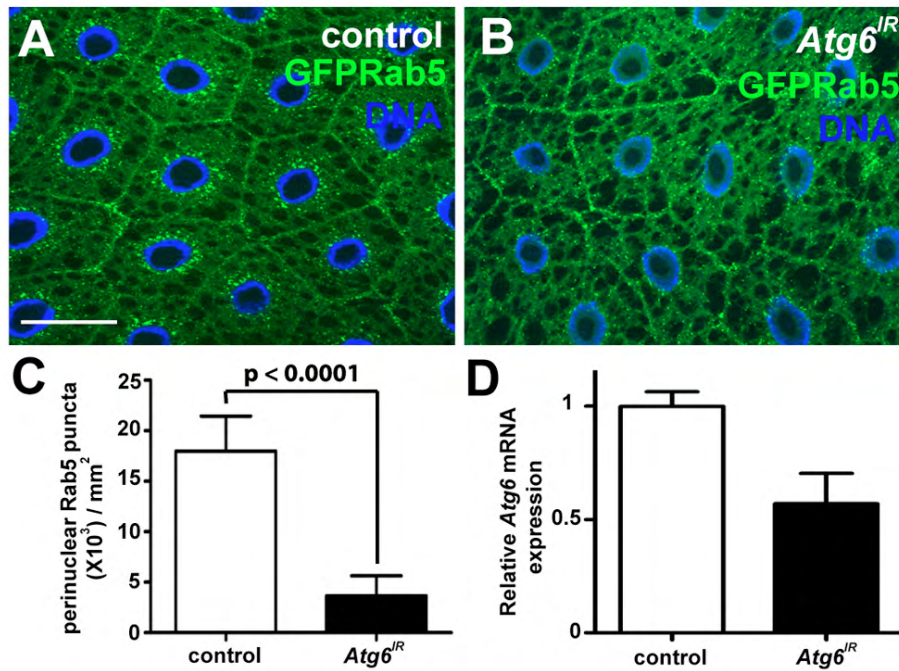
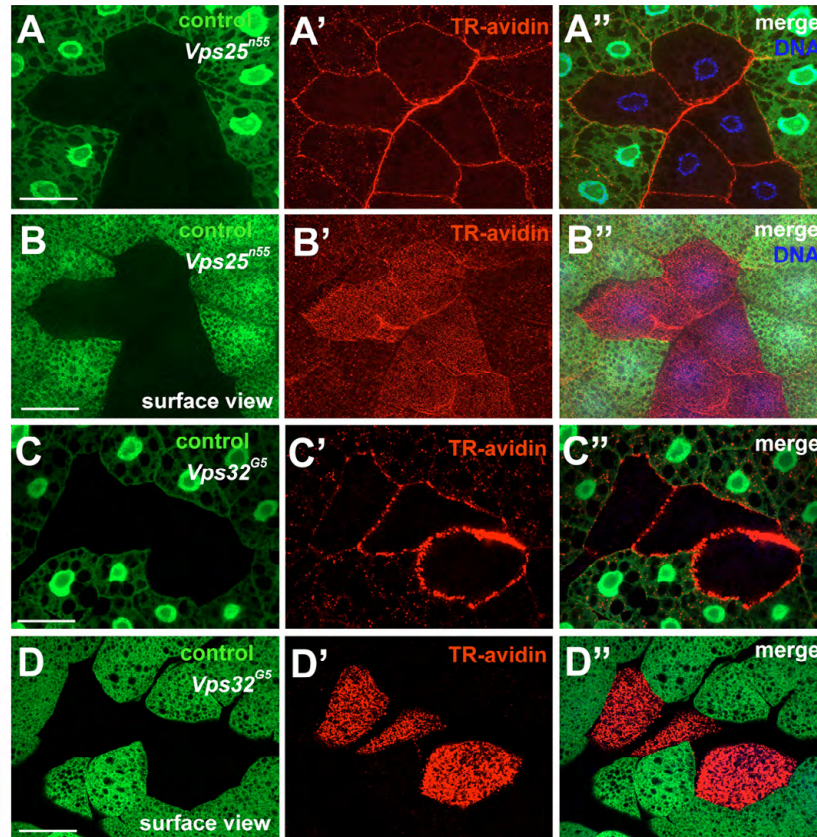


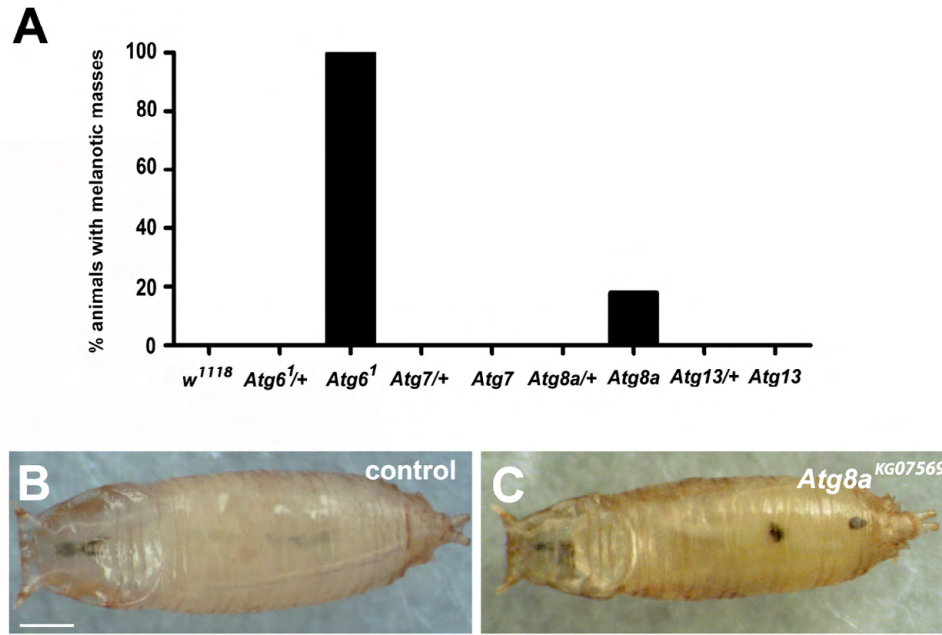
**Fig. S1 *Atg6* is required for development and autophagy.** (A) *Atg6* is required for development, as homozygous *Atg6* mutant animals die during development. Most homozygous *Atg6*<sup>1</sup> animal lethality occurs prior to puparium formation, and lethality can be rescued by expressing an *atg6* transgene ubiquitously in the whole animal. (B) Quantification of Ref(2)P (fly p62) puncta in control and homozygous *Atg6*<sup>1</sup> mutant fat body cells as in Fig. 1G. A two-tailed *t*-test was used for statistical analysis, and the *P*-value relative to control was  $5.07 \times 10^{-9}$ . The error bars represent s.d. (C-C'') Ref(2)P aggregates did not accumulate *Atg6*<sup>1</sup> mutant clone cells (mCherry negative) expressing a myc-tagged *Atg6* transgene in the entire fat body (*n*=7). Scale bar: 50  $\mu$ m.



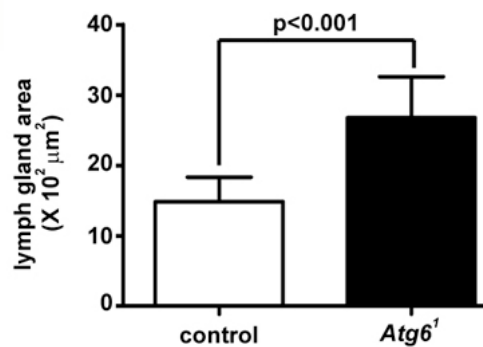
**Fig. S2. Loss of *Atg6* leads to defects in Rab5 localization.** (A) GFP-Rab5 distribution in control fed fat body cells is prominent at the plasma membrane and perinuclear region ( $n=11$ ). (B) Fat body cells expressing *Atg6<sup>IR</sup>* have fewer GFP-Rab5 puncta in the perinuclear region and at the plasma membrane ( $n=13$ ). (C) Quantification of GFP-Rab5 puncta in the perinuclear region (2  $\mu\text{m}$  diameter ring surrounding the nucleus). Two tailed  $t$ -test was used for statistical analysis, and the  $P$ -value relative to control was  $1.48 \times 10^{-8}$ . (D) Graph showing relative *Atg6* mRNA level normalized to *rp49* mRNA in control and *Atg6<sup>IR</sup>*. Error bars represent s.d. Scale bar: 50  $\mu\text{m}$ .



**Fig. S3. *Vps25* and *Vps32* mutant cells possess defects in fluid phase endocytosis.** (A-A'') Texas Red-avidin is endocytosed by control (GFP-positive) fat body cells, but is excluded from *Vps25<sup>n55</sup>* mutant cells (GFP negative) ( $n=10$ ). (B-B'') Surface view of the fat body depicted in A showing accumulation of Texas Red-avidin at the surface of the *Vps25<sup>n55</sup>* mutant cells but not in control cells. (C-C'') Texas Red-avidin is endocytosed by control (GFP-positive) fat body cells, but is excluded from *Vps32<sup>G5</sup>* mutant cells (GFP negative) ( $n=10$ ). (D-D'') Surface view of the fat body depicted in C showing accumulation of Texas Red at the surface of the *Vps32<sup>G5</sup>* cells but not in control cells. Scale bars: 50  $\mu\text{m}$ .



**Fig. S4. Mutations in core autophagy genes do not lead to melanotic mass formation.** (A) Graph showing percentage of larvae/pupae exhibiting melanotic tumor phenotype in *w<sup>1118</sup>* ( $n=145$ ) and *Atg6* ( $n=157$ ), *Atg7* ( $n=113$ ), *Atg8a* ( $n=101$ ), *Atg13* ( $n=99$ ) mutant animals. Note that melanotic tumors in *Atg8a* mutant are observed only after pupariation, and that all homozygous *Atg6* mutant larvae possess melanotic tumors. (B) Parental control *Atg8a<sup>KG07569/+</sup>* pupae do not have melanotic masses ( $n=94$ ). (C) *Atg8a<sup>KG07569/Y</sup>* ( $n=18/101$ ) pupae contains melanotic masses. Scale bar: 250  $\mu\text{m}$ .



**Fig. S5. *Atg6* mutants have enlarged lymph glands.** Quantification of lymph gland area in control ( $n=10$ ) and homozygous *Atg6<sup>1</sup>* ( $n=10$ ) mutants shown in Fig. 7F,G. A two-tailed *t*-test was used for statistical analysis, and the *P*-value relative to control was  $6.3 \times 10^{-5}$ . Error bars represent s.d.