Supplementary data for "Intraovarian transplantation of primordial follicles fails to rescue chemotherapy injured ovaries"

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Supplementary data include 2 Tables and 3 Figure

sTable 1. Ratio of ovarian follicle disrupted mouse at 5 weeks after busulfan treatment.

| | % of ovarian follicle disrupted mouse | | | | | | | | |
|-------------|---------------------------------------|-------|------|------|------|------|--|--|--|
| _ | 100% | 90% | 80% | 70% | 60% | 50% | | | |
| No. of mice | 13/28 | 11/28 | 3/28 | 0/28 | 0/28 | 1/28 | | | |

sTable 2. Primers and conditions used for semi-quantitative RT-PCR.

| Stage | Gene | Primer sequence | TM | Product size |
|-----------------------|--------|--------------------------|-------|---------------------|
| Progenitor | FIGLA | F: ACAGAGCAGGAAGCCCAGTA | 63°C | 205 bp |
| | | R: ACTCGCACAGCTGGTAGGTT | 03 C | |
| Primordial follicle | NOBOX | F: ACAAACGCCATGAGATTTCC | 63 °C | 215 bp |
| | | R: AACAGGGCCAGGTTCTAGGT | 03 C | |
| | LHX8 | F: CAGTTCGCTCAGGACAACAA | 58 °C | 207 bp |
| | | R: AGCCATTTCTTCCAACATGG | 36 C | |
| Duimour falliala | FOXO3A | F: GATGATGATGGACCCCTGTC | 56 °C | 247 bp |
| | | R: TCTTGGCGGTATATGGGAAG | 30 C | |
| Primary follicle | KIT | F: GTGGAGTGTAAGGCCTCCAA | 55 °C | 220 bp |
| | | R: CCTCGACAACCTTCCATTGT | 33 C | |
| | GDF9 | F: CCCCAAAACGAGTGTGAACT | 63 °C | 243 bp |
| Casandany falliala | | R: CACACTCAGGGGGCTGTACT | 03 C | |
| Secondary follicle | KITL | F: TCCGAAGAGGCCAGAAACTA | 55 °C | 238 bp |
| | | R: TGCGGCTTTCCTATTACTGC | 33 C | |
| Foult antual | DDR2 | F: CCGAAAGCTTCCAGAGTTTG | 55 °C | 249 bp |
| Early antral follicle | | R: TCTCCCAGCTTCTCCTTGAA | 33 C | |
| | AHR | F: ATGGTTCCTGTGTGCCCTACC | 63 °C | 465 bp |

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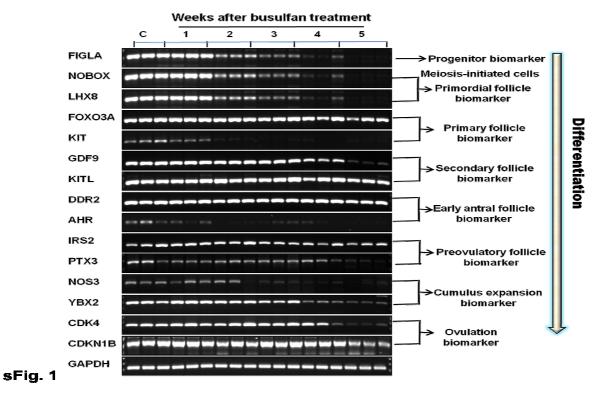
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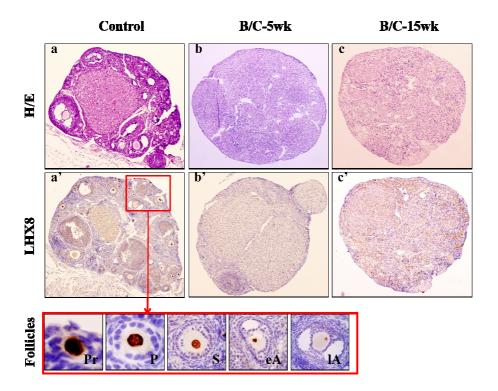
| | | R: GCTGCCTATAGTCGGGCTCTG | | |
|-----------------------|---------------|------------------------------|--------|----------------------|
| | SOX3 | F: GTCGTATTGCTTCTGTTTTCCTCT | 60.00 | 218 bp |
| Preovulatory follicle | | R: AATGCGTACACGAAATAGCAAATA | 60 °C | |
| | IRS2 | F: ACAACCTATCGTGGCACCTC | 62.00 | 223 bp |
| | | R: GGTGGTGGTAGAGGA AA | 62 °C | |
| Cumulus expansion | TNFAIP6 PTX3 | F: ATTTGAAGGTGGTCGTCTCG | (1.00 | 317 bp 274 bp |
| | | R: TGTGCCAGTAGCAGACCTGG | 61 °C | |
| | | F: GTGGGTGGAAAGGAGAACAA | 5.C.0C | |
| | | R: CCCAGATATTGAAGCCTGTGAT | 56 °C | |
| Ovulation | NANOS3 | F: CTGACAAGGCAAAGACACAGGA | (2.90 | 281 bp |
| | | R: AGGCAACACCACAAGTAGGG | 62 °C | |
| | YBX2 | F: ATGGGGTAGAACCCAAGGAG | 50 0C | 281 bp |
| | | R: GGAGGTCTCTGCGGCTATAG | 58 °C | |
| Luteal | CDK4 | F: GAAACTCTGAAGCCGACCAG | 57.0C | 244 bp |
| | | R: TTGTGCAGGTAGGAGTGCTG | 57 °C | |
| | CDIVIID | F: CAGAATCATAAGCCCCTGGA | 50.0C | 2241 |
| | CDKN1B | R: TCTGACGAGTCAGGCATTTG | 59 °C | 224 bp |
| | D52 | F: GGAGTATTTGGACGACCG | 56 °C | 350 bp |
| | P53 | R: TCAGTCTGAGTCAGGCCC | 30 C | 330 bp |
| | BCL-2 | F: TAAGCTGTCACAGAGGGGCT | 58 °C | 344 bp |
| | BCL-2 | R: TGAAGAGTTCCTCCACCACC | 30 C | |
| | BAX | F: CGAGCTGATCAGAACCATCA | 55 °C | 283 bp |
| Apoptosis-related | DAA | R: GAAAAATGCCTTTCCCCTTC | 33 C | |
| Apoptosis-related | FAS | F:GAGAATTGCTGAAGACATGACAATCC | 60 °C | 314 bp |
| | ras | R: GTAGTTTTCACTCCAGACATTGTCC | | |
| | FASL | F:TTAGCTTCTCTGGAGCAGTCAGCGTC | 62 °C | 320 bp |
| | | R:CCTTCTTCTTTAGAGGGGTCAGTGGC | | |
| | TNFaR55 | F: TGCTGCACCAAGTGCCACAAAG | 60 °C | 325 hn |
| | INFARSS | R: CACACGGTGTTCTGAGTCTCC | 00 C | 325 bp |
| | AKT | F: TCAAGAGGCAGGAAGAAGAGAC | 65 °C | 302 bp |
| | | R: AAGGAAGGGATGCCTAGAGTTC | 03 C | |
| | MEK | F: TACTCTGTGCAGTCGGACATCT | 62 °C | 344 bp |
| c-Kit signaling | | R: TGCTTCAGATCTGCTCTCTCTG | 02 C | |
| C-IXIt signating | mTOR | F: CTCCGATTGTGAAATTGTTTGA | 62 °C | 355 bp 377 bp |
| | miok | R: TGCTGGTAAATCAAAGGGTCTT | 02 C | |
| | SRC | F: TTGTCCTGATCATTTCAACACC | 62 °C | |
| | SKC | R: AGATGCCACAAATCATCAACTG | 02 C | |
| | Atg4A | F: CCCTCACACAACCCAGACTT | 55 °C | 287 bp |
| | Aig | R: CCCCTGTGGTTGTCACTTCT | | |
| | Atg4B | F: TGCTTTGAGAACCCAGACCT | 55 °C | 250 bp |
| | 7 Rig ID | R: CTCCTGACCCACTGCTCTTC | | |
| | Atg5 Atg6 LC3 | F: GGAGAGAGAGGAGCCAGGT | 55 ℃ | 227 bp 333 bp 251 bp |
| Autophagy-related | | R: TGTTGCCTCCACTGAACTTG | | |
| Autoplingy Telateu | | F: GGCCAATAAGATGGGTCTGA | 55 °C | |
| | | R: GCTTTTGTCCACTGCTCCTC | | |
| | | F: TTCTTCCTCCTGGTGAATGG | 55 °C | |
| | Gaparap | R: GTGGGTGCCTACGTTCTCAT | | 226 bp |
| | | F: CAGCAGGAGGGGTAATGGTA | 55 °C | |
| II amaalaa | | R: CCAATGTCAATCCCTTCCAC | | |
| Housekeeping | GAPDH | F: AGGTCGGTGTGAACGGTTTGA | 57 °C | 123 bp |
| gene | | R: TGTAGACCATGTAGTTGAGGTCA | | - |

sFigure 1. Female germ cell-specific gene expression levels in control and busulfan-treated mouse ovaries. FIGLA is expressed mainly from progenitor cells to primordial follicles, whereas NOBOX and LHX8 are

expressed mainly in primordial follicles. Both FOXO3A and c-kit are expressed in primary follicles and during the transition from primary to secondary follicles. GDF 9 and KITL expression is limited to secondary follicles. DDR2 and AHR are expressed primarily in early antral follicles, and PTX3 and IRS2 are expressed primarily in preovulatory follicles. YBX2 and CDK2 are involved in cumulus cell expansion, and CDK4 and CDKN1B are involved in ovulation. Time points after busulfan treatment are indicated above each panel.

sFigure 2. Expression of Lhx8 in control (upper left panel) and busulfan-treated mouse ovaries [5 (upper middle panel) and 15 (upper right panel) weeks]. Expression and localization of LHX8 protein were detected immunohistochemically using a rabbit anti-LHX8 polyclonal antibody. LHX8 protein was located in the nucleus of oocytes, and increasingly expressed during primordial (Pr), primary (P), secondary (S), early antrum (eA), and late antrum (IA) follicles in control ovary. However, any positive signal was not detected in both of 5 and 15 weeks ovaries after busulfan treatment. Method. Ovaries were fixed in 10% formaldehyde for 12–24 h before paraffin embedding. The samples were serially sectioned at 5µm and mounted on glass slides. Antigen retrieval was performed using 0.01M sodium citrate. The sections were blocked by BDT (3% BSA, 10% normal goat serum in TBS) for 30 min and incubated with rabbit anti-LHX8 polyclonal antibody at a dilution of 1: 200 (Abcam, Ab45118) overnight at 4°C. After rinsing thoroughly with TBS, the sections were incubated with HRP-conjugated goat anti-rabbit secondary antibody at a dilution of 1: 50 (Beyotime, A0208) for 30 min at 37°C. LHX8 expression in sectionswas detected by the reaction of peroxidase with 3,3′-diaminobenzidine tetrahydrochloride (DAB) and analyzed under Olympus BX51 fluorescence microscope.





sFig. 2