>2</sub>). <sup>13</sup>C NMR (400 MHz, D<sub>2</sub>O): δ 47.55 (<u>C</u>H<sub>2</sub>-N-<u>C</u>H<sub>2</sub>), 39.40 (2x <u>C</u>H<sub>2</sub>NH<sub>2</sub>), 26.60 (2x CH<sub>2</sub>-<u>C</u>H<sub>2</sub>-CH<sub>2</sub>).

The structure was confirmed via gCOSY, HSQCAD and gHMBCAD.

IR (solid): v (cm<sup>-1</sup>) 2954, 2894, 2841, 2751, 2702, 2502, 2415, 2025, 1607, 1526, 1489, 1462, 1404, 1356, 1281, 1165, 1055, 1027, 994, 930, 779, 436.

HR ESI-MS, m/z: 132.1504 [M+H]<sup>+</sup> (calc.: 132.1501)

Anal. calcd for C<sub>6</sub>H<sub>20</sub>N<sub>3</sub>Cl<sub>3</sub> (C<sub>6</sub>H<sub>17</sub>N<sub>3</sub> · 3HCl): C, 29.95; H, 8.38; N, 17.46; Cl, 44.21. Found: C, 30.12; H, 8.55; N, 17.18; Cl, 44.16.

## Preparation of Norspermidine disulfate hydrate



Norspermidine (700  $\mu$ L, 5.0 mmol) was dissolved in 10 mL ethanol and 8 mL of 20% sulfuric acid in ethanol was added slowly under ice cooling. The precipitate was sucked off and dissolved in hot ethanol/H<sub>2</sub>O. Addition of isopropanol precipitated amorphous material which was crystallized from H<sub>2</sub>O/acetone (yield 510 mg, 30%).

<sup>1</sup>H NMR (400 MHz, D<sub>2</sub>O):  $\delta$  3.18 ( $\Psi$ t, J = 7.8 Hz, 4H, C<u>H</u><sub>2</sub>-N-C<u>H</u><sub>2</sub>), 3.12 ( $\Psi$ t, J = 7.9 Hz, 4H, 2x C<u>H</u><sub>2</sub>NH<sub>2</sub>), 2.16-2.08 (m, 4H, 2x CH<sub>2</sub>-C<u>H</u><sub>2</sub>-CH<sub>2</sub>).

<sup>13</sup>C NMR (400 MHz, D<sub>2</sub>O): δ 47.63 (<u>C</u>H<sub>2</sub>-N-<u>C</u>H<sub>2</sub>), 39.50 (2x <u>C</u>H<sub>2</sub>NH<sub>2</sub>), 26.68 (2x CH<sub>2</sub>-<u>C</u>H<sub>2</sub>-CH<sub>2</sub>).

The structure was confirmed via gCOSY, HSQCAD and gHMBCAD.

IR (solid): v (cm<sup>-1</sup>) 3448, 2923, 1633, 1526, 1475, 1407, 1372, 1213, 1085, 996, 967, 895, 760, 595, 524, 420.

HR ESI-MS, m/z: 132.1505 [M+H]<sup>+</sup> (calc.: 132.1501).

Anal. calcd for  $C_6H_{23}N_3S_2O_9$  ( $C_6H_{17}N_3 \cdot 2H_2SO_4 \cdot H_2O$ ): C, 20.86; H, 6.71; N, 12.17; S, 18.57. Found: C, 21.02; H, 6.96; N, 12.16; S, 18.43.

## 2) Bacterial strains

*B. subtilis* strain NCBI 3610: a wild strain of *B. subtilis*, which is capable of forming robust biofilms.<sup>1</sup>

Staphylococcus aureus SC01 was obtained from the Kolter lab collection.<sup>2</sup>

*B. subtilis* strain NCBI 3610 and *S. aureus* SC01 were isolated as a single colonies growing overnight at 37°C on solid LB medium.

#### 3) Biofilm inhibition assay

*B. subtilis* strain NCBI 3610 was grown for 2 h with shaking at 37°C, then diluted 1:1000 in either 3 ml/well (12 well plates, polystyrene, VWR) or 1.5 ml/well (24 wells plates, polystyrene, VWR) of defined biofilm medium MSgg (pH 7.0).<sup>1</sup> Cells were grown without shaking for 3 days at 25°C. For pH experiments bacteria were additionally grown in MSgg at pH 6.0 and 8.0. Floating biofilms (pellicles) of *B. subtilis* were examined optically.

*S. aureus* strain SC01 was grown overnight with shaking at 37°C, then diluted 1:100 in 1ml/well of TSB medium, 0.5% glucose, 3% NaCl (pH 7.0). Cells were grown without shaking for 24 h at 37°C. For pH experiments bacteria were additionally grown in TSB at pH 6.0 and 8.0. Submerged biofilms of *S. aureus* were observed and quantified by crystal violet staining as blow.

**Crystal violet staining.** Crystal Violet (CV) staining was done as described previously except that the cells were grown in 24-well plates.<sup>3</sup> Wells were stained with 300  $\mu$ l of 1.0 % Crystal-violet dye, rinsed twice with 2 ml deionized water and thoroughly dried. For quantification, 0.5 ml of 95 % ethanol were added to each well. Plates were incubated for one hour at room temperature with shaking. CV solution was diluted and the OD at 595 nm was measured using an Ultraspec 2000 (Pharmacia Biotech).

#### 4) Growth data

*B. subtilis* was diluted from overnight LB cultures to MSgg, with and without the test compounds at 200  $\mu$ M and *S. aureus* was diluted from overnight LB culture to TSB medium, 0.5% glucose, 3% NaCl (pH 7.0). with and without the test compounds at 200  $\mu$ M. The strains were grown under shaking at 37°C and OD<sub>600</sub> was measured at various times. Each compound was tested with 3-4 replicates. Growth curves are given in figure S20.

#### 5) Solubility data (K<sub>s</sub> values)

Typically 20-30 mg of each compound was incubated with a volume of  $H_2O$  that was insufficient to fully dissolve the entire amount (typically 100-300  $\mu$ L) and incubated for at least 24 h at 20°C. The undissolved material was centrifuged off and a defined volume of the saturated was removed, lyophilized and the weight of was determined.

#### 6) pK<sub>a</sub> determination and ion speciation

For pH and pD measurements, a PHR-146B Micro combination pH electrode (Cole Parmer) was used. The electrode was calibrated for pH using commercial standard solutions of pH 4.0, 7.0 and 10.0 and for pD applying the relation<sup>4</sup> pD = pH + 0.40 to convert pD to pH which was additionally verified by two pD calibration standards of K<sub>2</sub>DPO<sub>4</sub>/ KD<sub>2</sub>PO<sub>4</sub> (0.025 M K<sub>2</sub>DPO<sub>4</sub> and 0.025 M KD<sub>2</sub>PO<sub>4</sub> in D2O) and K<sub>2</sub>CO<sub>3</sub>/KDCO<sub>3</sub> (0.025 M K<sub>2</sub>CO<sub>3</sub> and 0.025 M KDCO<sub>3</sub>) at  $25^{\circ}C.^{5}$ 

To measure  $pK_a$  values in the range of  $2 \le pH \le 12$  the strategy of Blagbrough et al. was applied. Titrations were conducted in D<sub>2</sub>O using <sup>1</sup>H or <sup>13</sup>C NMR as described before without correction for ionic strength.<sup>5</sup> Solutions of 25 mM of the compounds in D<sub>2</sub>O were prepared (~1 mL to compensate for losses during titration steps). Small amounts of DCl (35 wt.% in D<sub>2</sub>O, 99 atom% D, Sigma Aldrich) and KOD (40 wt.% in D<sub>2</sub>O, 98 atom% D, Sigma Aldrich) were added successively and pD values were measured followed by <sup>1</sup>H or <sup>13</sup>C NMR spectroscopy at 25°C.

Higher pK<sub>a</sub> values were determined by the method recommended by Popov et al. for  $12 \le pH \le 14$  in H<sub>2</sub>O by <sup>13</sup>C NMR at fixed ionic strength of I = 1 M with KOH/KCl using calculated pH values.<sup>4</sup> Compounds were dissolved in the pre-adjusted KOH/KCl solutions to give about 500 µL of 25 mM concentration. Corrections for the deprotonation of compound were applied as suggested. Close below pH 12.5, the calculated values were checked and if necessary corrected by pH measurements. In some cases, pH values below 12 were added to expand the pH range and directly measured by the microelectrode after compound addition.

Chemical shifts were plotted as function of pH or pD (Figure S2-S11) and cumulative association constants were calculated by HypNMR2008 (Protonic Software)<sup>6, 7</sup> with the setting  $pK_w = 13.80^8$  and using the relation pD = pH + 0.40 to finally convert  $pK_a$  (D<sub>2</sub>O) to  $pK_a$  (H<sub>2</sub>O).<sup>4</sup>

Ion speciations were calculated from  $pK_a$  values using the program HySS2009 - Hyperquad Simulation and Speciation (Protonic Software) and are given in Figure S12-S14.

#### 7) Crystal structures

Compounds 2a, 5a and 11a were re-crystallized from deionized water and structures were determined by X-ray diffraction at the Harvard University Center for Crystallographic Studies. Data were collected at 100 K from a crystal mounted on a Bruker APEX II CCD diffractometer (Mo<sub>Ka</sub> radiation,  $\lambda = 0.71073$  Å) equipped with an Oxford Cryosystems nitrogen flow apparatus. The collection method involved 0.5° scans in  $\omega$  at 28° in 2 $\theta$ . Data

integration down to 0.78 Å resolution was carried out using SAINT V7.46 A (Bruker diffractometer, 2009) with reflection spot size optimization. Absorption corrections were made with the program SADABS (Bruker diffractometer, 2009). The structure was solved by the direct methods procedure and refined by least-squares methods again  $F^2$  using SHELXS-97 and SHELXL-97<sup>9</sup> with OLEX 2 interface.<sup>10</sup> Non-hydrogen atoms were refined anisotropically, and hydrogen atoms were allowed to ride on the respective atoms. Crystal data as well as details of data collection and refinement are summarized in Table S4 and Figures S17-S19.

**Table S1.** Spacing of the repeating units of potential polysaccharides in the EPS. Distances of energy minimized structures of Chem3D Ultra 10.0.were measured in PyMOL as half medium linear distance across three sugar units in the secondary structure corresponding to the binding motif of norspermidine. For polyamines, shortest NH to NH bond distances are given.

Structure	Medium distance
Norspermidine	4.9 Å
Diethylenetriamine	3.7 Å
Spermidine (butyldiamine distance)	6.3 Å
Poly-α-1,4-D-glucuronic acid	5.3 Å
Poly-α-1,2-D-glucuronic acid	4.9 Å
Poly-α-1,6-D-glucose-2- sulfate	5.0 Å
Poly-α-1,6-D-glucose-β- 1,3-D-glucuronic acid	4.6 Å
Poly-β-1,6-N-acetyl-d- glucosamine (PGA)	5.2 Å
Helical poly(γ-D-glutamic acid)	~ 5 Å

		MBIC ( $\mu$ M) at pH 7.0				
Compound	1	B. subtilis	S. aureus <sup>a</sup>			
	base	37.5±12.5	175±25			
Norspermidine	sulfate	500	250			
-	chloride	150	500			
Norspermine	base	200	200			
Diethylenetriamine	base	>1000	>1000			
Biguanide	chloride	>1000	>1000			
Spermidine	base	1000	1000			
1	sulfate	>1000	>1000			
a	sulfate	>1000	750			
<sup>2</sup> b	base	1000	1000 (50*)			
3	chloride	>1000	750			
4	sulfate	>1000	50			
_ a	sulfate	500	75			
ъ <sub>b</sub>	base	375±125	400			
a	chloride	10*	>1000 (500*)			
6 b	base	10	50			
_ a	sulfate	5	55±15			
b	base	2	250			
a	sulfate	600	300			
8 b	base	>1000	100			
9	sulfate	100	500			
10	formate	30	20±10			
11 <b>a</b>	chloride	30	300			
ll b	base	7±3	750±250			
12	sulfate	>1000	>1000			
	1 1		C 1 · C 1 · C · · · · ·			

**Table S2.** Minimal biofilm inhibitory concentrations (MBIC) for all compounds.

<sup>a</sup>MBICs for *S. aureus* are based on 75% reduction of biofilm formation, \*incomplete inhibition.

Compound	<b>K</b> <sub>s</sub> (M)
5a	$(3.1 \pm 0.1) \cdot 10^{-2}$
6a	$1.58 \pm 0.01$
8a	$(6.3 \pm 0.3) \cdot 10^{-2}$
2a	$(5.34 \pm 0.03) \cdot 10^{-2}$
4	$(4.6 \pm 0.1) \cdot 10^{-2}$
9	$(4.1 \pm 0.1) \cdot 10^{-2}$
3	$(2.75 \pm 0.02) \cdot 10^{-1}$
1	$(6.4 \pm 0.2) \cdot 10^{-2}$
Norspermidine chloride	$2.84 \pm 0.03$
Norspermidine sulfate	$2.12 \pm 0.08$
<b>11a</b>	$(3.9 \pm 0.1) \cdot 10^{-1}$
7a	$(8.4 \pm 0.7) \cdot 10^{-2}$
12	$(3.0 \pm 0.2) \cdot 10^{-1}$

**Table S3.** Solubility data as  $K_s$  values for the compounds.

	Compound <u>2</u> Compound <u>5</u> Co		Compound <u>11</u>	
Crystal data			·	
Crystal system, space group	Monoclinic, $P2_1/n$	Monoclinic, C2/c	Triclinic, P-1	
a, b, c (Å)	14.6138 (10), 12.5995 (9), 14.617 (1)	9.0449 (13), 10.9984 (16), 36.497 (5)	9.3527 (11), 9.5924 (11), 11.4378 (14)	
α, β, γ (°)	93.700 (1)	94.140 (2)	82.994 (2), 71.145 (2), 73.734 (2)	
$V(\text{\AA}^3)$	2685.8 (3)	3621.2 (9)	931.65 (19)	
Ζ	8	8	2	
Radiation type	Μο Κα	Μο Κα	Μο Κα	
$\mu (mm^{-1})$	0.27	0.29	0.38	
Crystal size (mm)	$0.24 \times 0.16 \times 0.14$	$0.18 \times 0.10 \times 0.05$	$0.20\times0.16\times0.12$	
Data collection				
Absorption correction	Multi-scan SADABS	Multi-scan SADABS	Multi-scan SADABS	
$T_{\min}, T_{\max}$	0.938, 0.963	0.950, 0.986	0.928, 0.956	
No. of measured, independent and observed $[I > 2\sigma(I)]$ reflections	40375, 5934, 5362	26494, 3993, 3252	12063, 4113, 3599	
R <sub>int</sub>	0.046	0.053	0.026	
$(\sin \theta / \lambda)_{max} (\text{\AA}^{-1})$	0.641	0.641	0.641	
Refinement				
$R[F^2 > 2\sigma(F^2)], wR(F^2), S$	0.035, 0.090, 1.03	0.061, 0.167, 1.06	0.033, 0.086, 1.04	
No. of reflections	5934	3993	4113	
No. of parameters	446	305	305	
No. of restraints	26	26	20	
H-atom treatment	H atoms treated by a mixture of independent and constrained refinement			
$\Delta \rho_{\text{max}}, \Delta \rho_{\text{min}} (e \text{ Å}^{-3})$	0.66, -0.42	0.46, -0.52	0.37, -0.30	

Table S4.	Crystal	data,	and	data	on	collection	and	refinemen	ıt.



**Figure S1.** Structures of norspermidine and other polyamines and biofilm inhibition in *B. subtilis.* 



**Figure S2.** <sup>1</sup>H-NMR-titration curves of polyamines for A) spermidine, B) diethylenetriamine, C) norspermidine, D) norspermine.



**Figure S3.** A) and B) <sup>13</sup>C-NMR-titration curves of compound 1.



**Figure S4.** NMR-titration curves of compound **2**. A) <sup>1</sup>H-NMR titration in D<sub>2</sub>O. B) and C) <sup>13</sup>C-NMR titration in H<sub>2</sub>O for the upper pH range.



**Figure S5.** NMR-titration curves of compound 4. A), B) and C)  $^{13}$ C-NMR titration in H<sub>2</sub>O.



**Figure S6.** NMR-titration curves of compound **5**. A) and B)  $^{13}$ C-NMR for the upper pH range in H<sub>2</sub>O and C)  $^{1}$ H-NMR titration in D<sub>2</sub>O.



**Figure S7.** A) and B) <sup>13</sup>C-NMR titration curves of compound 6.



**Figure S8.** NMR-titration curves of compound 7. A) - C)  ${}^{13}$ C-NMR for the upper pH range in H<sub>2</sub>O and D) - E)  ${}^{1}$ H-NMR titration in D<sub>2</sub>O.



**Figure S9.** NMR-titration curves of compound **8**. A) and B) <sup>1</sup>H-NMR titration in D<sub>2</sub>O. C)-F) <sup>13</sup>C-NMR for the upper pH range in H<sub>2</sub>O.



**Figure S10.** NMR-titration curves of compound **9**. A) and B)  ${}^{13}$ C-NMR for the upper pH range in H<sub>2</sub>O and C)  ${}^{1}$ H-NMR titration in D<sub>2</sub>O.



**Figure S11.** <sup>13</sup>C-NMR-titration curves of compound **11**. A) and B) titration in  $D_2O$  and C) refinement in  $H_2O$  for the upper pH range.



**Figure S12.** Ion speciation data for A) norspermidine, B) norspermine, C) spermidine, and D) diethylenetriamine.



Figure S13. Ion speciation data for compounds  $\underline{1}$  (A),  $\underline{2}$  (B),  $\underline{4}$  (C),  $\underline{5}$  (D),  $\underline{6}$  (E), and  $\underline{7}$  (F).



Figure S14. Ion speciation data for compounds  $\underline{8}$  (A),  $\underline{9}$  (B), and  $\underline{11}$  (C).



Figure S15. Average degree of protonation in dependence of pH for compounds 1, 4 and 9.



Figure S16. Average degree of protonation in dependence of pH for spermidine and compounds 5, 8 and 11.



Figure S17. Crystal structure of compound <u>2</u>.



Figure S18. Crystal structure of compound <u>5</u>.



Figure S19. Crystal structure of compound <u>11</u>.



**Figure S20.** No significant inhibition of bacterial growth in A) *S. aureus* and B) *B. subtilis* was observed with 200  $\mu$ M of selected compounds.

## References

1. Branda, S. S.; Gonzalez-Pastor, J. E.; Ben-Yehuda, S.; Losick, R.; Kolter, R., Fruiting body formation by Bacillus subtilis. *Proc Natl Acad Sci U S A* **2001**, 98, (20), 11621-6.

2. Lopez, D.; Kolter, R., Functional microdomains in bacterial membranes. *Genes Dev* 2010, 24, (17), 1893-902.

3. O'Toole, G. A.; Kolter, R., Flagellar and twitching motility are necessary for Pseudomonas aeruginosa biofilm development. *Mol Microbiol* **1998**, 30, (2), 295-304.

4. Popov, K.; Rönkkömäki, H.; Lajunen, L. H. J., Guidelines for nmr measurements for determination of high and low pka values (IUPAC Technical Report). *Pure Appl. Chem.* **2006**, 78, (3), 663-675.

5. Blagbrough, I. S.; Metwally, A. A.; Geall, A. J., Measurement of polyamine pKa values. *Methods Mol Biol* **2011**, 720, 493-503.

6. Frassineti, C.; Ghelli, S.; Gans, P.; Sabatini, A.; Moruzzi, M. S.; Vacca, A., Nuclear magnetic resonance as a tool for determining protonation constants of natural polyprotic bases in solution. *Anal Biochem* **1995**, 231, (2), 374-82.

7. Frassineti, C.; Alderighi, L.; Gans, P.; Sabatini, A.; Vacca, A.; Ghelli, S., Determination of protonation constants of some fluorinated polyamines by means of 13C NMR data processed by the new computer program HypNMR2000. Protonation sequence in polyamines. *Anal Bioanal Chem* **2003**, 376, (7), 1041-52.

8. Kron, I.; Marshall, S. L.; May, P. M.; Hefter, G.; Königsberger, E., The Ionic Product of Water in Highly Concentrated Aqueous Electrolyte Solutions. *Monatsh Chem* **1995**, 126, 819-837.

9. Sheldrick, G. M., A short history of SHELX. *Acta Crystallogr A* 2008, 64, (Pt 1), 112-22.

10. Dolomanov, O. V.; Bourhis, L. J.; Gildea, R. J.; Howard, J. A. K.; Puschmann, H., OLEX2: a complete structure solution, refinement and analysis program. *J Appl Cryst* **2009**, 42, (229-341).