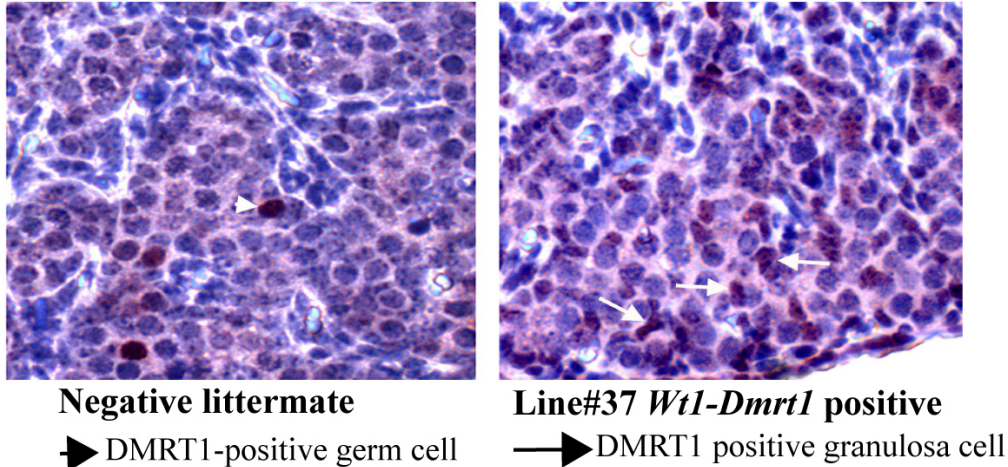


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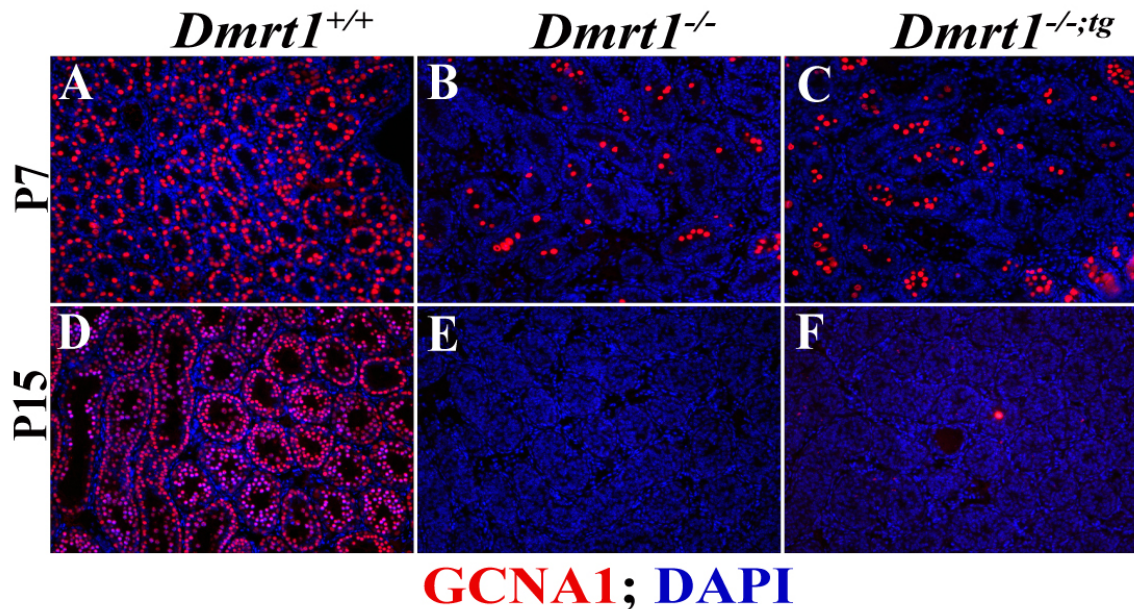
Supplementary Figure 1:

15.5 dpc ovary



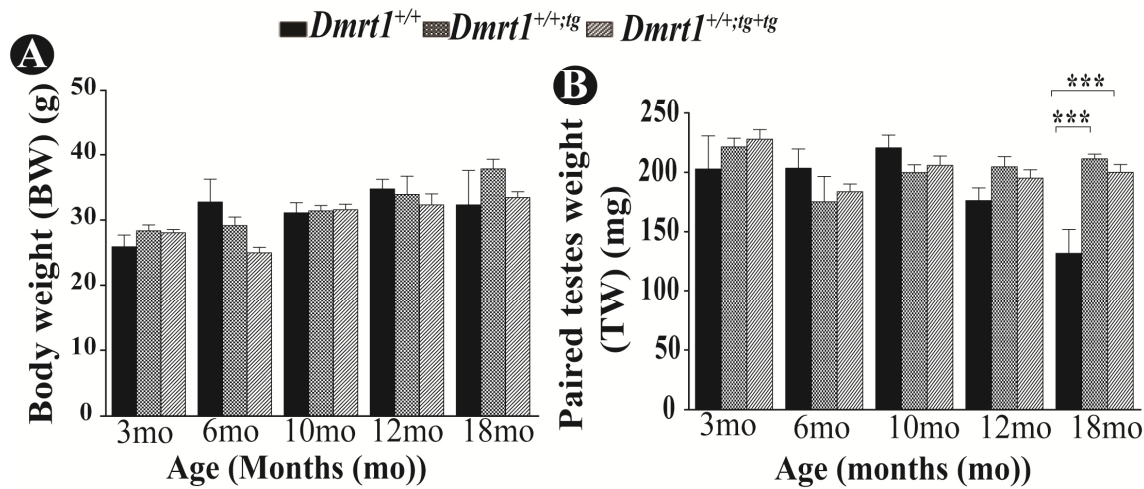
Supplemental Figure S1. Expression of transgenic DMRT1 in somatic cells of 15.5dpc ovary. Immunocytochemistry was used to examine DMRT1 expression in wild type (negative littermate, left) and transgenic ovaries isolated 15.5dpc embryos. In the wild type ovary (left) a few DMRT1-positive germ cells (arrowheads) were observed. As previously shown, DMRT1 was absent from ovarian somatic cells of 15.5dpc [1]. In the transgenic ovary (right), numerous DMRT1-positive somatic cells (some noted by arrows) were observed and few DMRT1-positive germ cells were present. Magnification x400.

Supplementary Figure 2:



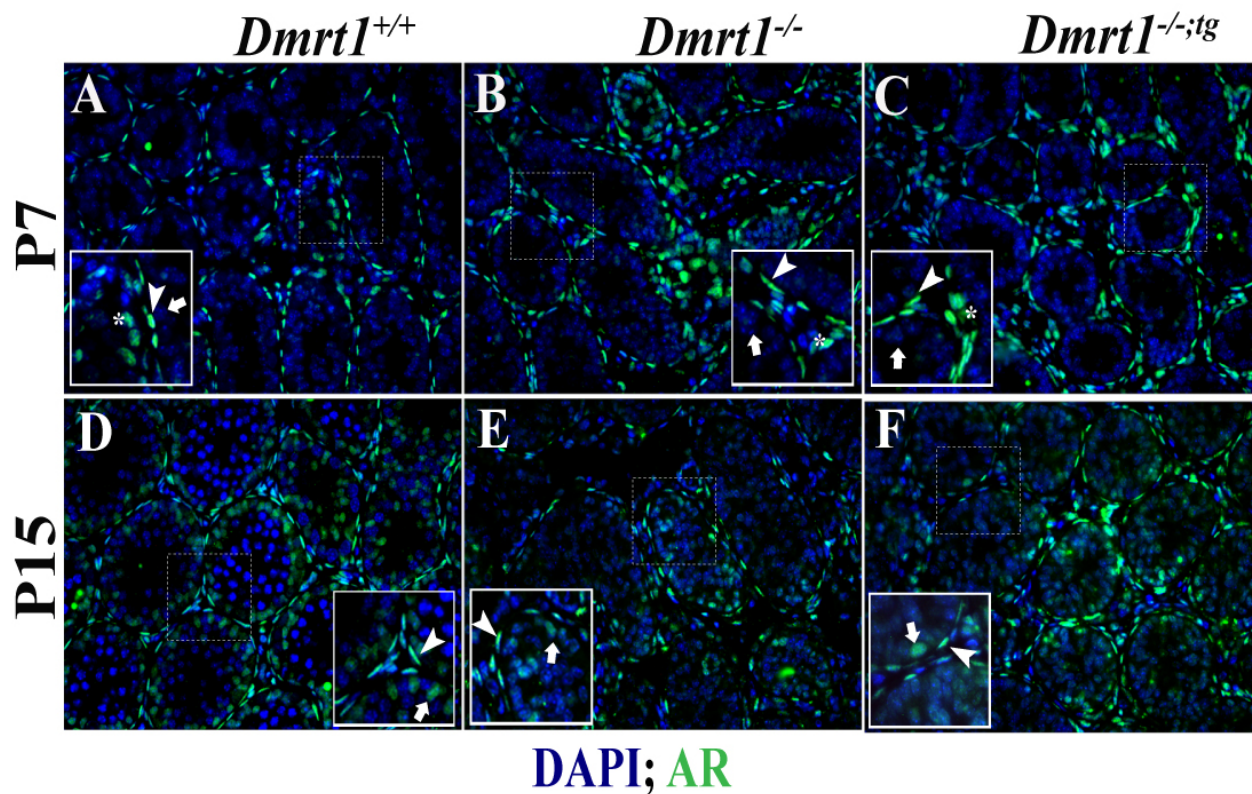
Supplemental Figure S2. Germ cell loss in *Dmrt1*^{-/-} and *Dmrt1*^{-/-;tg} mice testes at P7 and P15. GCNA1, a germ cell-specific protein, was examined to evaluate germ cell numbers in testes from different genotypes. GCNA1 immunofluorescence (red) was examined in testis sections from P7 (A-C) and P15 (D-F) wild type (A and D), *Dmrt1*^{-/-} (B and E) and *Dmrt1*^{-/-;tg} (C and F) mice. DAPI (blue) was used to stain nuclei. Final magnification for all micrographs is x400.

Supplementary Figure 3:



Supplemental Figure S3. Effect of exogenous *Dmrt1* on testes weights and sperm motility. Average body (A) and paired testes (B) weights were determined for wild type mice (*Dmrt1*^{+/+}; black bar) and mice with either one (*Dmrt1*^{+/+;tg}; stipple bar) or two (*Dmrt1*^{+/+;tg+tg}; stripe bar) copies of the transgene was determined at 3, 6, 10, 12, and 18 months of age. For each genotype, a minimum of three animals were used per time point. Bars represent mean ± SEM of three to 20 animals. ***p<0.0001 (Two tail Student T-Test).

Supplementary Figure 4:



Supplemental Figure S4. AR expression in P7 and P15 testes. AR immunofluorescence of P7 (A-C) and P15 (D-F) testes from *Dmrt1*^{+/+} (A & D), *Dmrt1*^{-/-} (B & E), and *Dmrt1*^{-/-;tg} (C & F) mice. Arrows denote AR-positive Sertoli cells. Asterisks denote AR-positive Leydig cells. Arrowheads denote AR-positive peritubular myoid cells. DAPI (blue) was used to stain for nuclei. Final Magnification x400.

Supplemental Tables

Supplemental Table S1. Primers used to PCR-amplify components of targeting vector.

Clone	Sequence (5'→3')	Direction	Product size (bp)	Polylinkers
Left arm (<i>Wt1</i> 5')	GCGCAAGCTTGAGCATCCTGGCTCCTCCTC	F	825	HindIII
	GCGCACGCGTCGTCGTTAGGCATGAGGTGCGGCTCGG	R		MluI
Right arm (<i>Wt1</i> 3')	GCGCCCGCGGTTCCGACGTGCGGGACCTG	F	776	SacII
	GCGCCCGCGGATCGATCCCTAAACCACAGCACCCTC	R		SacII
HPRT1	GCGCTCTAGAGTCGAGGACTTCAGGGATTG	F	1312	XbaI
	GCGCTCTAGAATTCAAAAAGTGGCGAATT	R		XbaI
LYS2	AGAGAGCGGCCGCGCACTTGCAATTACATAAAAAATTCCGG	F	4804	NotI
	AGAGAGCGGCCGCGCAAGTATTCATTTTAGACCCATG	R		NotI
<i>Dmrt1</i> <i>mMluI</i>	GCGCACGCGTTCGGCAAGCCCTCTGCACCG	F	237	MluI
	GCGCACGCGTAGCCGTGGTTCCTGCAGCGA	R		MluI

Supplemental Table S2. Primers used for genotyping.

	Primer Name	Sequence (5'→3')	Direction	Product size (bp)
Genotyping of Transgenic founders	<i>Wtl</i> -355	GCCTCAGAACCCAGGAGAG	F	873
	<i>Dmrt1</i> .16A	GTGAGGAACCTCCGTCGG	R	
Sex Genotyping	SRY Forward	AAGCGCCCCATGAATGCATT	F	250
	SRY Reverse	CGATGAGGCTGATATTTATA	R	
Genotyping of Transgenic pups	TGIF0088	GTCCTCTGAACCTAGCAGCTACG	F	689
	TGIF0090	CCATGGAGACTTTCTAACTGCTCCTG	R	
Transgene expression in Transgenic mice	<i>Dmrt1</i> 3'	CACAGGGTATTAGGAGGCTTG	F	434
	HPRT1	GGCCTATAGGCTCATAGTGCAA	R	
Genotyping of <i>Dmrt1</i> ^{+/+} allele	TGIF0105 (KOs 1N)	GATCTATCTGGAGCCAGGTGGTAG	F	277
	TGIF0107 (KOs 2N)	TGCACACGTGCACCCTCGCCATCG	R	
Genotyping of <i>Dmrt1</i> ^{-/-} allele	TGIF0105 (KOs 1N)	GATCTATCTGGAGCCAGGTGGTAG	F	420
	TGIF0106 (KOs 3N)	TCATGGCAGCTCTCCCAGTGGAGC	R	
Probe to identify positive YAC <i>Wtl</i> - <i>Dmrt1</i> clones	<i>Dmrt1</i> ΔMluI Up	GCGCACGCGTTCGGCAAGCCCTCTGCACCG	F	237
	<i>Dmrt1</i> ΔMluI Down	GCGCACGCGTAGCCGTGGTTCCTGCAGCGA	R	
Probe for Tg <i>Dmrt1</i> copy number determination	<i>Dmrt1</i> exon 5 forward	GCCCAGCAGTCAAGATTCTG	F	165
	<i>Dmrt1</i> exon 5 reverse	CGACTCAGTTCACAGGGTATT	R	

Supplemental Table S3. Primers used for quantitative PCR.

Transcript	Sequence	Tm	Direction	Product Length (bp)	Function	Reference
<i>Ar</i>	TGGCGGTCCTTCACTAATGTC	59	F	72	Marker of Somatic cells (Sertoli, Leydig & myoid cells) in the testis	[2]
	TGCGGTACTCATTGAAAACCAA	59	R			
<i>Gata1</i>	GTCAGAACCGGCCTCTCATC	58	F	59	Sertoli cell maturation marker (nuclear and cytoplasmic)	[3]
	TGCCTGCCCGTTTGCT	58	R			
<i>Krt18</i>	CTT GCT GGA GGA TGG AGA AG	58	F	72	Immaturity marker	[4]
	CTG CAC AGT TTG CAT GGA GT	58	R			
<i>Gata4</i>	GGGCCAACCCCTGGAAGAC	59	F	67	Sertoli cell nuclei marker	[5, 6]
	GACACACTCTCTGCCTTCTGAGAA	59	R			
<i>Espin</i>	TTACATGCAGACCAAGAACAAGCT	59	F	64	Adherens junction Protein	[7, 8]
	CCACCTTGGGCTCCTTGAG	59	R			
<i>TJP</i>	GCAATGGAGGAAACAGCTATATGG	60	F	61	Tight and adherens junction protein	[8, 9]
	AACCCAGGAGCCCTGTGAA	59	R			
<i>Ocln</i>	CTGGACATTTGCTCATCATAAAGA	58	F	102	Tight junction protein	[8, 9]
	GTTTGAATTCATCAGGTCTGTAAGGA	59	R			
<i>Cldn11</i>	GGACGAACTGGGCTCCAA	58	F	104	Tight junction protein	[8, 9]
	TGCACGTAGCCTGGAAGGA	59	R			
<i>Rpl7</i>	CAACAAGGCTTCAATTAACATGCT	59	F	59	Structural constituent of ribosome	[10]
	GGGTACCCCATGCAATG	59	R			

REFERENCES USED IN SUPPLEMENTAL DATA

1. Lei N, Hornbaker KI, Rice DA, Karpova T, Agbor VA, Heckert LL. Sex-specific differences in mouse DMRT1 expression are both cell type- and stage-dependent during gonad development. *Biol Reprod* 2007; 77:466-475.
2. Bremner WJ, Millar MR, Sharpe RM, Saunders PT. Immunohistochemical localization of androgen receptors in the rat testis: evidence for stage-dependent expression and regulation by androgens. *Endocrinology* 1994; 135:1227-1234.
3. Ketola I, Anttonen M, Vaskivuo T, Tapanainen JS, Toppari J, Heikinheimo M. Developmental expression and spermatogenic stage specificity of transcription factors GATA-1 and GATA-4 and their cofactors FOG-1 and FOG-2 in the mouse testis. *Eur J Endocrinol* 2002; 147:397-406.
4. Sasaki M, Yamamoto M, Arishima K, Eguchi Y. Effect of follicle-stimulating hormone on sertoli cell division in cultures of fetal rat testes. *Biol Neonate* 2000; 78:48-52.
5. Imai T, Kawai Y, Tadokoro Y, Yamamoto M, Nishimune Y, Yomogida K. In vivo and in vitro constant expression of GATA-4 in mouse postnatal Sertoli cells. *Mol Cell Endocrinol* 2004; 214:107-115.
6. McCoard SA, Wise TH, Fahrenkrug SC, Ford JJ. Temporal and spatial localization patterns of Gata4 during porcine gonadogenesis. *Biol Reprod* 2001; 65:366-374.
7. Grove BD, Vogl AW. Sertoli cell ectoplasmic specializations: a type of actin-associated adhesion junction? *J Cell Sci* 1989; 93 (Pt 2):309-323.
8. Mruk DD, Cheng CY. Tight junctions in the testis: new perspectives. *Philos Trans R Soc Lond B Biol Sci* 2010; 365:1621-1635.
9. Fahrioglu U, Murphy MW, Zarkower D, Bardwell VJ. mRNA expression analysis and the molecular basis of neonatal testis defects in *Dmrt1* mutant mice. *Sex Dev* 2007; 1:42-58.
10. Giallongo A, Yon J, Fried M. Ribosomal protein L7a is encoded by a gene (*Surf-3*) within the tightly clustered mouse *surfeit* locus. *Mol Cell Biol* 1989; 9:224-231.