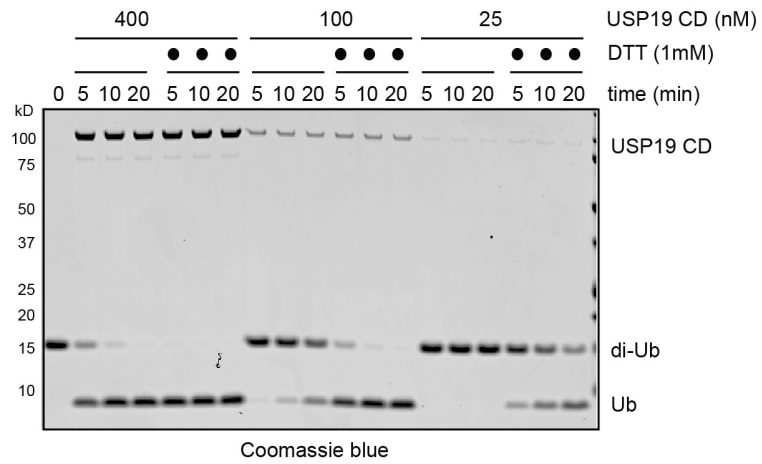


### Supplementary Figure S1 Purification of deubiquitinases

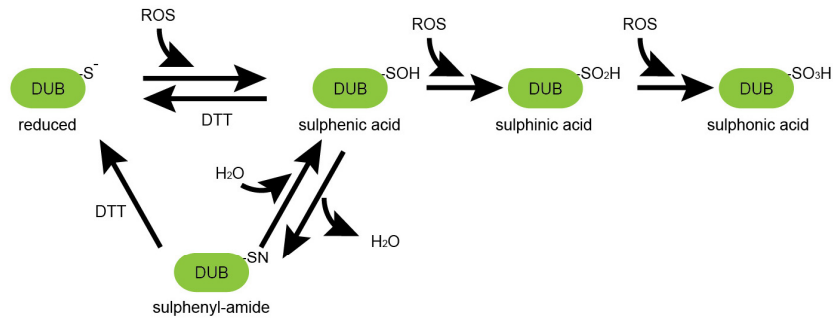
HEK293 cells were transfected with the indicated DUB-expressing plasmids. The cells were harvested 72 h after transfection. FLAG-tagged deubiquitinases were affinity-purified using FLAG-agarose beads. Purified proteins were analyzed by immunoblotting with the FLAG antibody.



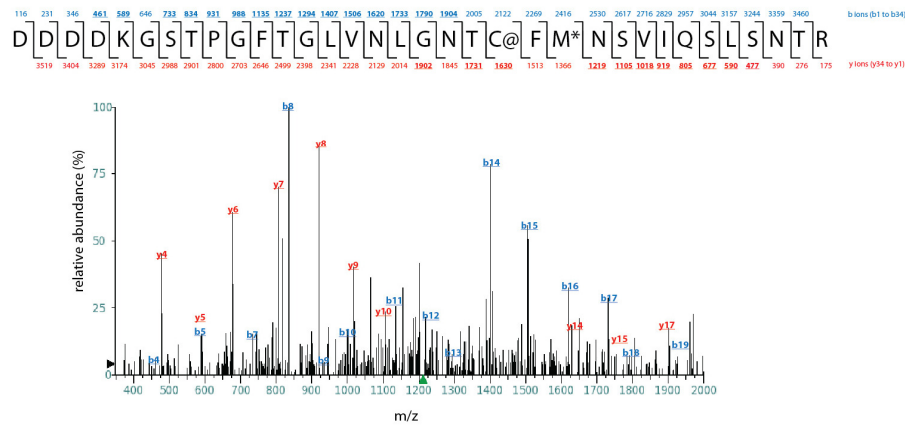
### Supplementary Figure S2 Activation of USP19 CD by DTT

Purified USP19 CD was treated with or without 1mM DTT and incubated with di-ubiquitin for the indicated time periods at 37 °C. The cleavage of di-ubiquitin was detected by SDS-PAGE electrophoresis followed by Coomassie blue staining.

a



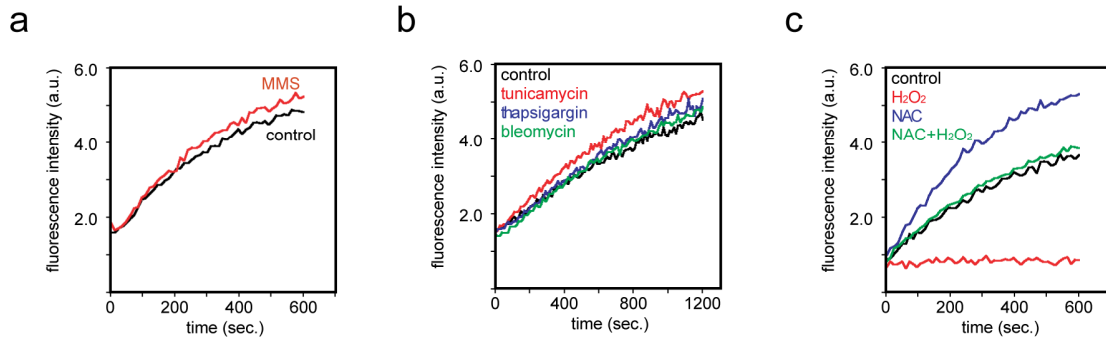
b



### Supplementary Figure S3 Oxidation of the catalytic cysteine of USP19 in the cell

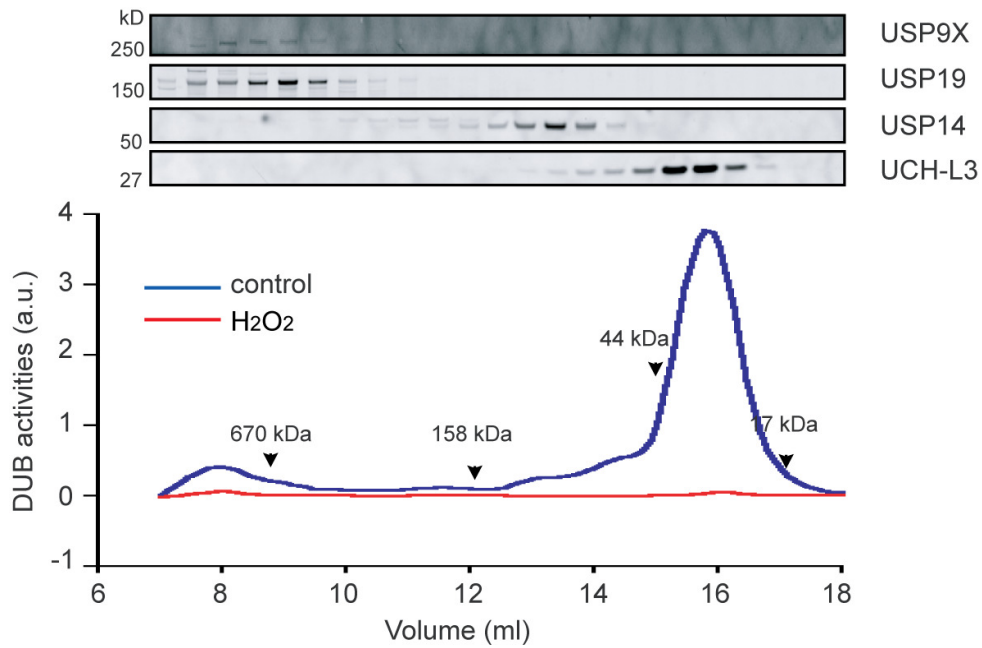
(a) The DUB redox species and their interrelationship. (b) The band indicated by the asterisk in Figure 3a was subject to trypsin digestion and mass spectrometry analysis. @ indicates the sulphenyl-amide modification, \* indicates oxidized methionine.





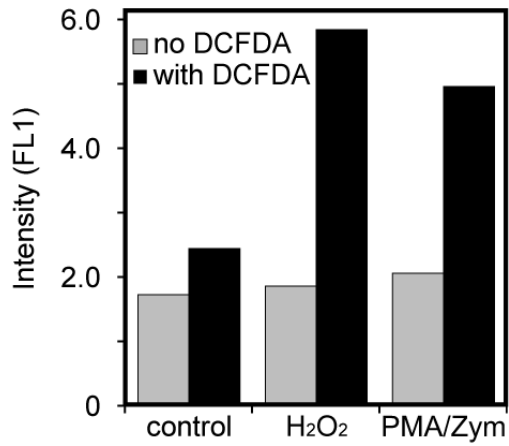
### Supplementary Figure S5 Oxidative stress reduces DUB activities in cells

(a) HEK293 cells were treated with or without 1mM MMS for 1 hour. Same number of cells (~3,000) was directly added to a deubiquitinating assay buffer containing 0.5% NP40 and Ub-AFC. DUB activities were assayed by monitoring the fluorescence. (b) HEK293 cells were treated with 5 $\mu$ g/ml tunicamycin, 1  $\mu$ M thapsigargin, or 200  $\mu$ g/ml bleomycin, or as a control with DMSO for 1 hour. The DUB activities were analyzed as in (a). (c) The anti-oxidant NAC prevents DUB inhibition by H<sub>2</sub>O<sub>2</sub>. HEK293 cells were pretreated with or without 20mM NAC for 30min before addition of 1mM H<sub>2</sub>O<sub>2</sub>. Where indicated, cells were incubated with H<sub>2</sub>O<sub>2</sub>-containing medium for 2min prior to the DUB activity assay. 2,000 cells were assayed for each sample.



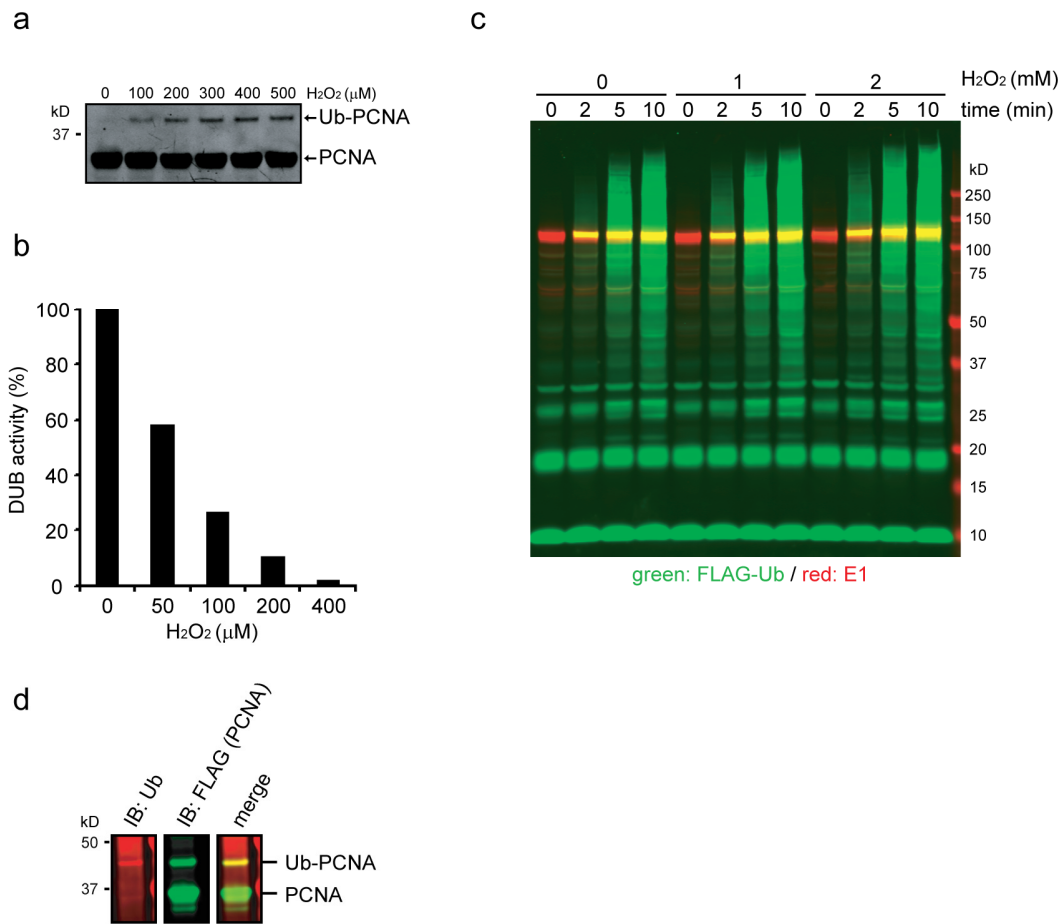
**Supplementary Figure S6 DUB inhibition by ROS occurs in a global manner**

Whole cell extract from HEK293 cells were subject to gel filtration analysis by a Superdex200 HR 10/30 column in a buffer containing 2mM DTT. The DUB activities in the resulting fractions were determined by incubating 2 $\mu$ l of each fraction in 100 $\mu$ l DUB assay buffer containing Ub-AFC. A portion of the samples (2 $\mu$ l) were treated with H<sub>2</sub>O<sub>2</sub> (0.5mM, 1min) before the addition of Ub-AFC. Another fraction of the samples were also analyzed by immunoblotting with antibodies against the indicated DUBs.



**Supplementary Figure S7 ROS generation in macrophage cells**

ROS generation was stimulated by an inflammatory response. RAW264.7 cells treated with 1mM H<sub>2</sub>O<sub>2</sub> or 500nM PMA plus 0.5mg/ml Zymosan for 1hr were stained with 10μM DCFDA. Unstained cells were used as a negative control. Shown are the mean values of fluorescence intensity.

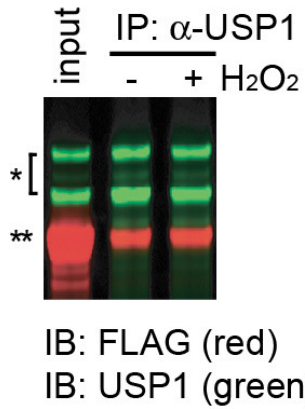


### Supplementary Figure S8 Oxidative stress inhibits DUBs to modulate PCNA ubiquitination

(a) PCNA ubiquitination is induced by H<sub>2</sub>O<sub>2</sub> in a dose dependent manner. HEK293 cells were treated with H<sub>2</sub>O<sub>2</sub> as indicated for 10min. Cells were lysed in the Laemmli buffer for immunoblotting analyses. (b) Dose dependent inhibition of DUBs by H<sub>2</sub>O<sub>2</sub>. (c) H<sub>2</sub>O<sub>2</sub> does not affect the ubiquitin conjugating system in HEK293 cells. Cell extracts from control or H<sub>2</sub>O<sub>2</sub>-treated cells were incubated with FLAG-tagged ubiquitin together with an energy regenerating system. *In vitro*-synthesized ubiquitin chains by endogenous enzymes were detected by immunoblotting. (d) Ubiquitination of PCNA *in vitro*. The purified PCNA-FLAG protein was incubated with UBE1 (E1), RAD6B (E2), RAD18 (E3), and bovine ubiquitin in the presence of ATP before immunoblotting analysis.



- \* Myc-USP1
- \*\* FLAG-UAF1



**Supplementary Figure S9 H<sub>2</sub>O<sub>2</sub> does not affect the interaction of USP1 with UAF1.**

The purified USP1-UAF1 complex was incubated in a buffer in the absence (-) or presence (+) of 1mM H<sub>2</sub>O<sub>2</sub> for 5min. The samples were then subject to immunoprecipitation using the USP1 antibody. The precipitated proteins were analyzed by immunoblotting (IB). A fraction of the untreated sample was included as the input.