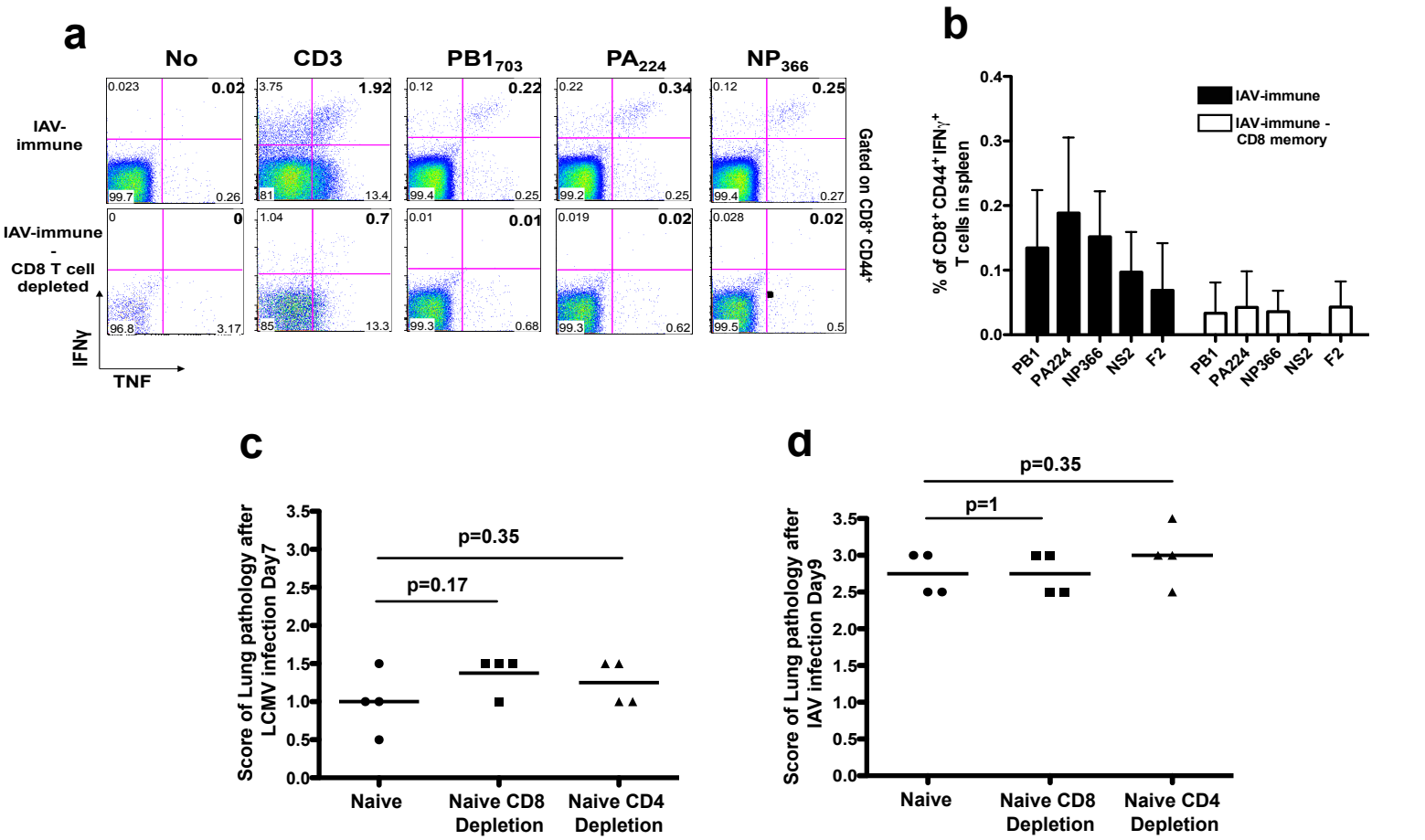
Supplemental figure 2



e **IAV-immune + LCMV**

groups	LCMV-specific CD8+ IFNg+	LCMV-specific CD4+ IFNg+
Naive	10.8% (+/- 1.7%)	1.2% (+/- 0.2%)
IAV-immune	16.8% (+/- 4.8%)	2.2% (+/- 0.8%)
IAV-immune CD8 depletion	13.5% (+/- 5.8%)	1.2% (+/- 0.5%)
IAV-immune CD4 depletion	16.2% (+/- 3.9%)	1.9% (+/- 0.8%)

f **Naive + LCMV**

groups	LCMV-specific CD8+ IFNg+	LCMV-specific CD4+ IFNg+
Naive	7.4% (+/- 5.7%)	2.9% (+/- 1.8%)
Naive CD8 depletion	12.8% (+/- 4.2%)	9.1% (+/- 5.2%)
Naive CD4 depletion	11.4% (+/- 5%)	3.8% (+/- 2.7%)

g **Naive + IAV**

groups	IAV-specific CD8+ IFNg+	IAV-specific CD4+ IFNg+
Naive	4.6% (+/- 2.9%)	1% (+/- 0.3%)
Naive CD8 depletion	4.5% (+/-1.7%)	0.7% (+/- 0.3%)
Naive CD4 depletion	1.6% (+/- 0.9%)	1% (+/- 0.3%)

Supplemental Figure S2: CD8 or CD4 depletion 4 weeks prior to LCMV or IAV infection in naïve mice does not alter either the immune response or the lung pathology. (**a,b**) CD8 memory T-cells were depleted in IAV-immune mice by i.p. injection of anti-CD8 every 3 days for 3 doses. Mice were rested for 3-4 wks to allow reconstitution of the naïve cell pool. Splenocytes from IAV-immune mice were stimulated with peptides or anti-CD3, and stained for IFNγ/TNF. Data are representative of 2 experiments (n=3-4 mice/gp/expt). (**c-g**) CD8 or CD4 T-cells were depleted in vivo by i.p. injection of either anti-CD8 or anti-CD4 mAb every 3 to 4 days for 3 doses, starting 6 weeks after immunization if mice were IAV-immune, and then rested at least 3 weeks for thymic reconstitution of T-cells. Lungs and splenocytes were harvested from control naïve, naïve CD8- or CD4-depleted mice on day 7 after LCMV-infection (**c,f**) or day 9 after IAV infection (**d,g**). Splenocytes were harvested from naïve control, IAV-immune CD4-memory -depleted and CD8-memory-depleted mice on day 7 after LCMV-infection (**e**). Lungs were stained with hematoxylin and eosin (**c,d**), and pathology was scored as described in Materials and Methods. Splenocytes were stained with CD8, CD4 and CD44 antibodies, and virus specific responses were measured by IFNγ and TNF production after virus-specific peptide stimulation in ICS assay. CD4 and CD8 frequencies were gated on live PBMC and CD8+ or CD4+ gates were used for the other frequencies shown.