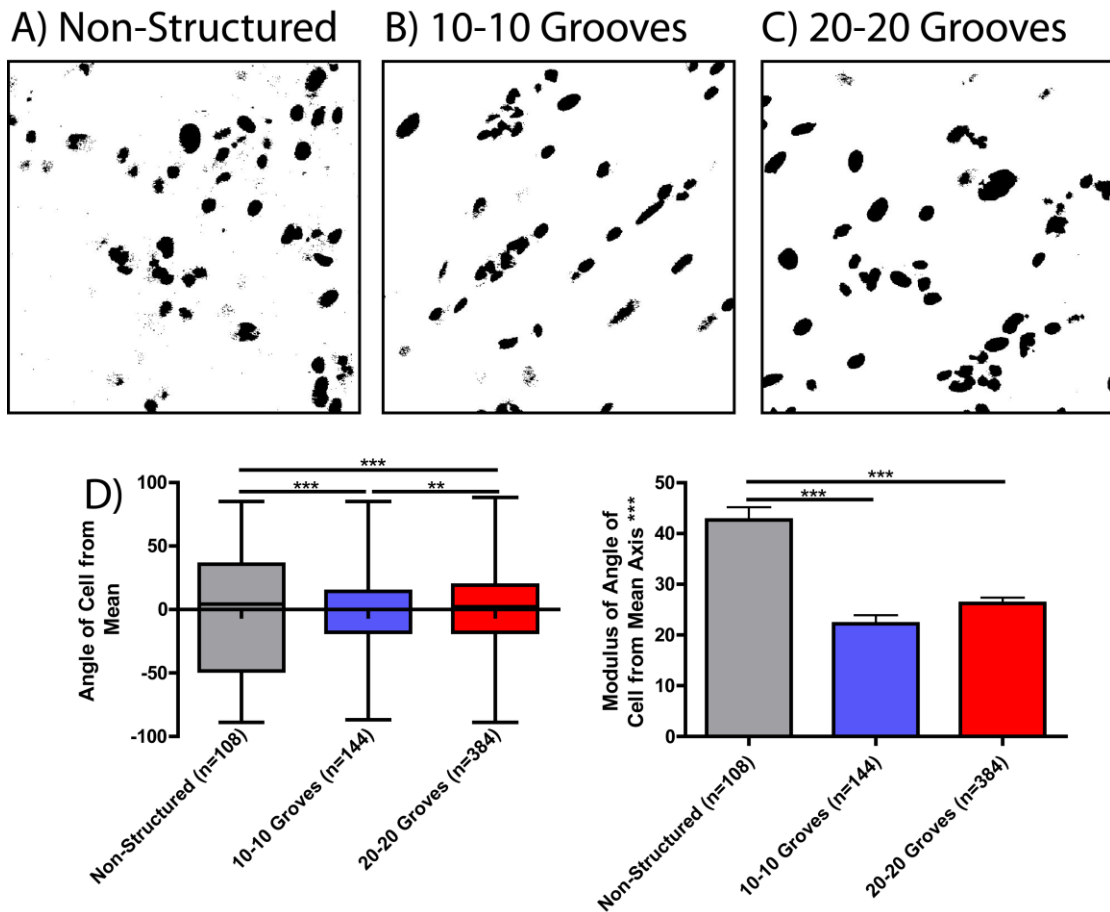


Supplemental material

Microgrooved Culture Substrates affect Calcium Cycling of Cardiac Myocytes

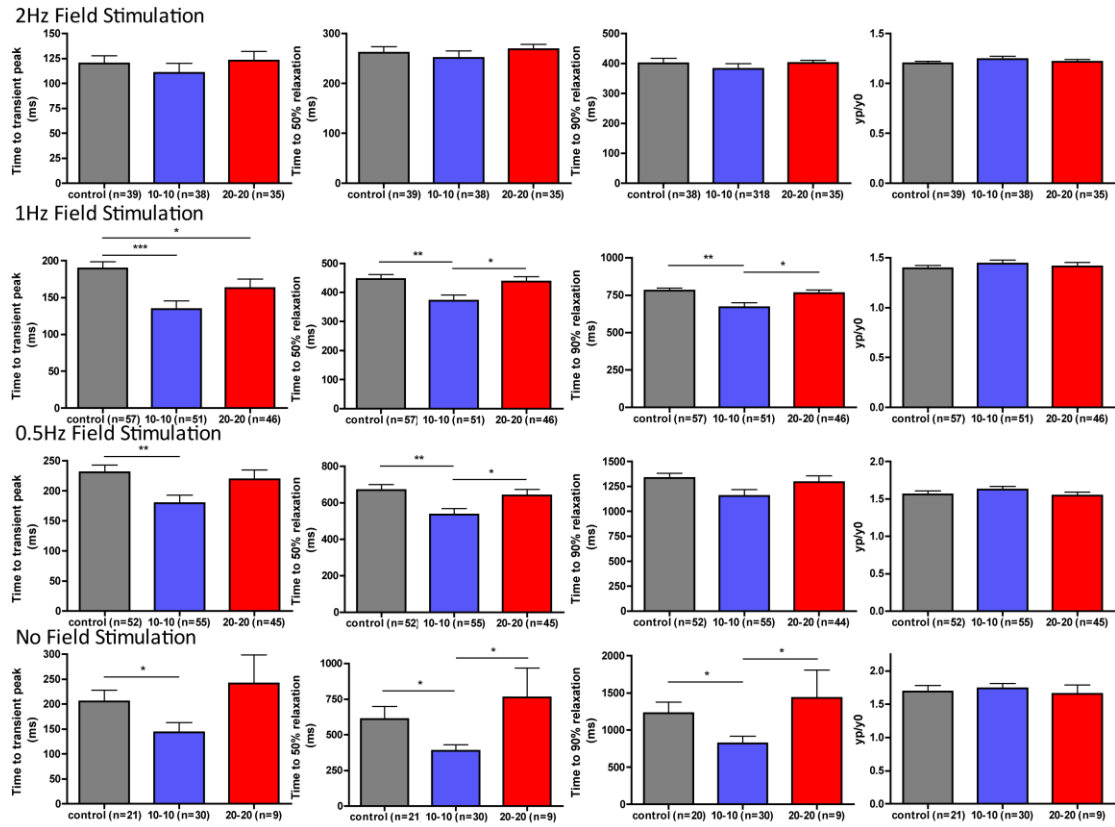
Derived from Human Induced Pluripotent Stem Cells

Rao et al., 2012



Supplemental Figure 1. Quantification of cell alignment of NRVM on microgrooved PDMS constructs. Immunohistochemistry DAPI images are converted into binary images (see representative examples A-C), alignment is quantified by measuring variation of the long-axis of each nucleus from the mean long-axis.

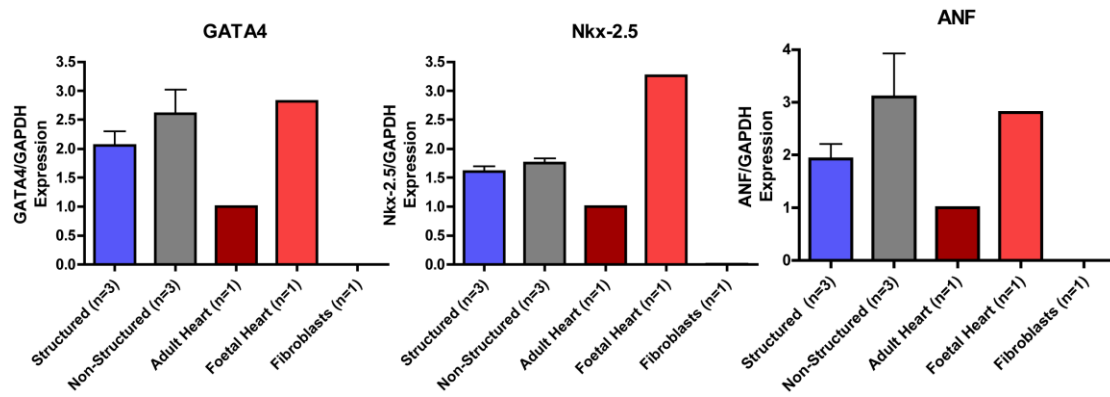
Effect of Microgrooved Tissue Culture Substrates on iPSC derived Cardiomyocytes



Supplemental Figure 2.

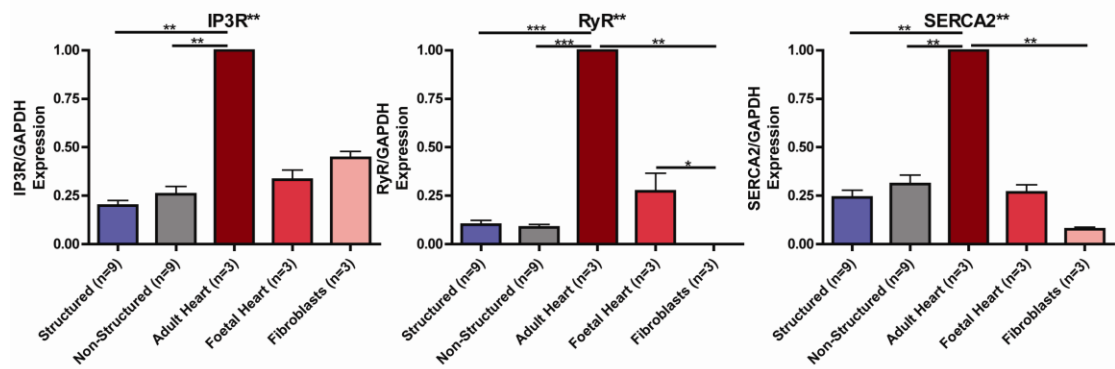
Time to Ca^{2+} transient peak (tP), 50% decay (t50), 90% decay (t90), and amplitude of fluorescence (fp/f0) of NRVM cultured on unstructured PDMS, 10-10 and 20-20 grooved constructs field-stimulated at 2Hz, 1Hz, 0.5Hz, and beating spontaneously

Effect of Microgrooved Tissue Culture Substrates on iPSC derived Cardiomyocytes

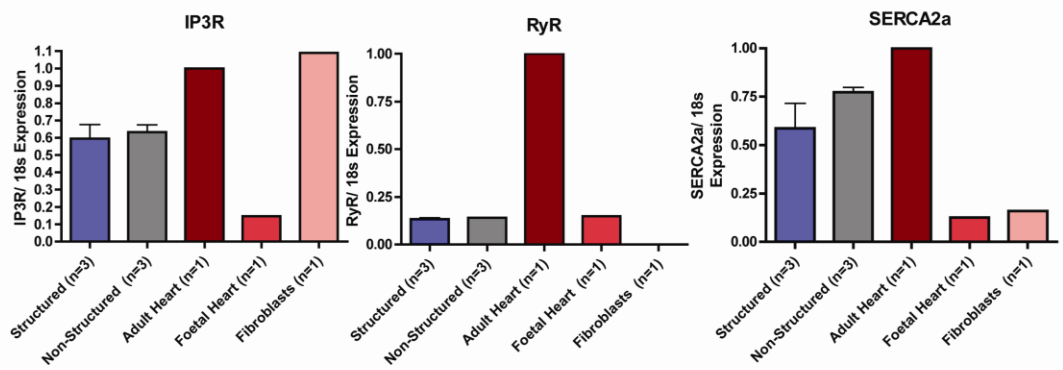


Supplemental Figure 3. Comparison of genetic markers of differentiation (GATA4, NKX2.5, and ANF) when normalized to GAPDH and expressed relative to adult heart tissue in iPSC-CM cultured on structured and non-structured PDMS, fibroblasts, adult heart and foetal heart tissue.

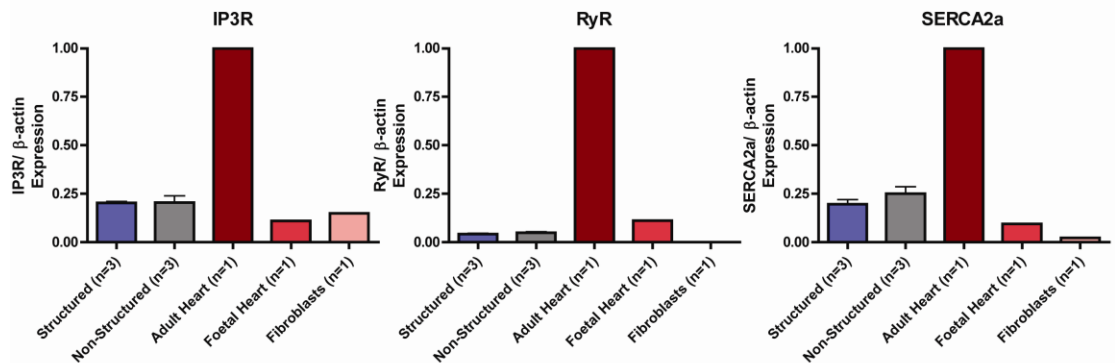
Normalized to GAPDH



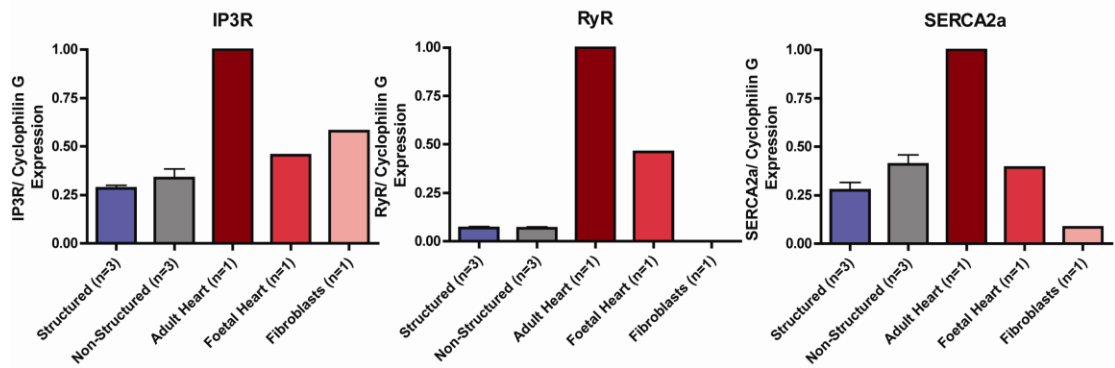
Normalized to 18s



Normalized to β -actin



Normalized to Cyclophilin G



Supplemental Figure 4. Comparison of IP3R, RyR and SERCA2a expression when normalized to GAPDH, 18s ribosomal RNA, Cyclophilin G, and β -actin, and expressed relative to adult heart tissue in iPSC-CM cultured on structured and non-structured PDMS, fibroblasts, adult heart and foetal heart tissue.