

Electronic Supplementary Information for
Organocatalytic Multicomponent Reaction for the Acquisition of a Potent and Selective Inhibitor of mPTPB, a Virulence Factor of Tuberculosis

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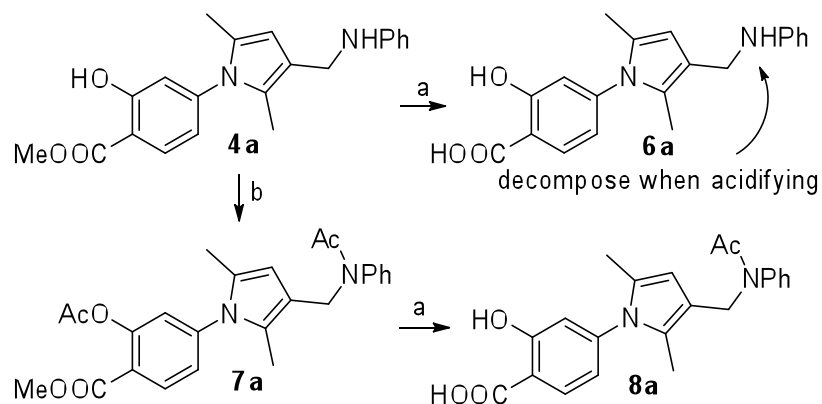
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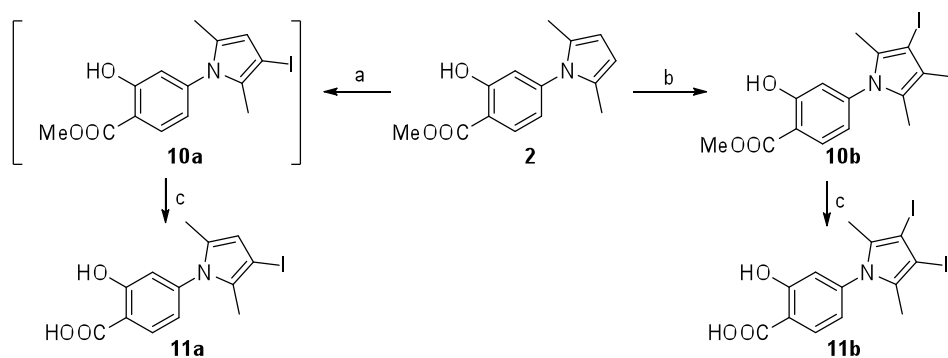
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Table S1. Complete list of optimization of reaction conditions

entry	solvent	catalyst	time	ratio of 4a/5/2, %	yield of 4a %	yield of 5 %
1	DMSO	HOAc (20 mol%)	24 h	no reaction		
2	DMF	HOAc (20 mol%)	24 h	no reaction		
3	THF	HOAc (20 mol%)	24 h	no reaction		
4	MeCN	HOAc (20mol%)	24 h	13/8/79		
5	Et ₂ O	HOAc (20 mol%)	24 h	13/6/81		
6	toluene	HOAc (20 mol%)	24 h	no reaction		
7	MeOH	HOAc (20 mol%)	24 h	9/8/91		
8	CH ₂ Cl ₂	HOAc (20 mol%)	24 h	52//22/26		
9	CHCl ₃	HOAc (20 mol%)	24 h	48//15/37		
10	CH ₂ Cl ₂	4-F-PhCO ₂ H(20 mol%)	24 h	47/5/48		
11	CH ₂ Cl ₂	Proline (20 mol%)	24 h	7/0/93		
12	CH ₂ Cl ₂	TFA (20 mol%)	24 h	decomposition		
13	CH ₂ Cl ₂	2 M HCl (20 mol%)	24 h	26/4/70		
14	CH ₂ Cl ₂	4-Me-PhCO ₂ H(20 mol%)	24 h	28/13/59		
15	CH ₂ Cl ₂	3-F-5-CF ₃ -PhCO ₂ H(20 mol%)	24 h	50/25/25		
16	CH ₂ Cl ₂	3,5-(CF ₃) ₂ -PhCO ₂ H(20 mol%)	24 h	46/17/37		
17	CH ₂ Cl ₂	MeOCH ₂ CO ₂ H (20 mol%)	24 h	45/36/19		
18	CH ₂ Cl ₂	PhCH ₂ CO ₂ H (20 mol%)	24 h	23/10/67		
19	CH ₂ Cl ₂	Cyc.C ₆ H ₁₁ CO ₂ H (20 mol%)	24 h	13/10/77		
20	CH ₂ Cl ₂	PTSA (20 mol%)	24 h	21/5/74		
21	CH ₂ Cl ₂	(3,5-(CF ₃) ₂ -PhNH) ₂ CS (20 mol%)	24 h	no reaction		
22	CH ₂ Cl ₂	diphenic acid (20 mol%)	24 h	47/23/30		
23	CH ₂ Cl ₂	HOAc (100 mol%)	24 h	59//17/24		
24	CHCl ₃	HOAc (100 mol%)	24 h	44/11/45		
25	CH ₂ Cl ₂	HOAc (100 mol%)	24 h	45/9/46	85	
26	CH ₂ Cl ₂	MeOCH ₂ CO ₂ H (100 mol%)	48 h	0/100/0		75

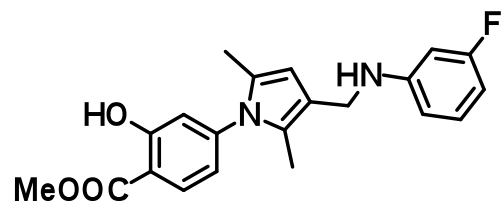
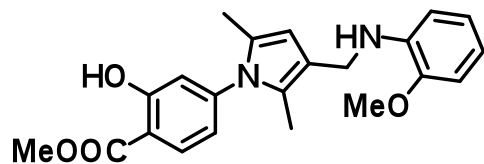
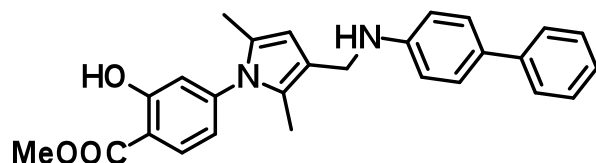
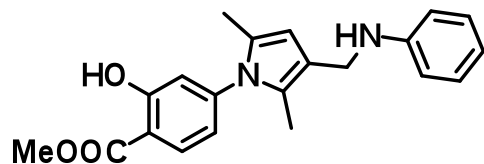
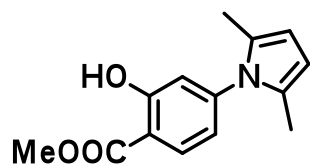


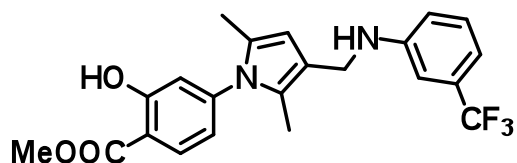
Scheme S1. Strategy to obtain stable hydrolyzed compound **8a**. a. 10% LiOH, MeOH, rt, 2 h, 95% yield. b. Ac₂O (3 equiv.), DMAP (0.1 equiv.), CH₂Cl₂, rt, 2 h, 95% yield.



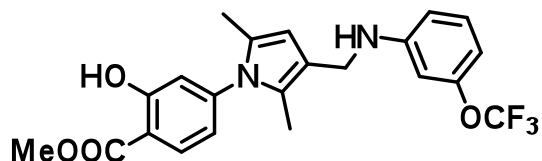
Scheme S2. Synthesis of **10a**, **10b**, **11a**, **11b**. a. I₂ (2.2 equiv.), NaHCO₃ (3 equiv.), ether, rt, 24 h, 60% yield. b. ICl (2.2 equiv.), NaHCO₃ (3 equiv.), ether, rt, 24 h, 70% yield. c. 10% LiOH, MeOH, rt, 2 h, 95% yield.

Characterizations of compounds **2**, **4a-4l**, **5**, **10a**, **10b**

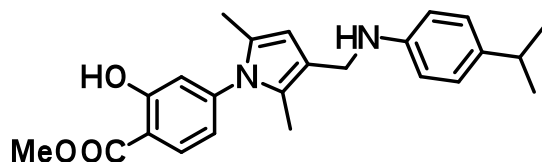




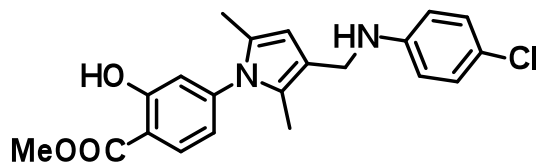
4f. ^1H NMR (500 MHz, CDCl_3) 10.93 (s, 1H), 7.95 (d, $J = 8.5$ Hz, 1H), 7.27 (d, $J = 5.0$ Hz, 1H), 6.94 (d, $J = 7.7$ Hz, 1H), 6.87-6.86 (m, 2H), 6.81-6.79 (m, 1H), 6.76-6.674 (m, 1H), 5.98 (s, 1H), 4.10 (s, 2H), 4.00 (s, 3H), 2.07 (s, 3H), 2.06 (s, 3H). ESI-MS Calcd. for $\text{C}_{22}\text{H}_{20}\text{F}_3\text{N}_2\text{O}_3$ (M-H^+): m/z 417.1432; found 417.1442.



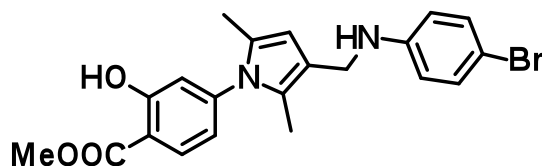
4g. ^1H NMR (500 MHz, CDCl_3) 10.93 (s, 1H), 7.95 (d, $J = 8.4$ Hz, 1H), 7.06 (d, $J = 8.5$ Hz, 2H), 6.86 (d, $J = 1.9$ Hz, 1H), 6.76-6.73 (m, 1H), 6.63-6.62 (m, 2H), 5.98 (s, 1H), 4.06 (s, 2H), 4.00 (s, 3H), 2.07 (s, 3H), 2.06 (s, 3H). ^{13}C NMR (125 MHz, CDCl_3) δ 170.1, 162.1, 147.4, 145.2, 140.2, 130.7, 128.1, 125.9, 122.4, 119.2, 117.1, 116.8, 112.8, 111.8, 107.5, 52.5, 40.8, 12.8, 10.7. ESI-MS Calcd. for $\text{C}_{22}\text{H}_{20}\text{F}_3\text{N}_2\text{O}_4$ (M-H^+): m/z 434.1465; found 434.1392.



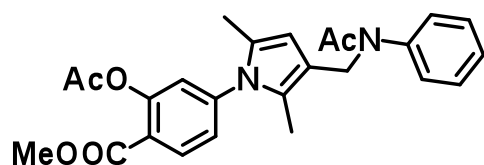
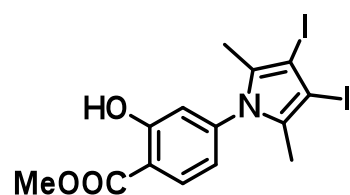
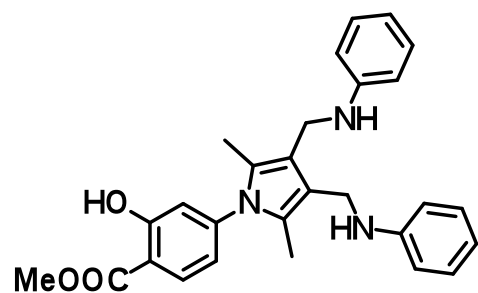
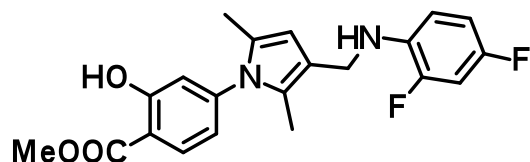
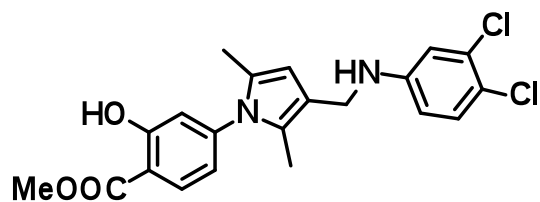
4h. ^1H NMR (500 MHz, CDCl_3) 10.92 (s, 1H), 7.94 (d, $J = 8.4$ Hz, 1H), 7.09-7.08 (m, 2H), 6.86 (d, $J = 2.0$ Hz, 1H), 6.76-6.74 (m, 1H), 6.66-6.64 (m, 2H), 5.99 (s, 1H), 4.07 (s, 2H), 4.00 (s, 3H), 2.83 (p, $J = 6.9$ Hz, 1H), 2.07 (s, 3H), 2.05 (s, 3H), 1.24 (s, 3H), 1.22 (s, 3H). ^{13}C NMR (125 MHz, CDCl_3) δ 170.1, 162.1, 146.7, 145.3, 137.7, 130.6, 127.9, 127.0, 125.8, 119.2, 117.4, 117.1, 112.7, 111.6, 107.6, 52.5, 40.9, 33.2, 24.3, 12.8, 10.7. ESI-MS Calcd. for $\text{C}_{24}\text{H}_{27}\text{N}_2\text{O}_3$ (M-H^+): m/z 391.2109; found 391.2036.



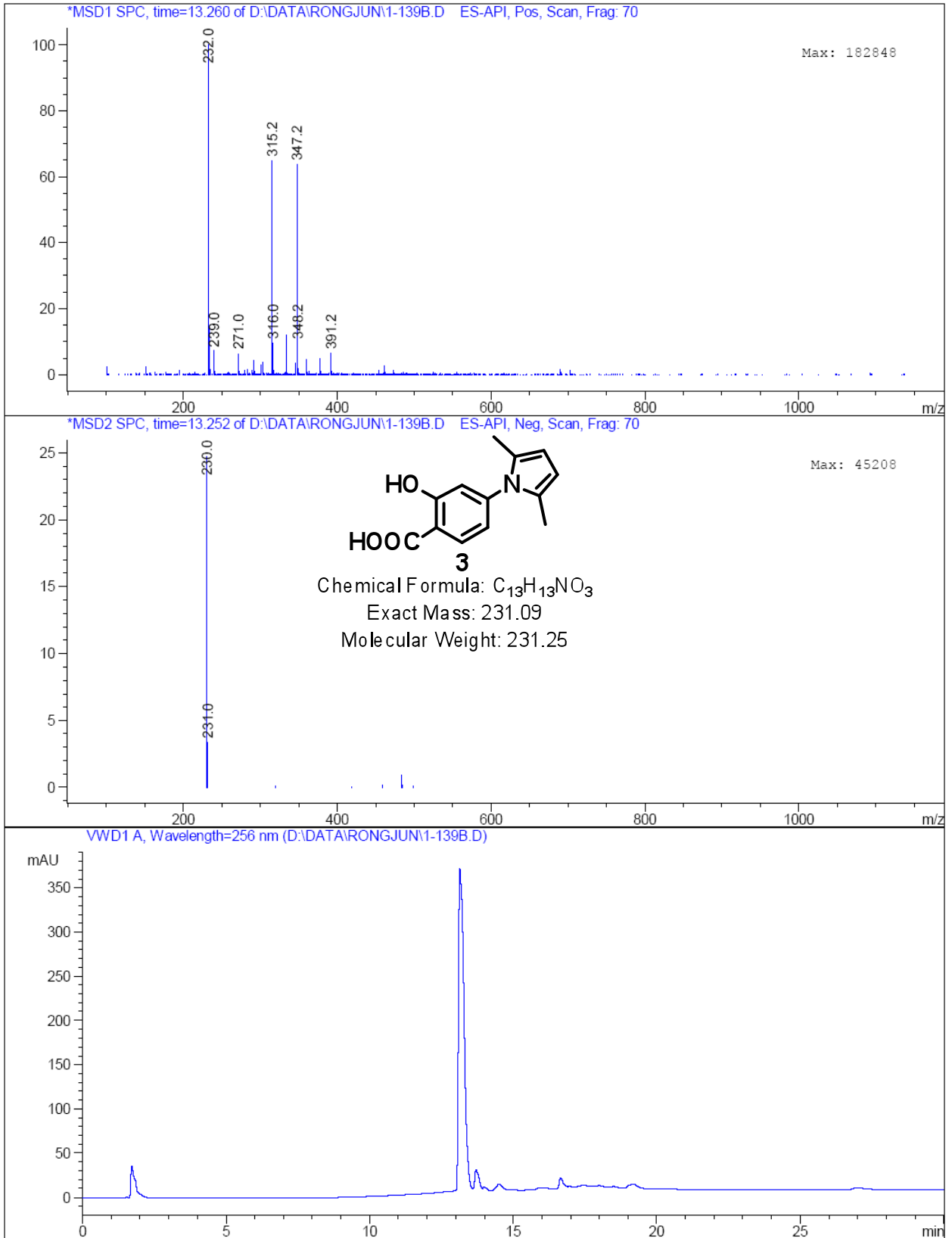
4i. ^1H NMR (500 MHz, CDCl_3) 10.93 (s, 1H), 7.94 (d, $J = 8.4$ Hz, 1H), 7.14 (dd, $J = 6.8, 2.1$ Hz, 2H), 6.85 (d, $J = 1.9$ Hz, 1H), 6.74 (dd, $J = 8.4, 2.0$ Hz, 1H), 6.60-6.58 (m, 2H), 5.97 (s, 1H), 4.05 (s, 2H), 4.00 (s, 3H), 2.07 (s, 3H), 2.05 (s, 3H). ^{13}C NMR (125 MHz, CDCl_3) δ 170.1, 162.1, 147.1, 145.2, 130.6, 129.0, 128.1, 125.8, 121.6, 119.2, 117.1, 116.8, 113.7, 111.7, 107.4, 52.5, 40.7, 12.8, 10.7. ESI-MS Calcd. for $\text{C}_{21}\text{H}_{21}\text{ClN}_2\text{O}_3$ (M-H^+): m/z 383.1168; found 383.1168.

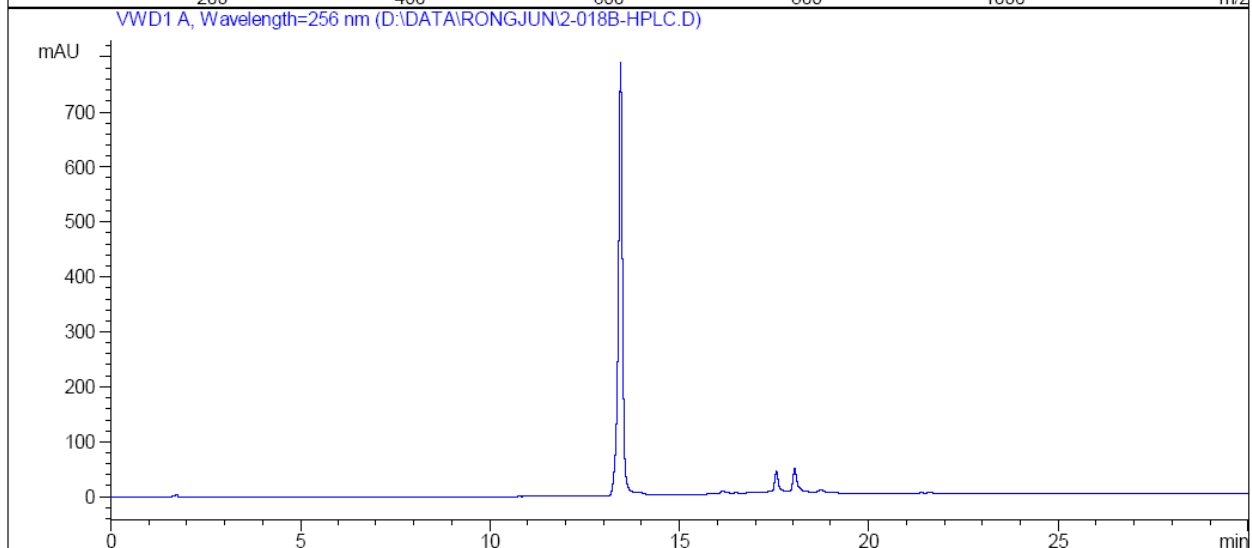
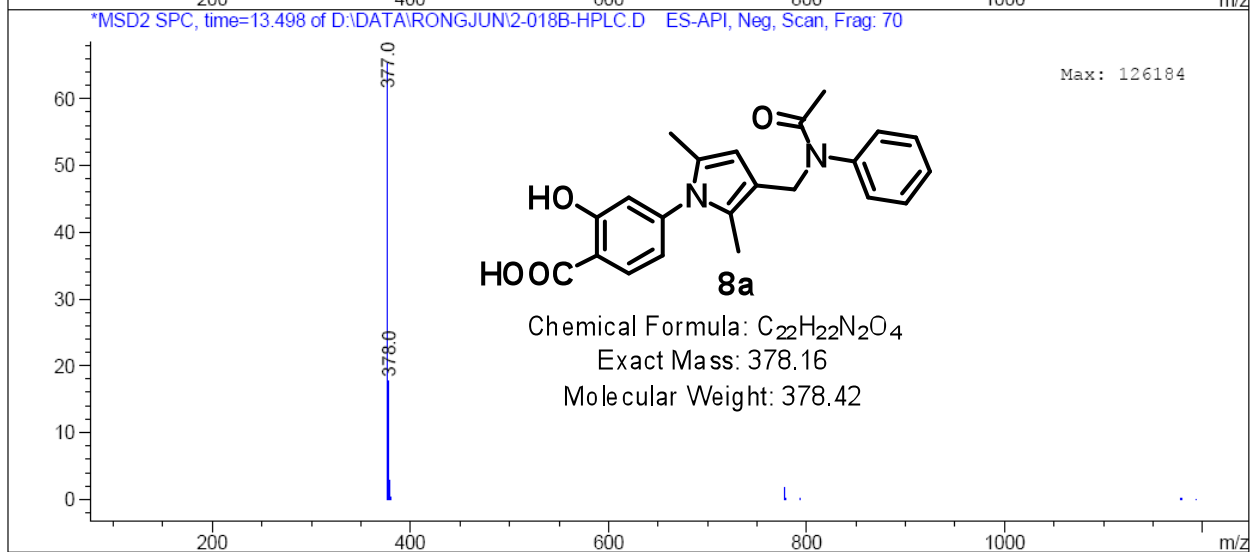
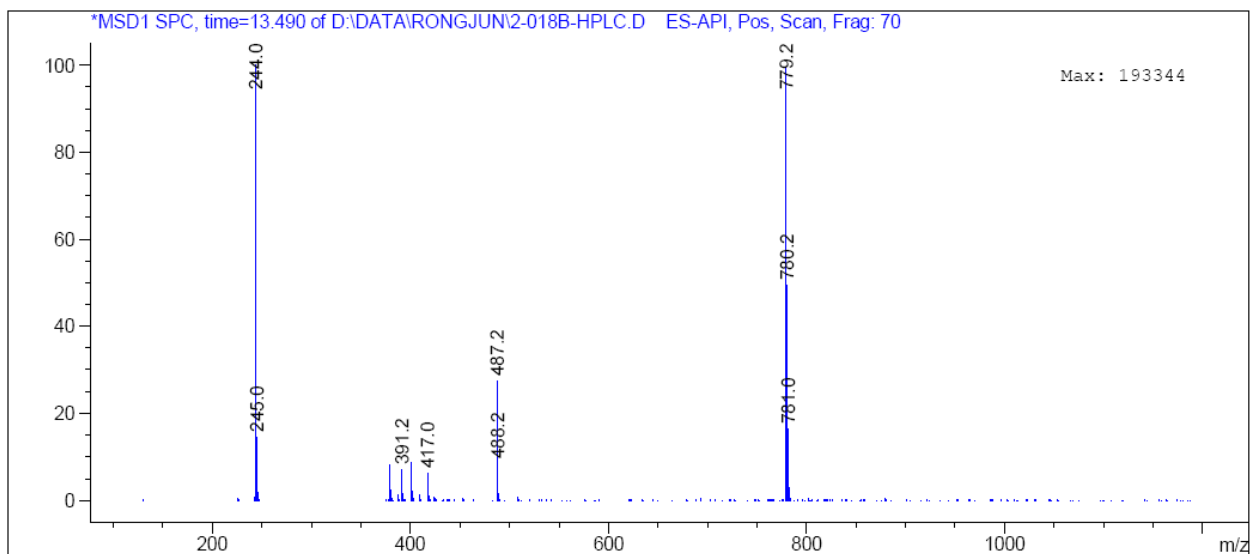


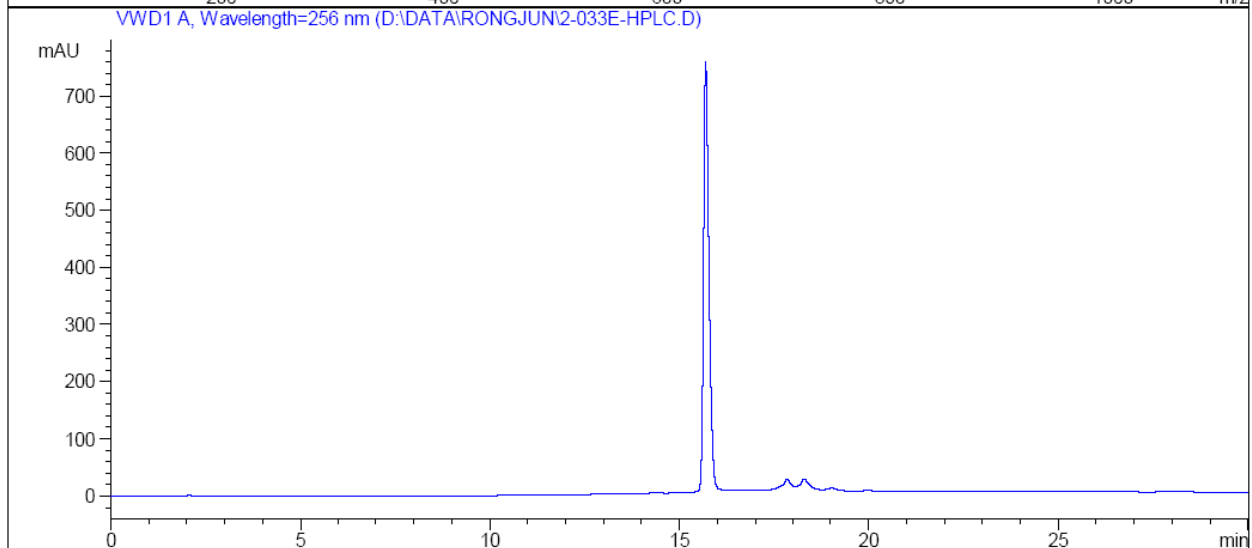
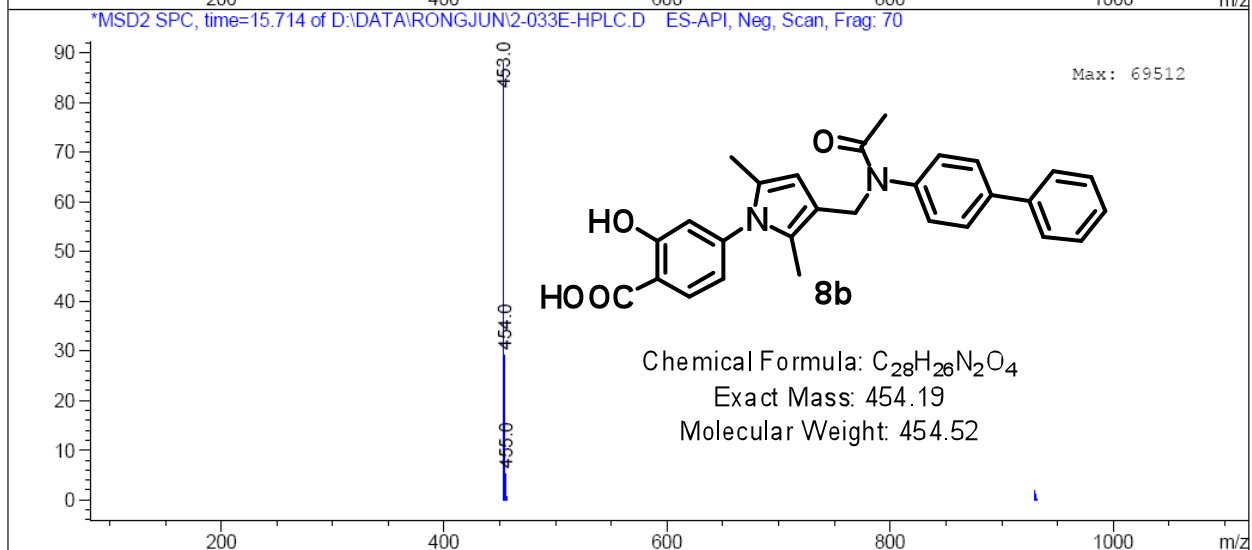
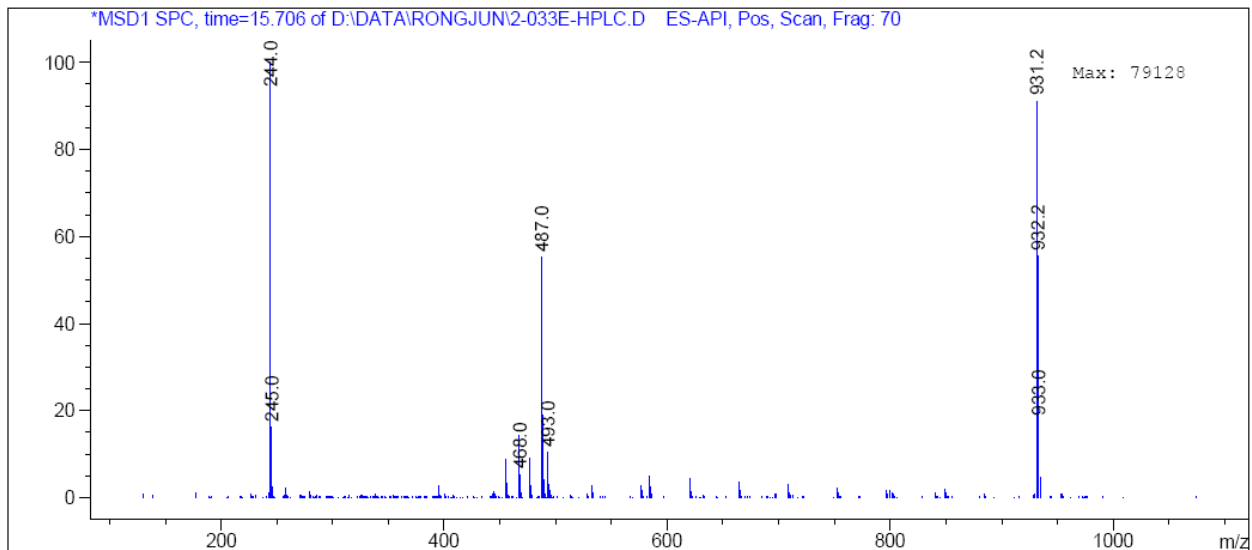
4j. ^1H NMR (500 MHz, CDCl_3) 10.92 (s, 1H), 7.93 (d, $J = 8.4$ Hz, 1H), 7.26 (d, $J = 8.8$ Hz, 2H), 6.84 (d, $J = 2.0$ Hz, 1H), 6.73 (dd, $J = 8.4, 2.0$ Hz, 1H), 6.54 (d, $J = 8.8$ Hz, 2H), 5.96 (s, 1H), 4.03 (s, 2H), 3.99 (s, 3H), 2.06 (s, 3H), 2.04 (s, 3H). ^{13}C NMR (125 MHz, CDCl_3) δ 170.0, 162.1, 147.5, 145.2, 131.8, 130.6, 128.1, 125.8, 119.2, 117.1, 116.8, 114.2, 111.7, 108.6, 107.4, 52.5, 40.6, 12.8, 10.7. ESI-MS Calcd. for $\text{C}_{21}\text{H}_{21}\text{BrN}_2\text{O}_3$ (M-H^+): m/z 427.0663; found 427.0660.

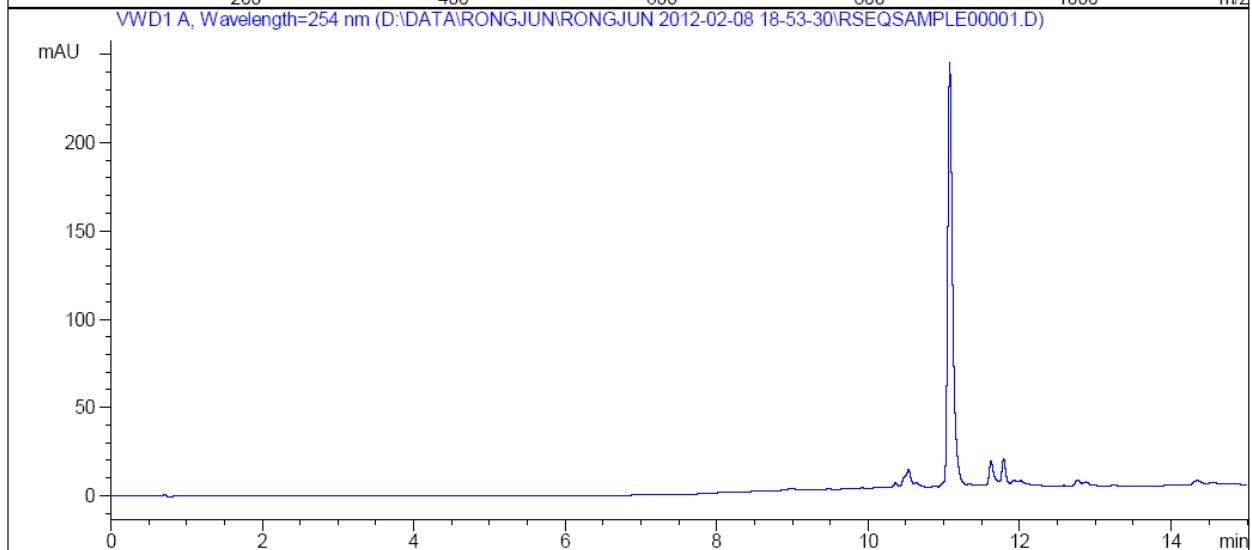
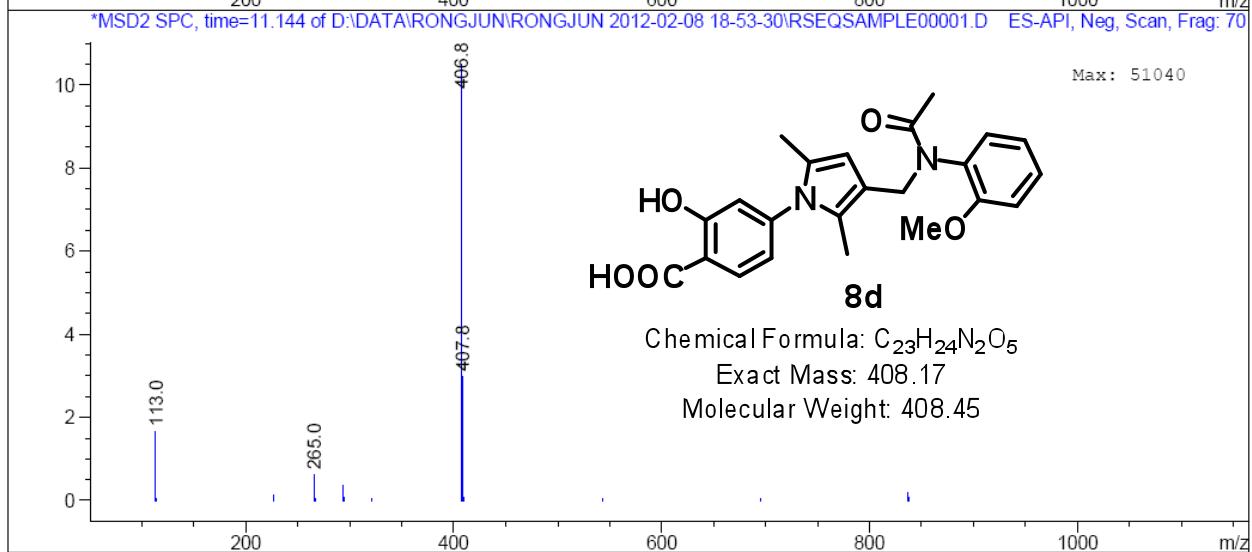
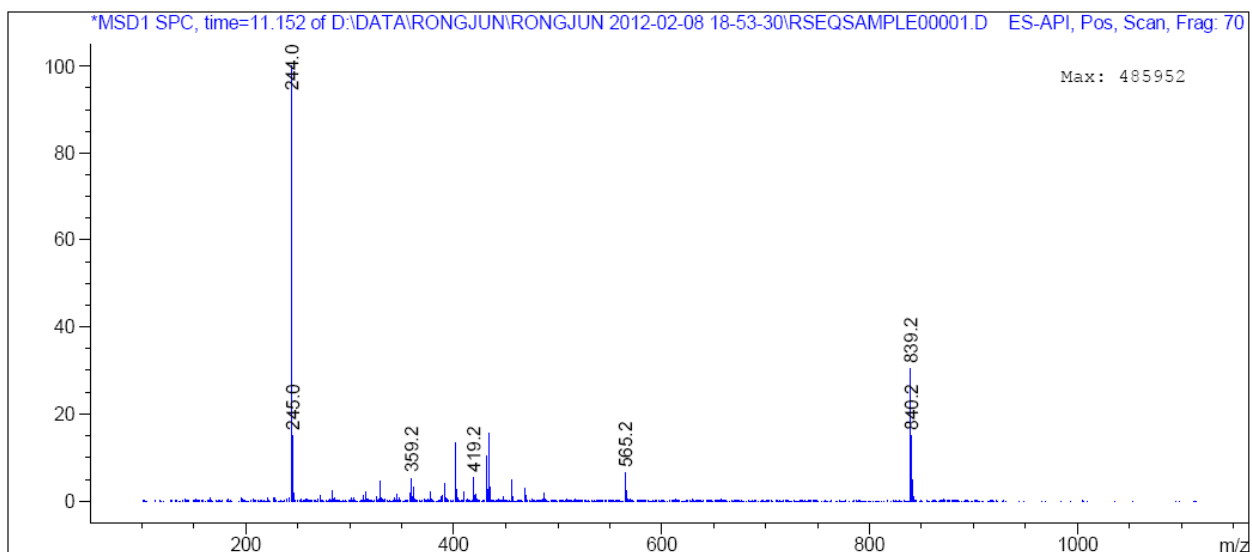


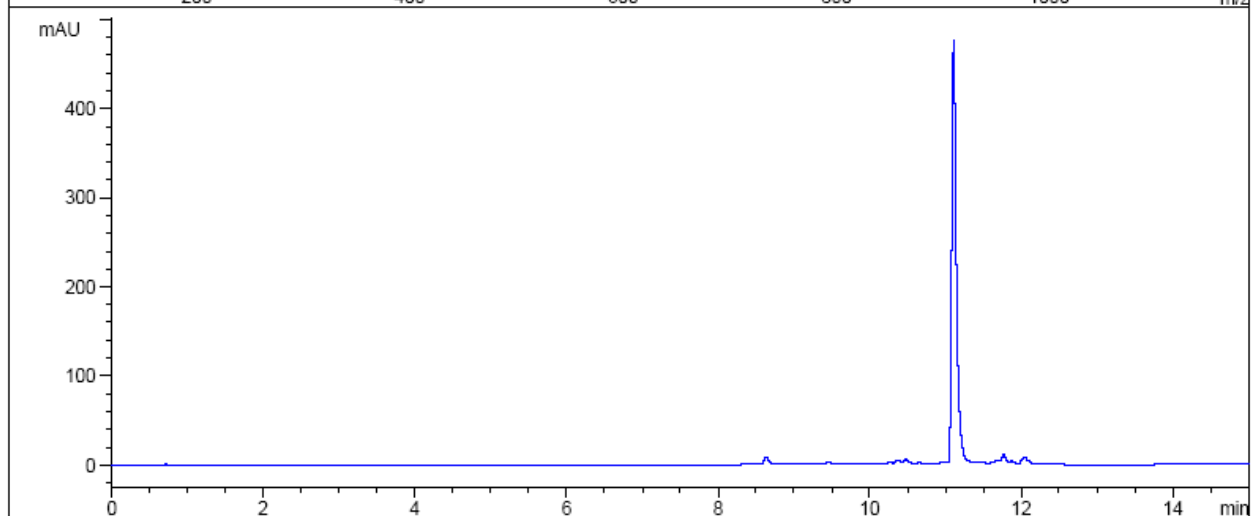
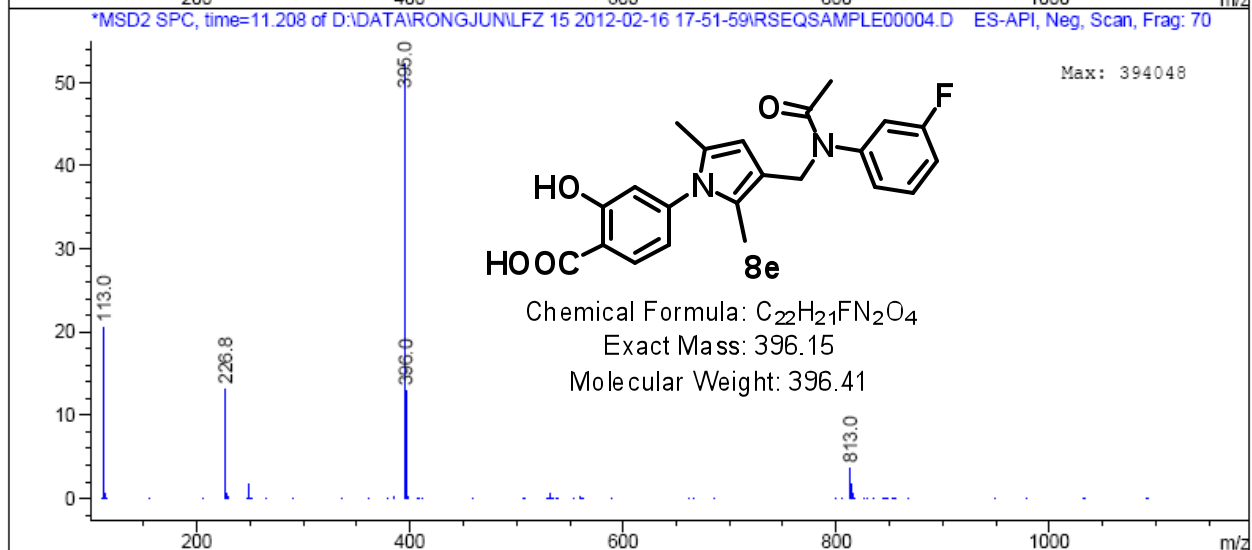
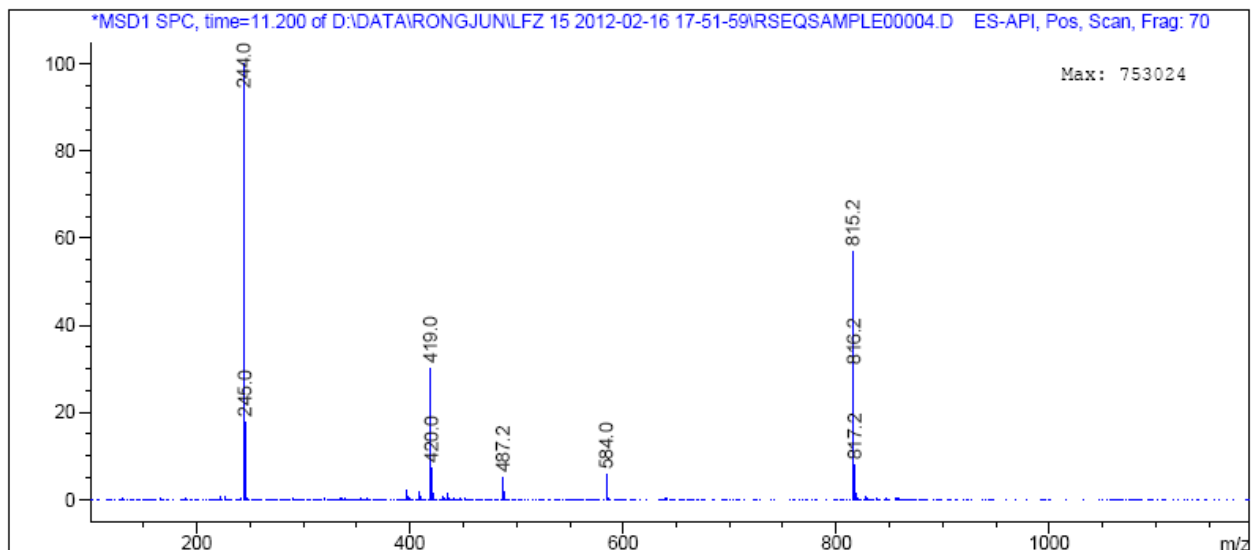
LC-MS studies of hydrolyzed compounds **3**, **8a-8l**, **9**, **11a**, **11b**

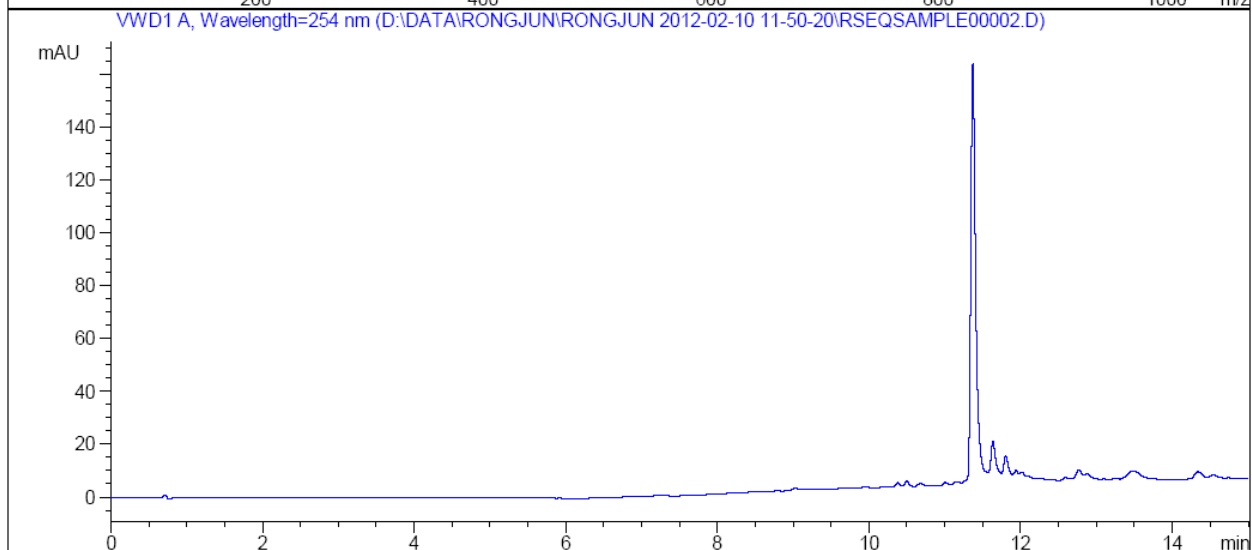
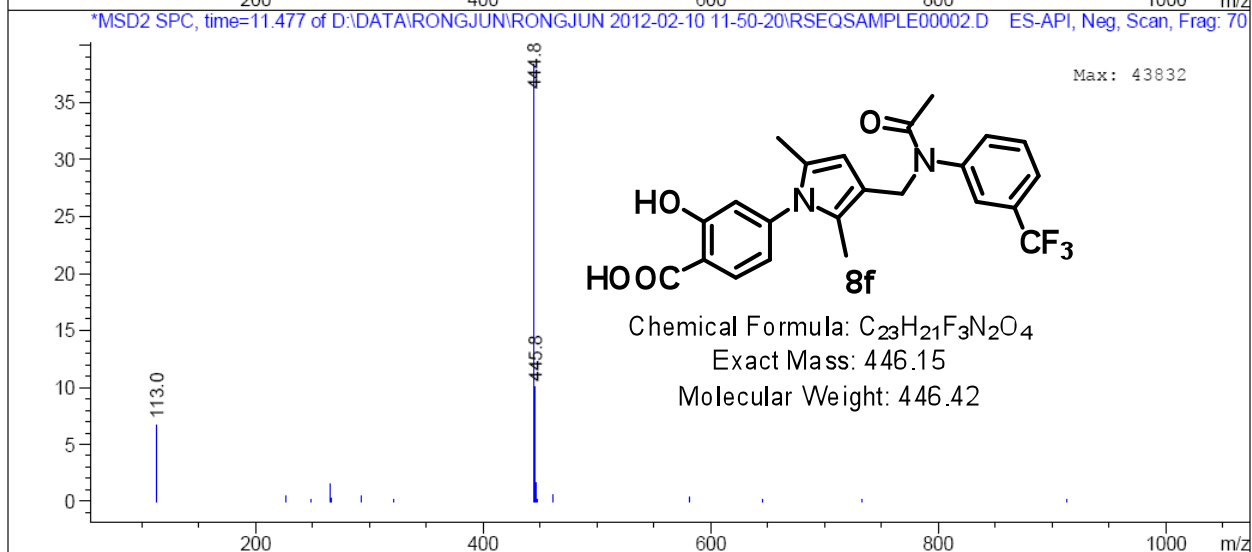
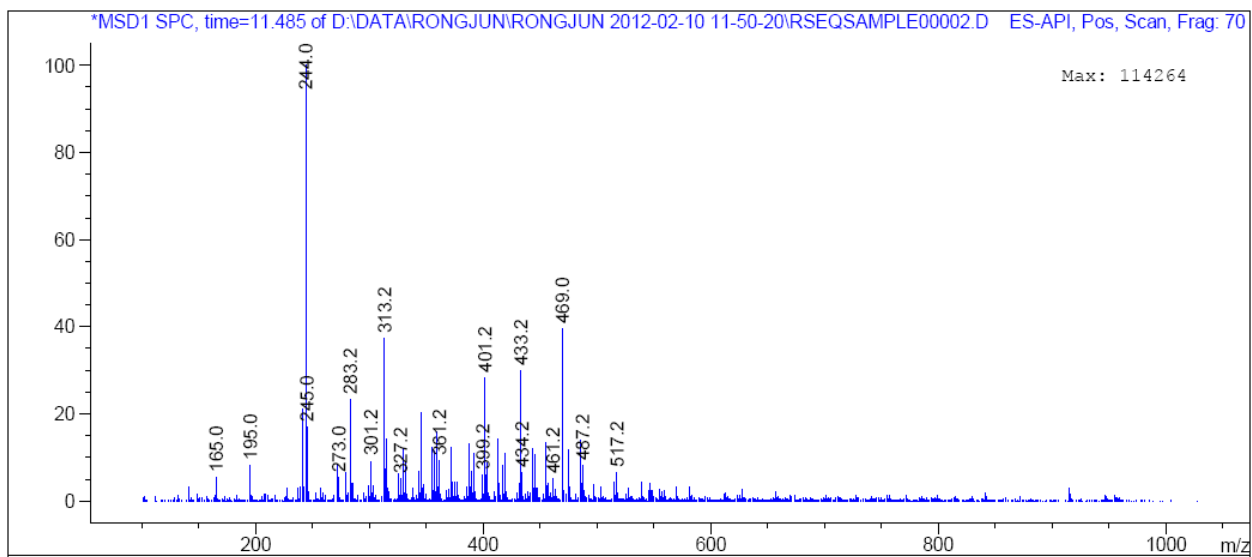


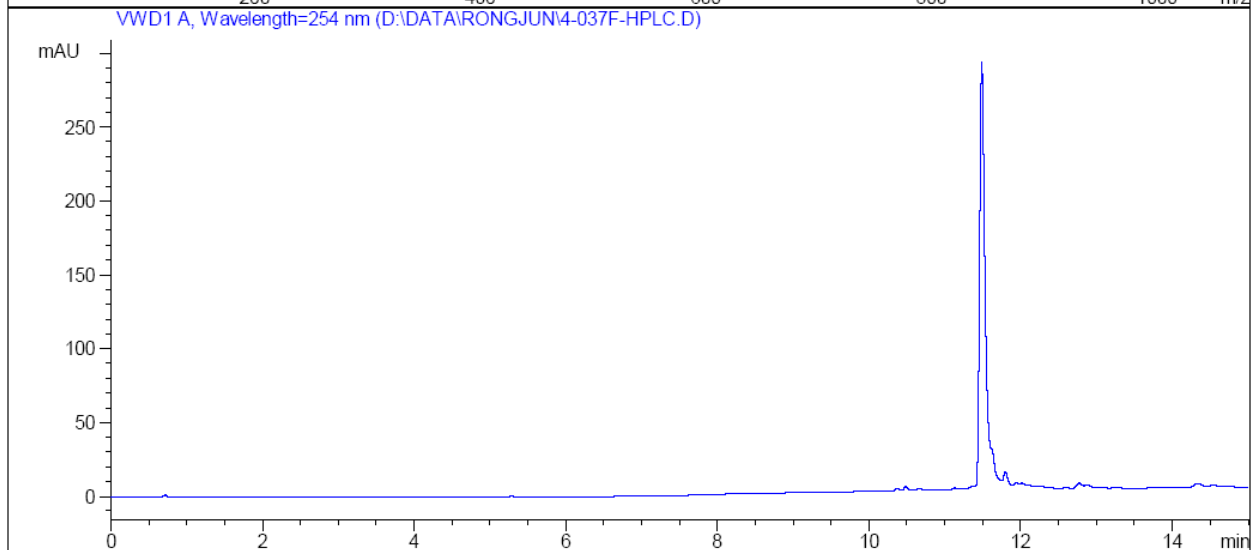
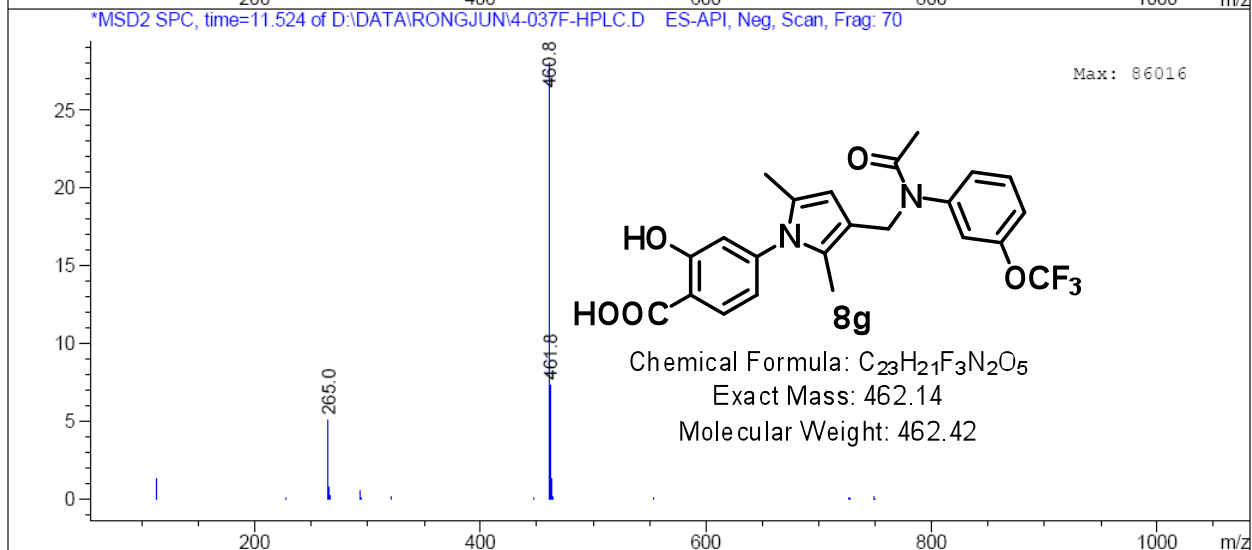
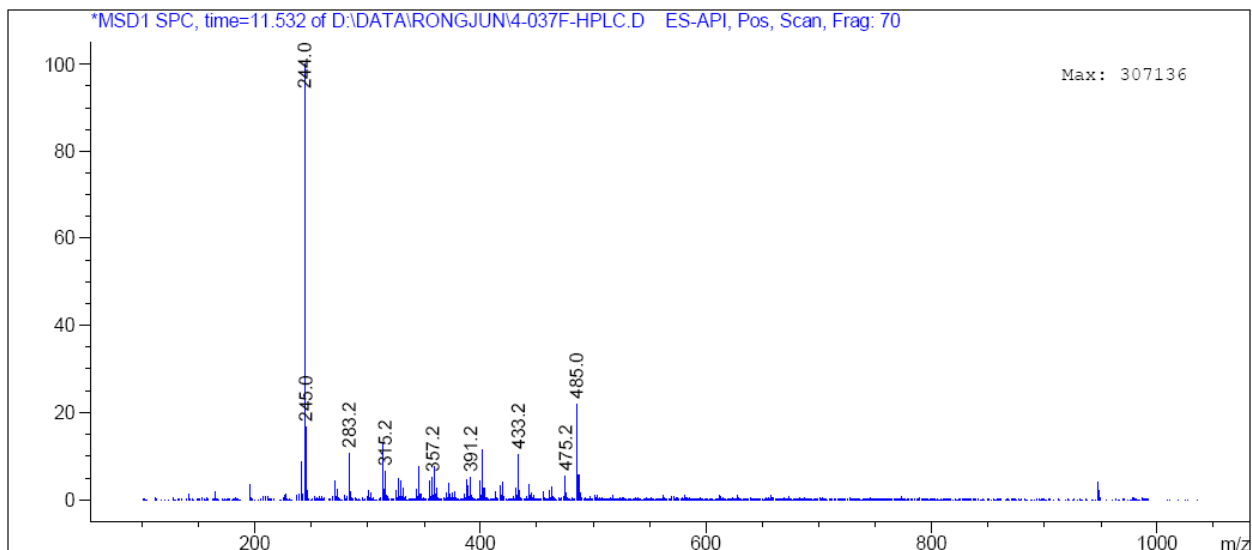


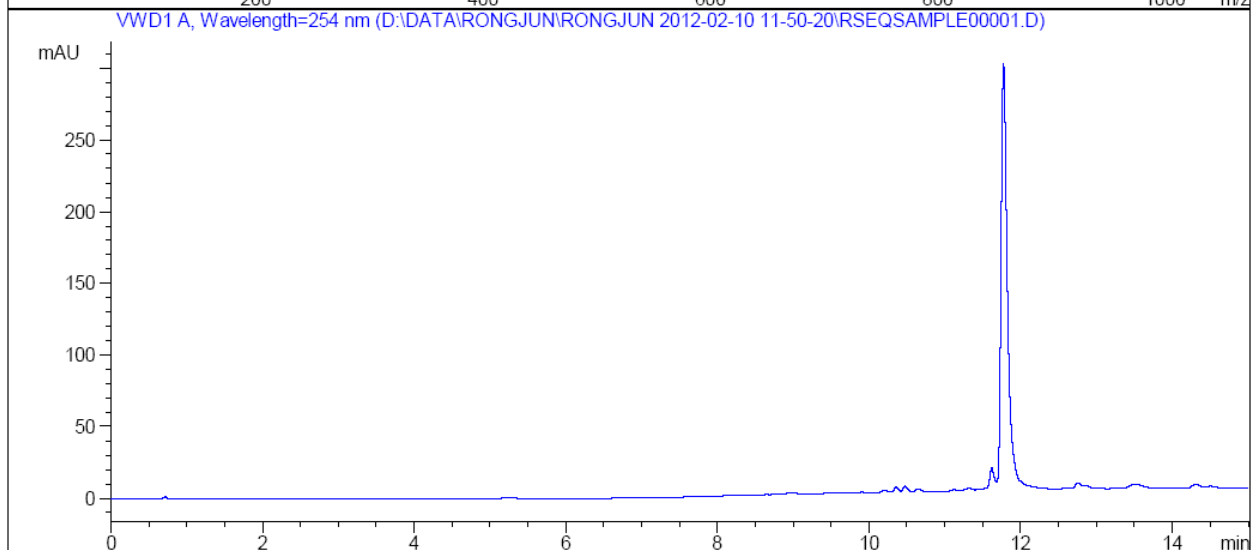
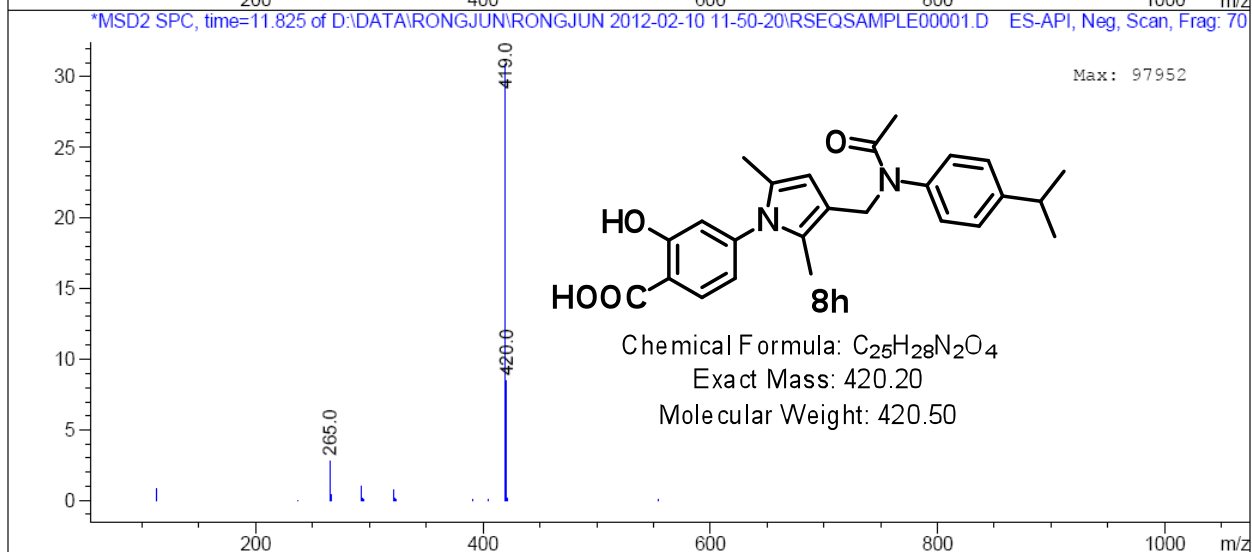
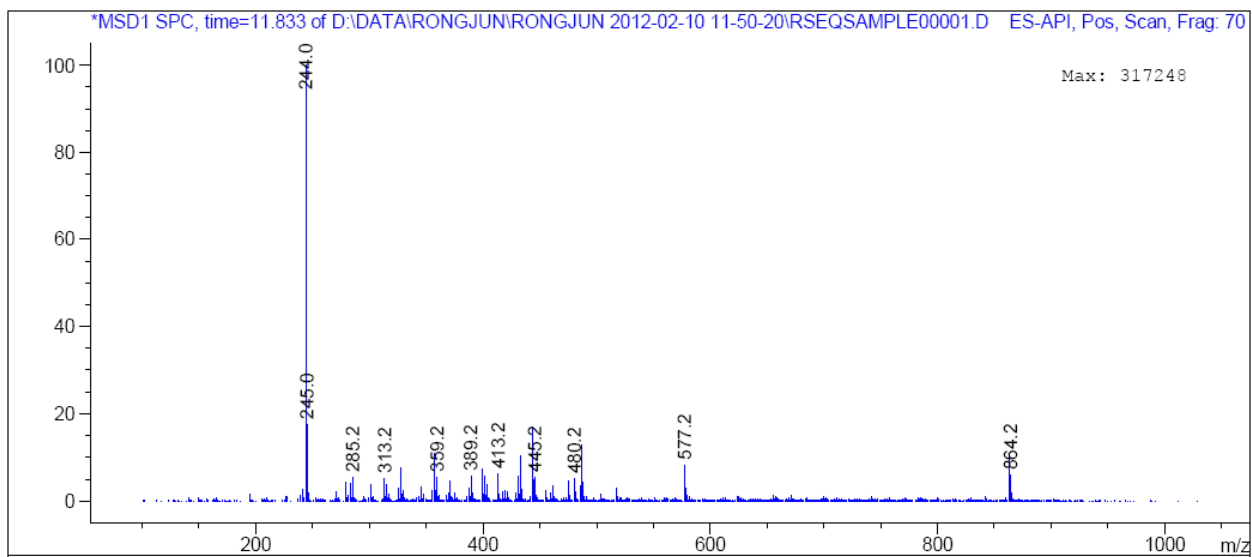


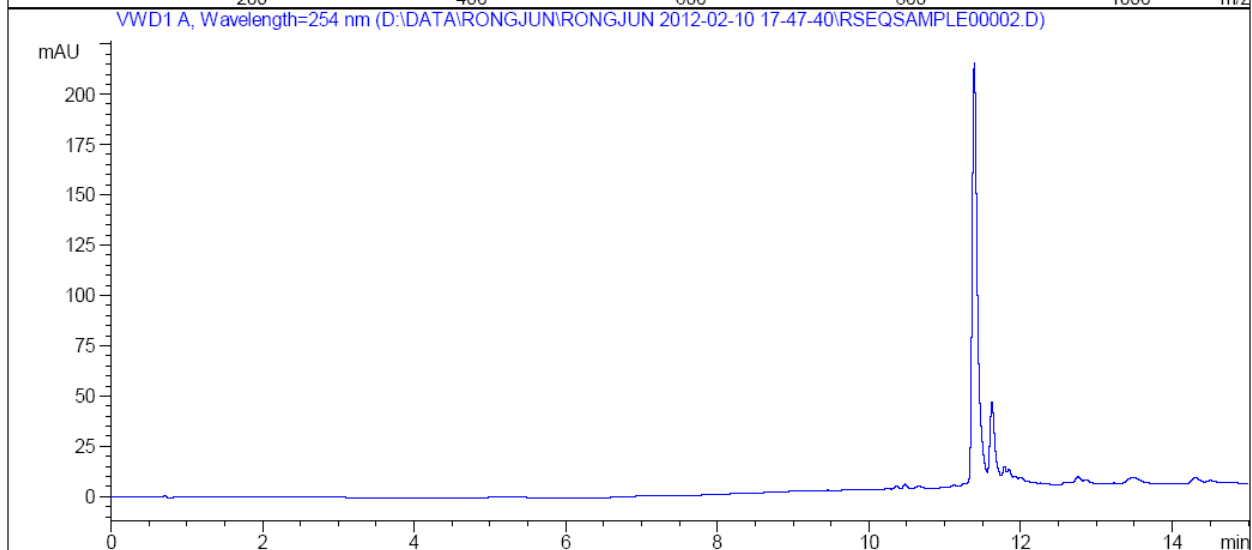
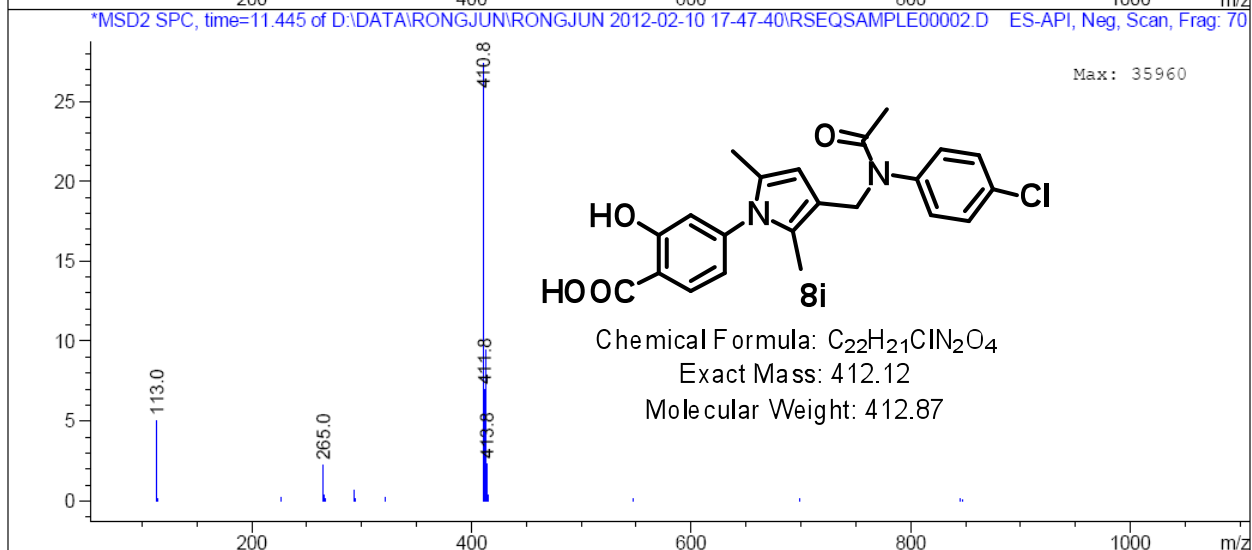
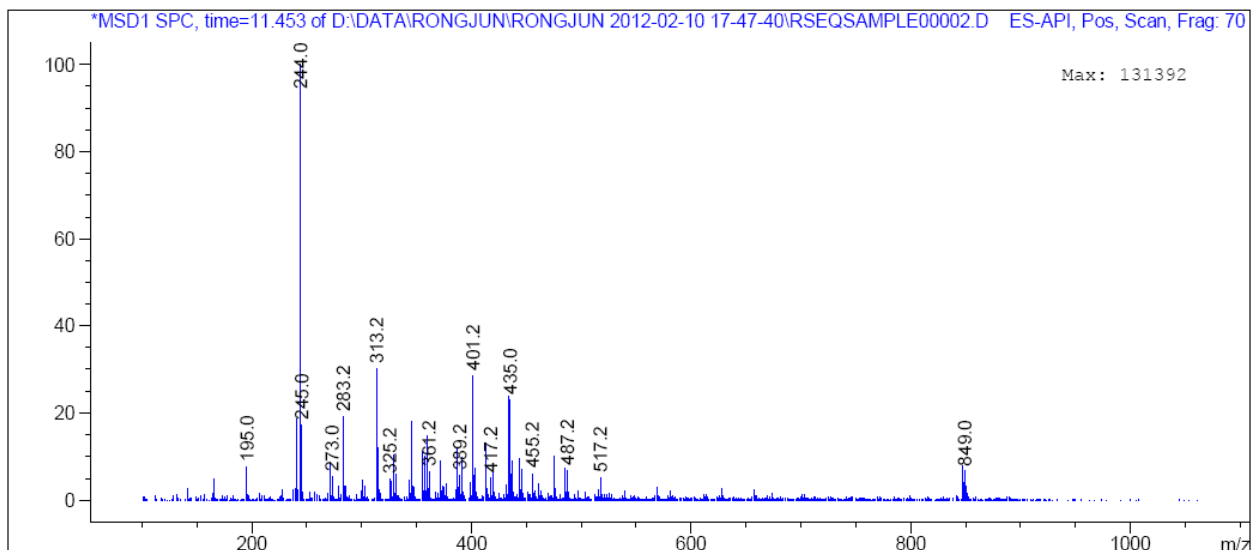


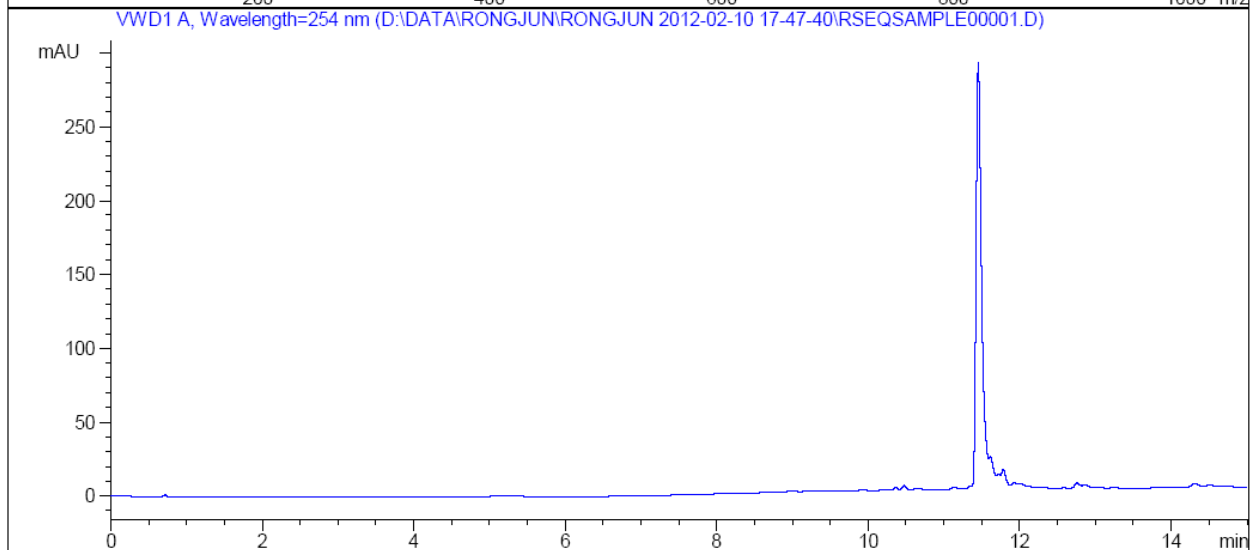
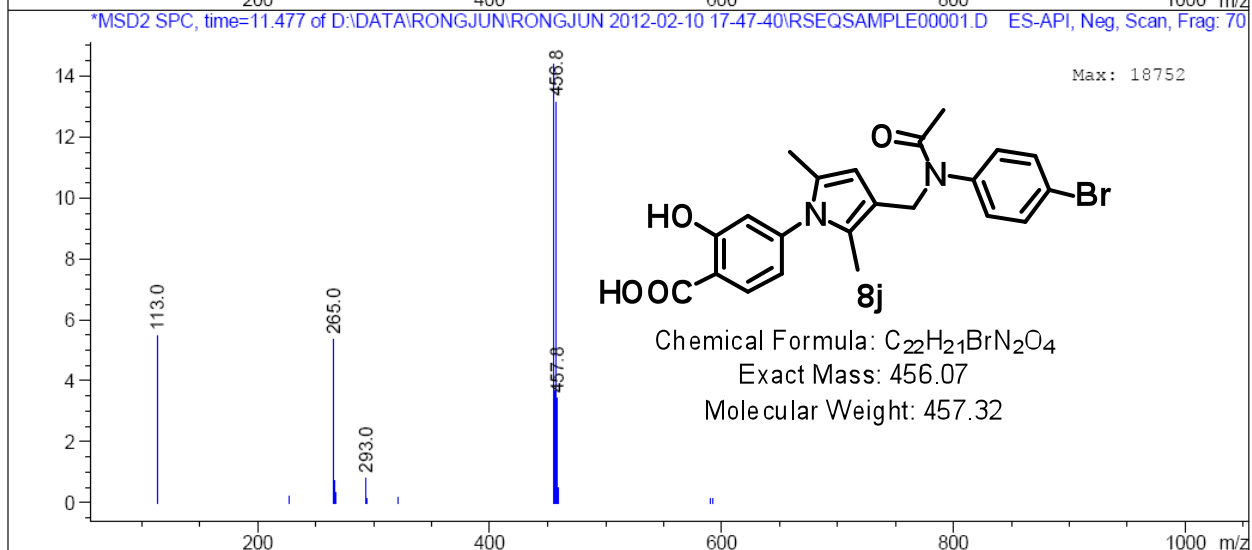
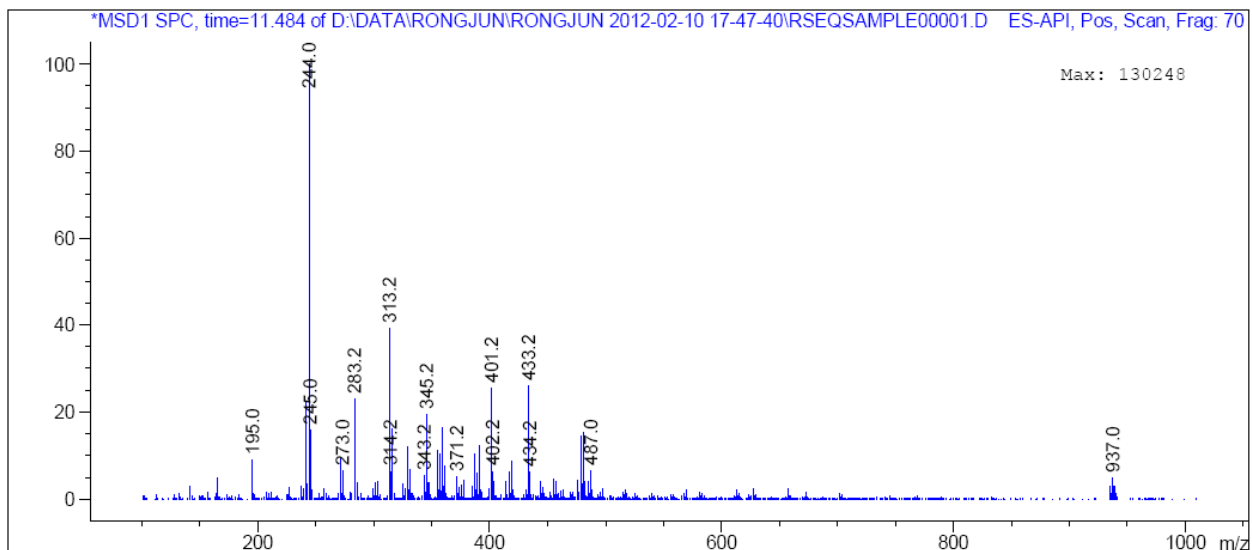


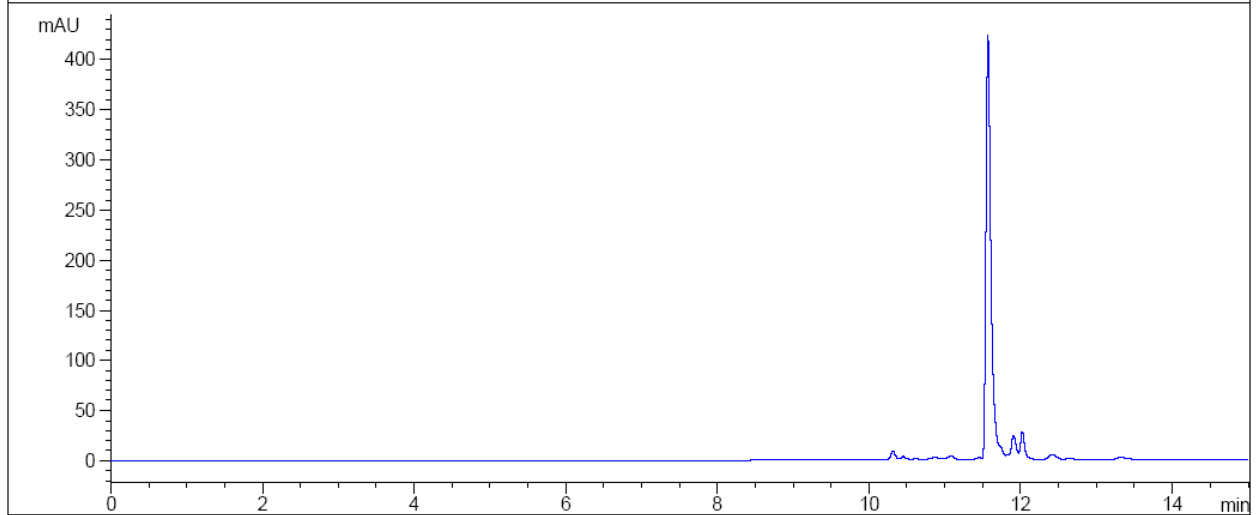
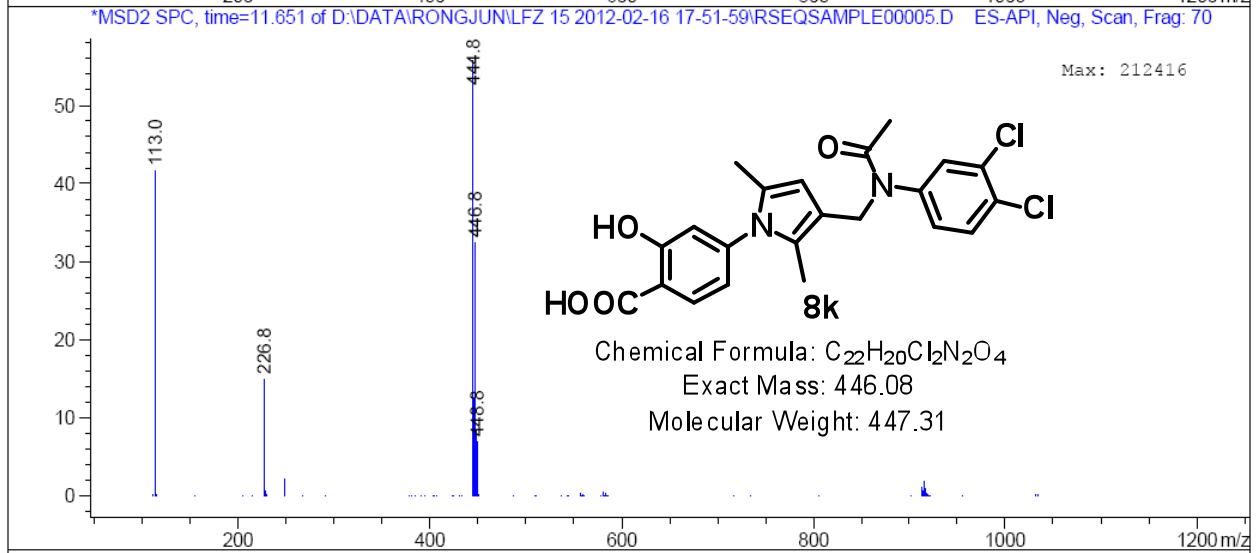
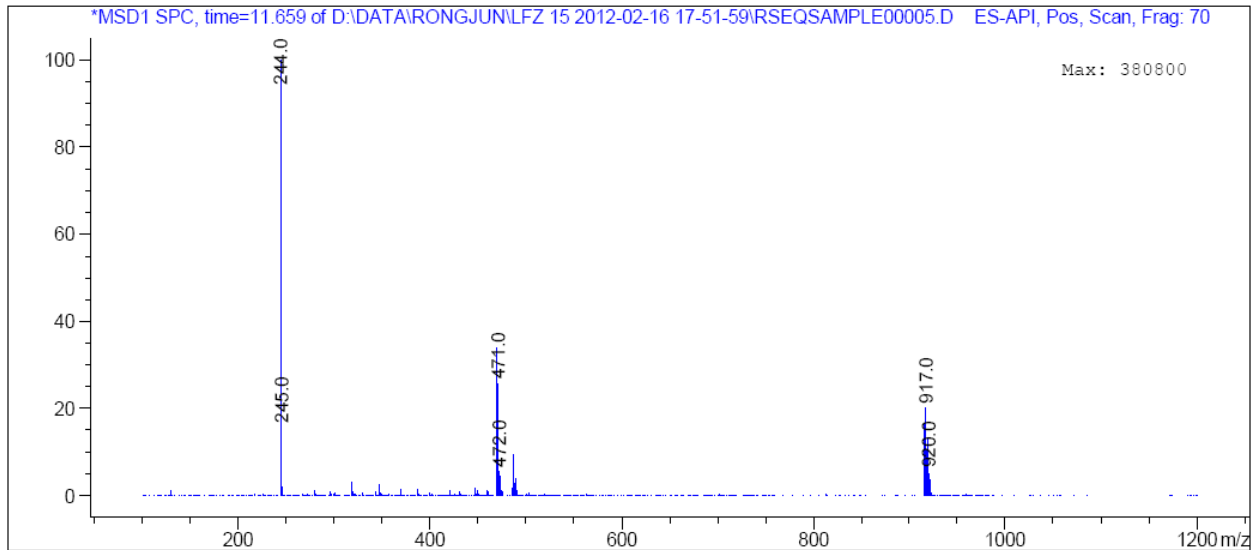


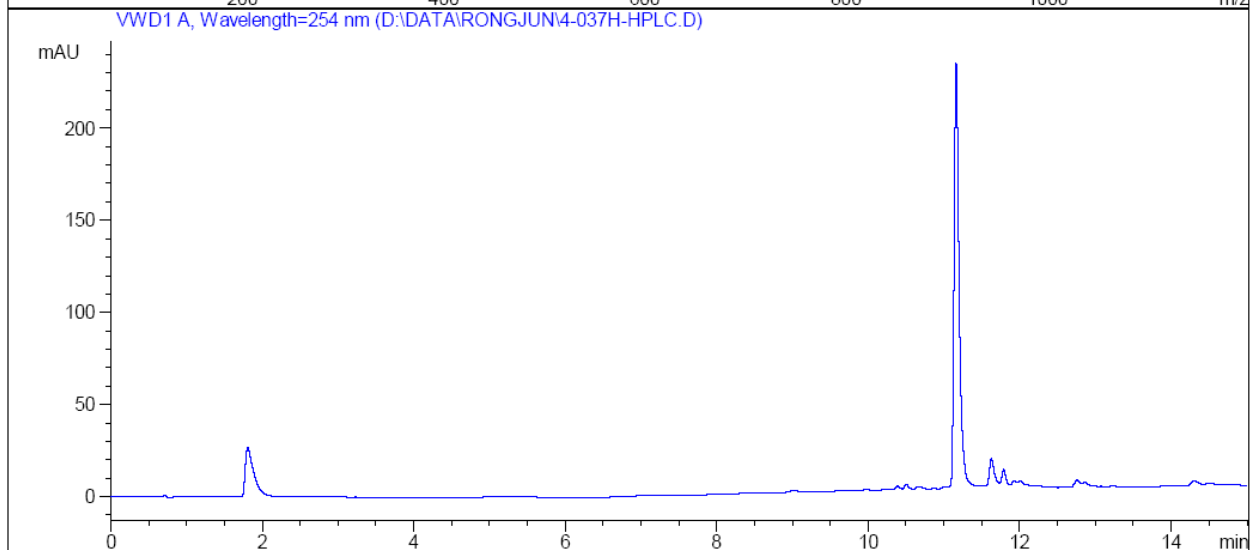
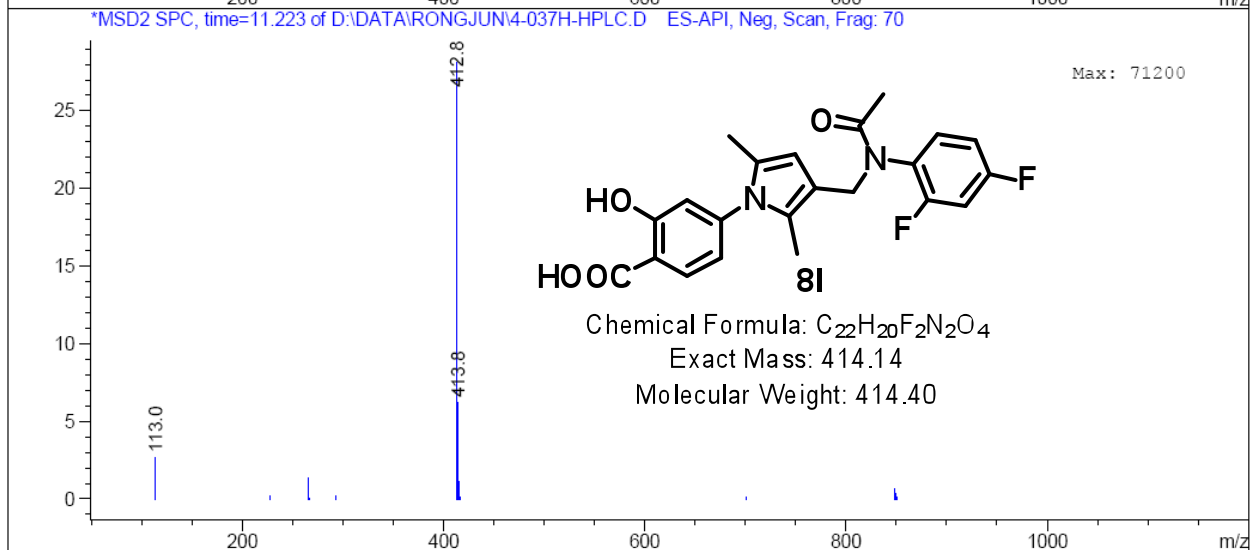
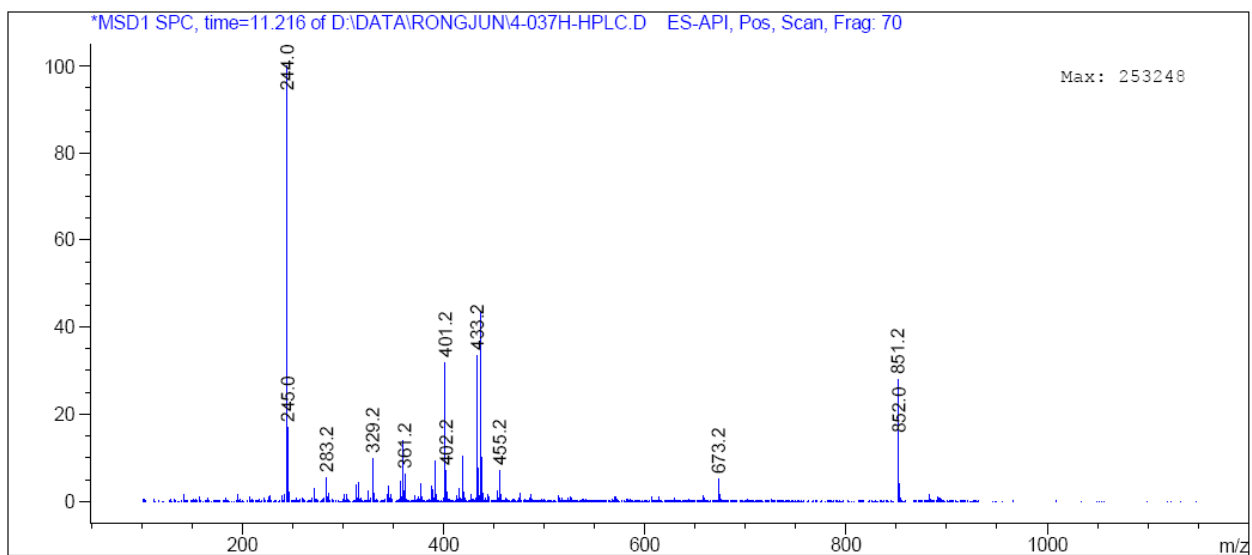


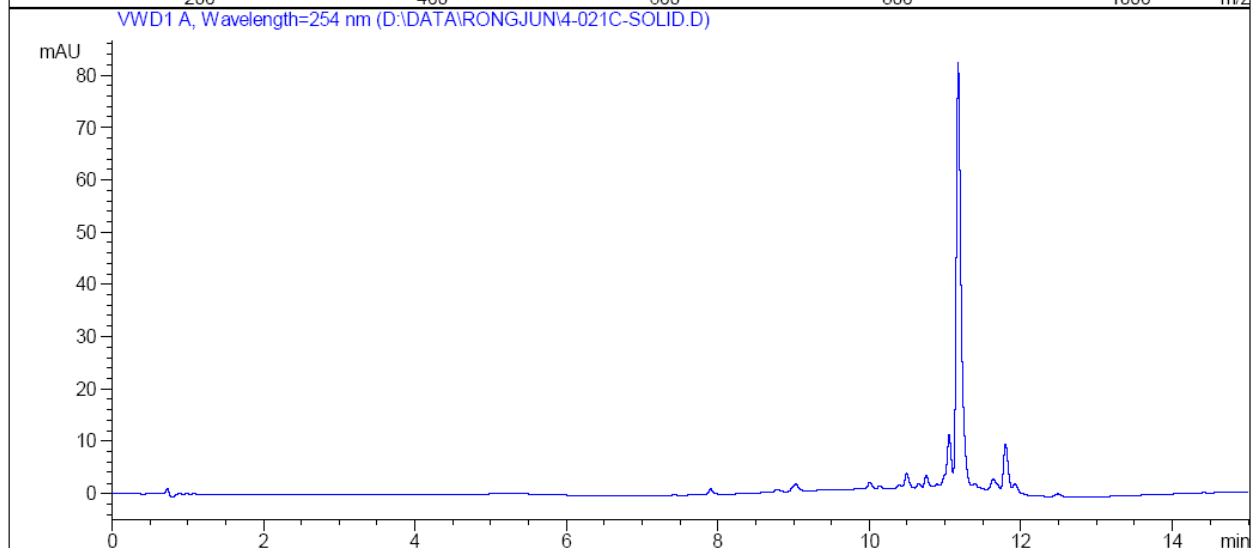
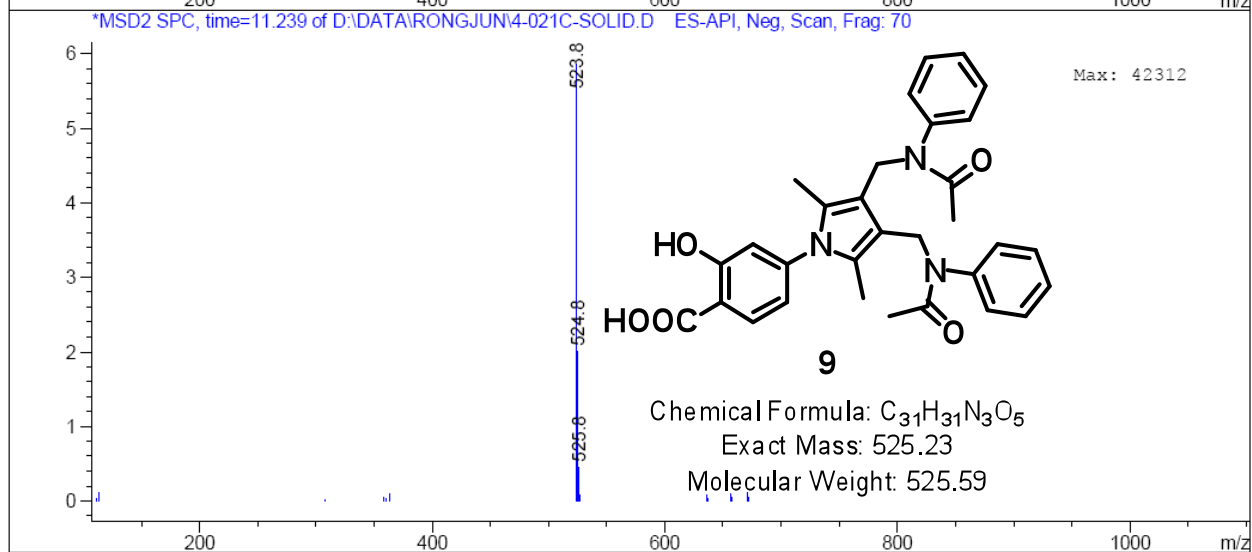
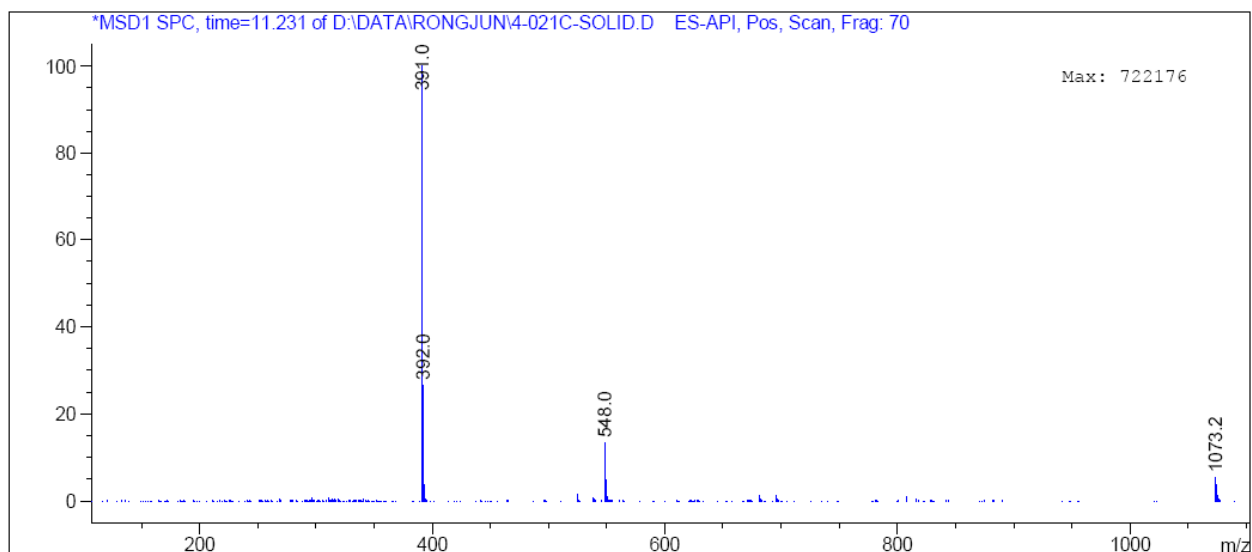


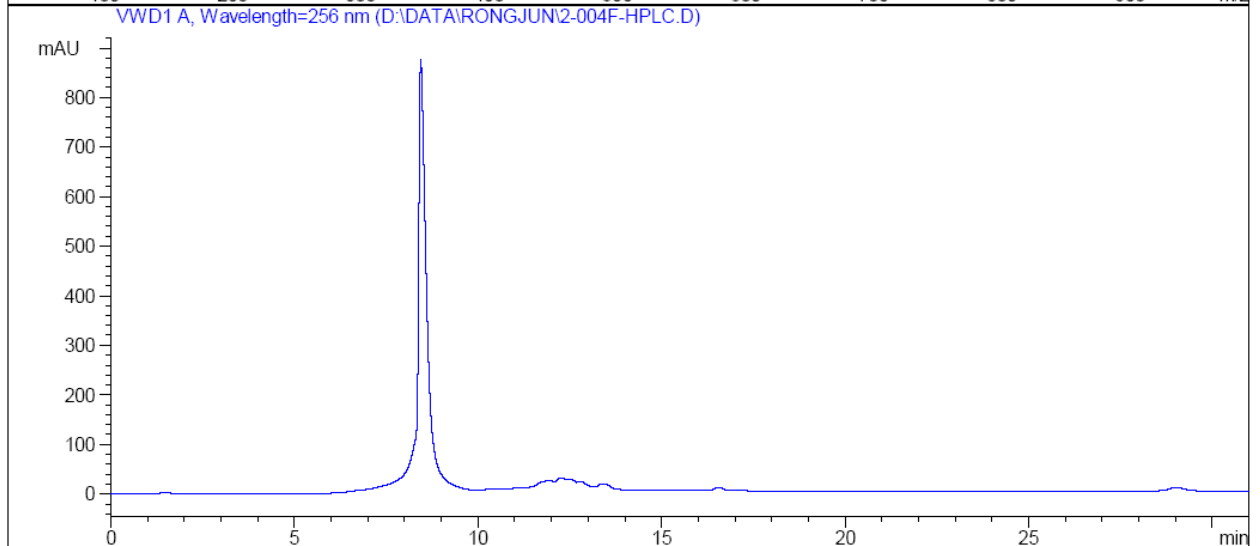
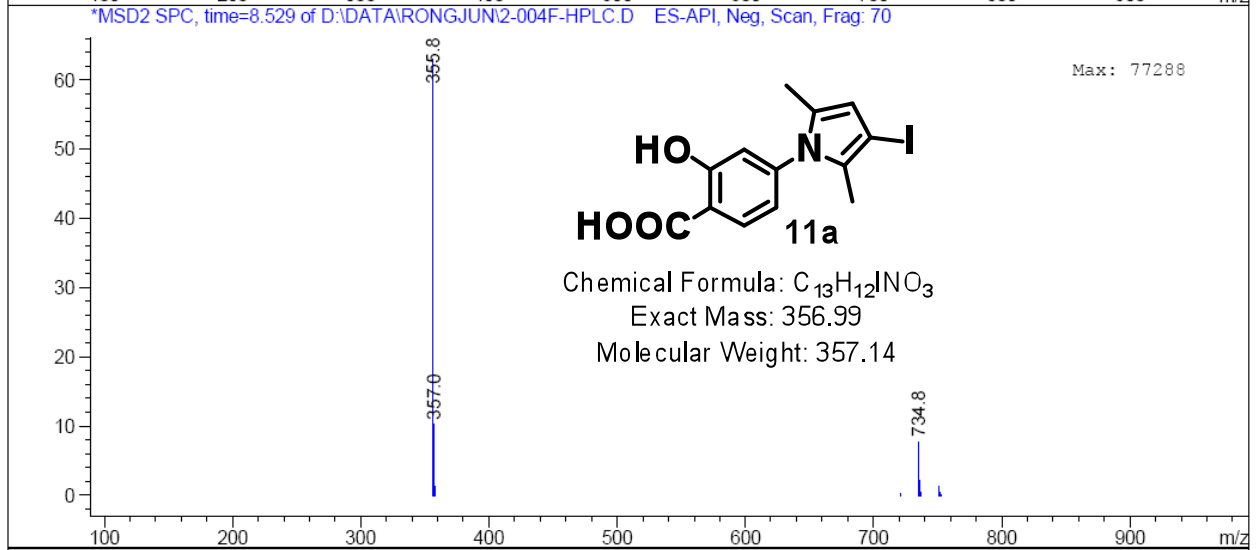
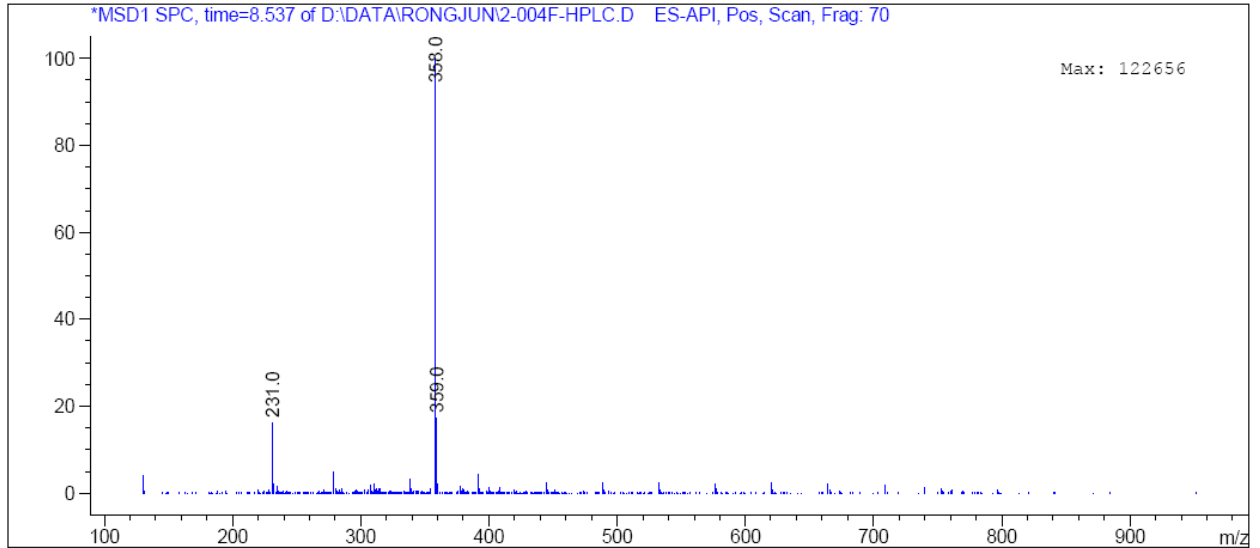


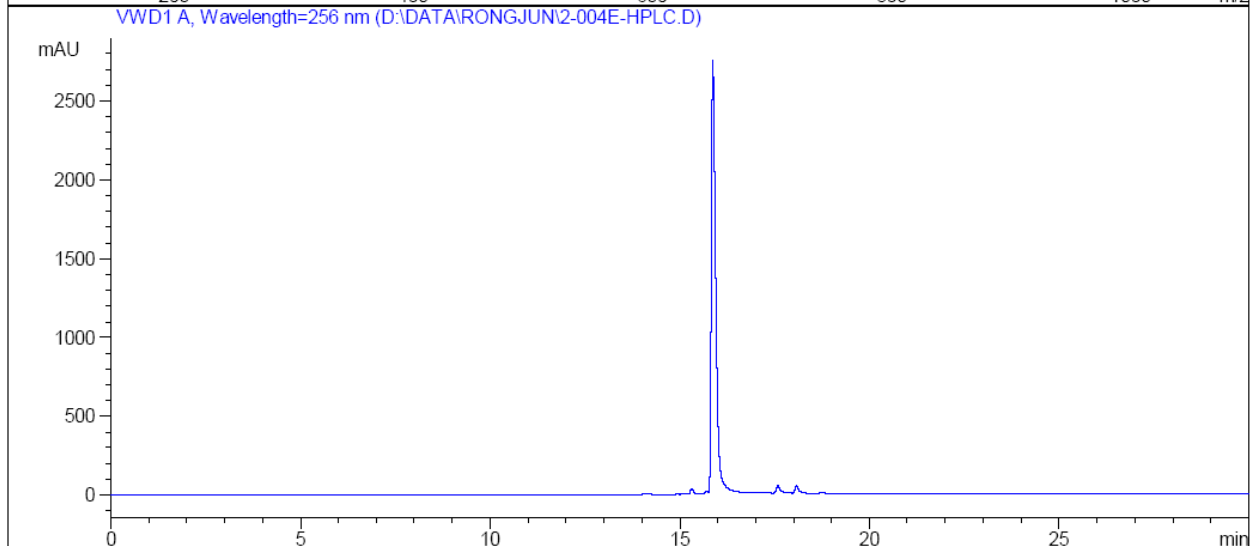
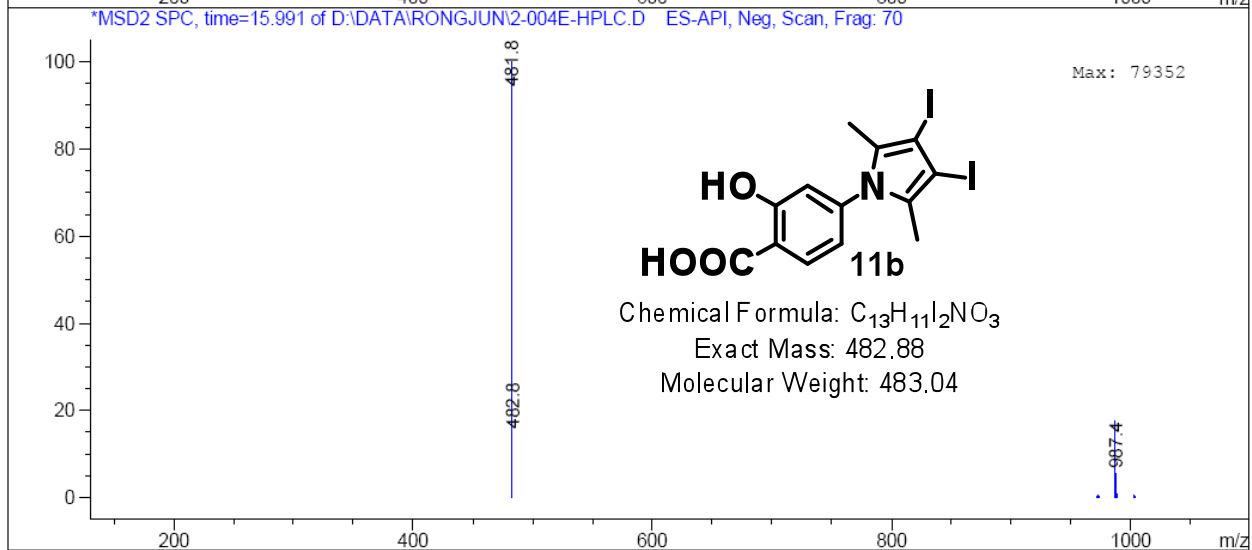
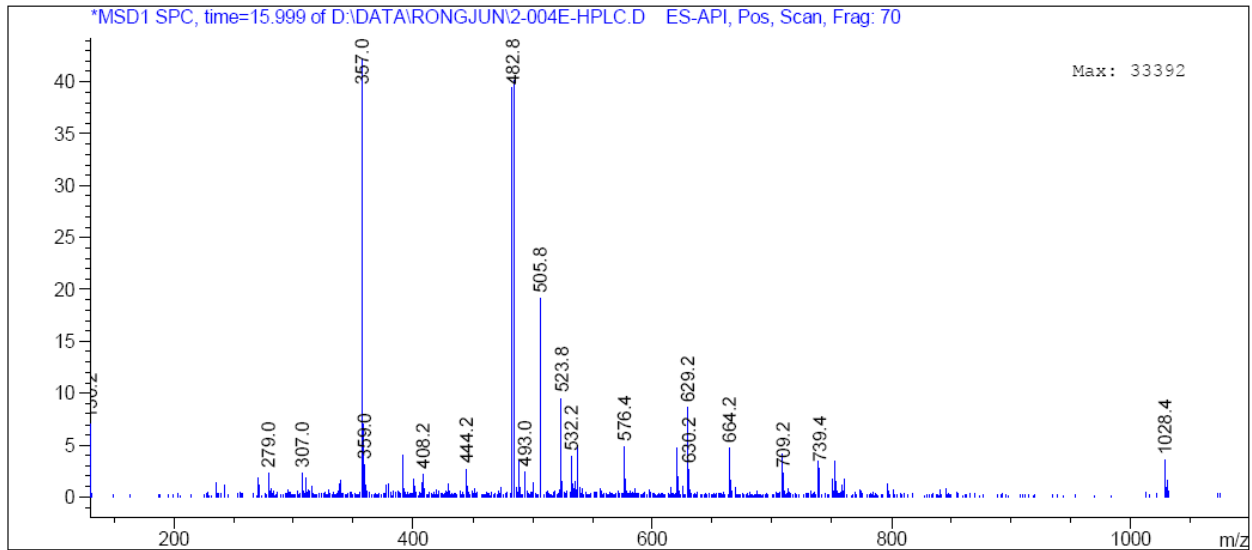




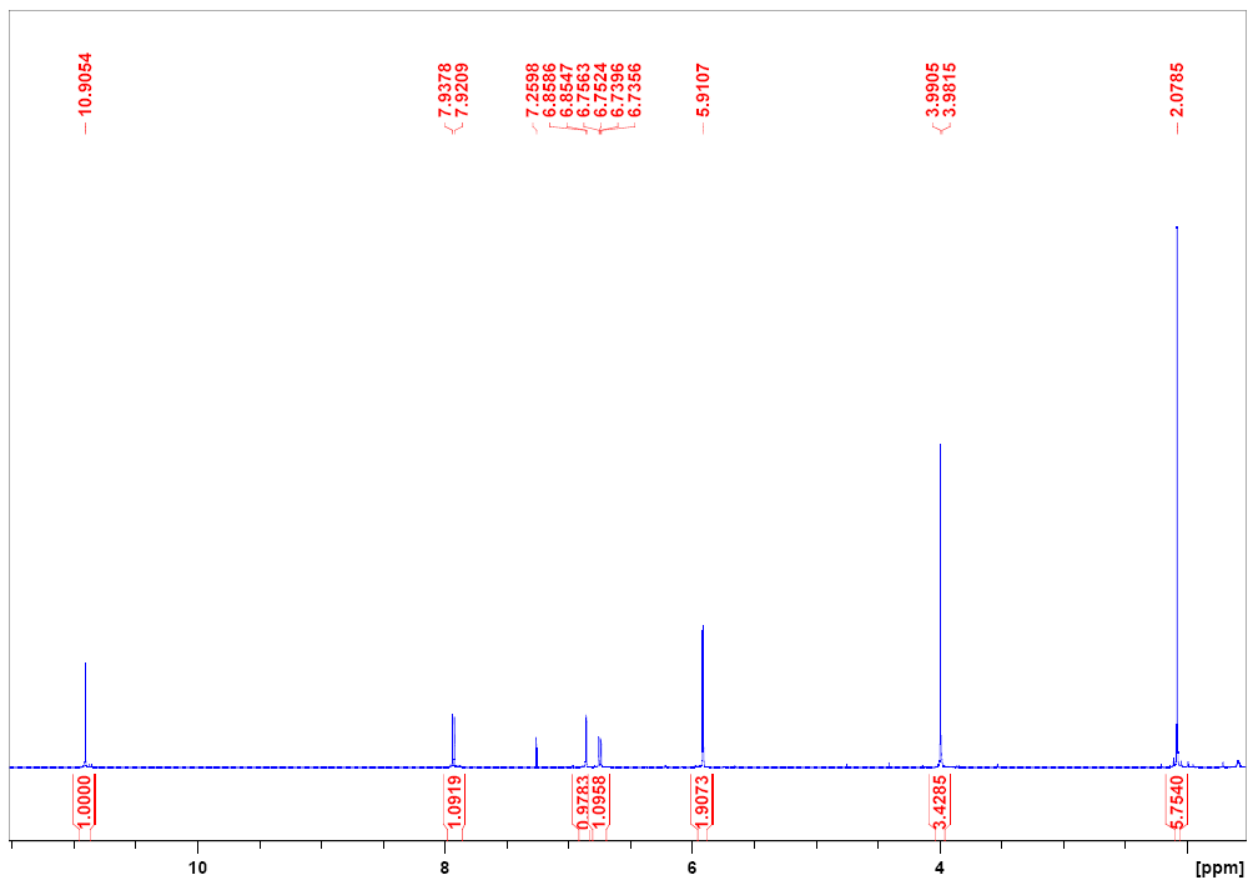
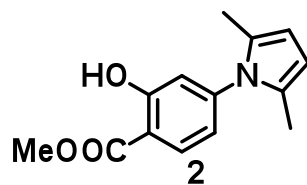


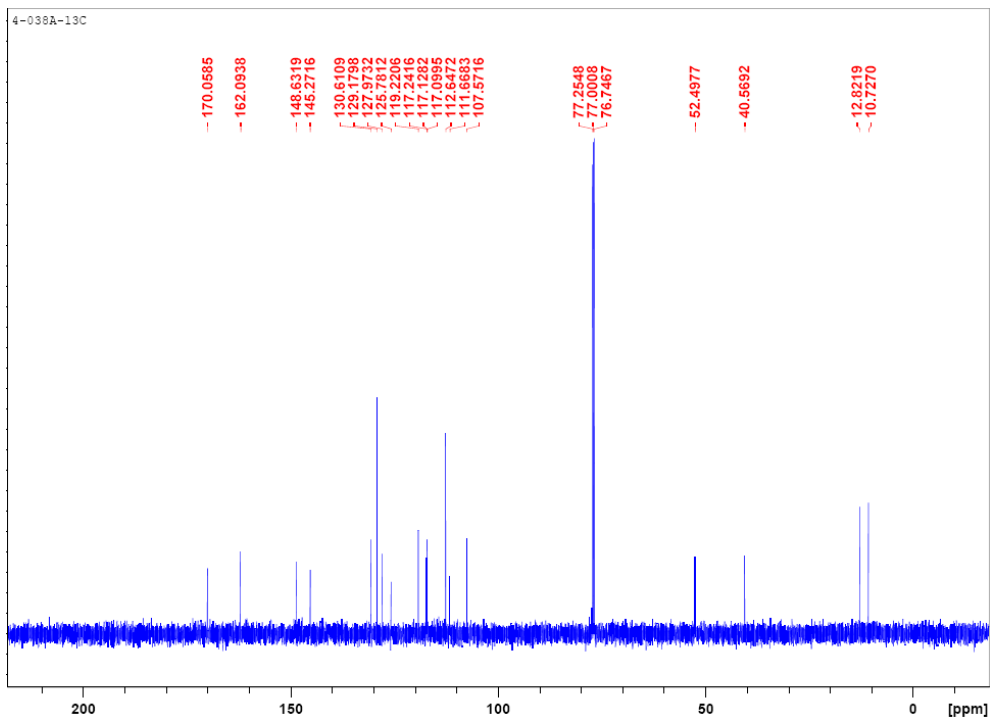
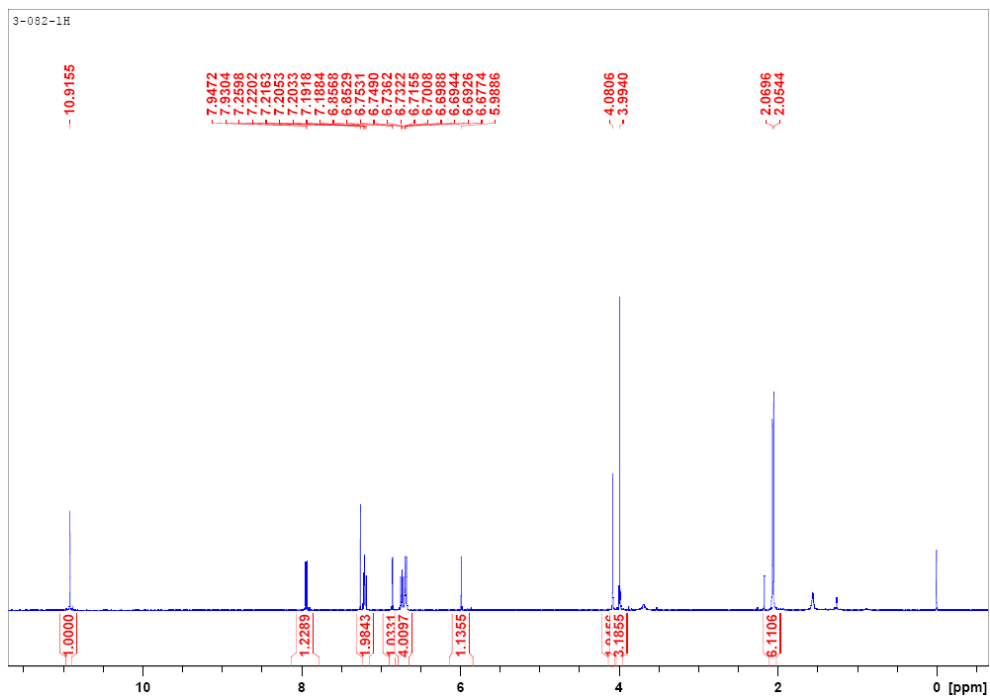
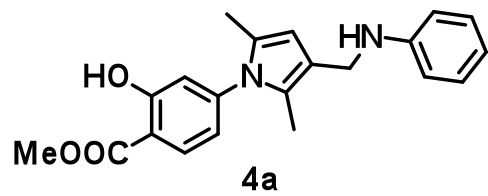


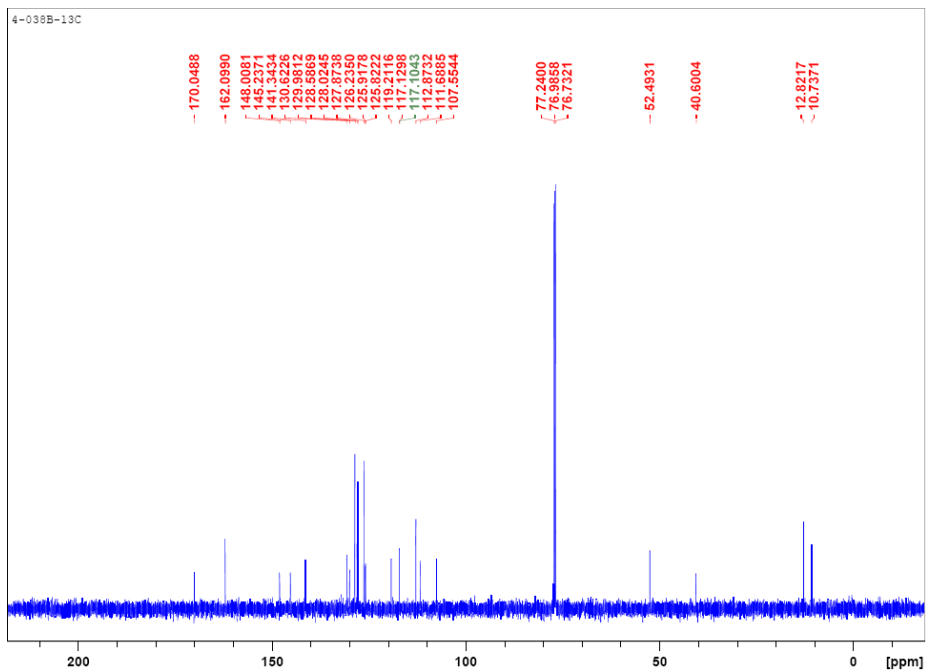
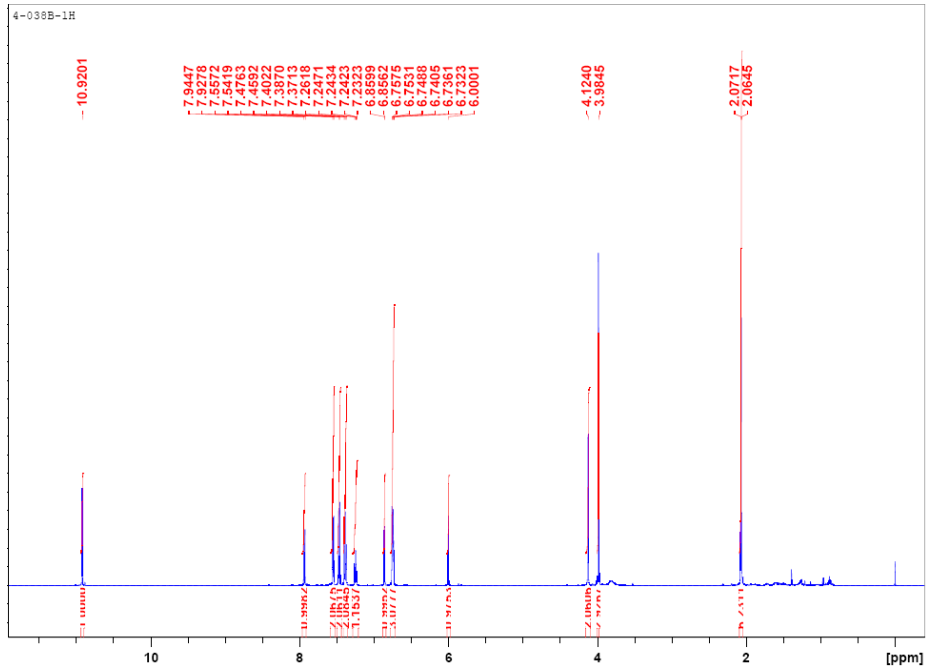
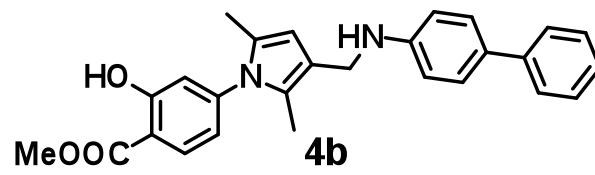


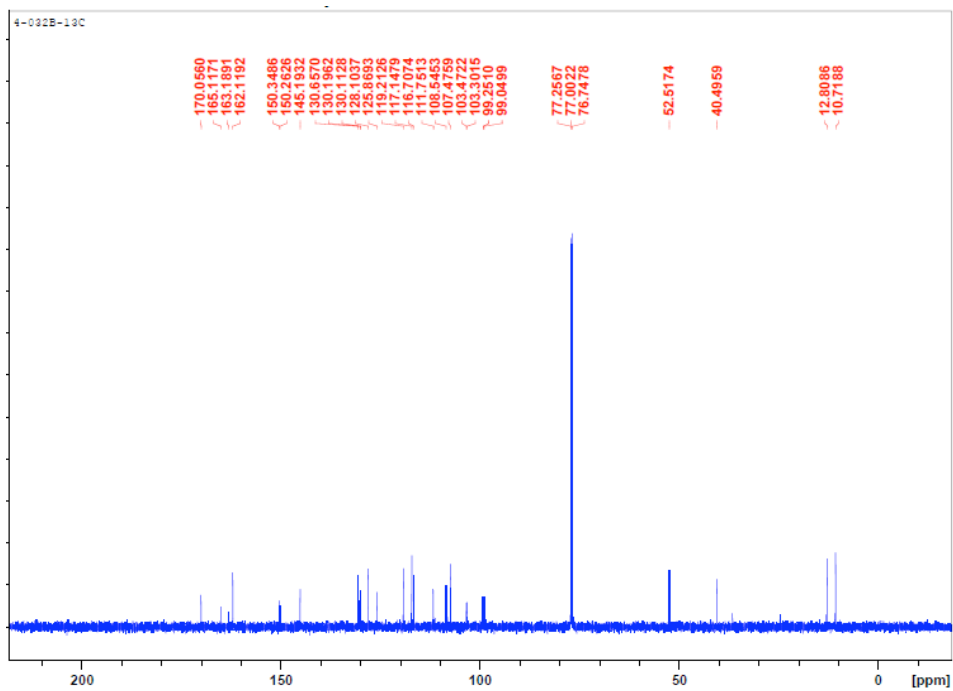
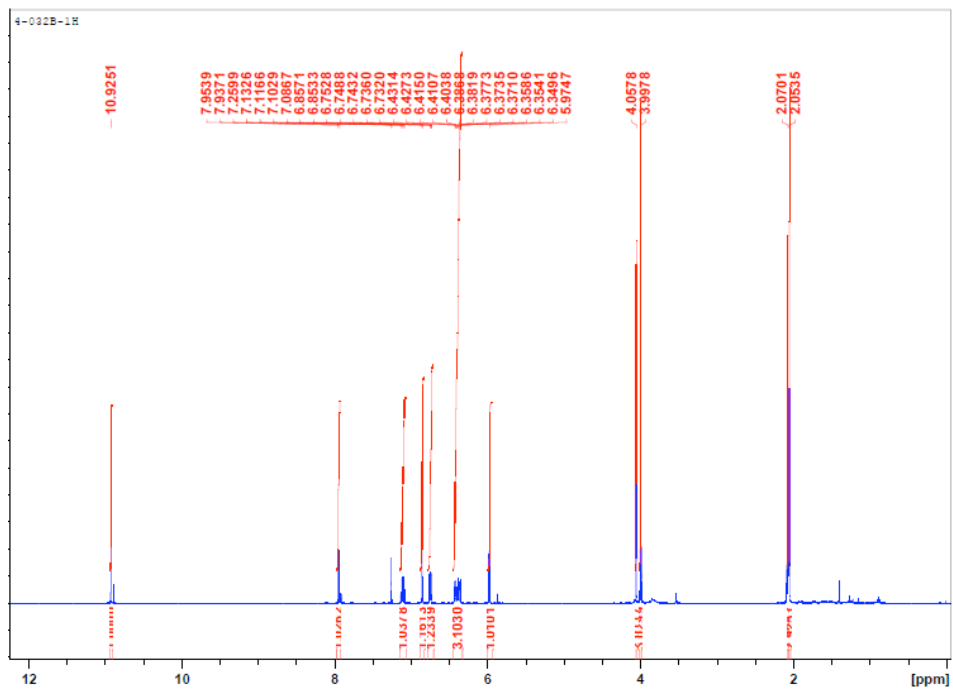
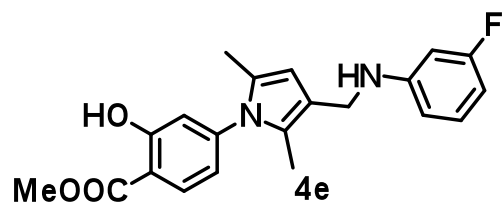


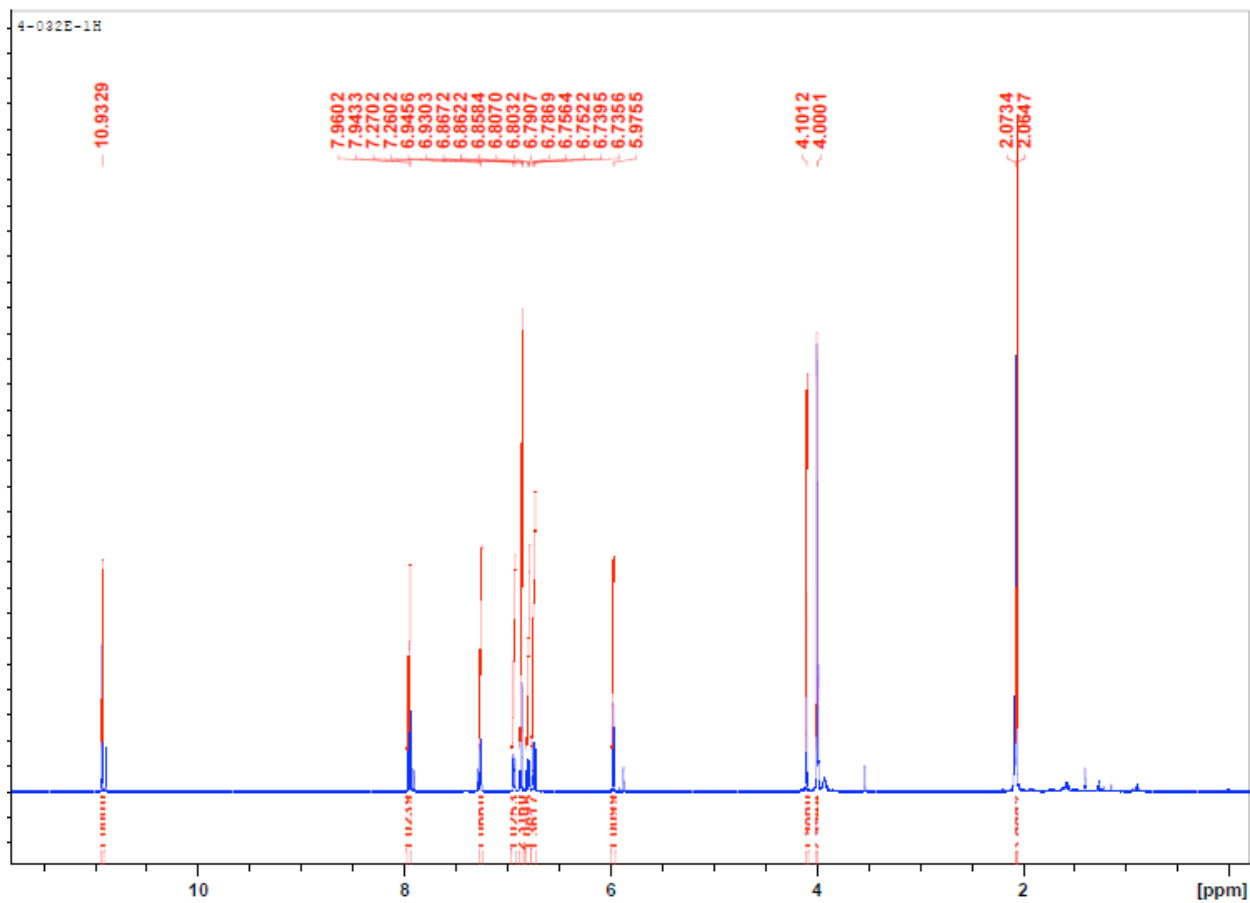
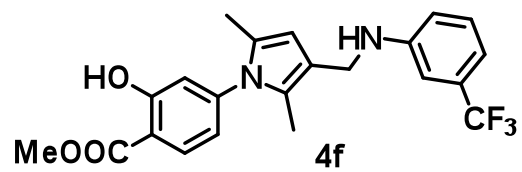
Copies of ^1H and ^{13}C NMR Spectra of compounds **2**, **4a-4l**, **5**, **7a**, **10b**

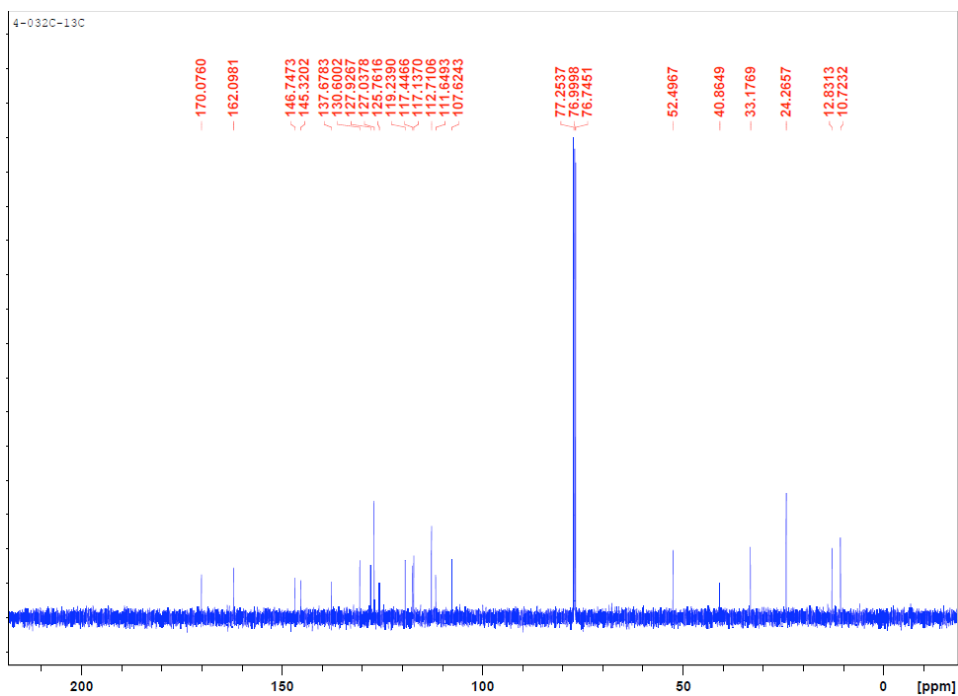
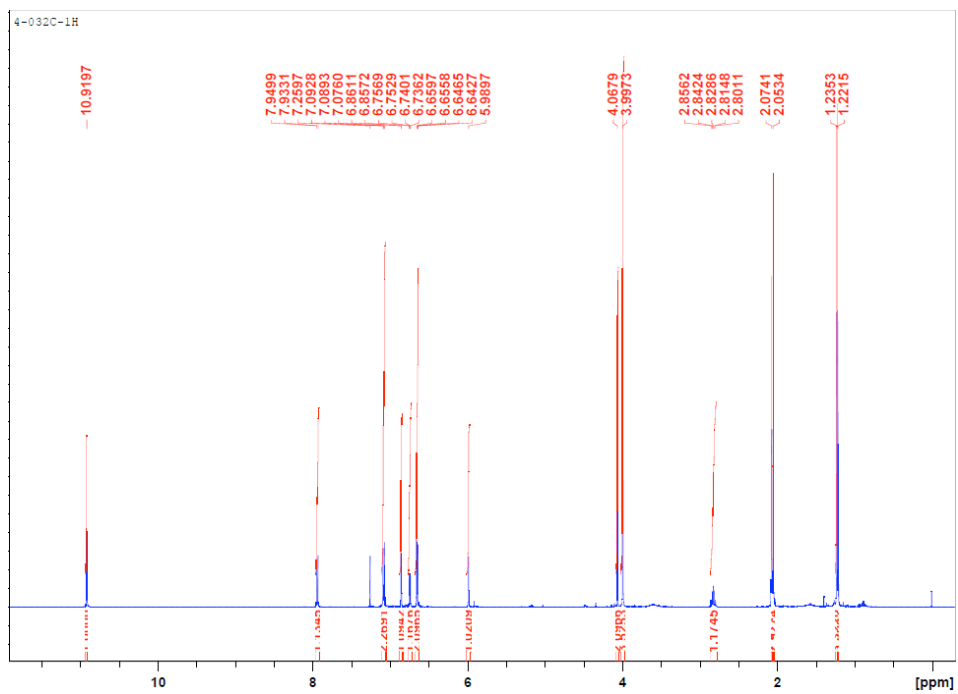
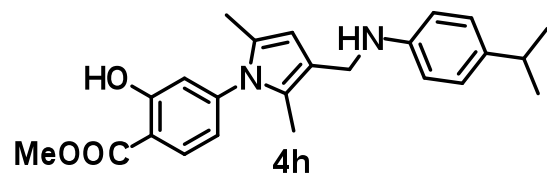


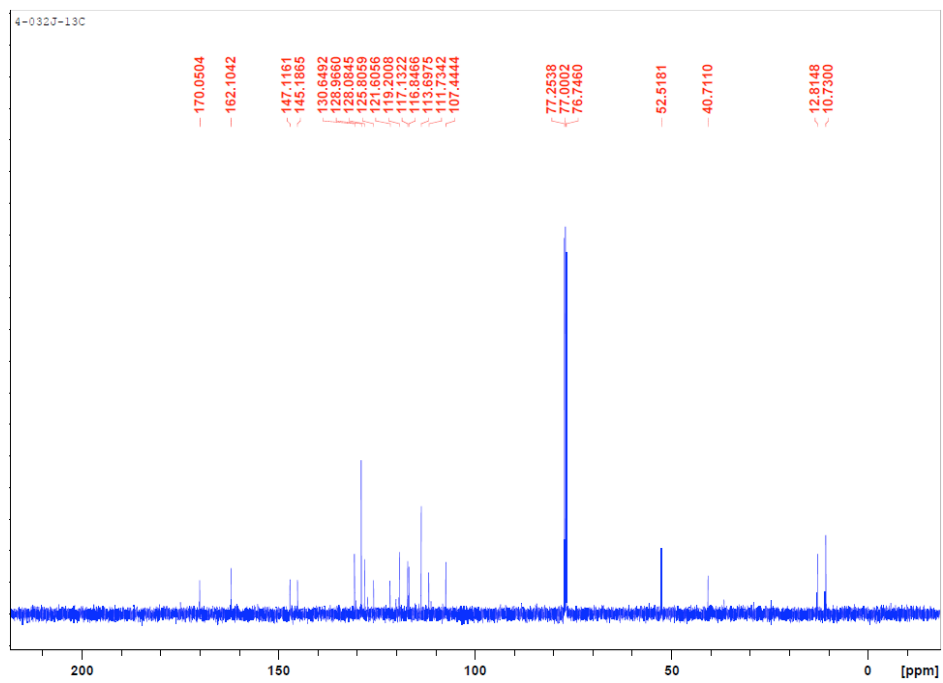
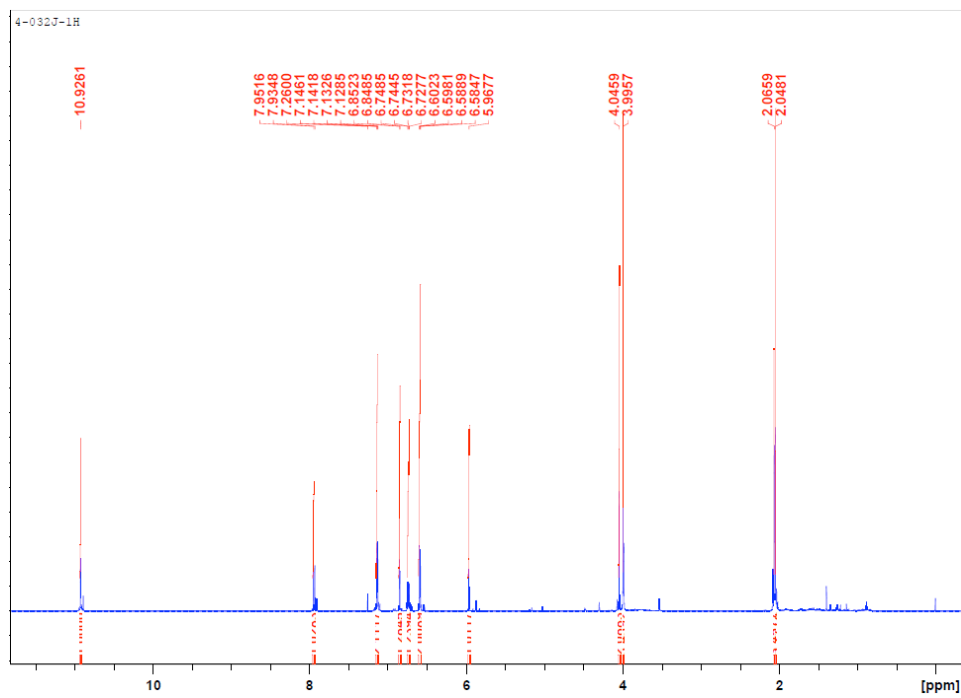
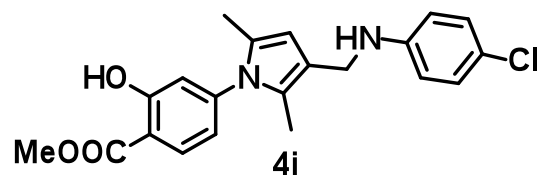


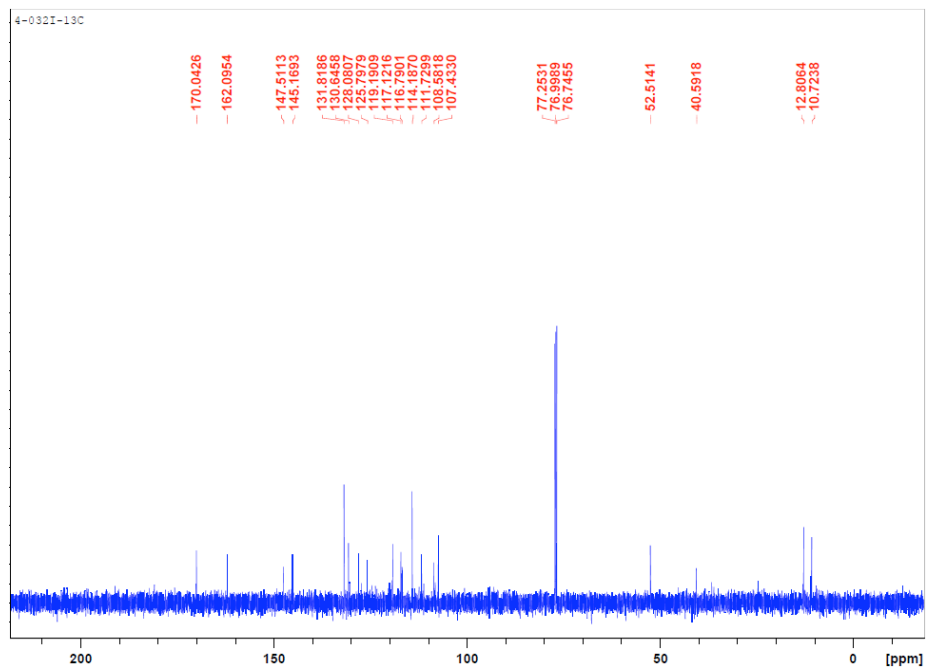
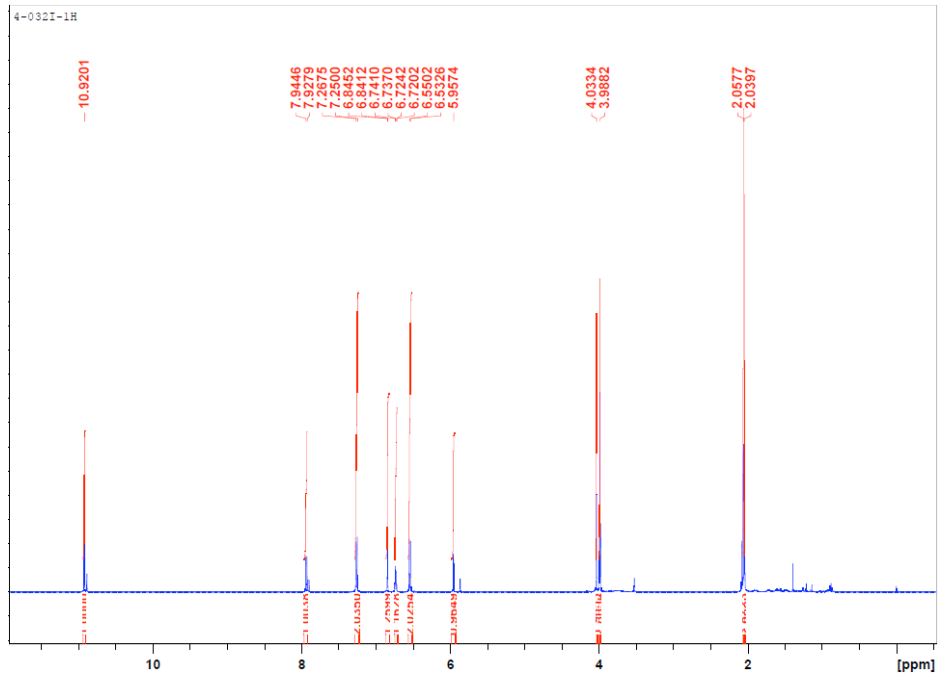
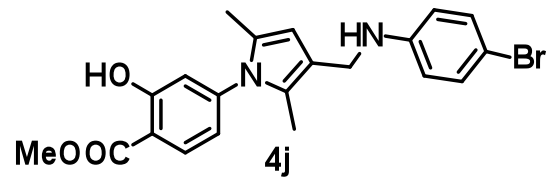


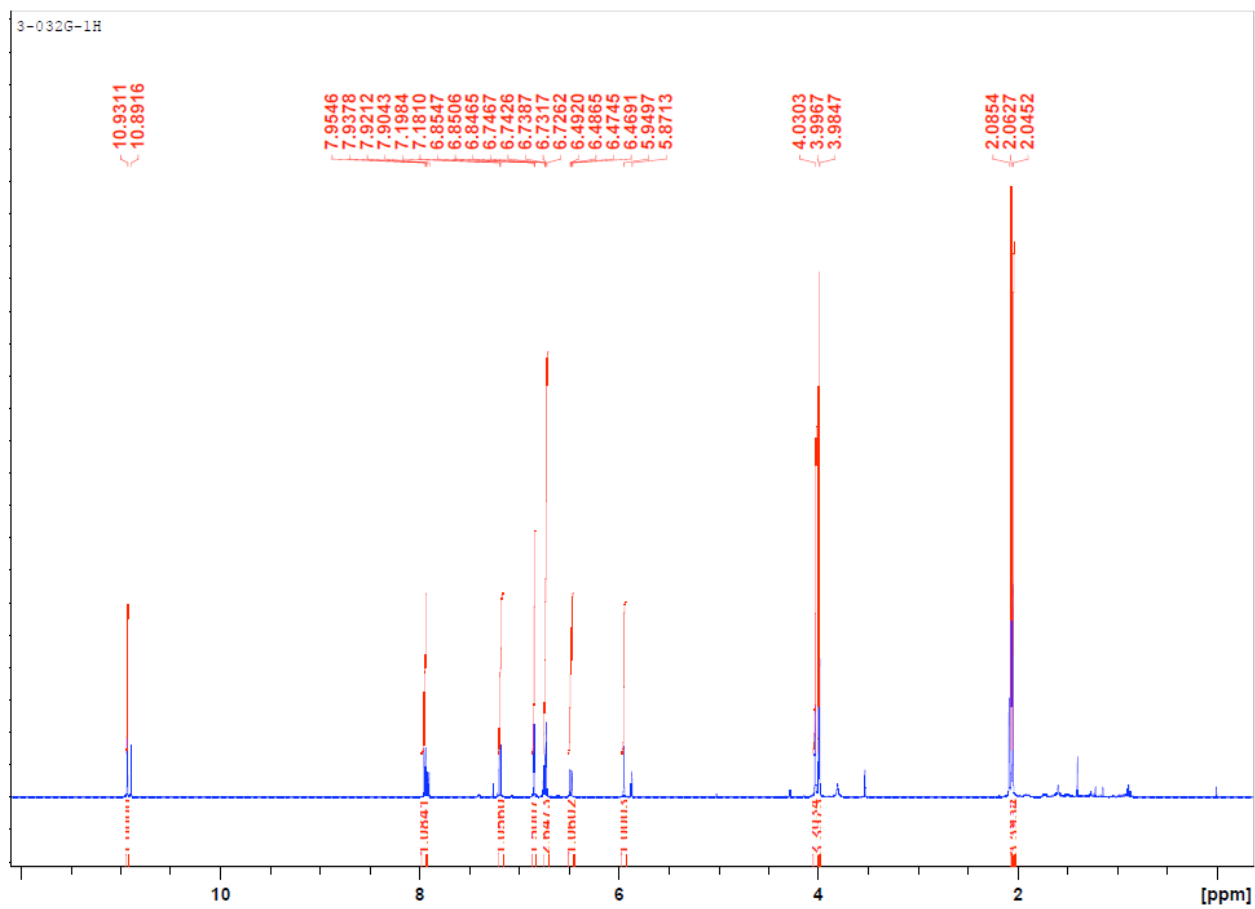
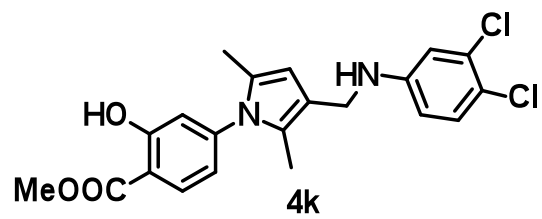


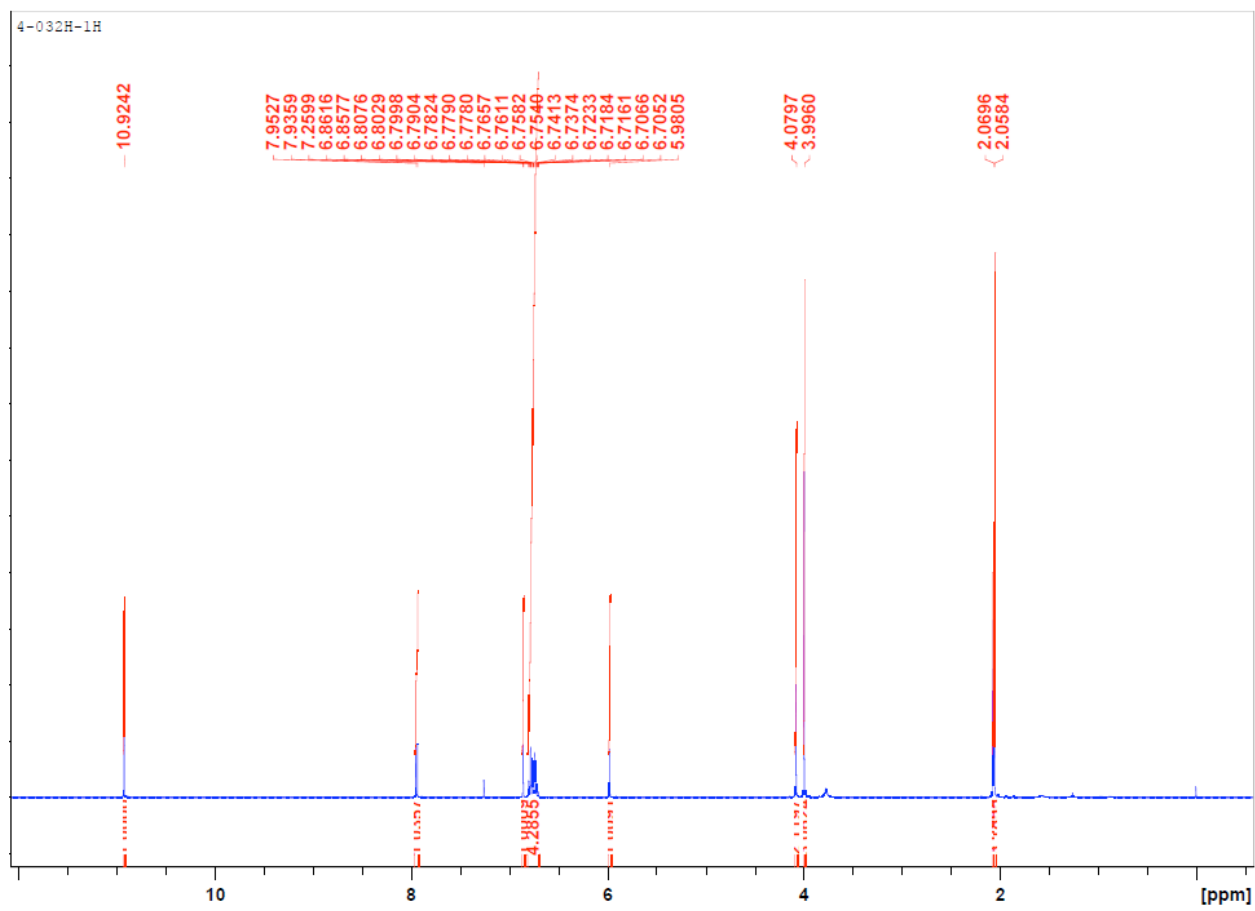
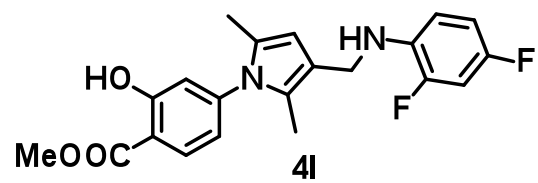


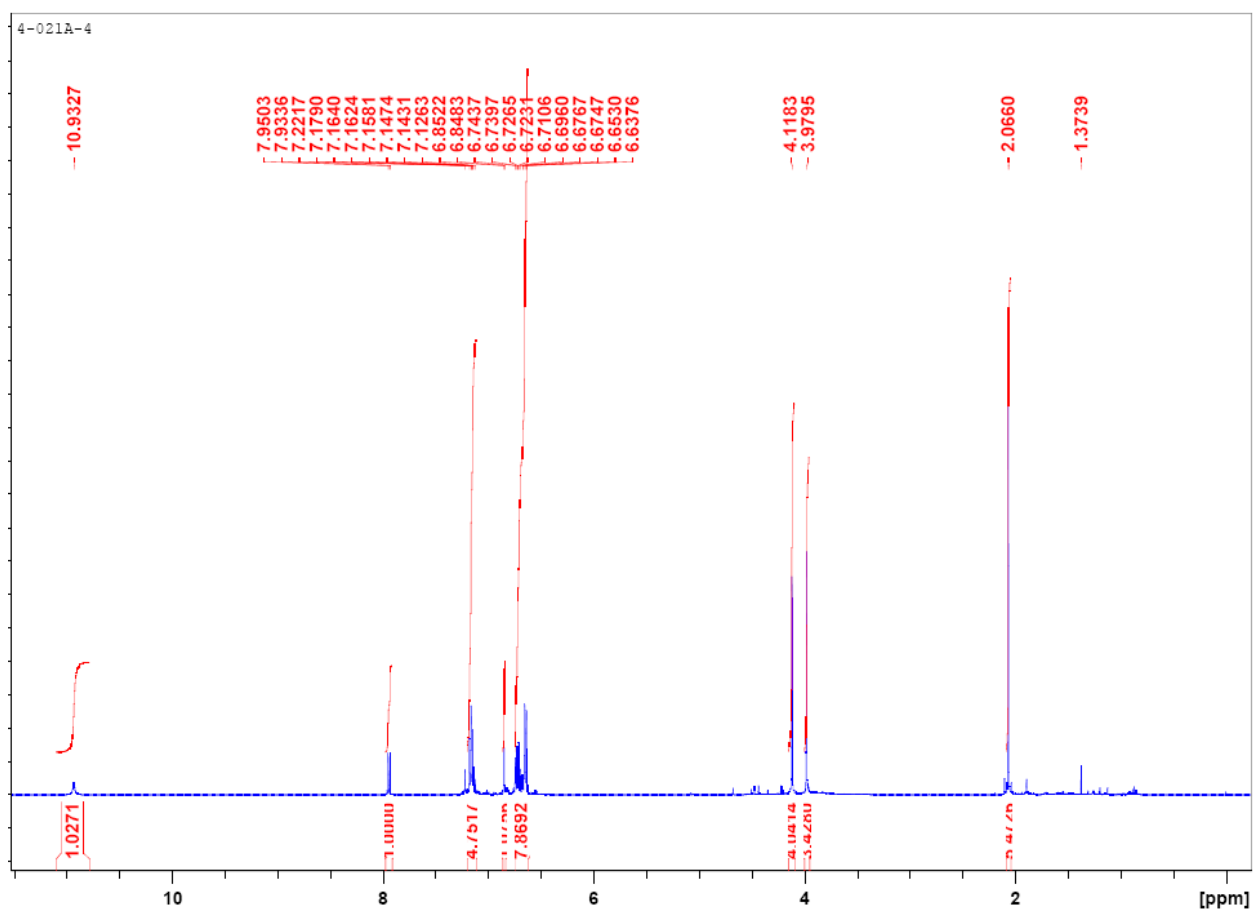
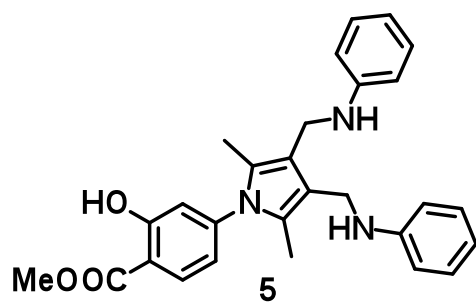


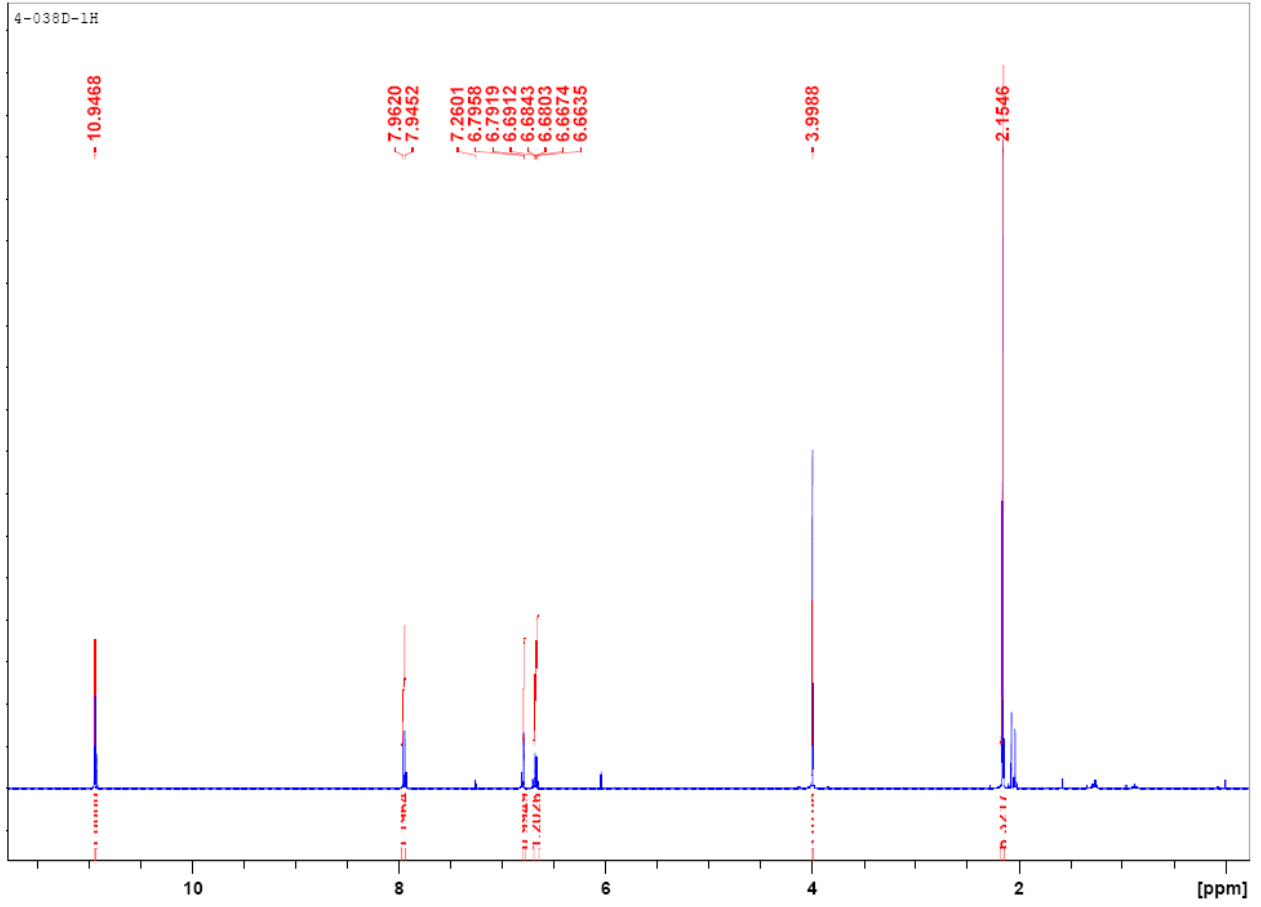
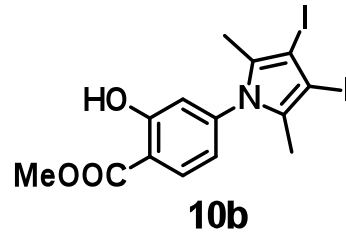


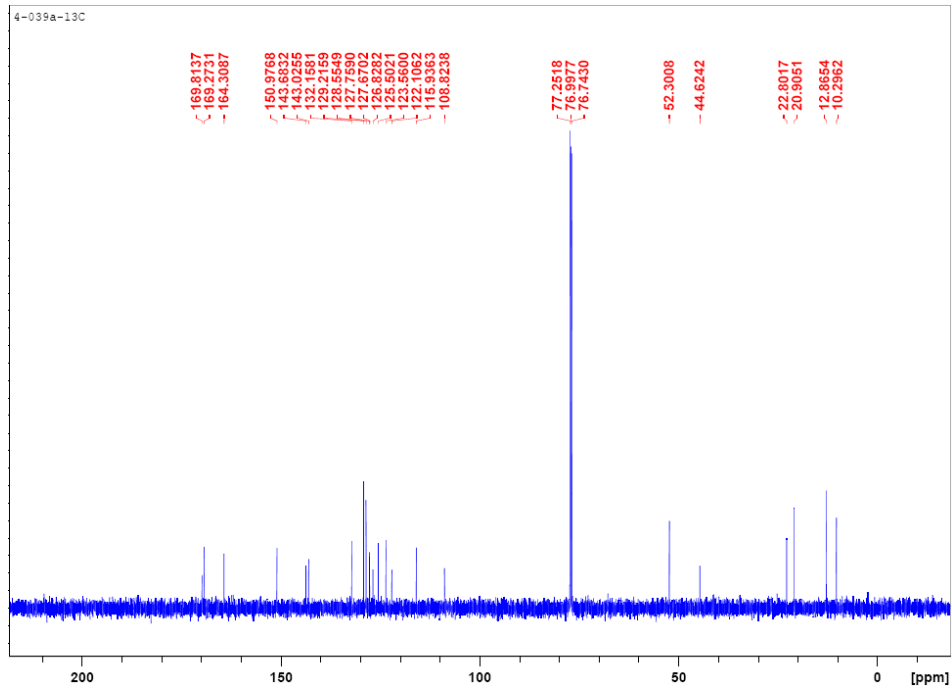
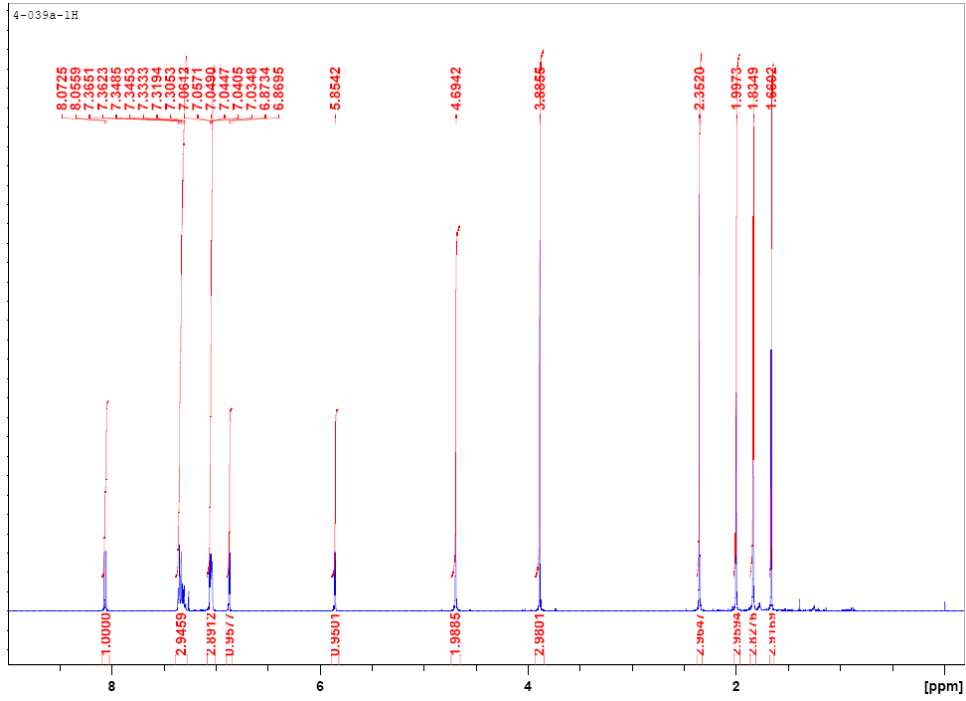
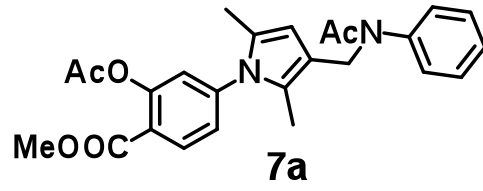












Inhibition studies for all compounds

The inhibition assays were performed on 96-well plates at 25°C in 50 mM 3,3-dimethylglutarate buffer, pH 7.0, containing 1 mM EDTA with an ionic strength of 0.15 M adjusted by NaCl. The reaction was started by the addition of 50 µl of the enzyme to 150 µl of reaction mixture containing *p*-nitrophenyl phosphate (*p*NPP) and various concentrations of the inhibitor [final concentration of mPTPB: 20 nM, final concentration of *p*NPP: 3 mM (the K_m value)]. The reaction was quenched after 10 min by the addition of 50 µl of 5 N NaOH. The absorbance at 405 nm was detected by a Spectra MAX340 microplate spectrophotometer (Molecular Devices). IC_{50} values were calculated by fitting the absorbance at 405nm *versus* inhibitor concentration to the following equation:

$$AI/A0=IC_{50}/(IC_{50}+[I])$$

where AI is the absorbance at 405 nm of the sample in the presence of inhibitor; A0 is the absorbance at 405 nm in the absence of inhibitor; and [I] is the concentration of the inhibitor.

Expression and purification of recombinant mPTPB were described previously.¹

[1] B. Zhou, Y. He, X. Zhang, J. Xu, Y. Luo, Y. Wang, S. G. Franzblau, Z. Yang, R. J. Chan, Y. Liu, J. Zheng, Z.-Y. Zhang, *Proc. Natl. Acad. Sci. USA* **2010**, *107*, 4573-4578.