

Suppl. Fig. 1. *Rap1* RNAi phenocopies the effects of *Rap1* mutants. A-D. Embryos stained with antibodies to Rap1. A. Rap1 is already localized to the plasma membrane during syncytial stages. B,C. Rap1 localizes smoothly to the membrane all along the apical basal axis (B,C), thus overlapping but not co-localizing with spot AJs (B'). D. *Rap1* RNAi effectively reduces Rap1 levels below detection. E-G. Single apical-basal sections (E vs F) or projections (G vs H) reveal that *Rap1* RNAi reduces apical Arm enrichment (brackets) without causing loss in basal junctions (arrowheads). I-N. Single apical-basal sections (I vs F), apical views (J vs. M) or projections (L vs N) reveal that *Rap1* RNAi reduces apical Baz restriction (L vs N, brackets). O,P. *Rap1* RNAi leads to more variable apical cell area.

Suppl. Fig. 2. *Rap1*^{MZ} mutants begin to lose epithelial integrity as development proceeds. A,B. Cuticle preparations of WT and *Rap1*^{MZ} mutant. C-J. Late germband extension (stage 8) embryos. In wild-type, groups of cells in the epidermis round up and undergo synchronous divisions—cortical levels of AJ proteins (C) and Baz (F) are reduced in these cells. D. In *cno*^{MZ} mutants we previously found that once cells round up, they have difficulty regaining columnar cell shape, and thus rounded up cells with reduced apical AJ proteins accumulate (D,D close-up, brackets). E,F. In WT, while Baz is planar-polarized in the ectoderm, it still forms continuous junctions all around the cell, even in cells undergoing division (H, arrows). G,H. In presumptive zygotically rescued *Rap1*^M mutants, while Baz continues to surround cells of the dorsal ectoderm, in the lateral ectoderm Baz at junctions becomes weak (red arrow) or fragmented (arrowheads). I,J. Even more severe defects are seen in presumptive *Rap1*^{MZ} mutants, where Baz accumulation becomes highly fragmented in the lateral ectoderm (arrowheads). Scale bars=10µm.

Suppl. Fig. 3. *Cno*, Baz, AJ proteins, and aPKC localize to the membrane as early as syncytial divisions. WT embryos. A,H,L. Syncytial divisions (cycle 12/13). (A) *Cno* shows robust localization to pseudocleavage furrows during syncytial divisions. Arm (A') and Baz (H) also localize to pseudocleavage furrows at lower levels. aPKC (L) also displays membrane localization during syncytial divisions, when processed by formaldehyde fixation. B-G, J-K, M-O. Mid (B-D, J-K) or late (F,G) cellularization. *Cno*, DEcad/Arm, and Baz localize to spot AJs (C,D,K, arrows), DEcad/Arm also localize to basal junctions (O,

arrowhead), and Cno also accumulates apically early (D, arrowhead). aPKC remains membrane associated but localizes uniformly all along the lateral membrane (O). Scale bars=10 μ m.

Suppl. Fig 4. Rap1 and Cno play roles in Baz localization during gastrulation but other cues partially restore apical Baz in their absence. A-D. Plots displaying the average Baz image intensity in embryos of different genotypes (apical is to the left) at the indicated stages.

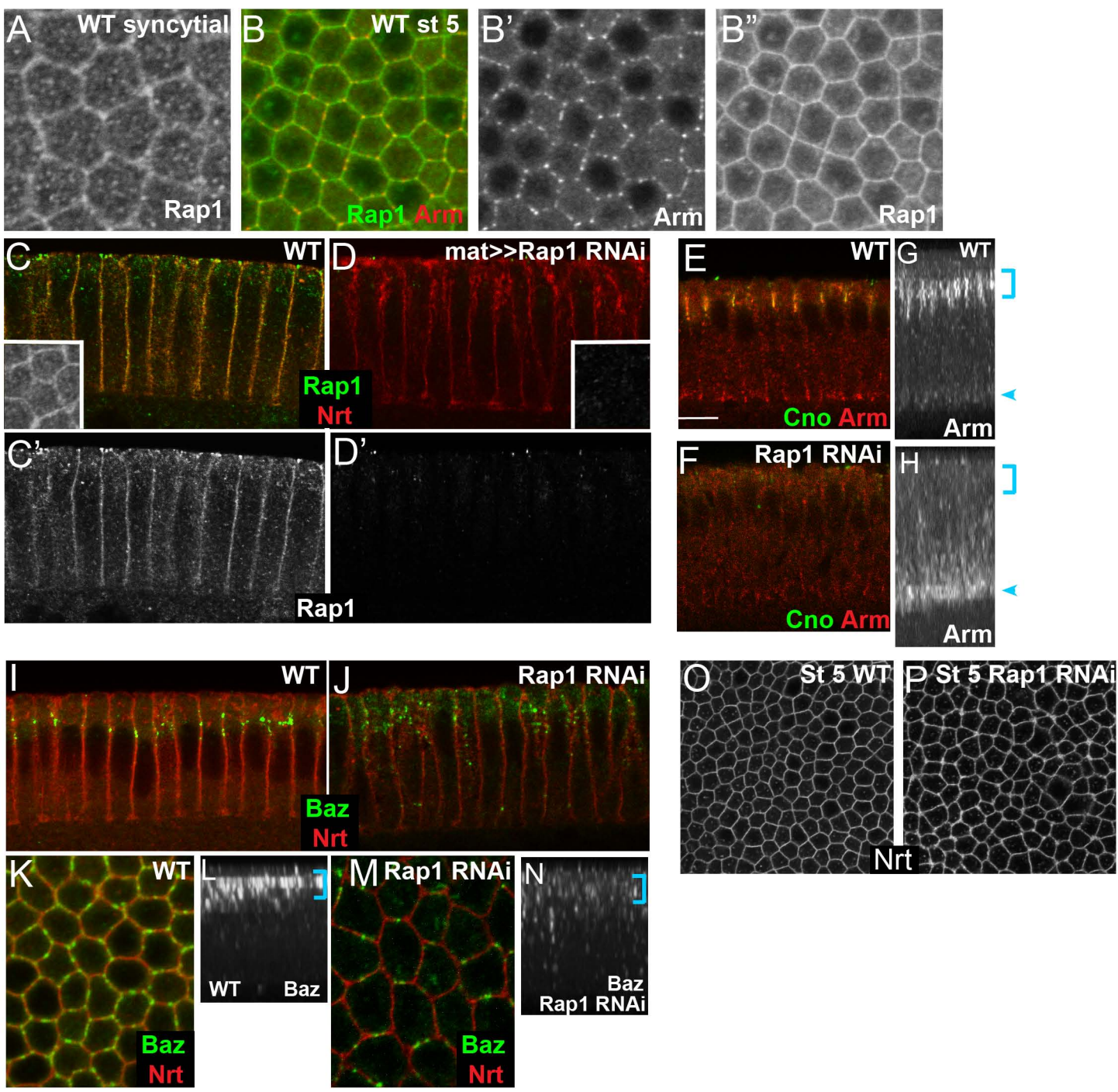
Suppl. Fig 5. Baz RNAi reduces Baz levels and mimics the effect of *baz* maternal and zygotic mutants. A-D. Maternal GAL4 driven *baz* RNAi effectively knocks down Baz, as assessed at gastrulation onset. In WT, Baz (A',C'') is apically enriched and co-localizes with AJs (A, A'' brackets, C, C''). Unlike Baz, AJs are also basally enriched (A, A'' arrows, C inset). *baz* RNAi reduces Baz staining to background levels (B' vs. A'; D' vs. C'') and disrupts apical accumulation of AJs (B'' bracket vs. A'' bracket). Scale bars=10 μ m.

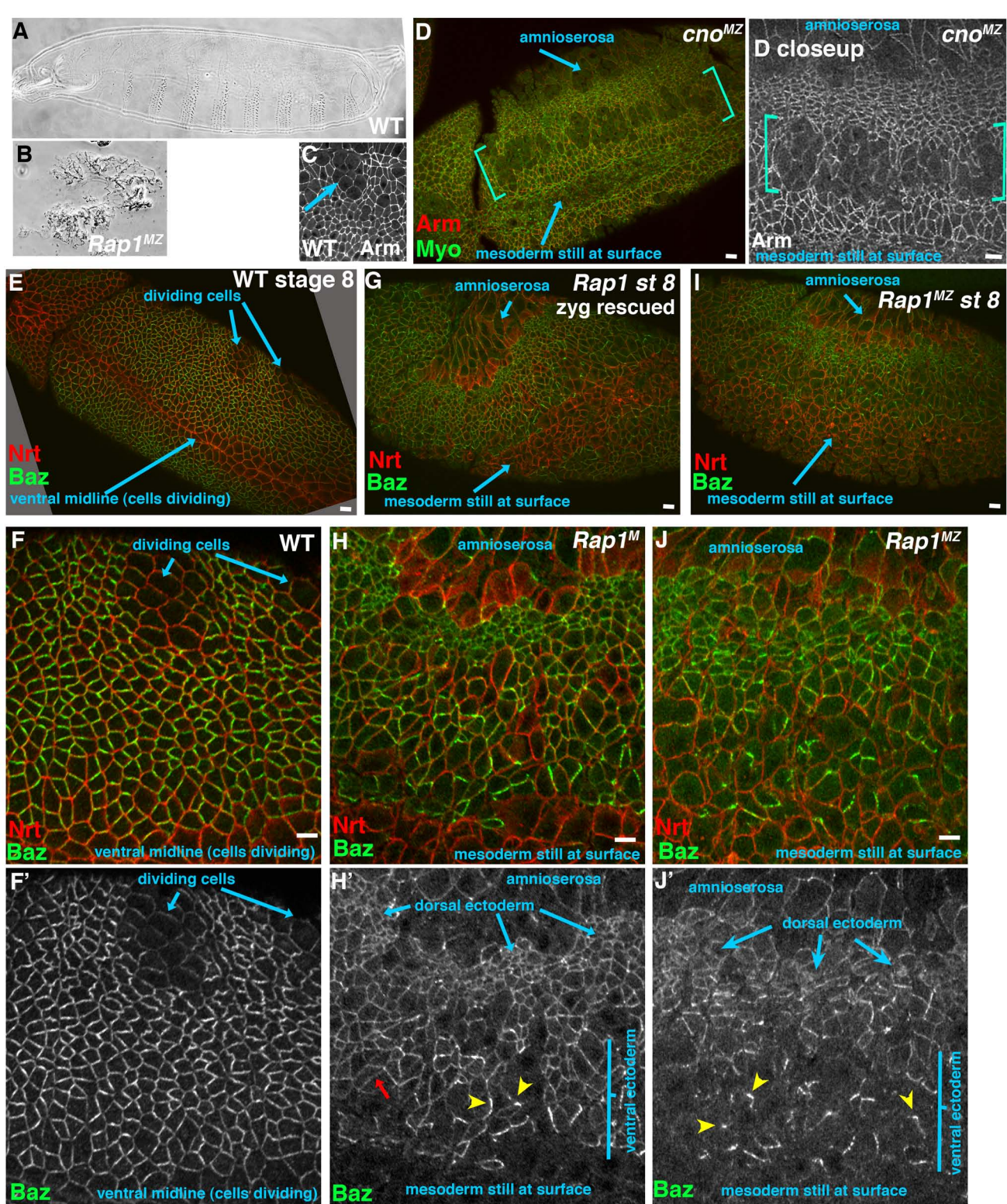
Suppl. Fig. 6. In the absence of aPKC, Cno is not removed from the apical domain during gastrulation. A,B. Gastrulation onset (stage 6). C-H. Germband extension (stage 7). A-D. Loss of aPKC (B, D) leads to failure to exclude Cno apically (compare arrows), and reduced Cno coalescence along the apical-basal axis (compare brackets). During germband extension (stage 7) apical accumulation of Cno in *aPKC* mutants becomes more accentuated (E vs. F). G,H. In *aPKC* mutants, the belt AJs seen in WT (G) do not form—instead AJ proteins accumulate in puncta at the dorsal and ventral borders of cells (H' arrows). While the majority of Cno is located on the apical membrane in *aPKC* mutants (F), a portion co-localizes with these dorsal and ventral AJ puncta (H'', arrows). Scale bars=10 μ m.

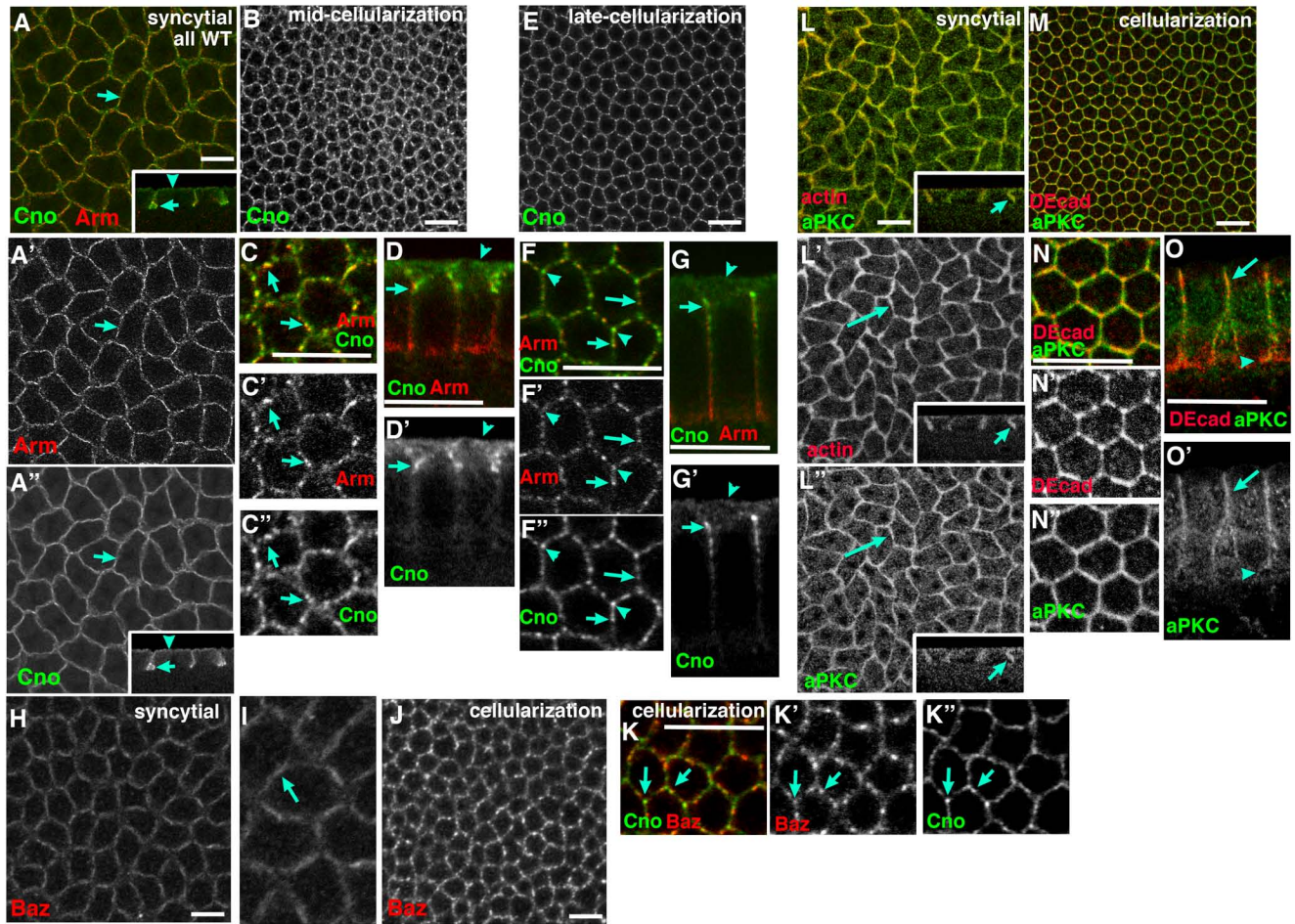
Suppl. Fig. 7. Rap1, Baz and aPKC play distinctive roles in maintaining columnar cell shape at gastrulation onset and during germband extension. Cell areas were measured at three different apical-basal depths during stage 6 or stage 7, as in Fig. 10, with mean CV values for each genotype and section. A-E. Representative stage 6 embryos, with mean CV values for each genotype and section. Arrows indicate examples of cell area variation. F-H. Bee-swarm scatter plots of cell areas. *Rap1*, *baz* RNAi, and *aPKC* all affect variability of apical cell area, while only *Rap1* significantly affects cell area variability in the most basal section. White lines=median value. I-M. Representative stage 7 embryos, with mean CV values. Arrows

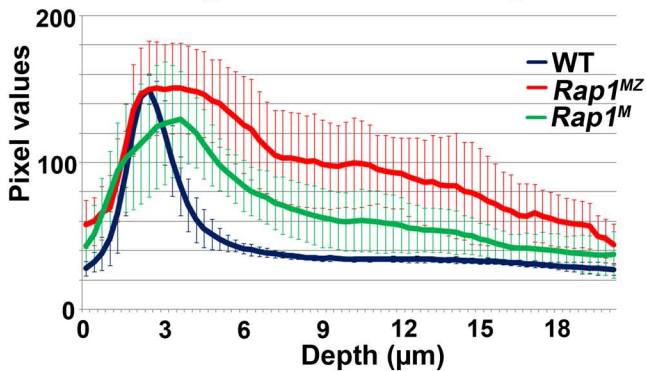
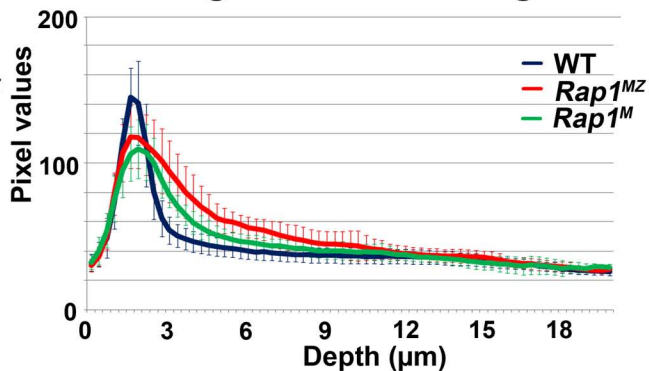
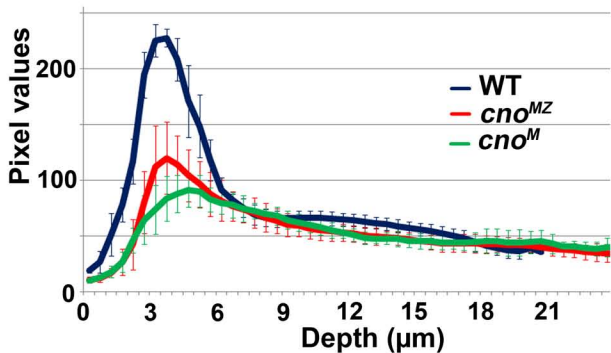
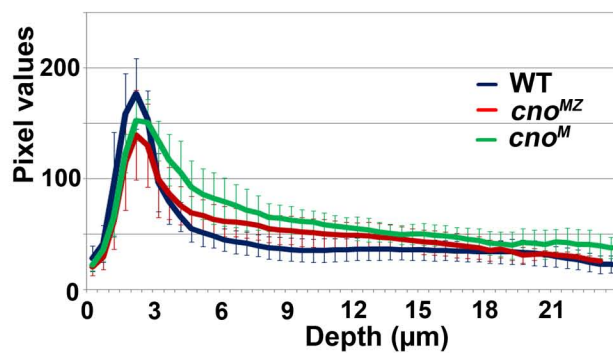
indicate examples of variable cell areas. N-P. Bee-swarm scatter plots of cell areas. *Rap1*, *baz* RNAi, and *aPKC* all significantly affect variability of apical cell area. Scale bar=10 μ m.

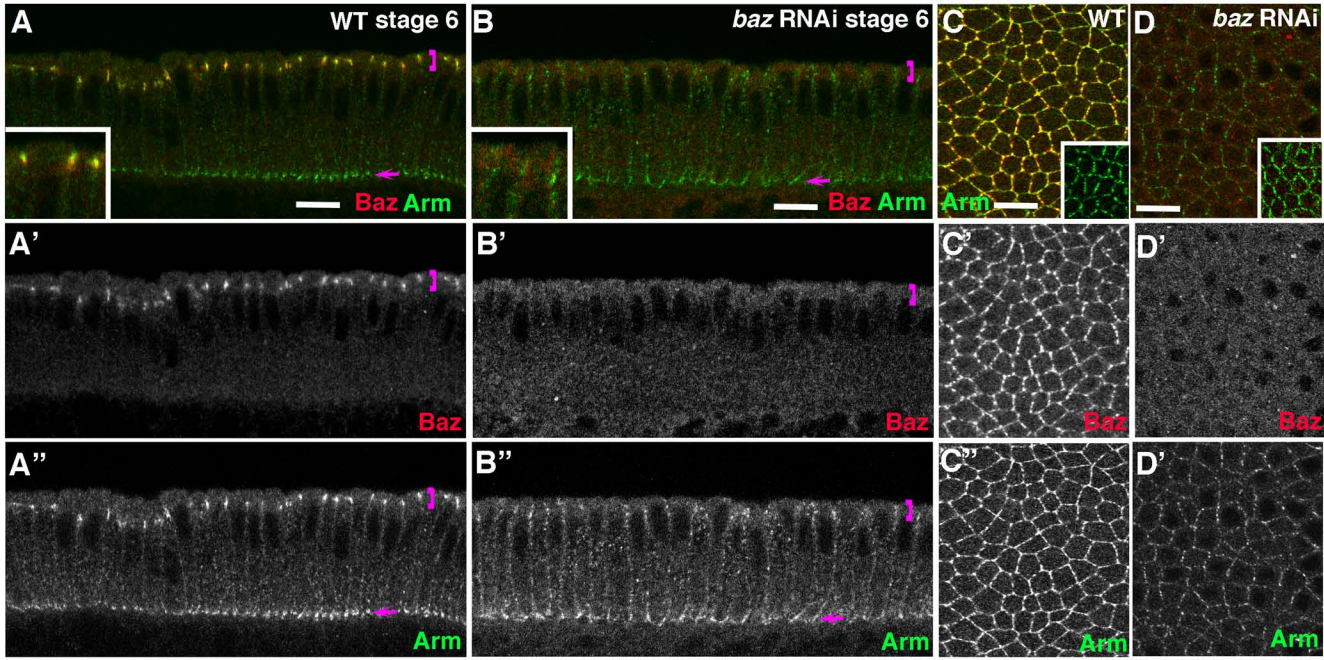
Suppl. Fig. 8. Old and new models of polarity establishment. See text for details.

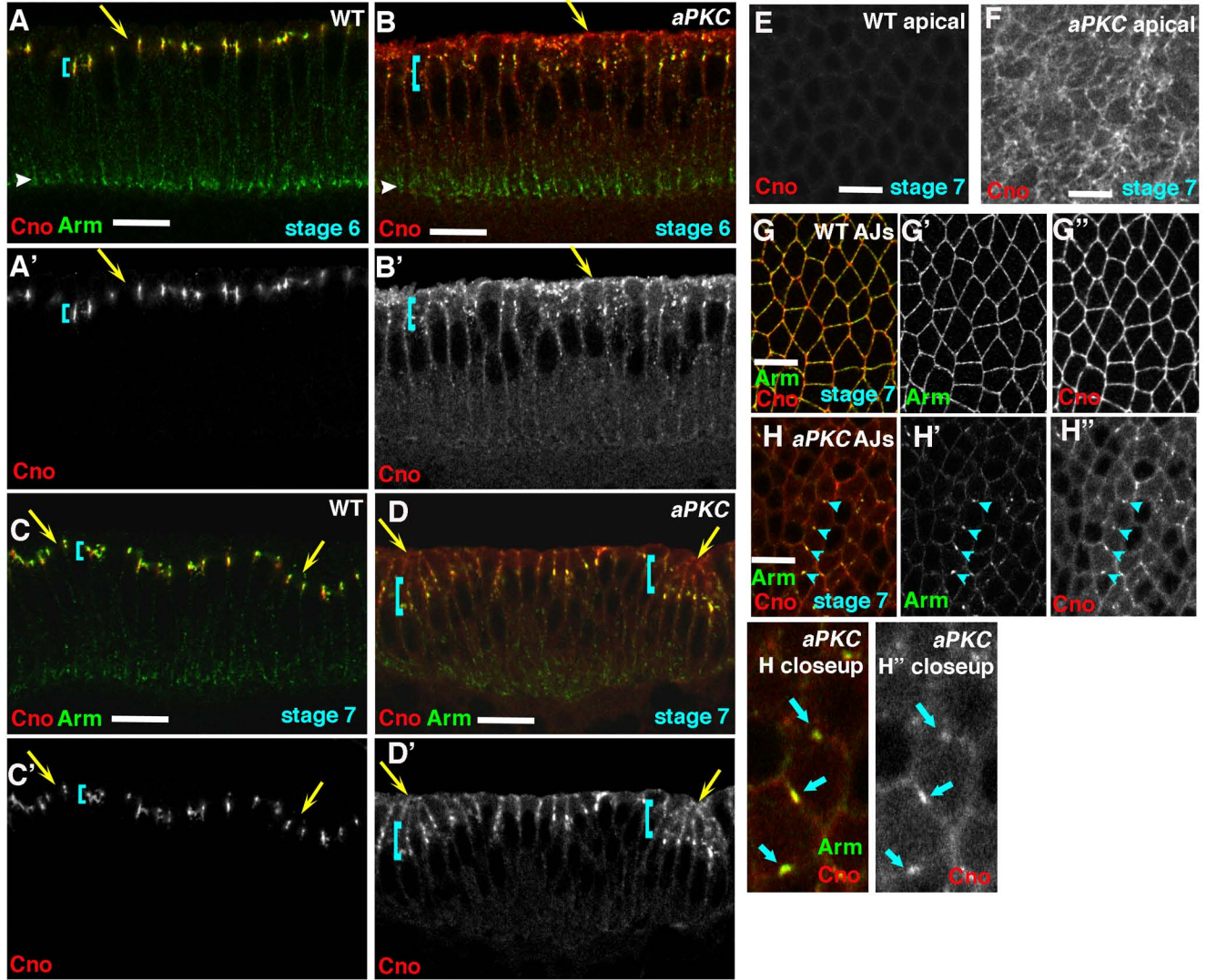




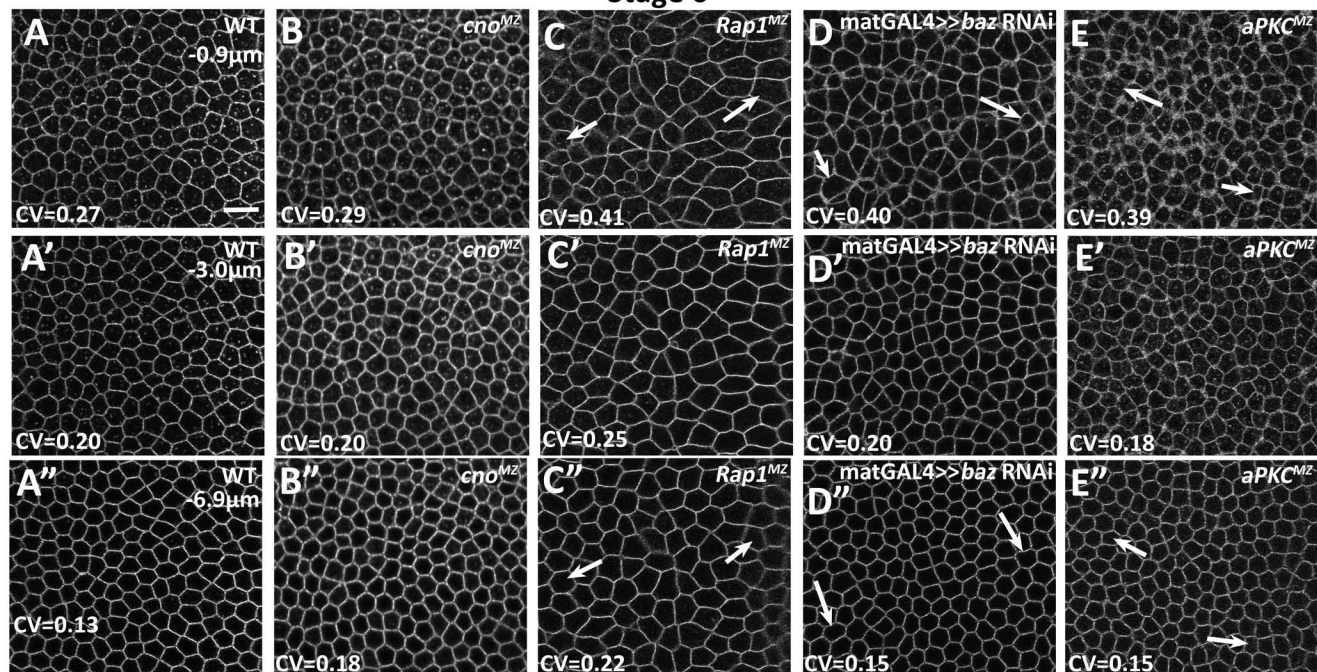


A Average Baz Levels stage 6**B Average Baz Levels stage 7****C****D**





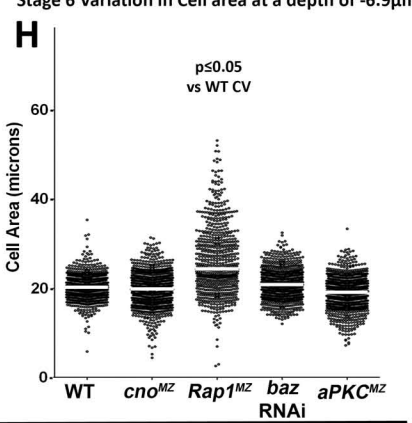
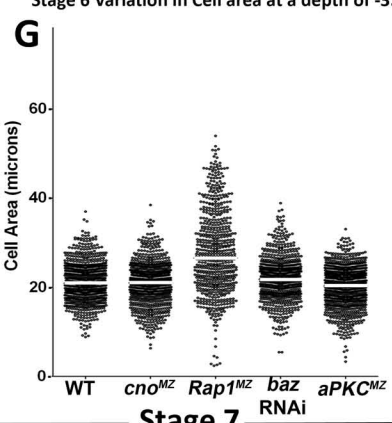
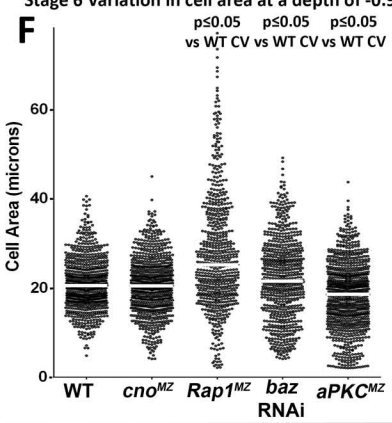
Stage 6



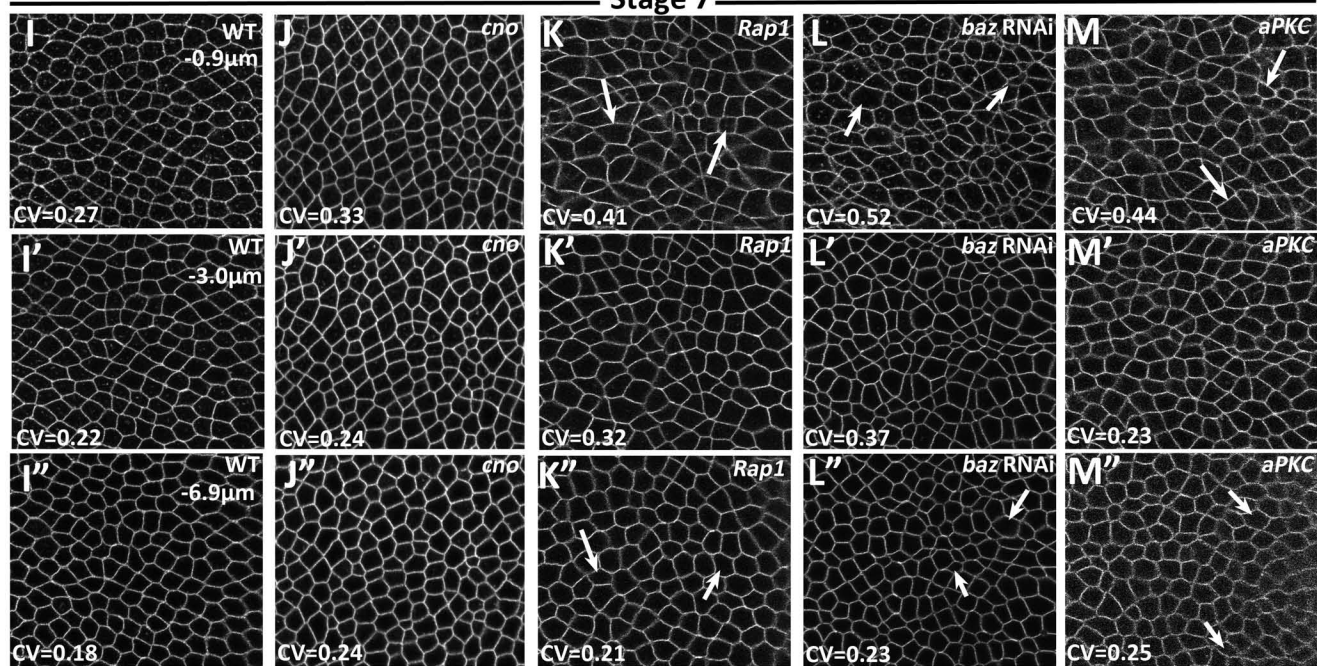
Stage 6 Variation in cell area at a depth of -0.9 μm

Stage 6 Variation in Cell area at a depth of -3.0 μm

Stage 6 Variation in Cell area at a depth of -6.9 μm



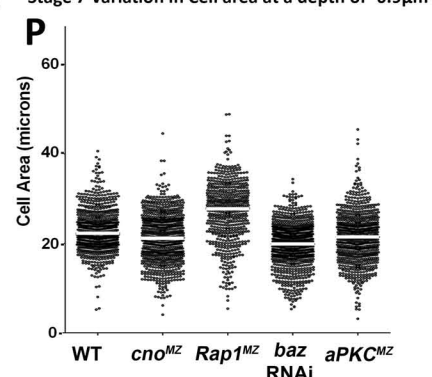
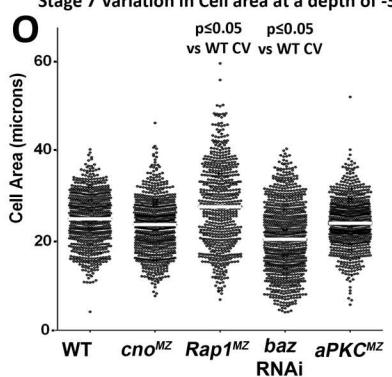
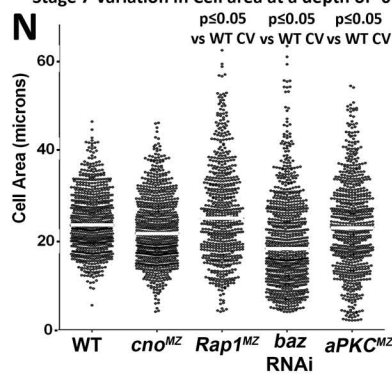
Stage 7

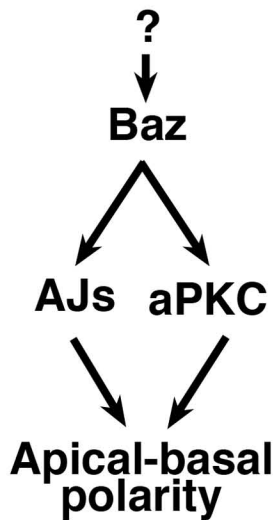


Stage 7 Variation in Cell area at a depth of -0.9 μm

Stage 7 Variation in Cell area at a depth of -3.0 μm

Stage 7 Variation in Cell area at a depth of -6.9 μm



A**B**