Supplementary Information

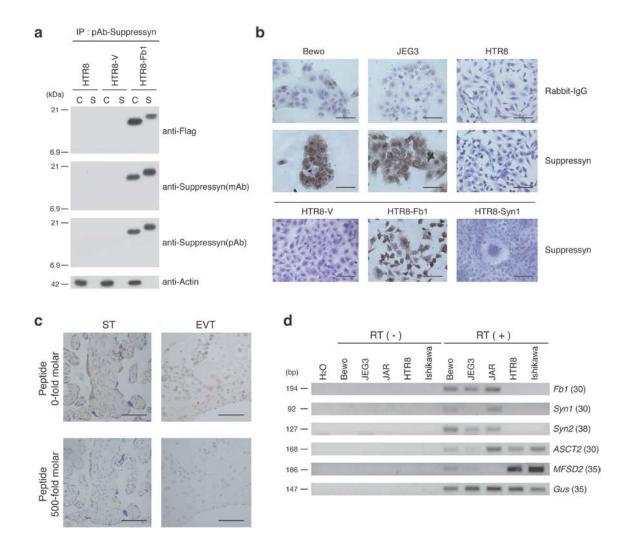
A novel human endogenous retroviral protein inhibits cell-cell fusion

Jun Sugimoto, Makiko Sugimoto, Helene Bernstein, Yoshihiro Jinno and Danny J. Schust

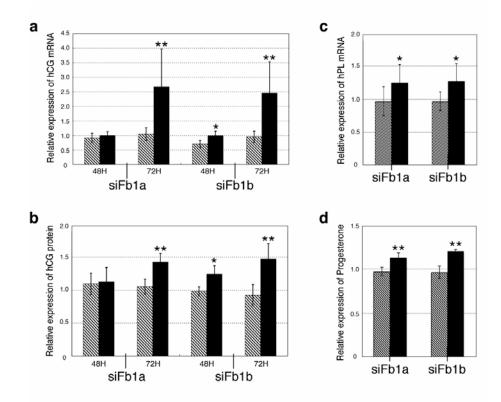
Supplementary Figures:

a			Nucleotide alignment	
			ATCTACCCAACCACTTTCTATACCTCTCTCCAACCAAAAGTCTTAATATGGGAATATCCCTCACCACGATCCTA	
			GTAGCTGTCCTGCTGTCCACAGCAGCCCCTCCGAGCTGCCGTGAGTGTTATCAGTCTTTGCACTACAGAGGGGAG	
			TACTTTACTTACCATACTCATATAGAAAGATCCTGTTATGGAAACTTAATCGAGGAATGTGTTGAATCAGGAAAG	
			AAAGTAAAGAATCTAGGAGTATGTGGCAGTCGTAATGGGGCTATTTGCCCCAGAGGGAAGCAGTGGCTTTGCTTC	
			GGACAATGGGGAGTAAACACTCAGGTGCTTGAGGACATAAAGAGAGAACAGATTATAGCCAAAGCCAAAGCCTCA	
			ACTCCCCCTGAAAATCGCCCGCGGCATTTCCATTCCTTTATACAAAAACTATAA	483 477
			Amino acid alignment	
			YTSLPTKSLNMGISLTTILILSVAVLLSTAAPPSCRECYQSLHYRGEMQQYFTYHTHIERSCYGNLIEECVESGKS	
			CGSRNGAICPRGKQWLCFTKIGQWGVNTQVLEDIKREQIIAKAKASKPTTPPENRPRHFHSFIQKL.	161 159
b	Suppressyn	40	APPSCRECYQSLHYRGEMQQYFTYHTHIERSCYGNLIEECVES	
	Syncytin-1	21	*** **	
	Suppressyn	84	KSYYKVKNLGVCGSRNGAICPRGKQWLCFTKIGQWGVNTQVLEDIKREQIIAKAKASKPT *. *. * * .* * * .* .* .*	т 144
	Syncytin-1	84	THYWTGKMINPSCPGGLGVTVCWTYFTQTGMSDGGGVQDQAREKHVKEVISQLTRVHGTS	S 144

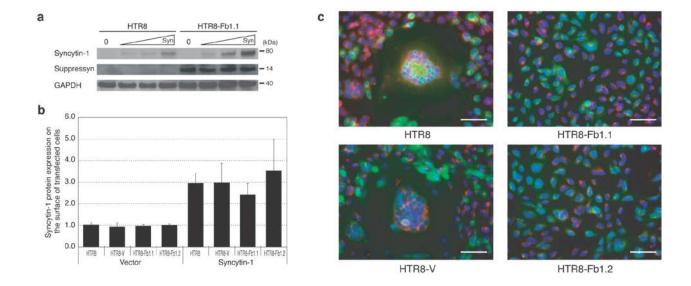
Supplementary Figure S1: Nucleotide and amino acid alignments for *HERV-Fb1* vs. a chimpanzee suppressyn homolog and suppressyn vs. syn1, respectively (a) Human and chimpanzee nucleotide and amino acid sequences are aligned to demonstrate homology. The chimpanzee amino acid sequence was derived from 28971776-28972252 (database; CGSC Build 2.2.3) genomic sequences on chromosome 21. (b) The 124 amino acid SU of syn1 is aligned with that of suppressyn. (*) identical amino acids; (.) similar amino acids. The reported ASCT-2 binding sites^{22, 27} are delimited by arrowheads. The conserved amino acid motif directly involved in syn1/ASCT-2 receptor interactions is within the box.



Supplementary Figure S2: Anti-suppressyn antibody specificity and suppressyn expression in human cell lines and primary tissues. (a) HTR8, HTR8-V and HTR8-Fb1 cell lysates (C) and supernatants (S) were immunoprecipitated with the polyclonal anti-suppressyn antibody. After separation over a 15% SDS-PAGE gel and protein transfer, suppressyn-specific detection was performed using anti-Flag, and monoclonal and polyclonal anti-suppressyn antibodies. (b) Trophoblast cell lines (Bewo, JEG3 and HTR8) were cultured under standard conditions and exposed to the suppressyn polyclonal antibody or to Rabbit-IgG (negative control) as primary reagents, then to a secondary, biotinylated anti-rabbit IgG antibody and finally to DAB for detection. Cells were counterstained with hematoxylin, mounted and visualized using standard microscopy. Transfected cells (HTR8-V, -Fb1 and –syn1) were used as additional positive and negative controls. Scale bar indicates 100μm. (c) The anti-suppressyn polyclonal antibody was incubated with 0- or 500-molar excess of a blocking peptide overnight. These antibody preparations were then used as primary antibodies for immunohistochemical analysis of third trimester villous and extravillous placental samples as in Figure 1g-1j. ST-syncytiotrophoblast, EVT-extravillous trophoblast. Scale bar indicates 100μm. (d) Conventional RT-PCR was performed using gene-specific primers (Supplementary Table 1). Reactions lacking reverse transcriptase (-) assess genomic DNA contamination. The number of amplification cycles used in a given amplification is provided in parentheses.



Supplementary Figure S3: *Fb1* **knock-down increases BeWo cell transcription and translation of secretory hormones.** hCG transcription (**a**) and secreted protein (**b**) were increased 2.5-fold (mRNA) and 1.5-fold (secreted protein) 72 hours after suppressyn knock-down when compared to parental cells or to those treated with control siRNA. Human placental lactogen (*hPL*) transcription (**c**) and progesterone secretion (**d**) were likewise increased at 72 hours post suppressyn siRNA transfection. Black bars- *Fb1* siRNA exposed; hatched bars-control siRNA exposed. Data in (**a, b, c, d**) are representative of three independent experiments performed in duplicate. Expression is normalized to unexposed samples cultured for similar time periods (48 or 72H), *p<0.05 and **p<0.01 when compared to matched siRNA control. Statistical comparisons used Kruskal-Wallis and Mann-Whitney U-testing without Bonferroni corrections.



Supplementary Figure S4: Suppressyn inhibits syn1-medited cell fusion and co-transfection does not inhibit suppressyn translation or syn1 surface expression. (a) HTR8 parent cells and HTR8 cells stably-transfected with a vector driving suppressyn expression (HTR8-Fb1.1) were transiently transfected with increasing amounts of a vector driving expression of syn1 for 72 hours. Lysates were separated by PAGE and immunoblotted with polyclonal antibodies specific for syn1 and suppressyn and a monoclonal antibody detecting GAPDH. (b) HTR8 parental cells and those stably-transfected with two independent vectors driving suppressyn expression (HTR8-Fb1.1 or -Fb1.2) or vector-only control (HTR8-V) were transiently transfected with syn1. Surface expression of syn1 was confirmed using a polyclonal antibody specific for syn1 and analyzed using flow cytometry. The relative syn1 expression ratio was normalized to that in non-transfected HTR8 cells. Data are representative of three independent experiments performed in duplicate. (c) Cells stably transfected with two independent vectors driving suppressyn expression (HTR8-Fb1.1 or -Fb1.2) or vector-only control (HTR8-V) were stained with either Cell Vue Claret (Red) or CFSE (Green) (MINCLARET and 21888; Sigma-Aldrich). Color-mixed cells were transiently transfected with a vector driving syn1 expression, fixed at 24 hrs and analyzed using a Leica DMI 6000 B fluorescence microscope. Scale bar indicates 100µm.

Supplementary Table 1: Primer and siRNA sequences.

< Cloning >					
Fb1-1st-S GATATCCAGGTGCTTATTAAAACA (21)					
Fb1-1st-AS	GTATCCATCGTGCCGCTGTAG (2315)				
Fb1flag-2nd-S	GGATATCCACAAGGAAGACTAACCACG (528)				
Fb1flag-2nd-AS	CGGATCCGGATATAGTTTTTGTATAAAGG (1040)				
Fb1-2nd-S	GGATATCCACAAGGAAGACTAACCACG (528)				
Fb1-2nd-AS	CGGGATCCCAGGAGGTTAACTGTAGTTT (2273)				
Syn1-1st-S	AACTGCGGTTAAAGTGGCTGGAGT (860)				
Syn1-1st-AS	TTGGTCAGGTGTGAGCTAAGTTGC (2835)				
Syn1-2nd-S	CGGATATCAGGATTTGCGCCTGCTCTTCAAAC (982)				
Syn1-2nd-AS	CGGGATCCCGTGTTTAAAGGTGGATGTGGT (2807)				
Syn2-1st-S	ACTTGTACACCACCAGGAGTTCCA(308)				
Syn2-1st-AS	AGCGGGTGACTTGAGAGATCCAAT(2751)				
Syn2-2nd-S	CGGGTACCATGGGCCTGCTCCTGCTGG(368)				
Syn2-2nd-AS	CCTGATCAAAGAAGGGTGACTCTTGA(1985)				
ASCT2-1st-S	AAGTTCCAGTCTCCAGGTGCTGTT(23)				
ASCT2-1st-AS	ACAGCAGGTATTTGTCCTCAGCCT(2338)				
ASCT2-2nd-S	CGGATATCATGGTGGCCGATCCTCCTC(138)				
ASCT2-2nd-AS	CCAGATCTATGACTGATTCCTTCTCA(1762)				
< Conventional and Real time PCR >					
Fb1-S	TCCGGGTTCCAACCAATGCAAGA				
Fb1-AS	TGTGCCAGTAGGCGAGATCAGT				
Gus-S	AGCAGTACCA TCTGGGTCTG				
Gus-AS	TTGGTTGTCT CTGCCGAGTG				
Syn1-S	CCACGAACGGACATCCAA				
Syn1-AS	TCCACTCCAGCCACTTTAAC				
Syn2-S	TCTCAAATGGTGCAGTGACTCGGA				
Syn2-AS	TGCTGGTTCTGGCTCTGGAGTTTA				
ASCT2-S	TCGATTCGTTCCTGGATCTTGCGA				
ASCT2-AS	ACACTACCAAGCCCAGGATGTTCA				
MFSD2-S	TCGCCTTATGCCCTGGATCATCTT				
MFSD2-AS	TCGGTGCTGATGAACATGGTGAGA				
hCG-S	CATCACCGTCAACACCACCATCT				
hCG-AS	AGGAGACCACGGGGTTCACG				
hPL-S	TGGACAGCTCACCTAGTGGCAAT				
hPL-AS	AAGCCTGGATAAGGGAACGGTTTG				
< siRNA >					
siFb1a	UGUAUCUACCCAACCACUUUCUAUA				
siFb1b	CCCGCGGCAUUUCCAUUCCUUUAUA				
siFb1a-C	Stealth RNAi negative low GC control : 12935-200				
siFb1b-C	CCCGGACUUCUACUUCCUUUCGAUA				

Numbers in parentheses indicate the 5' sequence positions of the respective genes. (HERV-Fb1: c21orf105, Syn1:

NM_014590.3, Syn2: BC068585, ASCT2: BC000062)