

# European Journal of Immunology

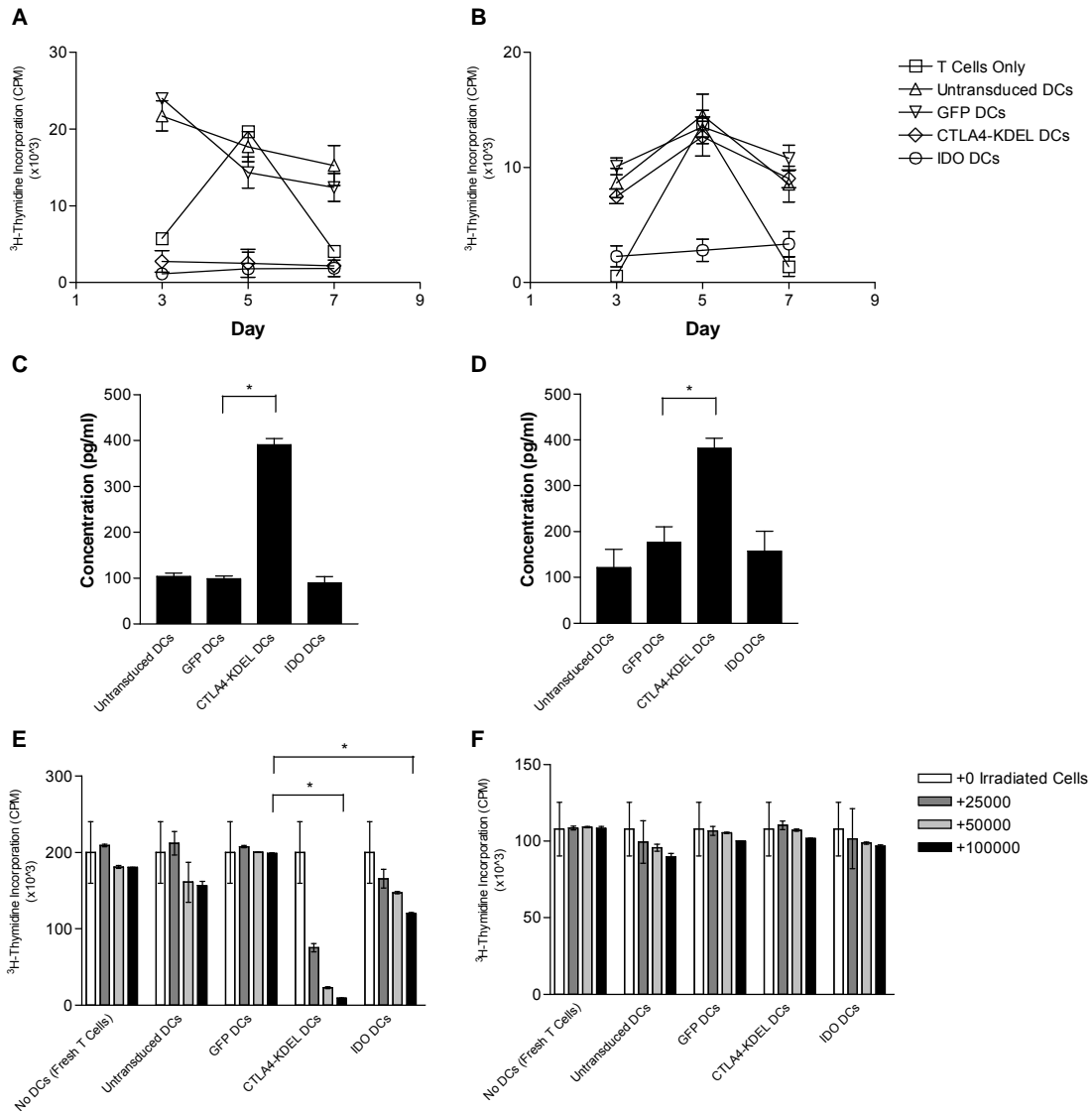
**Supporting Information**

**for**

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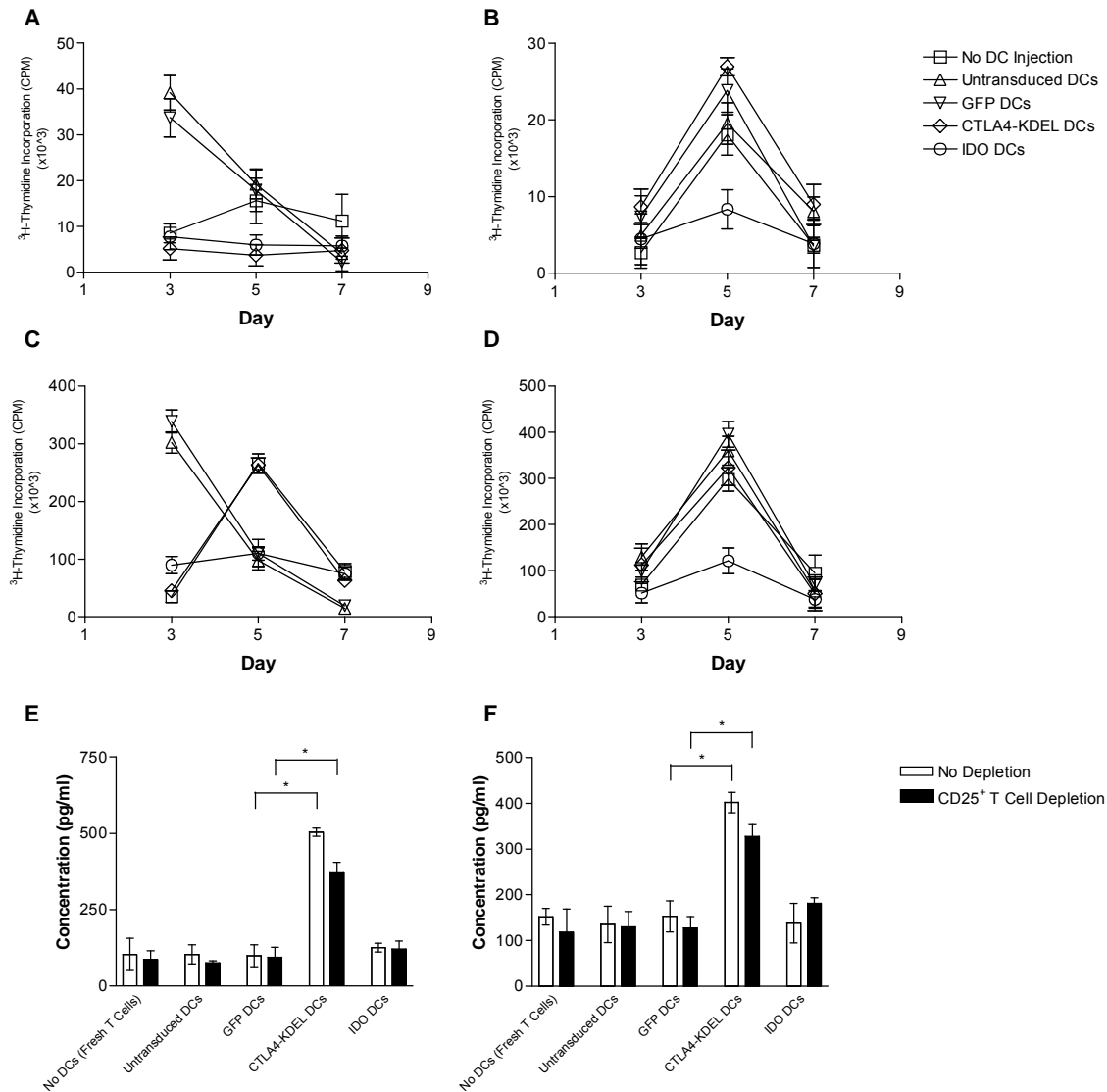
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and Andrew J. T. George

**Dendritic cell modification as a route to inhibiting corneal graft rejection by the  
indirect pathway of allorecognition**



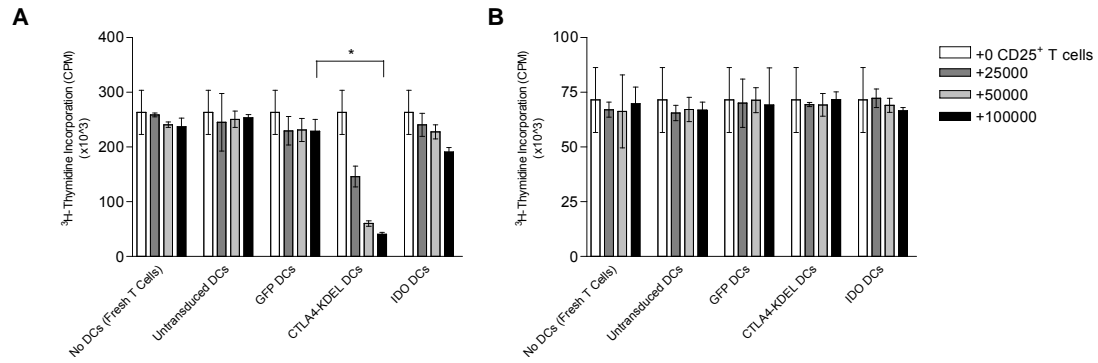
**SUPPORTING FIGURE 1. Induction of donor alloantigen-specific T-cell anergy and regulation in vitro.**

BALB/c DCs were transduced either with EIAV GFP (control), EIAV CTLA4-KDEL or EIAV IDO, followed by stimulation with LPS. The transduced (and untransduced) DCs were subsequently incubated in vitro with C3H/He-derived  $\text{CD4}^+$  T cells. After 10 days, the T cells were harvested and rechallenged in vitro with (A) donor BALB/c DCs or (B) third-party C57BL/6 DCs, and  $\text{CD4}^+$  T-cell proliferation assessed by thymidine incorporation on days 3, 5 and 7. (C-D) Culture supernatant from the donor DC rechallenge assay was harvested for detection of the immunoregulatory cytokines (C) TGF- $\beta$  and (D) IL-10 by ELISA. (E-F) In addition, after 10 days of the primary DC -  $\text{CD4}^+$  T-cell coculture, the T cells were harvested, irradiated and added (0- $10^5$   $\text{CD4}^+$  T cells) to a primary MLR between freshly-isolated C3H-derived  $\text{CD4}^+$  T cells and (E) donor BALB/c DCs or (F) third-party C57BL/6 DCs. T-cell proliferation was assessed by thymidine incorporation after 5 days. All results are shown as the mean  $\pm$  SD of triplicate wells and are representative of three independent experiments performed. \*  $p < 0.05$ , two-tailed  $t$ -test.



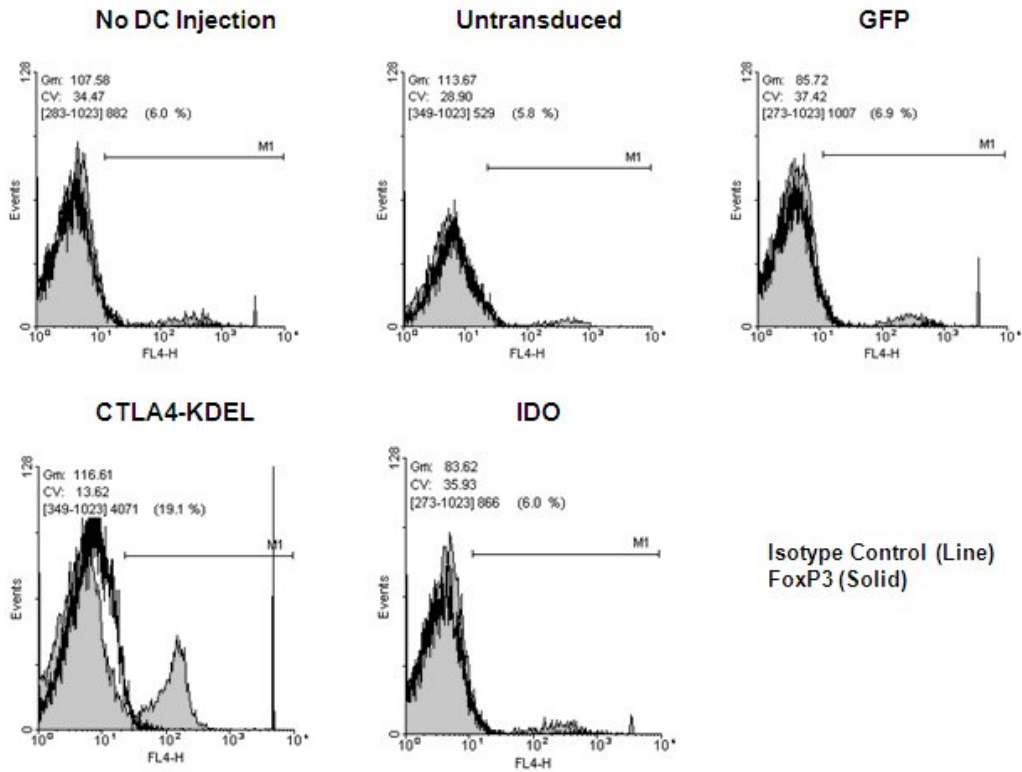
**SUPPORTING FIGURE 2. Induction of donor alloantigen-specific T-cell anergy in vivo and production of immunoregulatory cytokines.**

$2.5 \times 10^6$  BALB/c DCs (either untransduced, or transduced with EIAV GFP (control), EIAV CTLA4-KDEL or EIAV IDO) were injected intravenously into C3H/He mice. **(A-D)** After 10 days,  $CD4^+$  T cells purified from the spleens of injected mice were rechallenged in vitro with **(A)** donor BALB/c DCs or **(B)** third-party C57BL/6 DCs, and  $CD4^+$  T-cell proliferation assessed by thymidine incorporation on days 3, 5 and 7. The purified  $CD4^+$  T cells were also rechallenged with **(C)** donor BALB/c DCs or **(D)** third-party C57BL/6 DCs in the presence of 100 U/ml exogenous IL-2. **(E-F)** Culture supernatants from donor DC rechallenge assays (including those from  $CD4^+CD25^+$  T-cell depleted-rechallenge assays) were harvested on day 7 for detection of the immunoregulatory cytokines **(E)** TGF- $\beta$  and **(F)** IL-10 by ELISA. All results are shown as the mean  $\pm$  SD of triplicate wells and are representative of three independent experiments performed. \*  $p < 0.05$ , two-tailed *t*-test.



**SUPPORTING FIGURE 3. Generation of donor alloantigen-specific Treg cells in vivo with capacity for linked suppression.**

$2.5 \times 10^6$  BALB/c DCs (either untransduced, or transduced with EIAV GFP (control), EIAV CTLA4-KDEL or EIAV IDO) were injected intravenously into C3H/He mice. **(A-B)** After 10 days,  $CD4^+CD25^+$  T cells purified from the spleens of injected mice were irradiated and added in increasing numbers ( $0-10^5$   $CD4^+CD25^+$  T cells) to a primary MLR between freshly-isolated C3H-derived  $CD4^+$  T cells and **(A)** donor BALB/c DCs or **(B)** third-party C57BL/6 DCs. T-cell proliferation was assessed by thymidine incorporation after 5 days. All results are shown as the mean  $\pm$  SD of triplicate wells and are representative of three independent experiments performed. \*  $p < 0.05$ , two-tailed  $t$ -test.



**SUPPORTING FIGURE 4. Generation of FoxP3<sup>+</sup>CD4<sup>+</sup>CD25<sup>+</sup> T cells in vivo**

$2.5 \times 10^6$  BALB/c DCs (either untransduced, or transduced with EIAV-GFP (control), EIAV-CTLA4-KDEL or EIAV-IDO) were injected intravenously into C3H/He mice. After 10 days, CD4<sup>+</sup>CD25<sup>+</sup> T cells purified from the spleens of injected mice were stained and analysed by flow cytometry for expression of FoxP3. The results shown in are representative of three independent experiments.