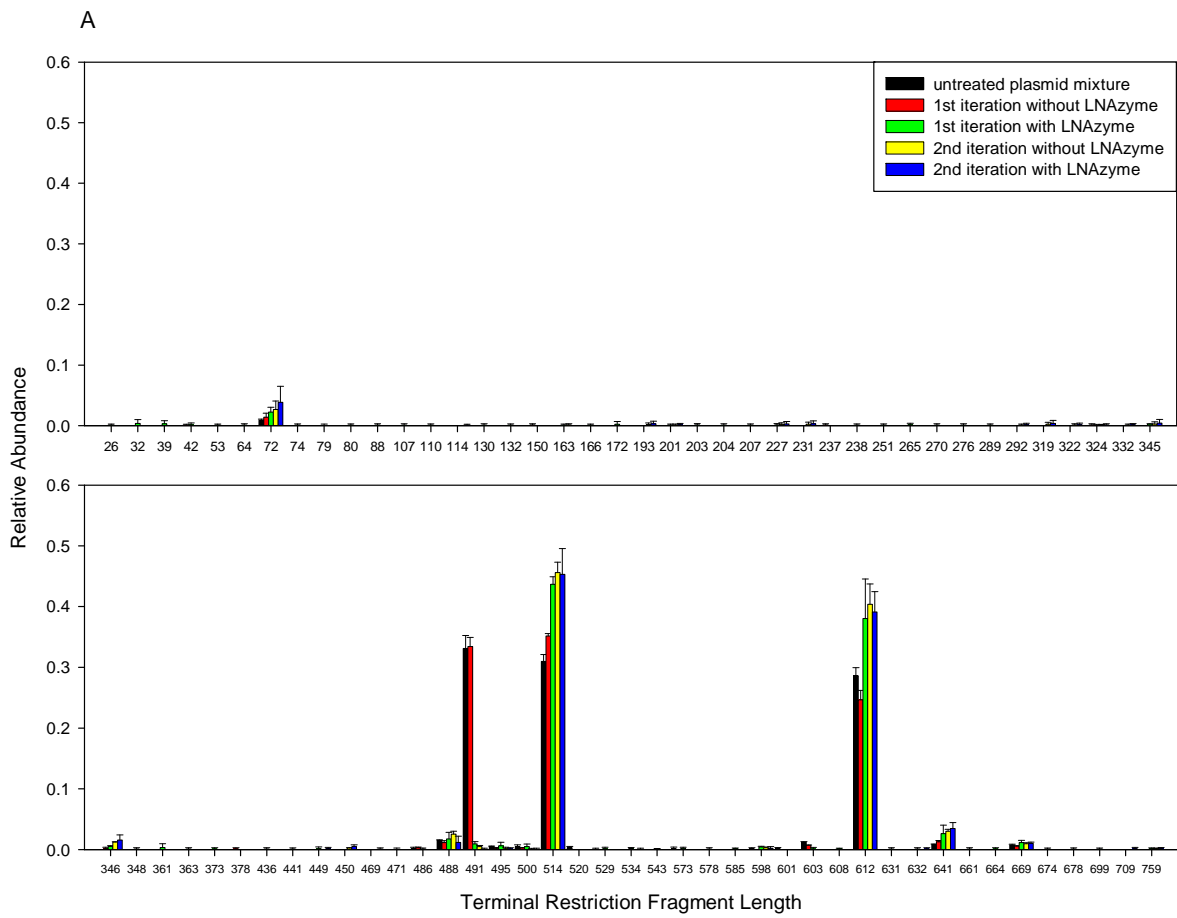
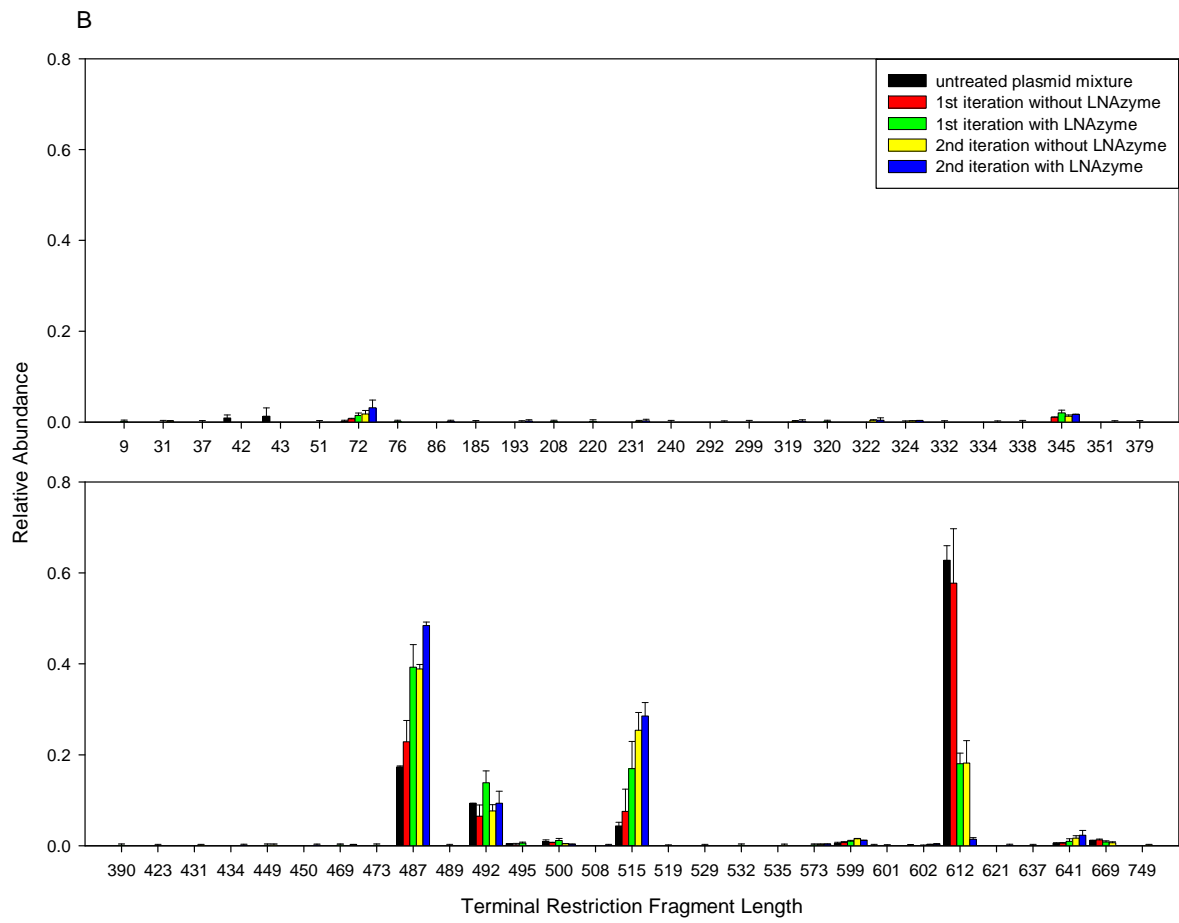


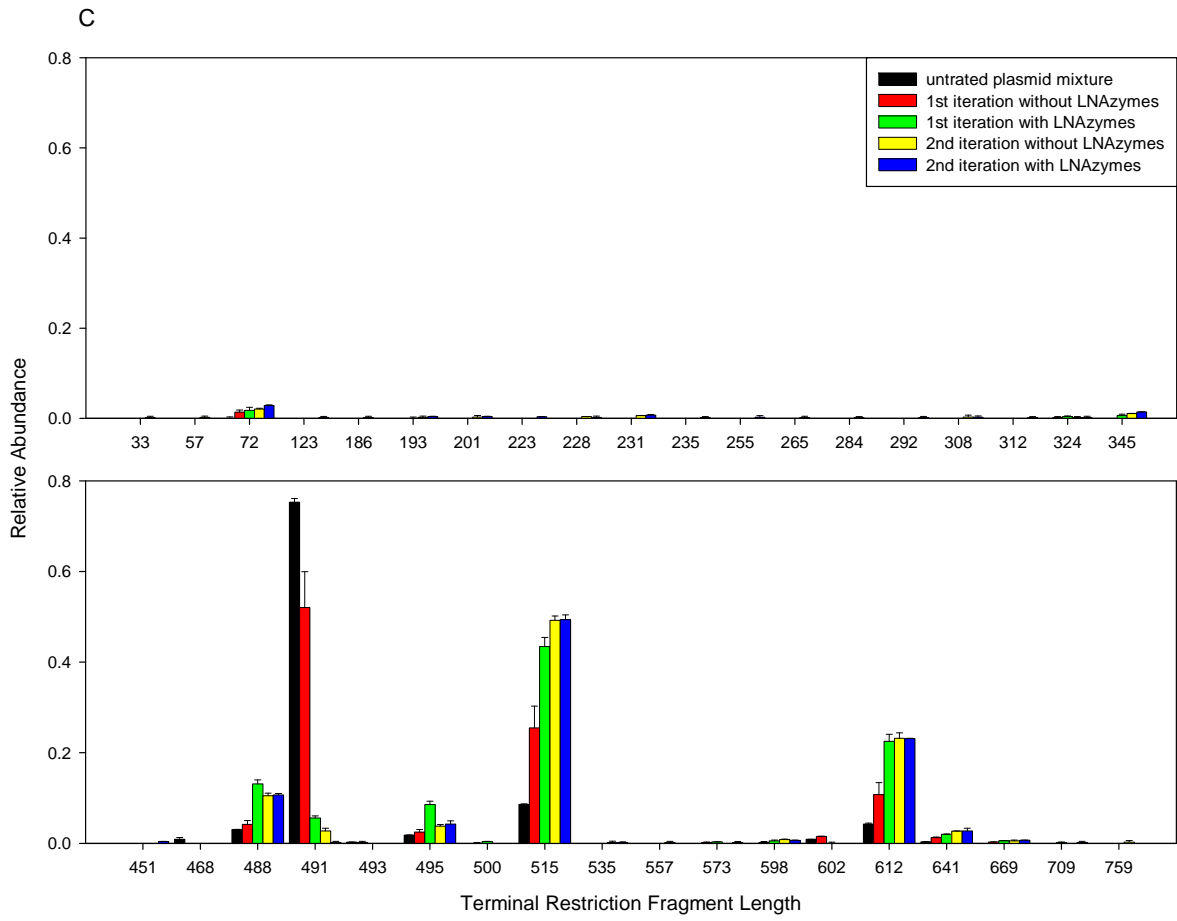
**Figure S2 A**



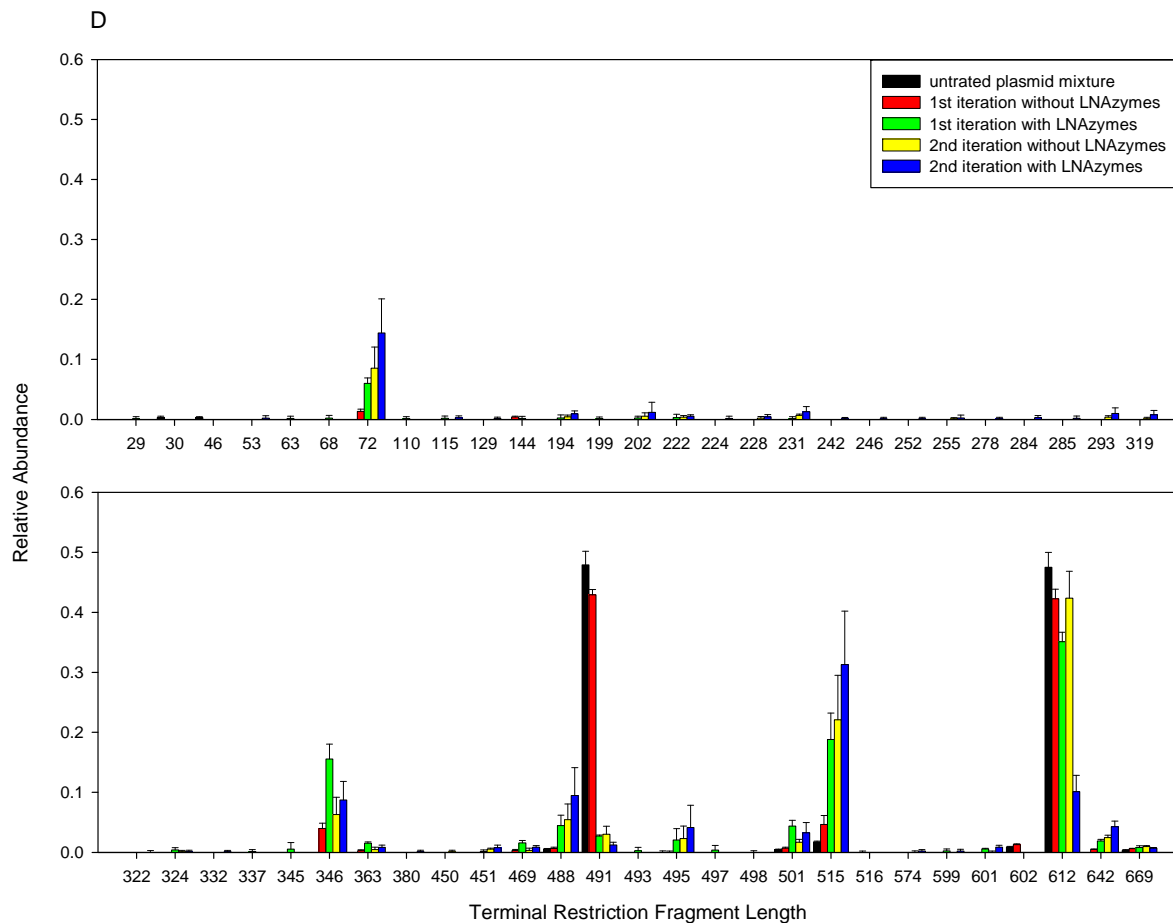
**Figure S2 B**



**Figure S2 C**



## Figure S2 D



**Fig. S2.** Evaluation of the LNAzyme cleavage protocol with artificial mixtures of cloned partial 16S rRNA genes (see Table 1 in the main text). The bars show the relative abundances of the rRNA genes in the mixtures before and after one or two iterations of the LNAzyme-mediated depletion of the *Nitrosomonas* and/or *Nitrospira* rRNA. Control experiments without added LNAzymes are also shown (the second iteration without LNAzyme was performed after a first iteration with LNAzyme). The relative abundances were determined by T-RFLP analysis. This figure shows the complete T-RFLP profiles, whereas Fig. 3 (main text) contains the relevant peaks only. Terminal restriction fragment lengths were determined to be 488 bases for *Commamonas*, 492 bases for *Nitrosomonas*, 515 bases for the *Chloroflexi*-related organism, and 612 bases for *Nitrospira*. Panels (A)-(D) show the same experiments as the respective panels of Fig. 3 (main text). The non-target peaks represent artifacts, which are common in T-RFLP analyses of mixed templates and may be caused by the presence of single-stranded PCR amplicons (1), the formation of heteroduplexes or chimeras (2), or by

incomplete or unspecific cleavage of the targets by the restriction enzyme (3). Error bars depict standard deviations of three replicate experiments.

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1. **Egert, M., and M. W. Friedrich.** 2003. Formation of pseudo-terminal restriction fragments, a PCR-related bias affecting terminal restriction fragment length polymorphism analysis of microbial community structure. *Appl. Environ. Microbiol.* **69**:2555-2562.
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3. **Osborn, A. M., E. R. B. Moore, and K. N. Timmis.** 2000. An evaluation of terminal-restriction fragment length polymorphism (T-RFLP) analysis for the study of microbial community structure and dynamics. *Environ. Microbiol.* **2**:39-50.