Supplementary Information

Discovery of Novel and Ligand-Efficient Inhibitors of Plasmodium

falciparum and Plasmodium vivax N-Myristoyltransferase

Mark D. Rackham[†], James A. Brannigan[‡], David K. Moss[§], Zhiyong Yu[†], Anthony J. Wilkinson[‡], Anthony A. Holder[§], Edward W. Tate[†], Robin J. Leatherbarrow[†]*

[†]Department of Chemistry, Imperial College London, South Kensington Campus, London. SW7 2AZ, UK.

[‡] York Structural Biology Laboratory, Department of Chemistry, University of York, York, YO10 5DD, U.K.

[§] Division of Parasitology, MRC National Institute for Medical Research, The Ridgeway, Mill Hill, London, NW7 1AA, U.K.

*r.leatherbarrow@imperial.ac.uk

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1. Additional Synthetic Schemes and Biological Results



Scheme S1. Variation of 3-Substituent of 2,3-Substituted Benzo[b]furans^a

^a Reagents and Conditions: (a) R-OH, diisopropyl azodicarboxylate, PPh₃, THF, rt, 18 h; b) 10% TFA

in DCM (v/v), rt, 2h; (c) Benzaldehyde, AcOH, NaBH(OAc)₃, rt, 18 h.

	Table S1. En	zyme Affinit	y of Various	Piperidine Re	placements
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No.	Structure	PfNMT ^{a} K _i (μ M)	$PvNMT^a$ K; (µM)	HsNMT ^{a} K _i (μ M)
S1b		>100	>100	>100
S2b		>100	nd ^b	nd ^b
S3b		>100	>100	>100

S4b	H_2N	>100	nd ^b	nd ^b
85	H C C C C C C C C C C C C C C C C C C C	>100	>100	nd ^b
S 6		>100	>100	nd ^b
S7		>100	nd ^b	nd ^b
S 8		>100	>100	nd ^b

^{*a*} Enzyme K_i values are calculated from the IC₅₀ values using the Cheng-Prusoff Equation (see Section 3.2). IC₅₀ values are the mean value of two or more determinations. Standard deviation is typically within 20% of the IC₅₀.

 b nd = not determined

2. Chemistry

2.1. General

All chemicals were purchased from Sigma-Aldrich Ltd (Gillingham, UK), Acros Organics (Geel, Belgium) and Alfa Aesar (Heysham, UK) and used without further purification. Moisture sensitive reactions were performed under nitrogen atmosphere using dried glassware, anhydrous solvents, and standard syringe/septa techniques.

Silica gel normal phase column chromatography was performed on an Isolera (Biotage, UK) automated apparatus with SNAP silica cartridges (Biotage, UK). Final compounds were purified on a Gilson semi-preparative Reverse Phase-HPLC system equipped with a HICHROM C₁₈ Column (250 x 21.2 mm), #306 pumps and a Gilson UV/Vis detector, detecting at 220 nm. The mobile phase consisted of H₂O + 0.1 % Formic acid (solvent A) and MeOH + 0.1% Formic Acid (solvent B), with an elution method of 0-2 min 50% B, 2-30 min 50%-98% B, 30-32 min 98%, 32-32.5 min 2% B at a flow rate of 12 mL/min. Following purification, the organic solvent was removed under reduced pressure and the compounds dried by lyophilisation. The purity of title compounds was verified by RP-HPLC/MS on a Waters 2767 system equipped with a photodiode array and an ESI mass spectrometer using a XBridge C18 (5 µm, 4.6 mm × 100 mm) column, equipped with an XBridge C18 guard column (5 µm, 4.6 mm × 20 mm). The following elution method was used: Gradient of solvent A and solvent B (as above): 0-10 min 5-98% B, 10-12 min 98% B, 12-13 min 98 to 5% B, 13-17 min 5% B. Flow rate: 1.2 mL/min. Purity of tested compounds was \geq 95%, unless specified.

¹H and ¹³C NMR spectra were recorded on 400 MHz and 101 MHz respectively Bruker AV instruments at room temperature and were referenced to residual solvent signals. Data are presented as follows: chemical shift in ppm, integration, multiplicity (br = broad, s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet) and coupling constants in Hz.

Mass spectra were obtained from the Mass Spectrometry Service of Department of Chemistry, Imperial College London.

2.2. Synthesis and Characterization of Compounds 2-5, 7-14b, S1-S8



<u>Ethyl-2-(2-ethoxy-2-oxoethoxy)benzoate 2</u> To a solution of ethyl-2-hydroxybenzoate (884 μ L, 6.02 mmol) in acetone (11 mL) was added potassium carbonate (2.50 g, 18.1 mmol) and ethyl bromoacetate (667 μ L, 6.02 mmol). The reaction mixture was stirred at 60 °C for 3 h. The white solid was removed by filtration and the filtrate concentrated under reduced

pressure, yielding the desired product **2** as an orange oil (1.46 g, 96%). ¹H NMR (CDCl₃, δ, ppm) 7.83 (1H, dd, *J* = 7.7, 1.8), 7.45 (1H, ddd, *J* = 8.4, 7.6, 1.8), 7.06 (1H, ddd, *J* = 7.7, 7.6, 0.7), 6.90 (1H, dd, *J* = 8.4, 0.7), 4.71 (2H, s), 4.38 (2H, q, *J* = 7.0), 4.28 (2H, q, *J* = 7.2), 1.40 (3H, t, *J* = 7.0), 1.30 (3H, t, *J* = 7.2).



Ethyl-3-hydroxybenzofuran-2-carboxylate **3** To a suspension of potassium *tert*-butoxide (1.80 g, 16.0 mmol) in tetrahydrofuran (60 mL) was added **2** (2.02 g, 8.01 mmol) in tetrahydrofuran (40 mL) slowly over 2 mins. The reaction mixture was stirred at room temperature for 15 mins, then quenched

with saturated ammonium chloride solution (100 mL). **3** was immediately extracted with 3×100 mL portions of ethyl acetate. The organic layers were combined, dried over magnesium sulfate and concentrated under reduced pressure to give desired product **3** as a yellow solid (1.56 g, 94%). ¹H NMR (CDCl₃, δ , ppm) 8.15 (1H, brs), 7.75 (1H, d, *J* = 7.9), 7.53–7.45 (2H, m), 7.31 (1H, ddd, *J* = 7.9, 6.1, 1.9), 4.50 (2H, q, *J* = 7.1), 1.47 (3H, t, *J* = 7.1).



tert-Butyl-4-((2-(ethoxycarbonyl)benzofuran-3-yl)oxy)piperidine-1carboxylate **4** To a solution of **3** (100 mg, 0.49 mmol) in tetrahydrofuran (3 mL) was added *tert*-butyl-4-hydroxypiperidine-1carboxylate (108 mg, 0.53 mmol) and triphenylphosphine (140 mg, 0.53 mmol). The reaction mixture was stirred at room temperature under nitrogen for 15 mins, and then diisopropyl azodicarboxylate (105 μL, 0.53 mmol) in tetrahydrofuran (0.5 mL) was added dropwise

over 2 mins. Reaction mixture was stirred at room temperature for 18 h, then concentrated under vacuum and crude product purified by flash chromatography to give **4** as a clear colourless oil (50 mg, 26%). ¹H NMR (CDCl₃, δ , ppm) 7.66 (1H, d, *J* = 7.9), 7.51 (1H, d, *J* = 8.4), 7.47–7.42 (1H, m), 7.31–7.24 (1H, m), 4.88–4.80 (1H, m), 4.44 (2H, q, *J* = 7.1), 3.83–3.74 (2H, m), 3.38–3.30 (2H, m), 2.00–1.80 (4H, m), 1.47 (9H, s), 1.43 (3H, t, *J* = 7.1).



<u>Ethyl 3-(piperidin-4-yloxy)benzofuran-2-carboxylate 5</u> To a solution of 4 (50 mg, 0.13 mmol) in dichloromethane (1 mL) was added trifluoroacetic acid (100 μ L) and the solution stirred at room temperature for 2 h. The reaction mixture was concentrated under reduced pressure and purified by HPLC yielding 5 as a pale yellow solid (36 mg, 97%). R_t = 8.5 min; ¹H NMR

(CDCl₃, δ , ppm) 9.07 (1H, s), 7.65 (1H, d, *J* = 7.9), 7.59–7.45 (2H, m), 7.32 (1H, apparent t, *J* = 7.9), 5.12–5.03 (1H, m), 4.46 (2H, q, *J* = 7.1), 3.71–3.54 (2H, m), 3.40–3.23 (2H, m), 2.35–2.14 (4H, m), 1.44 (3H, t, *J* = 7.1); ¹³C NMR (CDCl₃, δ , ppm) 159.39, 153.33, 146.22, 132.87, 128.97, 123.90, 122.62, 120.40, 113.25, 74.17, 61.58, 40.17, 27.56, 14.51; ESI HRMS, found 290.1399 (C₁₆H₂₀NO₄, [M+H]⁺, requires 290.1392).



<u>Ethyl-3-hydroxybenzo[*b*]thiophene-2-carboxylate 7</u> To a solution of methyl-2-mercaptobenzoate (1.63 mL, 11.9 mmol) and ethyl bromoacetate (1.32 mL, 11.9 mmol) in dry tetrahydrofuran (130 mL) at 0 °C was added potassium *tert*-butoxide (5.14 g, 71.3 mmol) gradually over 2 mins. The reaction mixture

was stirred and allowed to warm to room temperature over 15 mins, quenched with 2 M hydrochloric acid to pH 2 and diluted with 75 mL water. **7** was immediately extracted with 3×75 mL portions of ethyl acetate. The organic layers were combined, washed with 75 mL brine, dried over magnesium sulfate and concentrated under reduced pressure to give desired product **7** as a yellow solid (2.32 g, 88%). ¹H NMR (CDCl₃, δ , ppm) 10.21 (1H, s), 7.94 (1H, d, *J* = 8.0), 7.74 (1H, d, *J* = 8.0), 7.50 (1H, ddd, *J* = 8.0, 7.5, 1.4), 7.44–7.37 (1H, m), 4.43 (2H, q, *J* = 7.1), 1.43 (3H, t, *J* = 7.1).



<u>tert-Butyl-4-((2-(ethoxycarbonyl)benzo[b]thiophen-3-yl)oxy)</u> piperidine-1-carboxylate **8** To a solution of **7** (2.70 g, 12.2 mmol) in tetrahydrofuran (30 mL) was added *tert*-butyl-4-hydroxypiperidine-1carboxylate (4.89 g, 24.3 mmol) and triphenylphosphine (6.38 g, 24.3 mmol). The reaction mixture was stirred under nitrogen for 20 mins, and cooled to 0 °C and diisopropyl azodicarboxylate (4.79 mL, 24.3 mmol) in tetrahydrofuran (10 mL) was added dropwise

over 5 min. Reaction mixture was allowed to warm to room temperature and stirred for 1.5 h, then concentrated under reduced pressure and the crude product purified by flash chromatography to give **8** as a pink oil (4.77 g, 97%). ¹H NMR (CDCl₃, δ , ppm) 7.86 (1H, d, *J* = 7.9), 7.74 (1H, d, *J* = 8.0), 7.47 (1H, ddd, *J* = 8.0, 7.8, 0.8), 7.39 (1H, dd, *J* = 7.9, 7.8), 4.79 – 4.69 (1H, m), 4.38 (2H, q, *J* = 7.2), 4.01–3.86 (2H, m), 3.20–3.07 (2H, m), 2.05–1.95 (2H, m), 1.91–1.79 (2H, m), 1.48 (9H, s), 1.41 (3H, t, *J* = 7.2).



<u>Ethyl-3-(piperidin-4-yloxy)benzo[*b*]thiophene-2-carboxylate 9</u> To a solution of **8** (47.0 mg, 0.12 mmol) in dichloromethane (1.00 mL) was added trifluoroacetic acid (100 μ L) and the solution was stirred at room temperature for 2 h. The reaction mixture was concentrated under reduced pressure and purified by HPLC yielding **9** as a colourless oil (29 mg, 82%). R_t = 15.6 min;

¹H NMR (CDCl₃, δ , ppm) 8.73–8.36 (1H, s), 7.83–7.75 (2H, m), 7.52 (1H, ddd, *J* = 7.9, 7.1, 1.0), 7.44 (1H, ddd, *J* = 8.2, 7.1, 1.0), 5.00–4.94 (1H, m), 4.38 (2H, q, *J* = 7.1), 3.77–3.65 (2H, m), 3.39– 3.24 (2H, m), 2.36–2.17 (4H, m), 1.42 (3H, t, *J* = 7.1); ¹³C NMR (CDCl₃, δ , ppm) 161.9, 153.8, 138.2, 134.2, 128.4, 125.1, 123.3, 122.7, 116.3, 76.0, 61.6, 41.0, 28.2, 14.4; ESI HRMS, found 306.1173 (C₁₆H₂₀NO₃S, [M+H]⁺, requires 306.1164).



<u>3-((1-(*tert*-butoxycarbonyl)piperidin-4-yl)oxy)benzo[*b*]thiophene-2carboxylic acid **10** To a solution of **8** (2.50 g, 6.17 mmol) in methanol (30 mL) was added lithium hydroxide monohydrate (3.35 g, 79.8 mmol) and reaction mixture stirred at room temperature for 3 h. Reaction mixture was concentrated under reduced pressure, then dissolved in 100 mL water and acidifed to pH 2 with 2 M hydrochloric acid (30 mL). Precipitate</u>

removed by filtration and washed with 5×10 mL water, then dried overnight in a vacuum desiccator yielding **10** as an off-white solid (1.36 g, 58%). ¹H NMR (CDCl₃, δ , ppm) 7.87 (1H, d, J = 8.0), 7.81 (1H, d, J = 8.2), 7.53 (1H, ddd, J = 8.0, 7.0, 0.9), 7.44 (1H, dd, J = 8.2, 7.0), 4.82–4.73 (1H, m), 4.06–3.93 (2H, m), 3.15–3.03 (2H, m), 2.11–2.00 (2H, m), 1.94–1.80 (2H, m), 1.49 (9H, s).



<u>tert-butyl 4-((2-((benzyloxy)carbonyl)benzo[*b*]thiophen-3yl)oxy)piperidine-1-carboxylate **11a** To a solution of **10** (50 mg, 0.13 mmol) in dry acetonitrile (2 mL) was added hydroxybenzotriazole (27 mg, 0.20 mmol), *N*,*N*diisopropylethylamine (26 μ L, 0.16 mmol) and 1-ethyl-3-(3dimethylaminopropyl) carbodiimide hydrochloride (30 mg, 0.16 mmol) and reaction mixture stirred at room temperature</u>

for 15 mins. Phenylmethanol (17 µL, 0.16 mmol) was then added and reaction mixture stirred at room temperature for 18 h. The reaction mixture was concentrated under reduced pressure, dissolved in 10 mL saturated ammonium chloride solution, and **11a** extracted with 3×10 mL ethyl acetate. Combined organic layers were then washed with brine (10 mL), dried over magnesium sulphate, concentrated under reduced pressure and crude product purified by flash chromatography to give **11a** as a colourless oil (41 mg, 66%). ¹H NMR (CDCl₃, δ , ppm) 7.85 (1H, d, *J* = 8.0), 7.74 (1H, d, *J* = 8.2), 7.51 – 7.35 (7H, m), 5.37 (2H, s), 4.72 – 4.63 (1H, m), 3.96 – 3.83 (2H, m), 3.05 – 2.96 (2H, m), 1.97 – 1.86 (2H, m), 1.83 – 1.71 (2H, m), 1.48 (9H, s).



<u>Benzyl-3-(piperidin-4-yloxy)benzo[*b*]thiophene-2-carboxylate **12a 12a** was prepared as in **9**, replacing **8** with **11a** (40 mg, 0.09 mmol) and purified by HPLC yielding **12a** as a colourless oil (15 mg, 47%). $R_t = 10.5 \text{ min}$; ¹H NMR (CDCl₃, δ , ppm) 7.81 (1H, d, *J* = 8.0), 7.76 (1H, d, *J* = 8.1), 7.54 - 7.34 (7H, m), 5.36 (2H, s), 4.90 - 4.82 (1H, m), 3.58 - 3.48 (2H, m), 3.10 (2H, dd, *J* = 12.0, 6.0), 2.18 (4H, dd, *J* = 10.5, 5.3);</u>

¹³C NMR (CDCl₃, δ, ppm); 161.64, 154.12, 138.33, 135.61, 134.35, 128.87, 128.68, 128.44, 125.17, 123.22, 122.82, 116.00, 76.68, 67.13, 40.81, 28.24. ESI HRMS, found 368.1308 (C₂₁H₂₂NO₃S, [M+H]⁺, requires 368.1320).



tert-Butyl-4-((2-(((3-

<u>methoxybenzyl)oxy)carbonyl)benzo[*b*]thiophen-3yl)oxy)piperidine-1-carboxylate **11b** was prepared as in **11a**, replacing phenylmethanol with (3-methoxyphenyl)methanol (20 μ L, 0.16 mmol) and purified by flash chromatography yielding **11b** as a colourless oil (37 mg, 60%). ¹H NMR (CDCl₃,</u>

δ, ppm) 7.86 (1H, d, *J* = 8.0), 7.75 (1H, d, *J* = 8.2), 7.51 – 7.46 (1H, m), 7.43 – 7.37 (1H, m), 7.32 (1H, apparent t, *J* = 7.9), 7.04 (1H, d, *J* = 7.8), 7.02 – 7.00 (1H, m), 6.90 (1H, dd, *J* = 8.2, 2.3), 5.35 (2H, s), 4.74 – 4.66 (1H, m), 3.94 – 3.86 (2H, m), 3.84 (3H, s), 3.07 – 2.98 (2H, m), 1.98 – 1.88 (2H, m), 1.85 – 1.73 (2H, m), 1.48 (9H, s).



<u>3-Methoxybenzyl-3-(piperidin-4-yloxy)benzo[b]thiophene-2-</u> <u>carboxylate **12b**</u> was prepared as in **9**, replacing **8** with **11b** (34 mg, 0.07 mmol) and purified by HPLC, yielding **12b** as a colourless oil (5 mg, 18%). $R_t = 12.3$ min; ¹H NMR (CDCl₃, δ , ppm) 7.82 (1H, d, J = 8.0), 7.76 (1H, d, J = 8.1), 7.56 – 7.48 (1H, m), 7.47 – 7.40 (1H, m),

7.33 (1H, apparent t, J = 7.9), 7.04 (1H, d, J = 7.4), 7.01 – 6.98 (1H, m), 6.91 (1H, dd, J = 8.2, 2.4), 5.34 (2H, s,), 4.88 – 4.81 (1H, m), 3.84 (3H, s), 3.54 – 3.44 (2H, m), 3.11 – 3.01 (2H, m), 2.22 – 2.11 (4H, m).¹³C NMR (CDCl₃, δ , ppm); 161.48, 159.84, 153.90, 138.23, 136.98, 134.06, 129.82, 128.39, 125.07, 123.14, 122.61, 120.34, 115.87, 113.81, 113.79, 76.10, 66.82, 55.29, 41.01, 28.07. ESI HRMS, found 398.1425 (C₂₂H₂₄NO₄S, [M+H]⁺, requires 398.1426).



<u>tert-butyl 4-((2-(benzylcarbamoyl)benzo[b]thiophen-3-yl)oxy)piperidine-1-carboxylate 13a</u> To a solution of 10 (43 mg, 0.11 mmol) in dichloromethane (2 mL) was added N,N-diisopropylethylamine (22 µL, 0.13 mmol) followed by benzotriazol-1-yl-oxytripyrrolidinophosphonium hexafluorophosphate (65 mg, 0.13 mmol). After 30 mins stirring at room temperature, benzylamine (14 µL, 0.13 mmol) was added

and reaction mixture left stirring for 20 h. Reaction mixture was then diluted in 10 mL water, and **13a** was extracted with 3×10 mL dichloromethane. Combined organic layers were then washed with brine (10 mL), dried over magnesium sulphate and concentrated under reduced pressure and crude product purified by flash chromatography to give **13a** as a pale yellow oil (39 mg, 74%). ¹H NMR (CDCl₃, δ , ppm) 7.81 (1H, d, J = 7.5 Hz), 7.71 (1H, d, J = 7.3 Hz), 7.65–7.58 (1H, m), 7.47–7.29 (6H, m), 4.66 (2H, d, J = 5.5 Hz), 4.48–4.39 (1H, m), 4.00–3.82 (2H, m), 2.77–2.65 (2H, m), 1.94–1.82 (2H, m), 1.54–1.42 (11H, m).



N-Benzyl-3-(piperidin-4-yloxy)benzo[*b*]thiophene-2-carboxamide **14a**

14a was prepared as in **9**, replacing **8** with **13a** (38 mg, 0.08 mmol) and purified by HPLC yielding **14a** as a colourless oil (12 mg, 40%). $R_t = 11.2 \text{ min}$; ¹H NMR (CDCl₃, δ , ppm) 8.32 (1H, s), 7.82 (1H, d, J = 7.8 Hz), 7.69 (1H, d, J = 7.5 Hz), 7.50–7.28 (8H, m), 4.68 (2H, d, J = 5.9),

4.56–4.48 (1H, m), 3.38–3.27 (2H, m), 2.89–2.79 (2H, m), 2.16–1.99 (4H, m); ¹³C NMR (CDCl₃, δ, ppm) 161.66, 147.24, 138.21, 137.69, 133.21, 129.09, 128.40, 128.02, 127.23, 125.06, 123.85, 121.96, 77.37, 43.90, 41.61, 28.66, one aromatic sp² C not observed; ESI HRMS, found 367.1477 (C₂₁H₂₃N₂O₂S, [M+H]⁺, requires 367.1480).



<u>tert-butyl 4-((2-((3-methoxybenzyl)carbamoyl)benzo[*b*]thiophen-3yl)oxy)piperidine-1-carboxylate **13b 13b** was prepared as in **13a**, replacing benzylamine with 3-methoxybenzylamine (17 μ L, 0.13 mmol) and purified by flash chromatography yielding **13b** as a colourless oil (39 mg, 66%). ¹H NMR (CDCl₃, δ , ppm) 7.81 (1H, d, *J* = 7.8 Hz), 7.75 – 7.69 (1H, m), 7.62 (1H, t, *J* = 5.6 Hz), 7.47 – 7.35</u>

(2H, m), 7.32 – 7.27 (1H, m), 6.96 (1H, d, *J* = 7.6 Hz), 6.94 – 6.91 (1H, m), 6.85 (1H, dd, *J* = 8.3, 2.3 Hz), 4.64 (2H, d, *J* = 5.6 Hz), 4.49 – 4.38 (1H, m), 4.04 – 3.85 (2H, m), 3.81 (3H, s), 2.80 – 2.68 (2H, m), 1.98 – 1.85 (2H, m), 1.60 – 1.50 (2H, m), 1.47 (9H, s).

 $\begin{array}{l} \underset{N=0}{\overset{N-(3-Methoxybenzyl-3-(piperidin-4-yloxy)benzolb]}{\overset{N-(3-Methoxybenzyl-3-(piperidin-4-yloxybenzyl-3-(20-(2H, m), 6.98(1H, d, J = 7.6 Hz), 6.98(1H, d, J = 7.6 Hz), 6.96(2H, d, J = 5.9 Hz), 4.58 - 4.47(1H, m), 3.80(3H, s), 3.46 - 3.29(2H, m), 2.98 - 2.78(2H, m), 2.21 - 2.02(4H, m); {}^{13}C NMR (CDCl_3, \delta, ppm) \\ 161.61, 160.11, 147.24, 139.76, 137.61, 133.29, 130.16, 127.25, 125.09, 124.70, 123.80, 121.98, 120.48, 113.98, 113.19, 76.10, 55.40, 43.74, 41.66, 28.59; ESI HRMS, found 397.1581 \\ (C_{22}H_{25}N_2O_3S, [M+H])^+, requires 397.1586). \\ \end{array}{}$



<u>tert-Butyl 4-(((2-(ethoxycarbonyl)benzofuran-3-yl)oxy)methyl)piperidine-1-</u> <u>carboxylate **S1a**</u> **S1a** was prepared as in **4**, replacing *tert*-butyl-4hydroxypiperidine-1-carboxylate with *tert*-butyl 4-(hydroxymethyl)piperidine-1carboxylate (115 mg, 0.53 mmol), resulting in **S1a** as a clear colourless oil (65 mg, 33%). ¹H NMR (CDCl₃, δ , ppm) 7.71 (1H, d, *J* = 7.9), 7.50 (1H, d, *J* = 8.4), 7.47 – 7.41 (1H, m), 7.26 (1H, t, *J* = 7.9), 4.43 (2H, q, *J* = 7.1), 4.28 (2H, d, *J* = 6.3), 4.23 – 4.08 (2H, m), 2.82 – 2.68 (2H, m), 2.09 – 1.96 (1H, m), 1.92 –

1.84 (2H, m), 1.46 (9H, s), 1.41 (3H, t, *J* = 7.1), 1.39 – 1.27 (2H, m).



<u>Ethyl 3-(piperidin-4-ylmethoxy)benzofuran-2-carboxylate S1b</u> S1b was prepared as in **5**, replacing **4** with S1a (65 mg, 0.16 mmol), resulting in S1b as a clear colourless oil (48 mg, 71%). $R_t = 8.3 \text{ min;}^{1}\text{H} \text{ NMR} (\text{CDCl}_3, \delta, \text{ppm})$ 9.04 (1H, s), 8.61 (1H, s), 7.70 (1H, d, *J* = 7.9), 7.56 – 7.43 (2H, m), 7.34 – 7.25 (1H, m), 4.43 (2H, q, *J* = 7.1), 4.35 (2H, d, *J* = 5.4), 3.66 – 3.44 (2H, m), 3.11 – 2.92 (2H, m), 2.29 – 2.06 (3H, m), 1.91 – 1.72 (2H, m), 1.43 (3H, t, *J* =

7.1); ¹³C NMR (CDCl₃, δ, ppm) 159.58, 153.26, 148.64, 131.49, 128.81, 123.53, 122.25, 120.79, 113.07, 77.34, 61.35, 44.11, 34.85, 25.42, 14.49; ESI HRMS, found 304.1551 (C₁₇H₂₂NO₄, [M+H]⁺, requires 304.1549).



Ethyl 3-(2-((*tert*-butoxycarbonyl)amino)ethoxy)benzofuran-2-carboxylate **S2a** S2a was prepared as in 4, replacing *tert*-butyl-4-hydroxypiperidine-1carboxylate with *tert*-butyl (2-hydroxyethyl)carbamate (86 mg, 0.53 mmol), resulting in S2a as a clear colourless oil (60 mg, 35%). ¹H NMR (CDCl₃, δ, ppm) 7.72 – 7.68 (1H, d, J = 7.9 Hz), 7.53 – 7.47 (1H, m), 7.47 – 7.42 (1H, m), 7.31 – 7.24 (1H, m), 5.61 – 5.49 (1H, brs), 4.50 – 4.43 (4H, m), 3.58 – 3.48 (2H, q, *J* = 5.2 Hz), 1.48 – 1.43 (12H, m).



Ethyl 3-(2-aminoethoxy)benzofuran-2-carboxylate **S2b S2b** was prepared as in **5**, replacing **4** with **S2a** (58 mg, 0.17 mmol), resulting in **S2b** as a clear colourless oil (30 mg, 72%). $R_t = 8.0 \text{ min}$; ¹H NMR (CDCl₃, δ, ppm) 8.57 (2H, brs), 7.67 (1H, d, *J* = 7.9), 7.57 – 7.45 (2H, m), 7.36 – 7.28 (1H, m), 4.62 (2H, m), 4.41 (2H, q, *J* = 7.1), 3.39 (2H, m), 1.41 (3H, t, *J* = 7.1); ¹³C NMR (CDCl₃,

δ, ppm) 160.35, 153.42, 147.91, 133.11, 129.25, 124.01, 121.68, 120.30, 113.26, 70.53, 62.15, 39.41, 14.26; ESI HRMS, found 250.1083 (C₁₃H₁₆NO₄, [M+H]⁺, requires 250.1079).



Ethyl 3-(3-((*tert*-butoxycarbonyl)amino)propoxy)benzofuran-2carboxylate S3a S3a was prepared as in 4, replacing *tert*-butyl-4hydroxypiperidine-1-carboxylate with *tert*-butyl (2hydroxypropyl)carbamate (93 mg, 0.53 mmol), resulting in S2a as a clear yellow oil (77 mg, 44%). ¹H NMR (CDCl₃, δ , ppm) 7.77 – 7.71 (1H, d, J = 7.9 Hz), 7.54 – 7.49 (1H, d, J = 8.4 Hz), 7.48 – 7.42 (1H, m), 7.30 –

7.24 (1H, m), 5.36 – 5.27 (1H, m), 4.54 – 4.49 (2H, t, *J* = 5.8 Hz), 4.49 – 4.42 (2H, q, *J* = 7.2 Hz), 3.47 – 3.38 (2H, dt, *J* = 5.6 Hz), 2.08 – 2.00 (2H, m), 1.47 – 1.40 (12H, m).



<u>Ethyl 3-(2-aminopropoxy)benzofuran-2-carboxylate S3b</u> S3b was prepared as in 5, replacing 4 with S3a (73 mg, 0.20 mmol), resulting in S3b as a clear pale green oil (44 mg, 83%). $R_t = 7.9$ min; ¹H NMR (CDCl₃, δ , ppm) 8.31 (2H, brs), 7.73 (1H, d, J = 7.9), 7.59 – 7.45 (2H, m), 7.36 – 7.28 (1H, m), 4.62 (2H, t, J =5.3), 4.42 (2H, q, J = 7.1), 3.41 (2H, m), 2.43 – 2.24 (2H, m), 1.41 (3H, t, J =

7.1); ¹³C NMR (CDCl₃, δ, ppm) 160.29, 153.41, 148.84, 131.21, 129.09, 123.77, 121.19, 121.00, 113.35, 73.11, 61.91, 39.20, 26.92, 14.34; ESI HRMS, found 264.1228 (C₁₄H₁₈NO₄, [M+H]⁺, requires 264.1236).



Ethyl 3-(3-((tert-butoxycarbonyl)amino)butoxy)benzofuran-2-carboxylate

<u>S4a</u> S4a was prepared as in 4, replacing *tert*-butyl-4-hydroxypiperidine-1carboxylate with *tert*-butyl (2-hydroxybutyl)carbamate (101 mg, 0.53 mmol), resulting in **S4a** as a clear yellow oil (130 mg, 72%). ¹H NMR (CDCl₃, δ, ppm) 7.74 – 7.69 (1H, d, J = 7.9 Hz), 7.51 – 7.46 (1H, d, J = 8.5Hz), 7.46 – 7.40 (1H, m), 7.28 – 7.22 (1H, m), 4.81 – 4.72 (1H, m), 4.48 –

4.39 (4H, m), 3.26 - 3.17 (2H, m), 1.91 - 1.81 (2H, m), 1.79 - 1.70 (2H, m), 1.45 - 1.38 (12H, m).



<u>Ethyl 3-(2-aminobutoxy)benzofuran-2-carboxylate</u> **S4b** was prepared as in **5**, replacing **4** with **S4a** (135 mg, 0.358 mmol), resulting in **S4b** as a clear colourless oil (22 mg, 22%). $R_t = 8.0 \text{ min}$; ¹H NMR (CDCl₃, δ , ppm) 7.98 (2H, brs), 7.72 (1H, d, J = 7.9), 7.55 – 7.41 (2H, m), 7.33 – 7.24 (1H, m), 4.58 – 4.46 (2H, m), 4.39 (2H, q, J = 7.1), 3.19 (2H, m), 2.09 – 1.92 (4H, m), 1.40

(3H, t, J = 7.1); ¹³C NMR (CDCl₃, δ , ppm) 160.06, 153.35, 148.89, 130.85, 128.87, 123.55, 121.59, 121.23, 113.20, 73.71, 61.54, 39.76, 26.55, 24.65, 14.36; ESI HRMS, found 278.1392 (C₁₅H₂₀NO₄, [M+H]⁺, requires 278.1392).



<u>Ethyl 3-(3-(benzylamino)propoxy)benzofuran-2-carboxylate **S5** To a solution of **S3b** (20 mg, 0.08 mmol) in anhydrous tetrahydrofuran (1 mL) was added acid acetic (9 μ L, 0.15 mmol) and benzaldehyde (9 μ L, 0.09 mmol) and reaction mixture stirred at room temperature for 1.5 h. NaBH(OAc)₃ was then added (97 mg, 0.46 mmol) and suspension</u>

was stirred at room temperature for a further 18 h. Reaction mixture was diluted with 5% NaHCO₃ (aq) (5 mL) then extracted with 3 x 5 mL EtOAc. Combined organic layers were then washed with brine (10 mL), dried over sodium sulphate, concentrated under reduced pressure and crude product purified by HPLC to give **S5** as a white solid (18 mg, 67%). $R_t = 8.4$ min; ¹H NMR (CDCl₃, δ , ppm) 9.96 (1H, brs), 7.71 (1H, d, J = 8.0), 7.56 – 7.47 (4H, m), 7.41 – 7.35 (3H, m), 7.34 – 7.28 (1H, m), 4.55 (2H, t, J = 5.5), 4.40 (2H, q, J = 7.1), 4.33 (2H, brs), 3.41 – 3.32 (2H, m), 2.43 – 2.32 (2H, m), 1.43 (3H, t, J = 7.1); ¹³C NMR (CDCl₃, δ , ppm) 160.43, 153.49, 149.11, 131.29, 130.98, 130.39, 130.22, 129.56, 129.25, 129.22, 123.87, 120.95, 113.36, 72.88, 61.93, 52.00, 45.92, 25.99, 14.47; ESI HRMS, found 354.1708 (C₂₁H₂₄NO₄, [M+H]⁺, requires 354.1705).



Ethyl 3-((1-benzylpiperidin-4-yl)oxy)benzofuran-2-carboxylate S6 S6 was prepared as in 4, replacing *tert*-butyl-4-hydroxypiperidine-1carboxylate with 1-benzylpiperidin-4-ol (102 mg, 0.53 mmol) and purifying by HPLC, resulting in S6 as an orange oil (19 mg, 10%). $R_t =$ 8.6 min; ¹H NMR (CDCl₃, δ , ppm) 7.58 – 7.53 (2H, m), 7.52 – 7.37 (6H, m), 7.34 – 7.28 (1H, m), 5.11 (1H, s), 4.46 (2H, q, *J* = 7.1), 4.26

(2H, s), 3.51 - 3.36 (4H, m), 2.45 - 2.27 (2H, m), 2.26 - 2.12 (2H, m), 1.47 (3H, t,*J* $= 7.1); ¹³C NMR (CDCl₃, <math>\delta$, ppm) 159.33, 153.34, 146.07, 132.86, 131.18, 130.30, 129.52, 129.02, 128.48, 123.91, 121.98, 120.41, 113.44, 72.54, 61.47, 61.44, 47.09, 27.55, 14.63; ESI HRMS, found 380.1860 (C₂₃H₂₆NO₄, [M+H]⁺, requires 380.1862).



<u>Ethyl 3-(cyclohexyloxy)benzofuran-2-carboxylate S7</u> S7 was prepared as in 4, replacing *tert*-butyl-4-hydroxypiperidine-1-carboxylate with cyclohexanol (56 μ L, 0.53 mmol) and purifying by normal phase silica gel column chromatography, resulting in S7 as a colourless oil (28 mg, 20%). 50g SNAP column, 1 column volume 2% EtOAc in *n*-hexane, 10 column volumes 2%-

10% EtOAc, 2 column volume 10% EtOAc. $R_t = 5.1$ column volumes; ¹H NMR (CDCl₃, δ , ppm) 7.70 (1H, d, J = 7.8), 7.51 (1H, d, J = 8.5), 7.48 – 7.41 (1H, m), 7.31 – 7.24 (1H, m), 4.66 – 4.55 (1H, m), 4.45 (2H, q, J = 7.1), 2.11 – 1.98 (2H, m), 1.92 – 1.79 (2H, m), 1.76 – 1.63 (2H, m), 1.61 – 1.51 (1H, m), 1.44 (3H, t, J = 7.1), 1.41 – 1.31 (3H, m); ¹³C NMR (CDCl₃, δ , ppm) 159.67, 153.44, 147.55, 133.04, 128.31, 123.78, 123.26, 121.16, 112.92, 82.43, 61.10, 32.54, 25.59, 23.67, 14.55; ESI HRMS, found 289.1431 (C₁₇H₂₁O₄, [M+H]⁺, requires 289.1440).



(±)-*tert*-Butyl 3-((2-(ethoxycarbonyl)benzofuran-3-yl)oxy)piperidine-1carboxylate **S8a S8a** was prepared as in **4**, replacing *tert*-butyl-4hydroxypiperidine-1-carboxylate with *tert*-butyl-3-hydroxypiperidine-1carboxylate (107 mg, 0.53 mmol) resulting in **S8a** as a clear yellow oil (31 mg, 16%). **S8a** was carried through to **S8b** without further purificaton.



<u>Ethyl 3-(piperidin-3-yloxy)benzofuran-2-carboxylate **S8b**</u> **S8b** was prepared as in **5**, replacing **4** with **S8a** (30 mg, 0.08 mmol), resulting in **S8b** as a clear colourless oil (2.6 mg, 12%). $R_t = 8.3 \text{ min}$; ¹H NMR (CDCl₃, δ , ppm) 7.95 (1H, brs), 7.65 (1H, d, J = 7.8), 7.60 – 7.50 (2H, m), 7.35 (1H, dd, J = 7.8, 7.2), 4.94 (1H, brs), 4.47 (2H, q, J = 7.1), 3.62 – 3.47 (1H, m), 3.47 – 3.38 (1H, m), 3.37 – 3.17 (2H, m), 2.49 – 2.30 (2H, m), 2.09 – 1.85 (2H, m), 1.46 (3H, t, J = 7.1);

¹³C NMR (CDCl₃, δ, ppm) 160.72, 153.59, 146.02, 133.87, 129.41, 124.13, 121.78, 120.18, 113.50, 74.04, 62.48, 45.80, 43.94, 27.63, 17.92, 14.33; ESI HRMS, found 290.1389 ($C_{16}H_{20}NO_4$, [M+H]⁺, requires 290.1392).

3. Biochemical Data

3.1. Assay Information

All IC₅₀ determinations were carried out using a 7-diethylamine-3-(4'maleimidylphenyl)-4methylcoumarin (CPM) fluorescence assay, as described by Goncalves *et al.*^{1,2} The HsNMT1 and PvNMT assays are as described in the literature, for PfNMT the final enzyme concentration and peptide substrates are modified, see below:

PfNMT Final Concentration: 400 ng/mL

PfNMT Peptide Substrate: *Homo sapiens* $p60^{src}$ (2-16), final concentration 4.0 μ M, Sequence: GSNKSKPKDASQRRR-NH₂.

 IC_{50} values are the mean value of two or more determinations, standard deviation is typically within 20% of the IC_{50} . Data were elaborated using Microsoft Office Excel 2010 and IC_{50} values were determined using GraFit 7.0 (Erithacus Software Ltd, UK) by non-linear regression fitting.

3.2. K_i Calculations

 K_i values were calculated from the experimentally determined IC₅₀ values, the substrate concentration ([S]) and the Michaelis-Menten constant (K_m) as described by the Cheng-Prusoff equation:³

Equation 1. Cheng-Prusoff Equation for Determination of K_i from IC₅₀

$$K_i = \frac{IC_{50}}{1 + \frac{[S]}{K_m}}$$

For example, **9** had an experimentally determined IC₅₀ of $3.9 \pm 0.7 \mu$ M. The Michaelis Constant (K_m) was 2.0 μ M and the substrate concentration was 4.0 μ M, giving a K_i of 1.3 μ M.

3.3 P. falciparum Inhibition Assay

Measurement of the ability of compounds to kill parasites was performed in 96-well plates using a modification of the fluorescence-activated cell sorting (FACS) assay platform:^{4,5} each well (in total 100 μ L medium) contained synchronous cultures of late trophozoite-stage parasites (1% parasitemia and 2% hematocrit) and variable concentrations of an inhibitor in DMSO diluent or diluent alone at 0.5% final concentration. The mixture was incubated at 37 °C for a full 48-hour growth cycle. Aliquots of 50 μ L were removed from each well, added to 500 μ L freshly diluted hydroethidine (HE, 1:200 dilution of 10 mg/mL DMSO stock in PBS) and incubated for 20 min at 37°C. Samples were then diluted with 1 mL PBS to enable appropriate cell counts (50,000) and stored on ice. Parasites cultured in the absence of an inhibitor and non-infected red blood cells were used as positive and negative controls respectively. Parasitemia was measured using a FACS-Calibur flow cytometer. Growth inhibition at each concentration was calculated as % inhibition = [1-(readout-negative control) / (positive-negative control)] x 100%. To determine the EC₅₀ of an inhibitor, its activity at a range of concentrations from 10 μ M down to 0.31 μ M was measured and the 50% inhibitory concentration (EC₅₀) of an inhibitor was calculated by a nonlinear regression analysis using GraFit 7.0.1 version (Erithacus Software Limited, UK). All assays were carried out in triplicate.

4. Protein crystallization, X-ray data collection, processing and refinement

4.1 Experimental Procedure

Crystals of the ternary complex of the non-hydrolysable co-factor and compound 12b bound to PvNMT were obtained essentially as described previously.¹ X-ray diffraction data were collected on beamline ID14-1⁶ at the European Synchrotron Research Facility, Grenoble and processed using XDS^7 and $SCALA^8$ implemented within *xia2.*⁹ Data collection and refinement statistics are summarized in Supplementary Table S2. The data set is essentially complete (98%) to 1.9 Å resolution, tapering to 43 % completeness in the outer resolution shell at 1.63 Å spacing. For $R_{\rm free}$ calculations, 5% of the data were excluded. Rigid body refinement using maximum likelihood methods implemented in REFMAC¹⁰ using the protein chains of 4TST.pdb⁶ as a starting model was followed by refinement using anisotropic temperature factors interspersed with cycles of model building and adjustment using COOT.¹¹ Complete chains (corresponding to residues 27-410, numbering as in full-length protein) can be traced for two of the three molecules in the asymmetric unit. N-terminal residues (derived from the purification tag) in all three chains and loop residues 227-238 in chain C have not been modeled and these are assumed to be disordered. Electron density maps in the vicinity of the bound ligand are well-defined for the piperidine ring and the benzothiophene nucleus, but are less clear around the phenoxymethyl portion of the molecule. There appears to be alternate binding conformations for this part of the ligand, as reflected by the refined atomic B-factors (Table S3). The best fit to the electron density for this part of the molecule has been modeled in all three chains in the asymmetric unit and for purposes of illustration, the form bound to protein chain B is referred to in the main text. The final refined protein structure model displays good geometry with 96.2% of the residues in the preferred region of the Ramachandran plot and only 0.3% (corresponding to amino acid residue Phe336 in all three chains) as outliers. The coordinates and structure factor files have been deposited in the Protein Data Bank under the accession code 4BBH.

4.2. X-Ray Data Collection and Refinement Statistics

Table S2. X-ray data collection and refinement statistics

PDB accession code	4BBH
Cell dimensions a, b, c	57.47, 121.87, 178.34
Space Group	P2 ₁ 2 ₁ 2 ₁
Data collection	
Beamline / Wavelength	ESRF ID14-4 / 0.9393
Detector type	ADSC Q315r CCD
Images x oscillation (°)	360 x 0.5 & 180 x 1
Resolution (Å)	72–1.63 (1.72–1.63) ^a
R _{sym} (%) ^b	10.0 (34.7)
Ι/σΙ	15.4 (2.5)
Completeness (%)	83.7 (43.9) ^c
Redundancy	8.3 (2.8)
Refinement	
No. unique reflections	130696
$R_{\rm work} / R_{\rm free}^{d}$	21.4 / 26.7
No. atoms	10684
Protein	9597
Ligand	84
Co-factor	192
Water	788
B-factors (Å ²)	
All atoms	13.5
Protein (A/B/C)	13.2/12.3/13.2
Ligand (A/B/C)	26.4/28.5/26.9
Co-factor (A/B/C)	9.7/9.0/9.7
Water	19.5
R.m.s.deviations ^e	
Bond lengths (Å)	0.020
Bond angles (°)	2.179

PvNMT-NHM-**12b**

^aHighest resolution shell is shown in parentheses.

 ${}^{b}R_{sym} = \Sigma_{h}\Sigma_{l} |I_{h/r} < I_{h} > |/ \Sigma_{h}\Sigma_{l} < I_{h} >$, where I_{l} is the l^{th} observation of reflection h and $< I_{h} >$ is the weighted average intensity for all observations l of reflection h.

^c Data is 98% complete to 1.9 Å resolution

^d $R_{cryst} = \sum ||F_o| - |F_c|| / \sum |F_o|$ where F_o and F_c are the observed and calculated structure factor amplitudes, respectively. R_{free} is the R_{cryst} calculated with 5% of the reflections omitted from refinement.

^e Root-mean-square deviation of bond lengths or bond angles from ideal geometry.

Atom	Α	В	С
02	31.65	21.79	26.24
C7	25.96	23.54	26.80
01	32.04	27.64	36.34
C6	40.00	37.58	43.33
C5	47.00	48.70	42.75
C4	50.65	53.14	40.81
C3	47.04	51.59	44.65
C2	44.45	54.40	44.79
C21	48.42	57.91	46.92
C1	44.57	56.26	45.16
0	38.14	53.56	39.90
С	32.93	46.49	28.33
C8	21.23	19.53	22.32
C15	18.07	17.29	19.92
C14	15.48	15.71	19.82
C13	14.08	14.99	17.54
C12	13.42	13.77	17.64
C11	13.00	13.96	14.82
C10	14.40	14.71	16.26
C9	16.01	16.07	17.68
S	18.88	19.48	21.80
03	16.13	16.71	18.42
C16	16.21	17.16	16.71
C20	15.79	16.70	17.01
C19	15.32	15.66	16.85
Ν	15.29	17.05	17.20
C18	15.43	19.41	15.87
C17	17.31	15.91	16.38
Average	26.39	28.45	26.87



The B-factor (Å²) is given by $B = 8\pi^2 \langle \mu^2 \rangle$ where $\langle \mu^2 \rangle$ is the mean square displacement of the atom position. The higher values for atoms in the phenoxmethyl ring indicate that they are in a flexible portion of the molecule

5. References

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