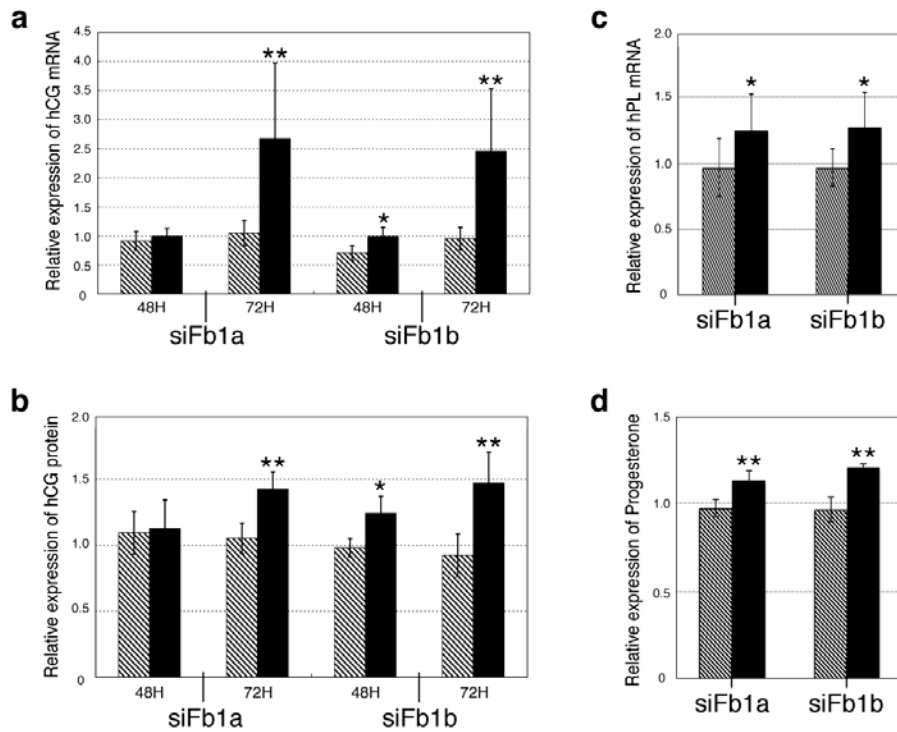
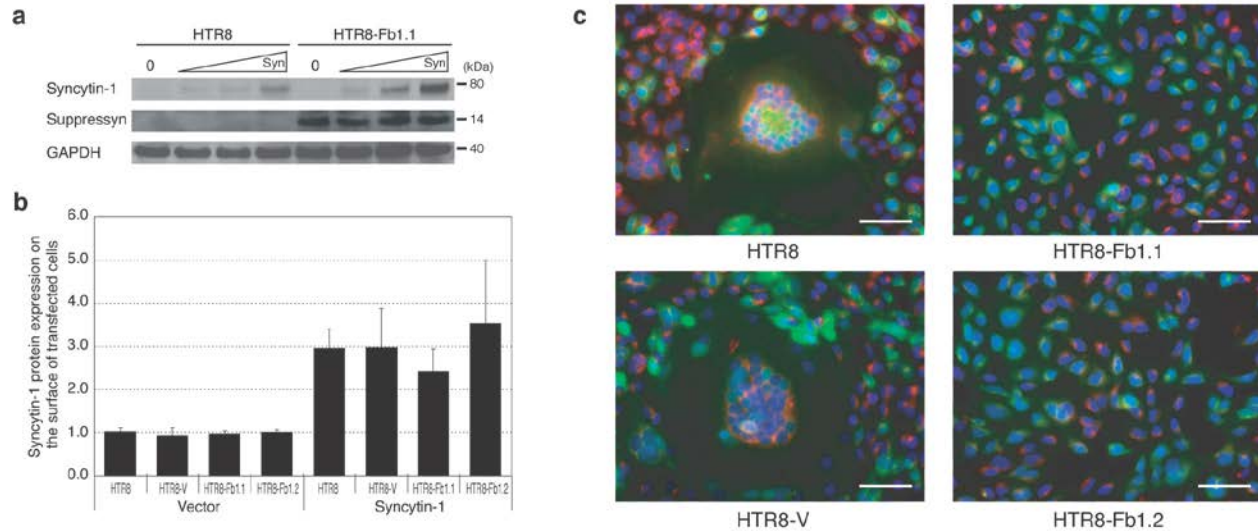


**Supplementary Figure S2: Anti-suppressyn antibody specificity and suppressyn expression in human cell lines and primary tissues.** (a) HTR8, HTR8-V and HTR8-Fb1 cell lysates (C) and supernatants (S) were immunoprecipitated with the polyclonal anti-suppressyn antibody. After separation over a 15% SDS-PAGE gel and protein transfer, suppressyn-specific detection was performed using anti-Flag, and monoclonal and polyclonal anti-suppressyn antibodies. (b) Trophoblast cell lines (Bewo, JEG3 and HTR8) were cultured under standard conditions and exposed to the suppressyn polyclonal antibody or to Rabbit-IgG (negative control) as primary reagents, then to a secondary, biotinylated anti-rabbit IgG antibody and finally to DAB for detection. Cells were counterstained with hematoxylin, mounted and visualized using standard microscopy. Transfected cells (HTR8-V, -Fb1 and -syn1) were used as additional positive and negative controls. Scale bar indicates 100µm. (c) The anti-suppressyn polyclonal antibody was incubated with 0- or 500-molar excess of a blocking peptide overnight. These antibody preparations were then used as primary antibodies for immunohistochemical analysis of third trimester villous and extravillous placental samples as in **Figure 1g-1j**. ST-syncytiotrophoblast, EVT-extravillous trophoblast. Scale bar indicates 100µm. (d) Conventional RT-PCR was performed using gene-specific primers (**Supplementary Table 1**). Reactions lacking reverse transcriptase (-) assess genomic DNA contamination. The number of amplification cycles used in a given amplification is provided in parentheses.



**Supplementary Figure S3: *Fb1* knock-down increases BeWo cell transcription and translation of secretory hormones.** hCG transcription (**a**) and secreted protein (**b**) were increased 2.5-fold (mRNA) and 1.5-fold (secreted protein) 72 hours after suppressyn knock-down when compared to parental cells or to those treated with control siRNA. Human placental lactogen (*hPL*) transcription (**c**) and progesterone secretion (**d**) were likewise increased at 72 hours post suppressyn siRNA transfection. Black bars- *Fb1* siRNA exposed; hatched bars-control siRNA exposed. Data in (**a**, **b**, **c**, **d**) are representative of three independent experiments performed in duplicate. Expression is normalized to unexposed samples cultured for similar time periods (48 or 72H), \* $p < 0.05$  and \*\* $p < 0.01$  when compared to matched siRNA control. Statistical comparisons used Kruskal-Wallis and Mann-Whitney U-testing without Bonferroni corrections.



**Supplementary Figure S4: Suppressyn inhibits syn1-mediated cell fusion and co-transfection does not inhibit suppressyn translation or syn1 surface expression.** (a) HTR8 parent cells and HTR8 cells stably-transfected with a vector driving suppressyn expression (HTR8-Fb1.1) were transiently transfected with increasing amounts of a vector driving expression of syn1 for 72 hours. Lysates were separated by PAGE and immunoblotted with polyclonal antibodies specific for syn1 and suppressyn and a monoclonal antibody detecting GAPDH. (b) HTR8 parental cells and those stably-transfected with two independent vectors driving suppressyn expression (HTR8-Fb1.1 or -Fb1.2) or vector-only control (HTR8-V) were transiently transfected with syn1. Surface expression of syn1 was confirmed using a polyclonal antibody specific for syn1 and analyzed using flow cytometry. The relative syn1 expression ratio was normalized to that in non-transfected HTR8 cells. Data are representative of three independent experiments performed in duplicate. (c) Cells stably transfected with two independent vectors driving suppressyn expression (HTR8-Fb1.1 or -Fb1.2) or vector-only control (HTR8-V) were stained with either Cell Vue Claret (Red) or CFSE (Green) (MINCLARET and 21888; Sigma-Aldrich). Color-mixed cells were transiently transfected with a vector driving syn1 expression, fixed at 24 hrs and analyzed using a Leica DMI 6000 B fluorescence microscope. Scale bar indicates 100 $\mu$ m.

**Supplementary Table 1: Primer and siRNA sequences.**

< Cloning >	
Fb1-1st-S	GATATCCAGGTGCTTATTAACA (21)
Fb1-1st-AS	GTATCCATCGTGCCGCTGTAG (2315)
Fb1flag-2nd-S	GGATATCCACAAGGAAGACTAACCACG (528)
Fb1flag-2nd-AS	CGGATCCGGATATAGTTTTGTATAAAGG (1040)
Fb1-2nd-S	GGATATCCACAAGGAAGACTAACCACG (528)
Fb1-2nd-AS	CGGGATCCCAGGAGGTTAACTGTAGTTT (2273)
Syn1-1st-S	AACTGCGGTTAAAGTGGCTGGAGT (860)
Syn1-1st-AS	TTGGTCAGGTGTGAGCTAAGTTGC (2835)
Syn1-2nd-S	CGGATATCAGGATTTGCGCCTGCTCTCAAAC (982)
Syn1-2nd-AS	CGGGATCCCGTGTAAAGGTGGATGTGGT (2807)
Syn2-1st-S	ACTTGACACCACCAGGAGTTCCA(308)
Syn2-1st-AS	AGCGGGTGACTTGAGAGATCCAAT(2751)
Syn2-2nd-S	CGGGTACCATGGGCCTGCTCCTGCTGG(368)
Syn2-2nd-AS	CCTGATCAAAGAAGGGTGACTCTTGA(1985)
ASCT2-1st-S	AAGTTCAGTCTCCAGGTGCTGTT(23)
ASCT2-1st-AS	ACAGCAGGTATTTGTCTCAGCCT(2338)
ASCT2-2nd-S	CGGATATCATGGTGGCCGATCCTCCTC(138)
ASCT2-2nd-AS	CCAGATCTATGACTGATTCTTCTCA(1762)
< Conventional and Real time PCR >	
Fb1-S	TCCGGGTTCCAACCAATGCAAGA
Fb1-AS	TGTGCCAGTAGGCGAGATCAGT
Gus-S	AGCAGTACCA TCTGGGTCTG
Gus-AS	TTGGTTGTCT CTGCCGAGTG
Syn1-S	CCACGAACGGACATCCAA
Syn1-AS	TCCACTCCAGCCACTTTAAC
Syn2-S	TCTCAAATGGTGCAGTGACTCGGA
Syn2-AS	TGCTGGTTCTGGCTCTGGAGTTA
ASCT2-S	TCGATTTCCTGGATCTTGCGA
ASCT2-AS	ACACTACCAAGCCCAGGATGTTCA
MFSD2-S	TCGCCTTATGCCCTGGATCATCTT
MFSD2-AS	TCGGTGCTGATGAACATGGTGAGA
hCG-S	CATCACCGTCAACACCACCATCT
hCG-AS	AGGAGACCACGGGTTACAG
hPL-S	TGGACAGCTCACCTAGTGGCAAT
hPL-AS	AAGCCTGGATAAGGGAACGGTTTG
< siRNA >	
siFb1a	UGUAUCUACCCAACCACUUUCUAUA
siFb1b	CCCGCGCAUUUCCAUUCUUUAUA
siFb1a-C	Stealth RNAi negative low GC control : 12935-200
siFb1b-C	CCCGGACUUCUACUUCUUUCGAUA

Numbers in parentheses indicate the 5' sequence positions of the respective genes. (*HERV-Fb1*: c21orf105, *Syn1*: NM\_014590.3, *Syn2*: BC068585, *ASCT2*: BC000062)