ON THE COMPOSITION OF HYPER-CONSERVATION SCORE (HCS)

We defined the sequence conservation measure, which we call Hyper Conservation Score (HCS), by first selecting one of the two conservation measures defined for the 44 way alignments available at the UCSC genome browser (33). We choose as Sequence Conservation Score (SCS) the *phastCons*-derived metric (34) instead of the *phyloP* one (105), as the former considers neighboring bases in determining a base score, being thus sensible to stretches of conserved bases: this fact makes it more suitable for identifying conserved elements than *phyloP*, which instead computes conservation independently at each position. *phastCons* takes into account the phylogenetic tree to estimate the probabilities for bases to be conserved or not in the HMM models it is based upon. Nevertheless, being our aim to identify exceptionally conserved sequence stretches because of their potential functional meaning as cis components of core posttranscriptional networks, we estimated as essential the requirement for sharing of the sequences among the different vertebrate species considered. To put more weight on the phylogenetic distance, we included in our metric the Branch Length Score (BLS) as introduced in a comparison between close *Drosophila* species (35). This measure is the proportion of the distance covered by the branches of the phylogenetic tree by the alignment of a particular sequence, thus giving more importance to elements conserved across a wide range of species than to the ones restricted to a group of closely related species. We argued that, while phylogenetic information are already included in SCS, BLS would have been not redundant. To verify this we computed the Pearson correlation coefficient between SCS and BLS, obtaining a value of 0.48, which indicates only a moderate correlation of the two components of our HCS. This result confirms that the BLS usefully complements the SCS.

We further had to find a convenient measure of relative weight of SCS and BLS in HCS. We performed several runs of our pipeline, varying the SCS-BLS score composition from SCS only (100%-0%) to BLS only (0%-100%), through five intermediate proportions (80%-20%; 60%-40%; 50%-50%; 40%-60%; 20%-80%). What we obtain as result is a progressive increase in HCE sizes in parallel with a marked reduction of their total number. While more than 120000 HCEs are produced in the first two runs (100%-

0%, 80%-20%), only 3149 are retained in the half-half proportion (50%-50%), and this number goes down to just 232 HCEs for the BLS-only run. Median and average HCE lengths increase respectively from 62 and 17 bases to 114 and 249 bases: the 50%- 50% case has a median length of 23 bases and an average length of 100 bases. We selected the 50% SCS and 50% BLS composition as our final conservation measure, because of the number of selected HCEs identified a small percentage of the total UTR space (0.47%) and a corresponding small percentage of mRNAs (1.8%). With this choice we believed to have greatly reduced the number of false positives HCEs in our final dataset.

SUPPLEMENTARY REFERENCES

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Supplementary Table 1: List of the 23 HCE-containing RRM-type RBP identified by our pipeline. Listed are gene symbol, name, Uniprot gene function description and whether the protein contains only RRM or also other domains.

Figure S1. HCEs cluster in genes belonging to three different biological functions.

Ontology enrichment analysis of HCEs-containing genes highlights three groups of genes corresponding to three different biological functions. Multiple ontologies were used to infer possible functional groupings: the top results are a most significant group composed of genes involved in chromosomes assembly, a significant set consisting of 23 genes coding for RRM-containing genes for RBPs and a third, less significant group of genes playing a role in transcription. Here the ontology terms clusters giving rise to these groups are shown, along with their enrichment p-value and the final list of involved genes.

ARID1A, ARID4A, CHD1, CHD2, EPC1, EZH2, FBXL10 HAT1, HDAC9 HIST1H1A, HIST1H1B, HIST1H1C, HIST1H1D, HIST1H1E, HIST1H1T HIST1H2AB.HIST1H2AC.HIST1H2AD.HIST1H2AH.HIST1H2AJ HIST1H2AK, HIST1H2AL, HIST1H2AM, HIST1H2BB, HIST1H2BC HIST1H2BD, HIST1H2BE, HIST1H2BF, HIST1H2BG, HIST1H2BH HIST1H2BI.HIST1H2BJ.HIST1H2BL.HIST1H2BM.HIST1H2BN HIST1H3B.HIST1H3C.HIST1H3D.HIST1H3E.HIST1H3F.HIST1H3G HIST1H3H, HIST1H3I, HIST1H4A, HIST1H4B, HIST1H4C, HIST1H4D HIST1H4E, HIST1H4F, HIST1H4G, HIST1H4H, HIST1H4I, HIST1H4J HIST1H4K.HIST1H4L.HIST2H2AB.HIST2H2AC.HIST3H2A HIST3H2BB, HIST4H4, HLTF, HNRNPD, ING2, MYCN, PAM, PBX1 RAD51, RGS12, SATB1, SMARCA5, SMARCB1, SUGT1, TAF1, TBPL1 TCF7L2,TOP1,TOP2B,ZNF238

CPEB1.CUGBP1.EIF4B.ELAVL4.EWSR1.FUS.HNRNPA1 HNRNPA3, HNRNPD, HNRNPM, MSI2, PABPC1, PPARGC1B PTBP2,RBM15,RBM16,RBM26,RBMS1,RBMS3,SFRS11 SFRS15, SYNCRIP, THOC4

ACID1, ADORA2A, ALX1, ARID1A, ARID4A, ARNT, ATF2, ATPIF1 BAZ2B, BCL11A, BLZF1, BNC2, CAMK2A, CCNL1, CHD1, CHD2 CNOT2, CRX, CTCF, DKFZp434D2111, DKFZp686H10114 DKFZp686K08123,DMRTA1,EBF3,EDF1,EED,ELP4,EPC1 ESRRA, ETV1, EU036692, EWSR1, EZH2, FBXL10, FIST3, FOXP2 FRY, FST, FUBP1, GLI2, HDAC9, HES1, HIF1A, HIPK3, HIVEP1 HLTF, HNRNPD, HOXA5, HOXD1, HOXD8, IKZF5, ING2, IRX3, IRX5 KCNH5,KCNH7,KHDRBS1,KHDRBS2,KHDRBS3,KIAA0569 KIAA0916,KIAA0968,KLF10,KLF4,LHX2,LMO4,MCM5,MED25 MEIS1, MEIS2, MITF, MLL and 65 more

Figure S2. HCEs in 3'UTR of chromosome assembly genes identify SLBP binding sites.

A significant fraction of HCEs found in the 3'UTR of genes belonging to the chromosome assembly functional group was noticed to harbor a sequence corresponding to the binding motif of the stem-loop binding protein (SLBP), which is known to bind to the 3'UTR of histone genes and to stabilize the mRNA in order to compensate for the absence of a poly(A) tail. This stabilization mechanism is known to be heavily conserved and can thus be considered as a benchmark for our HCE identification method. Here the ClustalW2 alignment of these HCEs with the SLBP binding motif (the first sequence in the alignment) is displayed.

Figure S3. ¹H-NMR spectrum of SL (top) and NF (bottom). Only the downfield portion of the spectra are displayed for better visualization of the imino/amino-proton region. Spectra were acquired at 5 °C on 0.1 mM samples in 10 mM phosphate buffer, pH 7, and 10 mM NaCl. Imino proton peaks are labeled by numbers.

Figure S4. Temperature dependence of the ¹H-NMR spectrum of SL. Only the iminoproton region is displayed. Spectra were acquired with the same acquisition parameters on 0.1 mM samples in 10 mM phosphate buffer, pH 7, and 10 mM NaCl.

Figure S5. Concentration dependence of the ¹H-NMR spectrum of SL. Only the downfield portion of the spectra are displayed for better visualization of the imino/aminoproton region. Spectra were acquired at 5 °C on 0.1 mM (top) and 1.0 mM (bottom) samples in 10 mM phosphate buffer, pH 7, and 10 mM NaCl.