

Generation of Candidate Ligands for Nicotinic Acetylcholine Receptors via *In Situ* Click Chemistry with a Soluble Acetylcholine Binding Protein Template

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Supporting Information

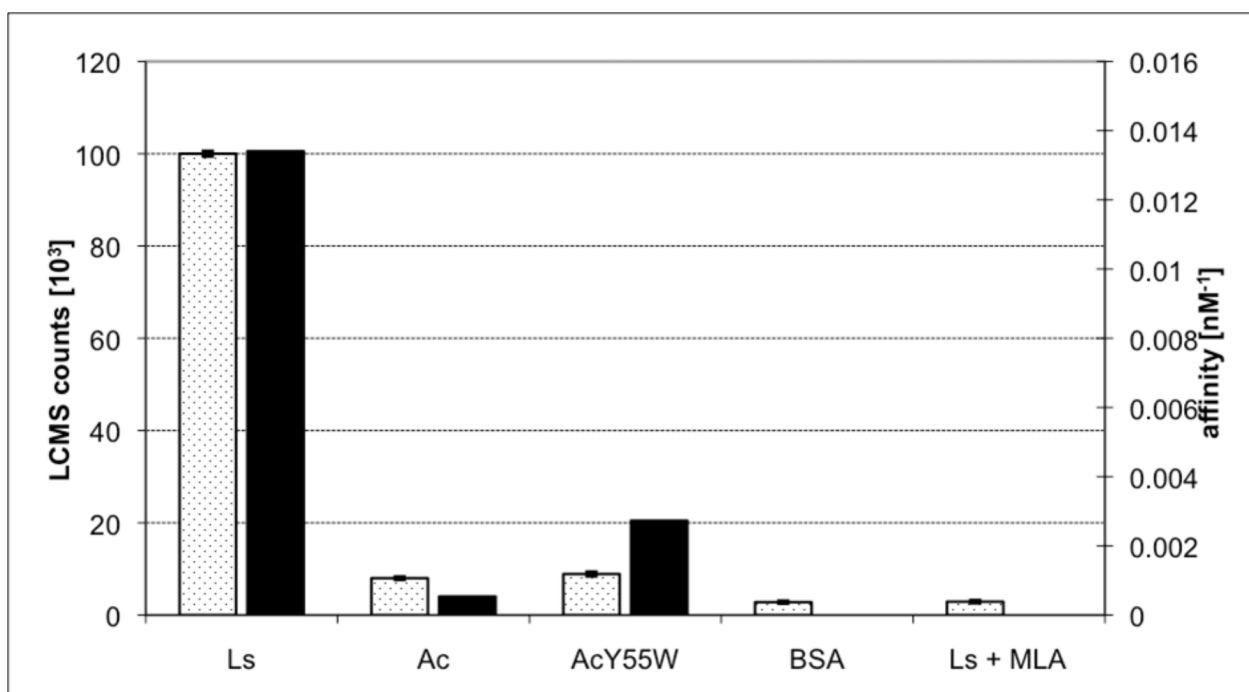
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In situ click screens

Table S1: LCMS data – in situ click chemistry proof of concept run

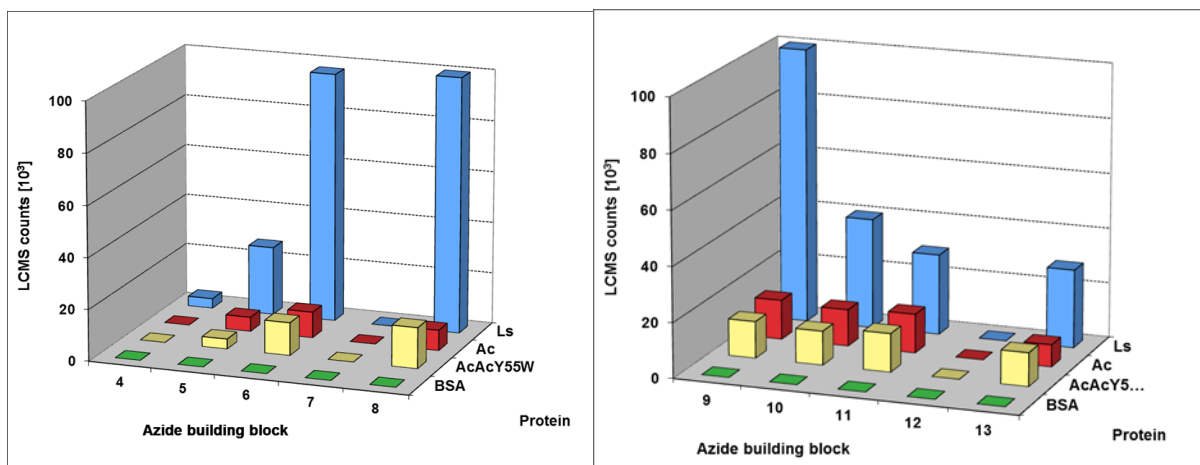
Protein	Cmpd #	Run 1 [counts]	Run2 [counts]	Run3 [counts]	Average [counts]	Standard Deviation	Standard error	Standardized [counts]	±
Ls	2	396748	395310	392222	394760	2313	1335	100.0	0.3
Ac	2	32164	31753	31058	31658	559	323	8.0	0.1
AcY55W	2	36796	33579	35472	35282	1617	933	8.9	0.2
BSA	2	11145	10662	11165	10991	285	164	2.8	0.0
Ls + MLA	2	10745	11620	11394	11253	454	262	2.9	0.1



Standardized LCMS counts (dotted) compared to affinity towards the respective AChBP.

Table S2: LCMS data- *in situ* click chemistry screen of library 1a

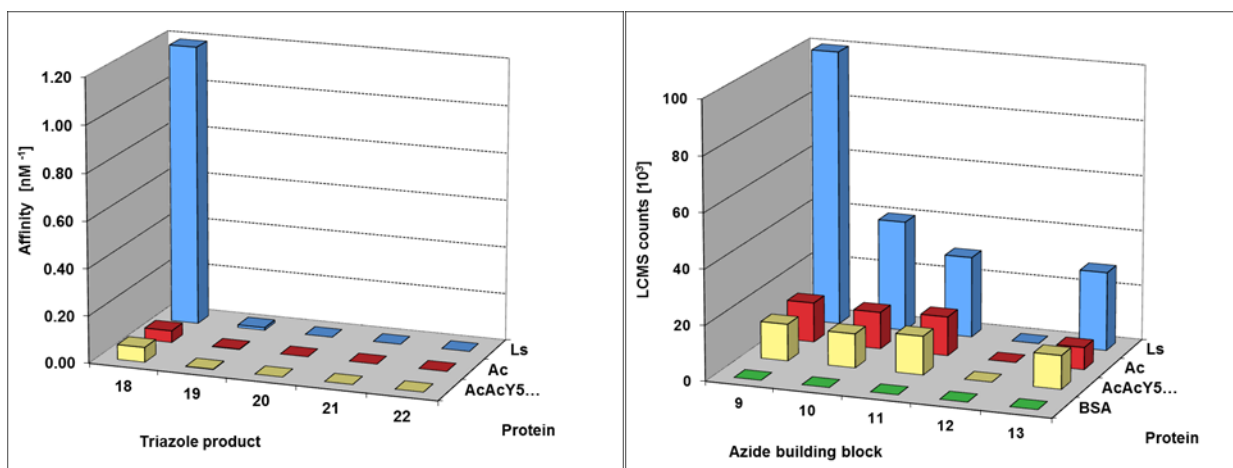
Protein	Cmpd #	Run 1 [counts]	Run 2 [counts]	Run 3 [counts]	Average [counts]	Standard Deviation	Standard error	Standardized [counts]	±
Ls	2	16265	15088	16791	16048	872	503	3.8	0.1
Ac	2	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
AcY55W	2	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
BSA	2	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
Ls	14	441443	410258	411660	421120	17614	10169	100.0	2.4
Ac	14	34251	32840	33599	33563	706	408	8.0	0.1
AcY55W	14	66415	67796	66470	66894	782	451	15.9	0.1
BSA	14	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
Ls	15	429438	416561	382930	409643	24013	13864	97.3	3.3
Ac	15	42763	42740	42121	42541	364	210	10.1	0.0
AcY55W	15	52099	55744	53859	53901	1823	1052	12.8	0.2
BSA	15	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
Ls	16	115527	107630	113806	112321	4153	2398	26.7	0.6
Ac	16	24040	22750	23576	23455	653	377	5.6	0.1
AcY55W	16	2350	22555	24468	16458	12255	7075	3.9	1.7
BSA	16	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
Ls	17	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
Ac	17	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
AcY55W	17	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
BSA	17	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a



Comparison of LCMS counts of *in situ* click screen with AChBP affinities of the triazole products – library 1a

Table S3: LCMS data- *in situ* click chemistry screen of library 1b

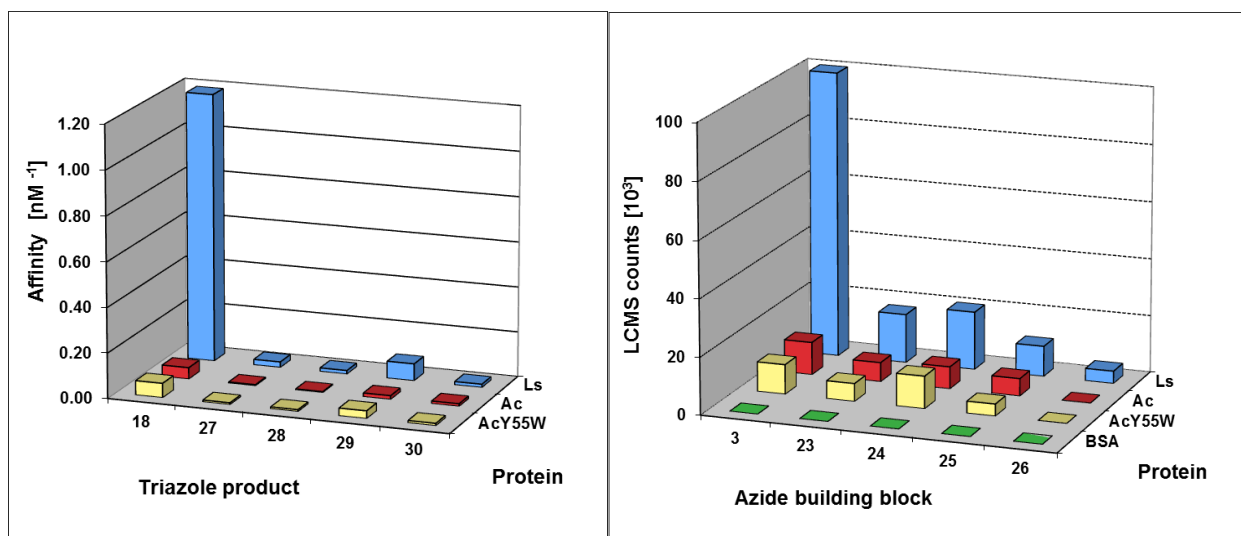
Protein	Cmpd #	Run 1 [counts]	Run 2 [counts]	Run 3 [counts]	Average [counts]	Standard Deviation	Standard error	Standardized [counts]	±
Ls	18	296691	296691	300771	298051	2356	1360	100.0	0.5
Ac	18	42574	42617	43031	42741	252	146	14.3	0.0
AcY55W	18	38150	40327	40691	39723	1374	793	13.3	0.3
BSA	18	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
Ls	19	123595	123595	110082	119091	7802	4504	40.0	1.5
Ac	19	38205	42986	38088	39760	2795	1614	13.3	0.5
AcY55W	19	35820	36079	38307	36735	1367	789	12.3	0.3
BSA	19	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
Ls	20	90441	90441	80586	87156	5690	3285	29.2	1.1
Ac	20	40907	41469	44052	42143	1677	968	14.1	0.3
AcY55W	20	40497	41143	43258	41633	1444	834	14.0	0.3
BSA	20	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
Ls	21	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
Ac	21	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
AcY55W	21	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
BSA	21	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
Ls	22	86823	86823	80722	84789	3522	2034	28.4	0.7
Ac	22	23015	24760	24011	23929	875	505	8.0	0.2
AcY55W	22	36921	34330	37269	36173	1606	927	12.1	0.3
BSA	22	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a



Comparison of LCMS counts of *in situ* click screen with AChBP affinities of the triazole products – library 1b

Table S4: LCMS data- *in situ* click chemistry screen of alkynes versus azide 10

Protein	Cmpd #	Run 1 [counts]	Run 2 [counts]	Run 3 [counts]	Average [counts]	Standard Deviation	Standard error	Standardized [counts]	±
Ls	18	235245	227407	217881	226844	8696	5020	100.0	2.2
Ac	18	24008	26823	26466	25766	1533	885	11.4	0.4
AcY55W	18	24516	23189	22508	23404	1021	590	10.3	0.3
BSA	18	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
Ls	27	38984	39170	37786	38647	751	434	17.0	0.2
Ac	27	14019	16280	15198	15166	1131	653	6.7	0.3
AcY55W	27	14140	13515	14673	14109	580	335	6.2	0.1
BSA	27	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
Ls	28	47103	44413	46436	45984	1401	809	20.3	0.4
Ac	28	19838	16470	16036	17448	2081	1202	7.7	0.5
AcY55W	28	24750	25039	27016	25602	1233	712	11.3	0.3
BSA	28	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
Ls	29	24019	21630	26540	24063	2455	1418	10.6	0.6
Ac	29	15480	12790	13027	13766	1489	860	6.1	0.4
AcY55W	29	10071	8890	9407	9456	592	342	4.2	0.2
BSA	29	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
Ls	30	9930	9929	9487	9782	255	148	4.3	0.1
Ac	30	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
AcY55W	30	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
BSA	30	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a

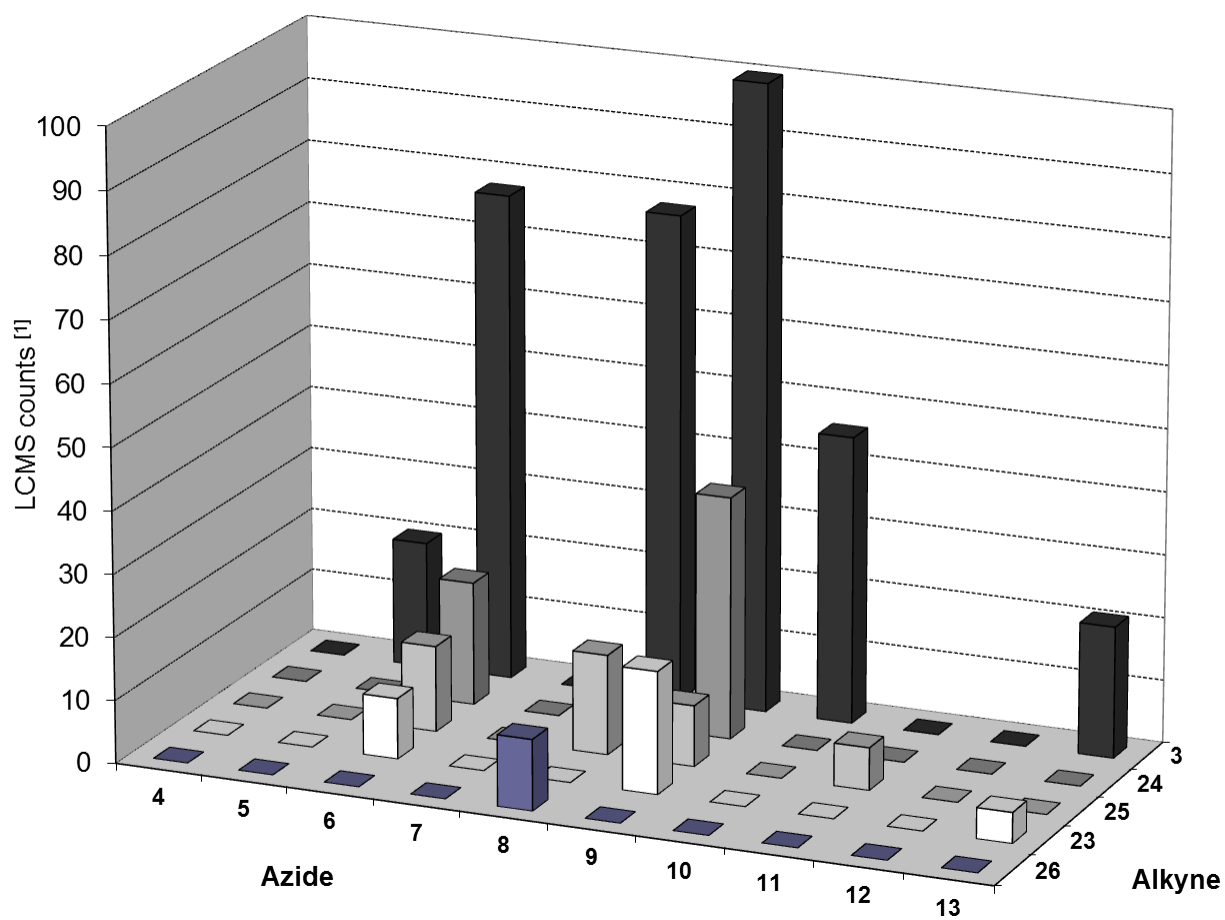


Comparison of LCMS counts of *in situ* click screen with AChBP affinities of the triazole products – library 2

Table S5: LCMS data- *in situ* click chemistry screen of alkyne-azide pool against *Ls*

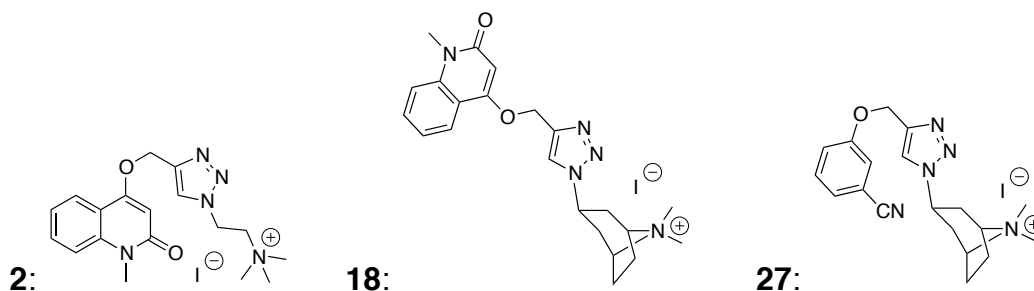
Alkyne #	Azide #	Run 1 [counts]	Run 2 [counts]	Run 3 [counts]	Average [counts]	Standard Deviation	Standard error	Standardized [counts]	±
3	4	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
3	5	9170	9348	9920	9479	392	226	20.1	0.5
3	6	36836	37065	36199	36700	449	259	77.7	0.5
3	7	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
3	8	37990	37305	39059	38118	884	510	80.7	1.1
3	9	47573	47313	46858	47248	362	209	100.0	0.4
3	10	21953	19361	23536	21617	2108	1217	45.8	2.6
3	11	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
3	12	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
3	13	9189	10595	9836	9873	704	406	20.9	0.9
23	4	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
23	5	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
23	6	4690	4269	4703	4554	247	143	9.6	0.3
23	7	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
23	8	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
23	9	9139	9286	9191	9205	75	43	19.5	0.1
23	10	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
23	11	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
23	12	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
23	13	2219	2348	2339	2302	72	42	4.9	0.1
24	4	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
24	5	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
24	6	9164	9439	9160	9254	160	92	19.6	0.2
24	7	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
24	8	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
24	9	17626	17835	19142	18201	822	474	38.5	1.0
24	10	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
24	11	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
24	12	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
24	13	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
25	4	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
25	5	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
25	6	6095	6361	6863	6440	390	225	13.6	0.5
25	7	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
25	8	7707	7292	7514	7504	208	120	15.9	0.3
25	9	4678	4439	4620	4579	125	72	9.7	0.2
25	10	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
25	11	3104	3426	3053	3194	202	117	6.8	0.2
25	12	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
25	13	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
26	4	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
26	5	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
26	6	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
26	7	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
26	8	5621	4881	5519	5340	401	232	11.3	0.5
26	9	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a

Alkyne #	Azide #	Run 1 [counts]	Run 2 [counts]	Run 3 [counts]	Average [counts]	Standard Deviation	Standard error	Standardized [counts]	±
26	10	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
26	11	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
26	12	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a
26	13	n.d.	n.d.	n.d.	n.a	n.a	n.a	n.d	n.a



LCMS counts of triazoles (standardized to largest peak) formed *in situ* click in screen of azides (4-13) and alkynes (3 and 24-26) against *Ls*

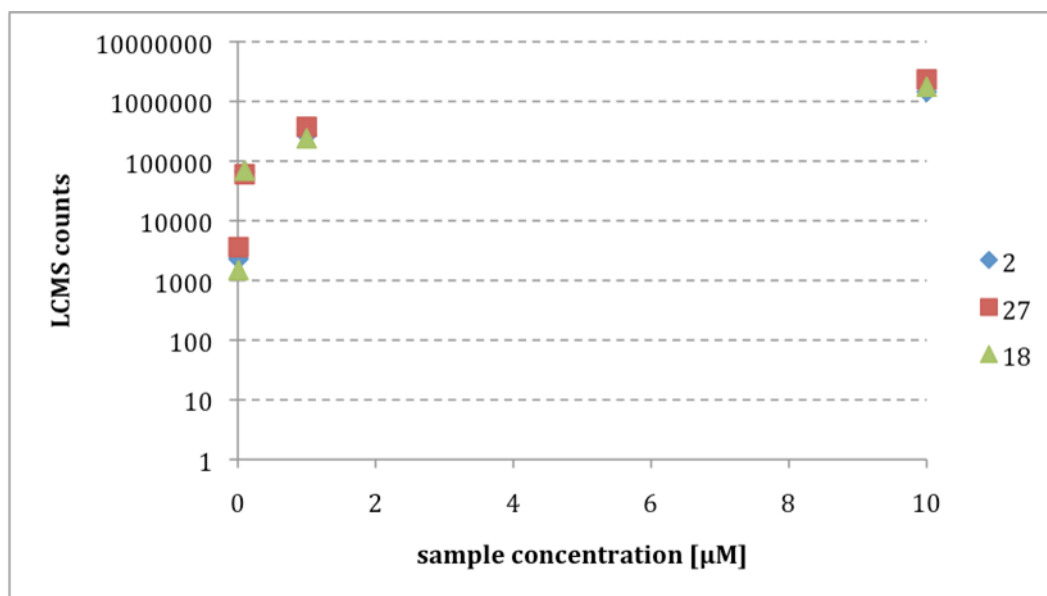
Comparison of Mass Spectrum Response



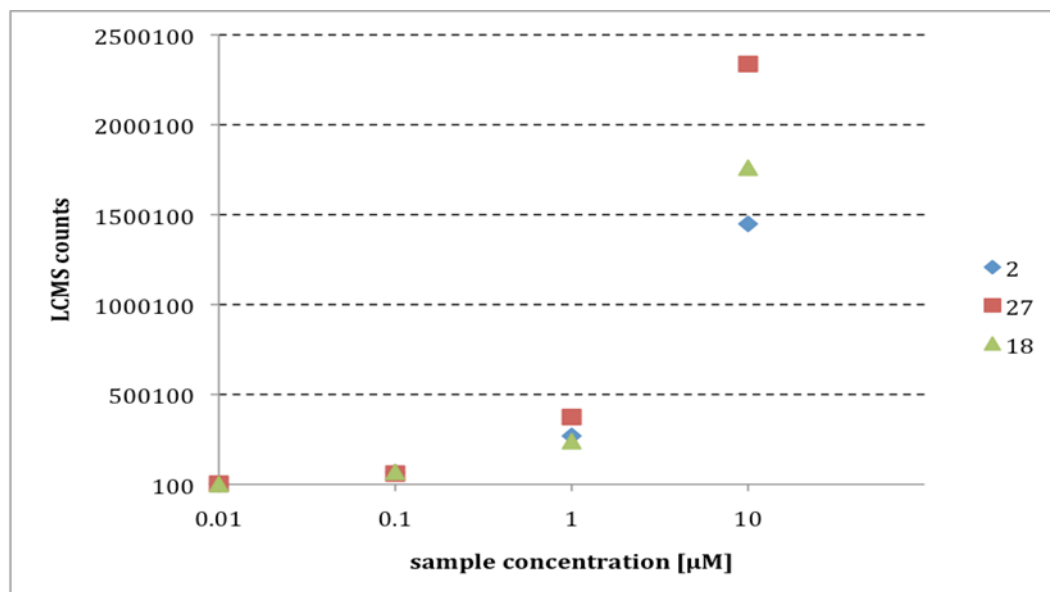
Compounds **2** (starting compound), **18** (varying azide portion) and **27** (varying alkyne portion) were dissolved in water at 1 mM. The solutions were combined and serially diluted to yield the following concentrations: 10 μM , 1 μM , 0.1 μM , 0.01 μM and 0.001 μM with respect to each component. Three samples (25 μL) at each concentration were directly injected into the LC/MS instrument to perform LC/MS-SIM analysis: Zorbax, 4.6 mm x 3 cm, SB-C18 (rapid resolution) reverse phase column, preceded by a Phenomenex C18 guard column, flow rate 0.5 mL/min, using a gradient elution as follows: (H_2O + 0.05% TFA):(MeCN + 0.05 % TFA) 100: 00 to 0:100 over 8 min; then 100 % MeCN + 0.05 % TFA for 2 min with a post run time of 4 min using the starting solvent ratio. Detection was by ESI-MS using positive selected-ion monitoring, tuned to the molecular weights of **2**, **18** and **27** (M^+). The mass response for each peak was integrated (Table S6) and graphed against the concentration (see below). It should be noted that, although the desired peaks were visible to the naked eye at 0.001 μM , they were too small to be integrated accurately, and therefore were not included. These data demonstrate that for these closely related compounds the mass spec response can be used to gauge the amount of compound present in the *in situ* reaction mixtures.

Table S6: LCMS data – compounds 2, 18 and 27 at different concentrations

Concentration [μM]	LCMS counts			Average [LCMS counts]	Compound
10	1476750	1404670	1464700	1448706.667	2
1	272439	262922	273307	269556	2
0.1	75681	59282	65099	66687.33333	2
0.01	2064	1937	2874	2291.666667	2
0.001	N/A	N/A	N/A	0	2
	A	B	C	Average	
10	2430050	2288850	2295840	2338246.667	27
1	382072	374389	366273	374244.6667	27
0.1	66523	57867	55889	60093	27
0.01	3985	3282	3582	3616.333333	27
0.001	N/A	N/A	N/A	0	27
	A	B	C	Average	
10	1703600	1843820	1736990	1761470	18
1	251013	236817	231924	239918	18
0.1	85989	60709	60601	69099.66667	18
0.01	1349	1344	1767	1486.666667	18
0.001	N/A	N/A	N/A	0	18



LCMS counts (logarithmic scale) of triazoles 2, 18 and 27 as a function of concentration



LCMS counts of triazoles 2, 18, and 27 as a function of concentration (logarithmic scale)

Experimental

General Information

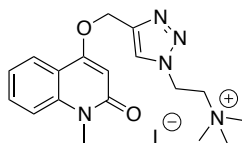
^1H and ^{13}C NMR spectra were recorded on either a Varian Mercury-300 or a Bruker DRX-600 equipped with a DCH cryoprobe and calibrated using residual undeuterated solvent as an internal reference. The following notation is used: br – broad, s – singlet, d – doublet, t – triplet, q – quartet, quin – quintet, m – multiplet, dd – doublet of doublets, ddd – doublet of doublets of doublets, dt – doublet of triplets, td – triplet of doublets, app – apparent. Infrared spectra were recorded on a Nicolet Avatar 370 Fourier transform infrared spectrometer. LCMS analysis was performed on an Agilent 1100 LC/MSD with an Agilent 1100SL mass spectrometer (ES) eluting with 0.1% trifluoroacetic acid in H_2O and 0.05% trifluoroacetic acid in CH_3CN . High resolution mass spectra (HRMS) were recorded on an Agilent Mass spectrometer using ESI-TOF (electrospray ionization-time of flight). Melting points were recorded using a Thomas Hoover Capillary Melting Point apparatus and are reported uncorrected. Unless otherwise noted, analytical grade solvents and commercially available reagents were used without further purification.

General Procedure for Copper Catalyzed Alkyne Azide Cycloaddition

Amino-azide (1 eq, 0.5 mmol) and alkyne (1 eq, 0.5 mmol) were stirred together in *t*-butanol/water (2:1, yielding a final azide/alkyne concentration of 0.1 M). To this suspension was added aq. copper sulfate (0.5 M, 1 mol %) and aq sodium ascorbate (0.1 M, 10 mol %). The reaction was stirred at room temperature until LCMS analysis showed complete consumption of the starting materials. The reaction was subsequently basified with sat aq K_2CO_3 , diluted with water and extracted with ethyl acetate (x3). The organic layers were combined, dried over MgSO_4 , filtered and evaporated under reduced pressure. The resulting residue was dissolved in methyl iodide (1 M) and stirred at room temperature until LC-MS analysis showed complete conversion of the amine intermediate to the desired quaternary ammonium derivative. The reaction was subsequently evaporated to dryness under reduced pressure and the residue purified by

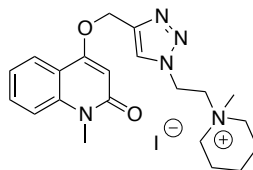
reverse phase column chromatography (Biotage Snap Cartridge KP-C18-HS) to yield the desired salt.

***N,N,N*-trimethyl-2-(4-(((1-methyl-2-oxo-1,2-dihydroquinolin-4-yl)oxy)methyl)-1*H*-1,2,3-triazol-1-yl)ethanaminium iodide (2)**



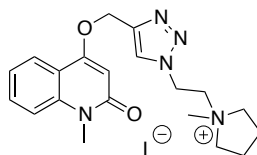
2: (199 mg, 85%) white solid; mp = 161-163 °C; ¹H NMR (300 MHz, D₂O) δ ppm 8.22 (s, 1H), 7.10-7.02 (br m, 2H), 6.69-6.66 (m, 2H), 5.17 (s, 1H), 5.07 (t, *J* = 6.7 Hz, 2H), 4.57 (s, 2H), 4.08 (t, *J* = 6.7 Hz, 2H), 3.28 (s, 9H), 2.85 (s, 3H); ¹³C NMR (75 MHz, D₂O) δ ppm 163.8, 160.9, 143.2, 137.6, 131.6, 131.5, 125.3, 122.48, 114.6, 114.3, 95.2, 63.9, 62.1, 54.0, 44.4, 29.3; FTIR 3450, 1634, 1582, 1234, 1154, 1117, 750 cm⁻¹; HRMS *m/z* 342.1932 [M]⁺, calculated for C₁₈H₂₄N₅O₂⁺ 342.1025.

1-methyl-1-(2-(4-(((1-methyl-2-oxo-1,2-dihydroquinolin-4-yl)oxy)methyl)-1*H*-1,2,3-triazol-1-yl)ethyl)piperidin-1-ium iodide (15)



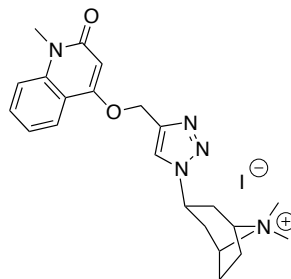
15: (198 mg, 73%) white solid; mp = 150-152 °C; ¹H NMR (300 MHz, DMSO-*d*₆) δ ppm 8.48 (s, 1H), 7.82 (d, *J* = 7.5 Hz, 1H), 7.69 (t, *J* = 8.5 Hz, 1H), 7.54 (d, *J* = 7.5 Hz, 1H), 7.25 (t, *J* = 8.5 Hz, 1H), 6.30 (s, 1H), 5.39 (s, 2H), 5.01 (t, *J* = 6.9 Hz, 2H), 3.95 (t, *J* = 6.9 Hz, 2H), 3.57 (s, 3H), 3.40 (t, *J* = 5.5 Hz, 4H), 3.11 (s, 3H), 1.86-1.71 (m, 4H), 1.61-1.46 (m, 2H); ¹³C NMR (150 MHz, DMSO-*d*₆) δ ppm 162.1, 160.3, 142.2, 139.4, 131.6, 125.6, 122.7, 121.5, 115.4, 114.8, 97.6, 61.8, 60.1, 42.6, 40.4, 40.1, 28.5, 20.5, 19.2; FTIR 3442, 1637, 1580, 1217, 689 cm⁻¹; HRMS *m/z* 382.2251 [M+H]⁺, calculated for C₂₁H₂₈N₅O₂⁺ 382.2238.

1-methyl-1-(2-(4-(((1-methyl-2-oxo-1,2-dihydroquinolin-4-yl)oxy)methyl)-1*H*-1,2,3-triazol-1-yl)ethyl)pyrrolidin-1-ium iodide (16)



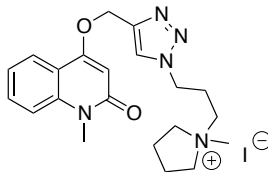
16: (202 mg, 82%) off-white solid; mp = 148-151 °C; ¹H NMR (600 MHz, D₂O) δ ppm 8.34 (s, 1H), 7.68 (dd, *J* = 8.1, 1.5 Hz, 1H), 7.55 (td, *J* = 8.6, 1.5 Hz, 1H), 7.30 (app d, *J* = 8.6 Hz, 1H), 7.18 (app t, *J* = 8.1 Hz, 1H), 5.86 (s, 1H), 5.22 (s, 2H), 5.10 (t, *J* = 6.4 Hz, 2H), 4.05 (t, *J* = 6.4 Hz, 2H), 3.53-3.46 (m, 4H), 3.38 (s, 3H), 3.11 (s, 3H), 2.17 (m, 4H); ¹³C NMR (150 MHz, D₂O) δ ppm 165.5, 162.6, 143.9, 139.2, 132.8, 126.6, 123.7, 123.5, 116.4, 115.7, 96.7, 66.0, 62.8, 62.2, 48.6, 45.7, 30.1, 21.9; FTIR 3362, 1630, 1576, 1236, 1117, 751 cm⁻¹; HRMS *m/z* 368.2089 [M]⁺, calculated for C₂₀H₂₆N₅O₂⁺ 368.2081.

8,8-dimethyl-3-(4-(((1-methyl-2-oxo-1,2-dihydroquinolin-4-yl)oxy)methyl)-1*H*-1,2,3-triazol-1-yl)-8-azabicyclo[3.2.1]octan-8-ium iodide (18)



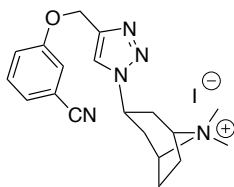
18: (229 mg, 88%) white solid; mp = 186-188 °C; ¹H NMR (600 MHz, DMSO-*d*₆ + 1 drop CDCl₃) δ ppm 8.65 (s, 1H), 7.85 (app d, *J* = 8.0 Hz, 1H), 7.64 (app t, *J* = 8.5 Hz, 1H), 7.51 (app d, *J* = 8.5 Hz, 1H), 7.23 (app t, *J* = 8.0 Hz, 1H), 6.28 (s, 1H), 5.33 (s, 2H), 4.85 (m, 1H), 3.94 (m, 2H), 3.55 (s, 3H), 3.20 (s, 3H), 3.01 (s, 3H), 2.85-2.81 (m, 4H), 2.23-2.15 (m, 2H), 1.75-1.69 (m, 2H); ¹³C NMR (150 MHz, DMSO-*d*₆ + 1 drop CDCl₃) δ 163.5, 161.7, 143.7, 140.7, 132.9, 126.0, 124.1, 122.9, 116.7, 116.1, 99.0, 67.4, 63.3, 54.0, 51.1, 48.9, 44.8, 31.1, 25.0; FTIR 3448, 1637, 1577, 1240, 765, 750 cm⁻¹; HRMS *m/z* 394.2245 [M]⁺, calculated for C₂₂H₂₈N₅O₂⁺ 394.2243.

1-methyl-1-(2-(4-(((1-methyl-2-oxo-1,2-dihydroquinolin-4-yl)oxy)methyl)-1*H*-1,2,3-triazol-1-yl)ethyl)pyrrolidin-1-ium iodide (19)



19: (218 mg, 86%) white solid; mp = 156-158 °C; ¹H NMR (600 MHz, D₂O) δ ppm 8.21 (s, 1H), 7.58 (dd, *J* = 8.1, 1.4 Hz, 1H), 7.47 (td, *J* = 8.6, 1.48 Hz, 1H), 7.20 (app d, *J* = 8.1 Hz, 1H), 7.13 (app t, *J* = 8.6 Hz, 1H), 5.78 (s, 1H), 5.13 (s, 2H), 4.61 (t, *J* = 6.5 Hz, 2H), 3.52-3.36 (m, 6H), 3.30 (s, 3H), 3.03 (s, 3H), 2.54-2.45 (m, 2H), 2.14-2.03 (m, 4H); ¹³C NMR (150 MHz, D₂O) δ ppm 165.4, 162.4, 143.3, 139.1, 132.7, 126.1, 123.5, 123.4, 116.3, 115.6, 96.5, 65.3, 62.2, 61.5, 48.8, 48.0, 30.0, 24.9, 21.9; FTIR 3367, 1636, 1578, 1461, 1237, 751 cm⁻¹; HRMS *m/z* 382.2243 [M+H]⁺, calculated for C₂₁H₂₈N₅O₂⁺ 382.2246.

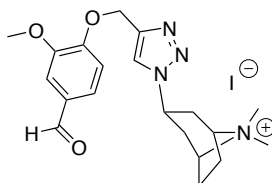
3-(4-((3-cyanophenoxy)methyl)-1*H*-1,2,3-triazol-1-yl)-8,8-dimethyl-8-azabicyclo[3.2.1]octan-8-ium iodide (27)



27: (214 mg, 92%) white solid; mp = 225-228 °C; ¹H NMR (600 MHz, DMSO-*d*₆ + 1 drop CDCl₃) δ ppm 8.41 (s, 1H), 7.58-7.56 (m, 1H), 7.52 (dd, *J* = 7.9, 1.1 Hz, 1H), 7.45 (app dt, *J* = 7.6, 1.1 Hz, 1H), 7.39 (ddd, *J* = 8.3, 2.5, 0.8 Hz, 1H), 5.25 (m, 3H), 4.06 (br s, 2H), 3.32 (s, 3H), 3.10 (s, 3H), 2.65-2.60 (m, 2H), 2.42-2.40 (m, 2H), 2.36-2.31 (m, 2H), 2.22-2.19 (m, 2H); ¹³C NMR (150 MHz, DMSO-*d*₆ + 1 drop CDCl₃) δ ppm 158.1, 141.9, 130.9, 124.9, 124.2, 120.5, 118.7, 117.5, 112.2, 68.0, 60.9, 50.2, 48.3, 44.2, 31.8, 24.4;

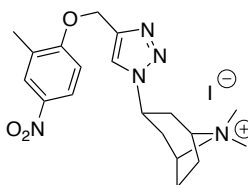
FTIR 3026, 2239, 1678, 1593, 1255, 1025, 797 cm^{-1} ; HRMS m/z 338.1985 $[\text{M}]^+$, calculated for $\text{C}_{19}\text{H}_{24}\text{N}_5\text{O}^+$ 338.1975.

8,8-dimethyl-3-(4-(((1-methyl-2-oxo-1,2-dihydroquinolin-4-yl)oxy)methyl)-1*H*-1,2,3-triazol-1-yl)-8-azabicyclo[3.2.1]octan-8-ium iodide (28)



28: (268 mg, 93%) white solid; mp = 210-212 $^{\circ}\text{C}$; ^1H NMR (600 MHz, $\text{DMSO-}d_6$ + 1 drop CDCl_3) δ ppm 9.86 (s, 1H), 8.41 (s, 1H), 7.57 (dd, J = 6.0, 1.8 Hz, 1H), 7.42 (s, 1H), 7.41 (d, J = 6.0 Hz, 1H), 5.28 (s, 2H), 5.26-5.21 (m, 1H), 4.04 (br s, 2H), 3.81 (s, 3H), 3.12 (s, 3H), 3.09 (s, 3H), 2.64-2.60 (m, 2H), 2.42-4.40 (m, 2H), 2.34-2.31 (m, 2H), 2.21-2.19 (m, 2H); ^{13}C NMR (150 MHz, $\text{DMSO-}d_6$ + 1 drop CDCl_3) δ ppm 191.5, 152.8, 149.3, 141.9, 129.9, 124.6, 112.5, 109.7, 67.9, 61.7, 55.6, 49.9, 48.2, 47.5, 43.7, 31.9, 24.6; FTIR 2959, 1678, 1586, 1514, 1271, 1151, 995 cm^{-1} ; HRMS m/z 371.2090 $[\text{M}]^+$, calculated for $\text{C}_{20}\text{H}_{27}\text{N}_4\text{O}_3^+$ 371.2078.

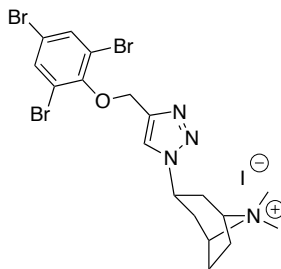
8,8-dimethyl-3-(4-((2-methyl-4-nitrophenoxy)methyl)-1*H*-1,2,3-triazol-1-yl)-8-azabicyclo[3.2.1]octan-8-ium iodide (29)



29: (280 mg, 89%) yellow solid; mp = 200-202 $^{\circ}\text{C}$; ^1H NMR (600 MHz, $\text{DMSO-}d_6$ + 1 drop CDCl_3) δ ppm 8.41 (s, 1H), 8.15 (dd, J = 9.0, 2.8 Hz, 1H), 8.11 (d, J = 2.8 Hz, 1H), 7.40 (d, J = 9.0 Hz, 1H), 5.37 (s, 2H), 5.22 (m, 1H), 4.05 (br s, 2H), 3.31 (s, 3H), 3.09 (s, 3H), 2.65-2.60 (m, 2H), 2.43-2.39 (m, 2H), 2.34-2.30 (m, 2H), 2.22 (s, 3H), 2.20-2.17 (m, 2H); ^{13}C NMR (150 MHz, $\text{DMSO-}d_6$ + 1 drop CDCl_3) δ ppm 161.5, 141.9, 140.4, 127.5, 125.7,

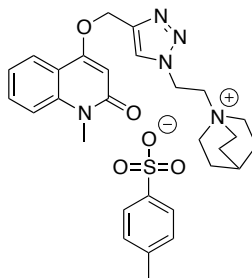
124.3, 123.6, 111.7, 67.9, 62.1, 50.0, 48.2, 43.8, 31.9, 24.4, 16.0; FTIR 3453, 1590, 1521, 1334, 1274, 1094, 749 cm^{-1} ; HRMS m/z 372.2037 $[\text{M}]^+$, calculated for $\text{C}_{19}\text{H}_{26}\text{N}_5\text{O}_3^+$ 372.2030.

8,8-dimethyl-3-(4-((2,4,6-tribromophenoxy)methyl)-1*H*-1,2,3-triazol-1-yl)-8-azabicyclo[3.2.1]octan-8-ium iodide (30)



30: (255 mg, 76%) tan solid; mp = 223-225 $^{\circ}\text{C}$; ^1H NMR (600 MHz, $\text{DMSO-}d_6$ + 1 drop CDCl_3) δ ppm 8.47 (s, 1H), 7.98 (s, 2H), 5.28-5.22 (m, 1H), 5.11 (s, 2H), 4.05 (br s, 2H), 3.32 (s, 3H), 3.10 (s, 3H), 2.67-2.59 (m, 2H), 2.42-2.39 (m, 2H), 2.33-2.28 (m, 2H), 2.22-2.18 (m, 2H); ^{13}C NMR (150 MHz, $\text{DMSO-}d_6$ + 1 drop CDCl_3) δ ppm 151.6, 141.4, 135.0, 124.8, 119.1, 117.4, 68.0, 66.1, 50.2, 47.9, 44.1, 31.7, 24.3; FTIR 3055, 1438, 1234, 949, 734 cm^{-1} ; HRMS m/z 546.9344 $[\text{M}]^+$, calculated for $\text{C}_{18}\text{H}_{22}\text{Br}_3\text{N}_4\text{O}^+$ 546.9338.

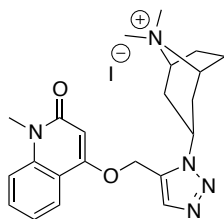
1-(2-(4-(((1-methyl-2-oxo-1,2-dihydroquinolin-4-yl)oxy)methyl)-1*H*-1,2,3-triazol-1-yl)ethyl)quinuclidin-1-ium 4-methylbenzenesulfonate (14)



2-Azidoethyl 4-methylbenzenesulfonate (120 mg, 0.5 mmol) and alkyne **3** (106 mg, 0.5 mmol) were stirred together in *t*-butanol/water (2:1, 5 mL). To this suspension was added copper sulfate (0.5 M in water, 12.5 μL , 1 mol%) and aq. sodium ascorbate (0.1 M, 500 μL , 10 mol%). The reaction was stirred at room temperature for 16 h. Water (20

mL) was added and the mixture was extracted with ethyl acetate (X3, 20 mL). The organic layers were combined, dried over MgSO₄, filtered and evaporated under reduced pressure. The residue was suspended in toluene (10 mL) and quinuclidine (56 mg, 0.5 mmol) added. The reaction was stirred at reflux for 2 h, then allowed to cool to room temperature and evaporated under reduced pressure. The residue was purified by reverse phase column chromatography (Biotage Snap Cartridge KP-C18-HS) to yield the salt **14** as an off white solid (259 mg, 92%); mp = 202-205 °C; ¹H NMR (600 MHz, D₂O) δ ppm 8.24 (s, 1H), 7.57 (d, *J* = 8.2 Hz, 2H), 7.43 (d, *J* = 8.0 Hz, 1H), 7.38 (t, *J* = 8.5 Hz, 1H), 7.17 (d, *J* = 8.2 Hz, 2H), 7.04 (d, *J* = 8.5 Hz, 1H), 7.02 (t, *J* = 8.0 Hz, 1H), 5.64 (s, 1H), 5.01 (s, 2H), 4.96 (t, *J* = 6.7 Hz, 2H), 3.74 (t, *J* = 6.7 Hz, 2H), 3.42-3.34 (m, 9H), 3.19 (s, 3H), 2.23 (quin, *J* = 3.3 Hz, 1H), 1.98-1.90 (m, 6H); ¹³C NMR (150 MHz, D₂O) δ ppm 165.2, 162.2, 143.6, 142.9, 140.4, 138.9, 132.6, 130.1, 126.5, 126.0, 123.4, 123.2, 116.6, 116.0, 93.4, 62.4, 62.2, 56.1, 44.2, 29.9, 24.0, 21.2, 19.5; FTIR 3489, 1638, 1589, 1209, 1186, 1010, 678 cm⁻¹; HRMS *m/z* 394.2243 [M]⁺, calculated for C₂₂H₂₈N₅O₂⁺ 394.2243.

8,8-dimethyl-3-(5-(((1-methyl-2-oxo-1,2-dihydroquinolin-4-yl)oxy)methyl)-1*H*-1,2,3-triazol-1-yl)-8-azabicyclo[3.2.1]octan-8-ium iodide (18a)

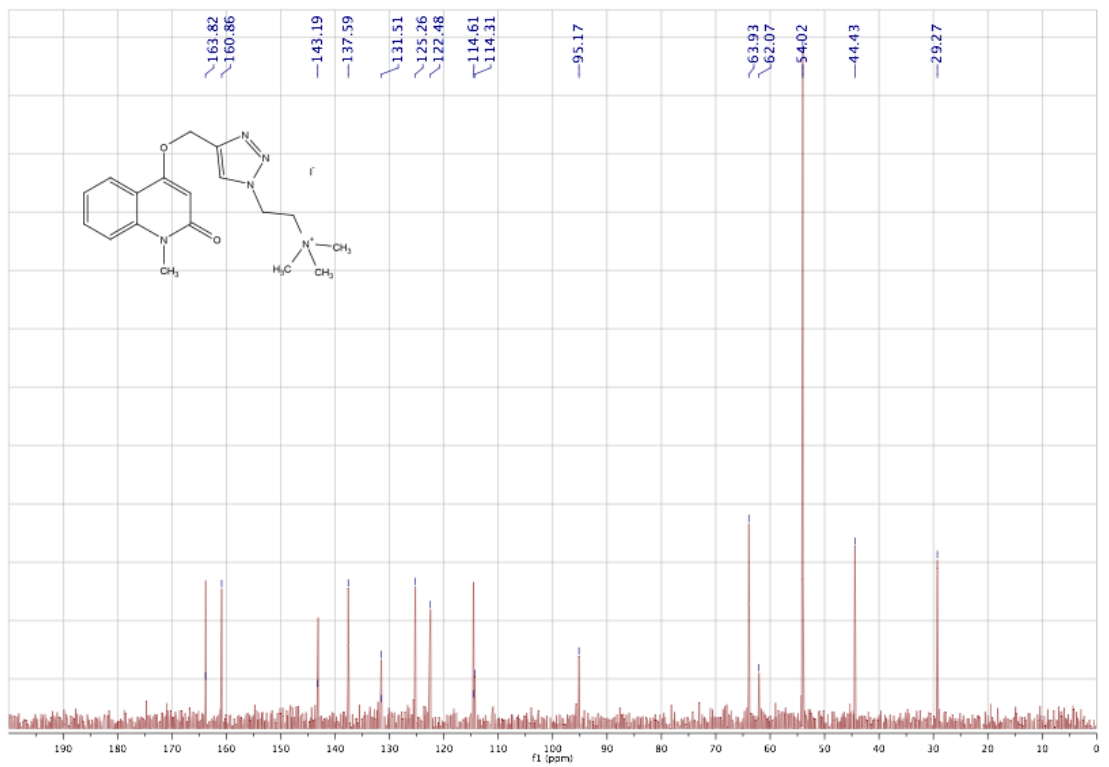
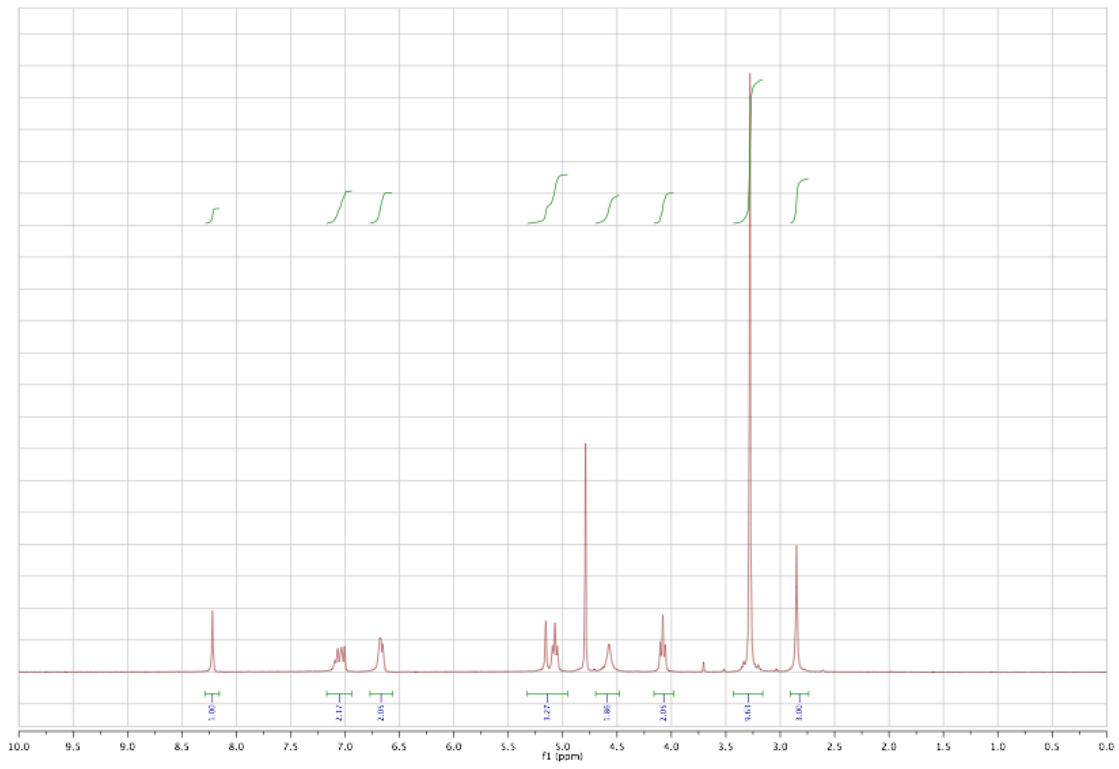


A solution of alkyne **3** (106 mg, 0.5 mmol) and azide **31** (83 mg, 0.5 mmol) in dioxane (600 μL) was added to Cp^{*}RuCl(PPh₃)₂ (12 mg, 0.015 mmol) dissolved in dioxane (3 mL). The vial was purged with nitrogen, sealed and heated at 60 °C. After 16 h, water (20 mL) was added and the mixture was extracted with ethyl acetate (X3, 10 mL). The organic layers were combined, dried over MgSO₄, filtered and evaporated under reduced pressure. The resultant residue was dissolved in methyl iodide (500 μL) and stirred at room temperature for 1 hour, then evaporated to dryness under reduced pressure and purified by reverse phase column chromatography (Biotage Snap

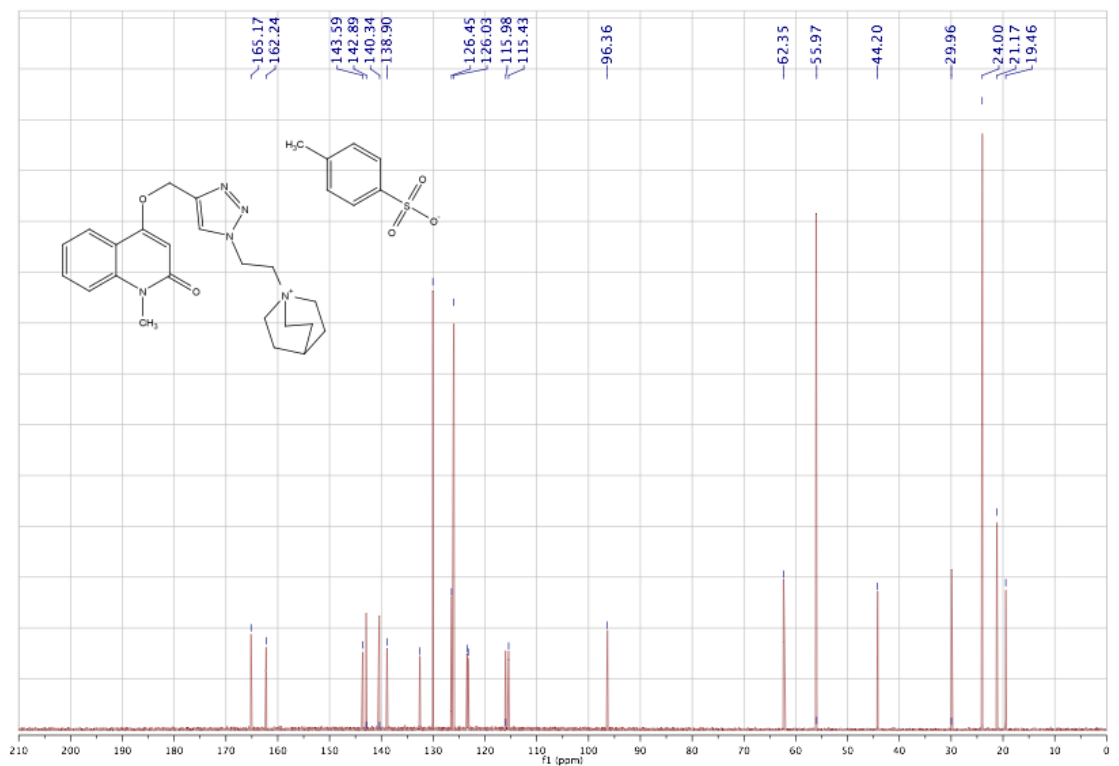
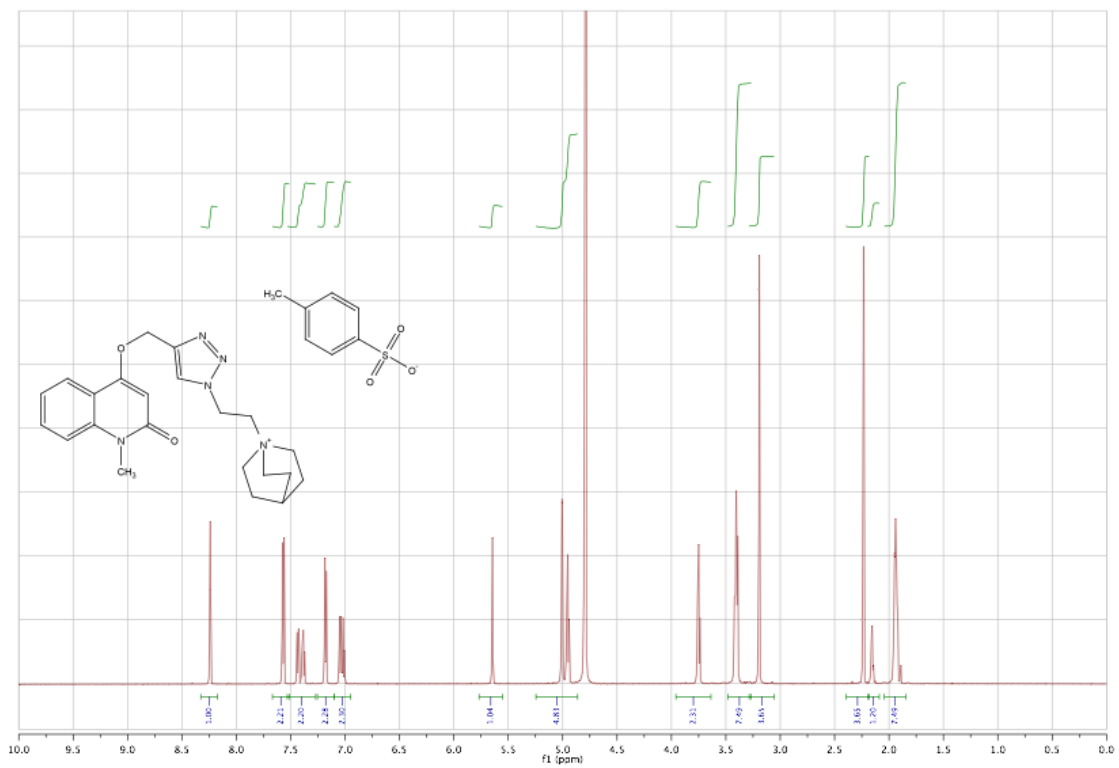
Cartridge KP-C18-HS) yielding **18a** as a brown solid (187 mg, 72%); mp = 194-196 °C; ¹H NMR (600 MHz, DMSO-*d*₆) δ ppm 8.02 (s, 1H), 7.90 (app d, *J* = 7.9 Hz, 1H), 7.67 (app t, *J* = 8.4 Hz, 1H), 7.53 (app d, *J* = 8.9 Hz, 1H), 7.27 (app t, *J* = 7.4 Hz, 1H), 6.36 (s, 1H), 5.62 (s, 2H), 5.11 (m, 1H), 4.08 (m, 2H), 3.57 (s, 3H), 3.37 (s, 3H), 3.10 (s, 3H), 2.75 (t, *J* = 12.24 Hz, 2H), 2.40 (m, 2H), 2.30 (m, 2H), 2.219 (m, 2H); ¹³C NMR (150 MHz, DMSO-*d*₆) δ ppm 161.4, 161.2, 140.8, 133.9, 133.0, 123.8, 123.1, 116.4, 116.2, 99.3, 69.1, 60.4, 51.2, 47.7, 45.1, 33.7, 32.3, 30.1, 25.6; FTIR 3407, 1631, 1580, 1457, 1231, 1114, 753 cm⁻¹; HRMS *m/z* 394.2245 [M]⁺, calculated for C₂₂H₂₈N₅O₂⁺ 394.2248.

NMR Spectra

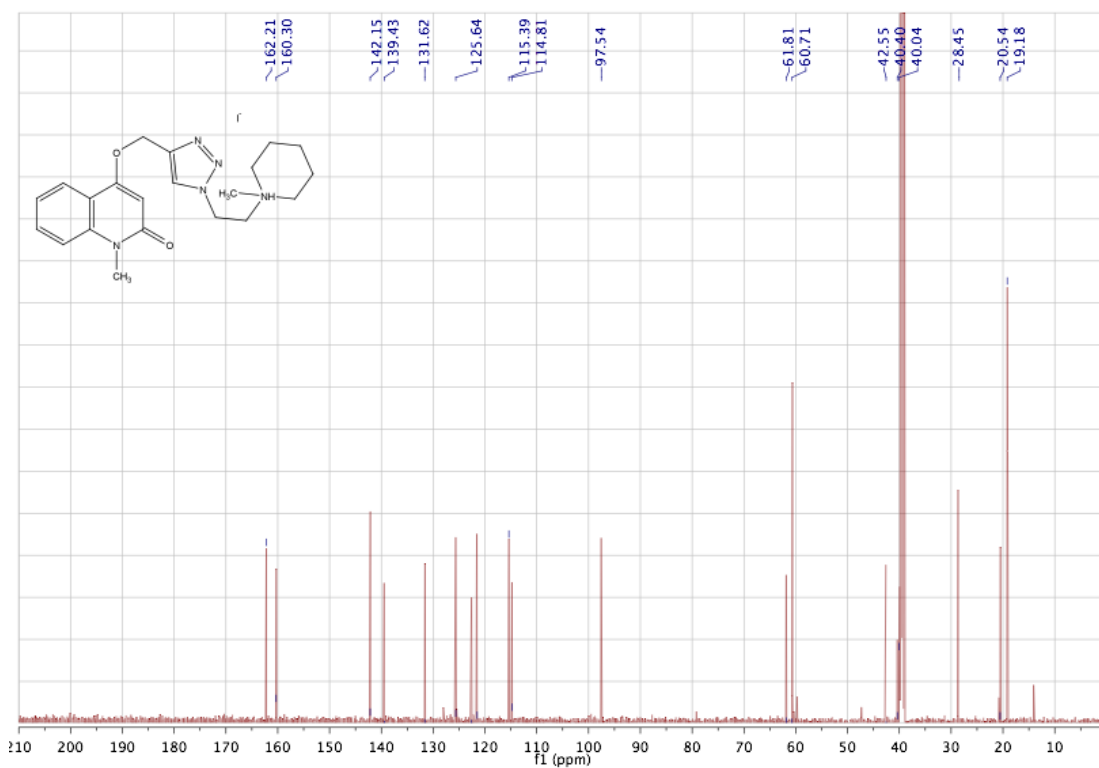
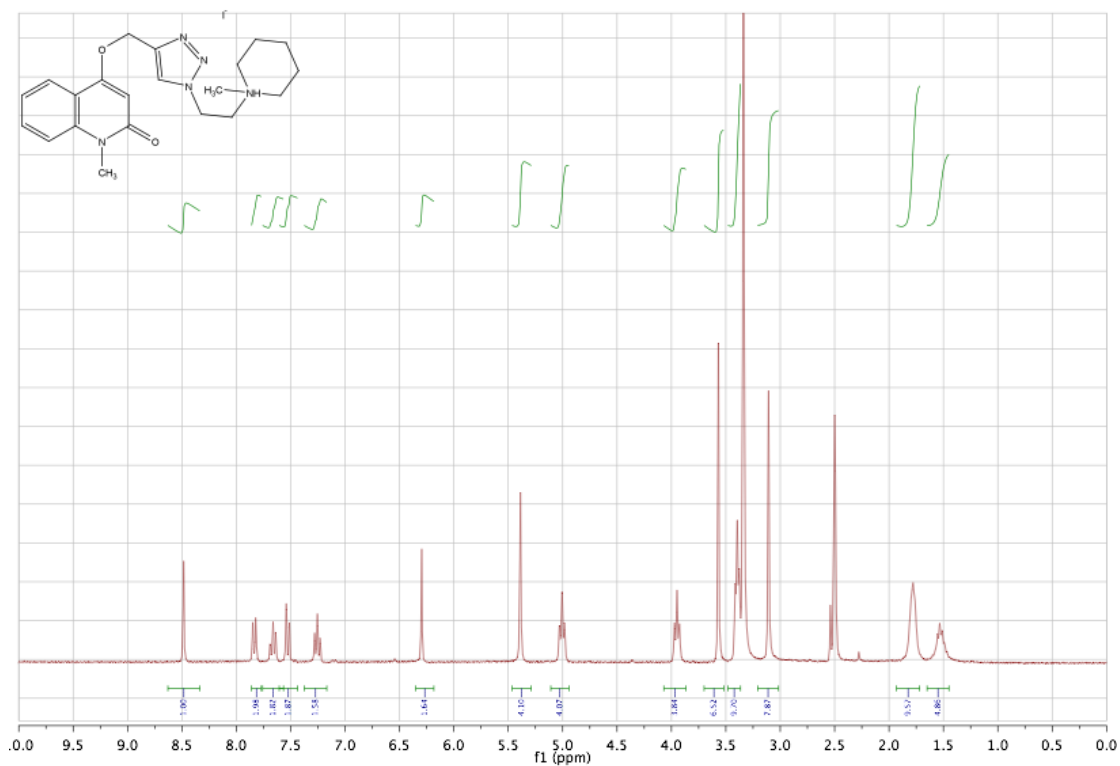
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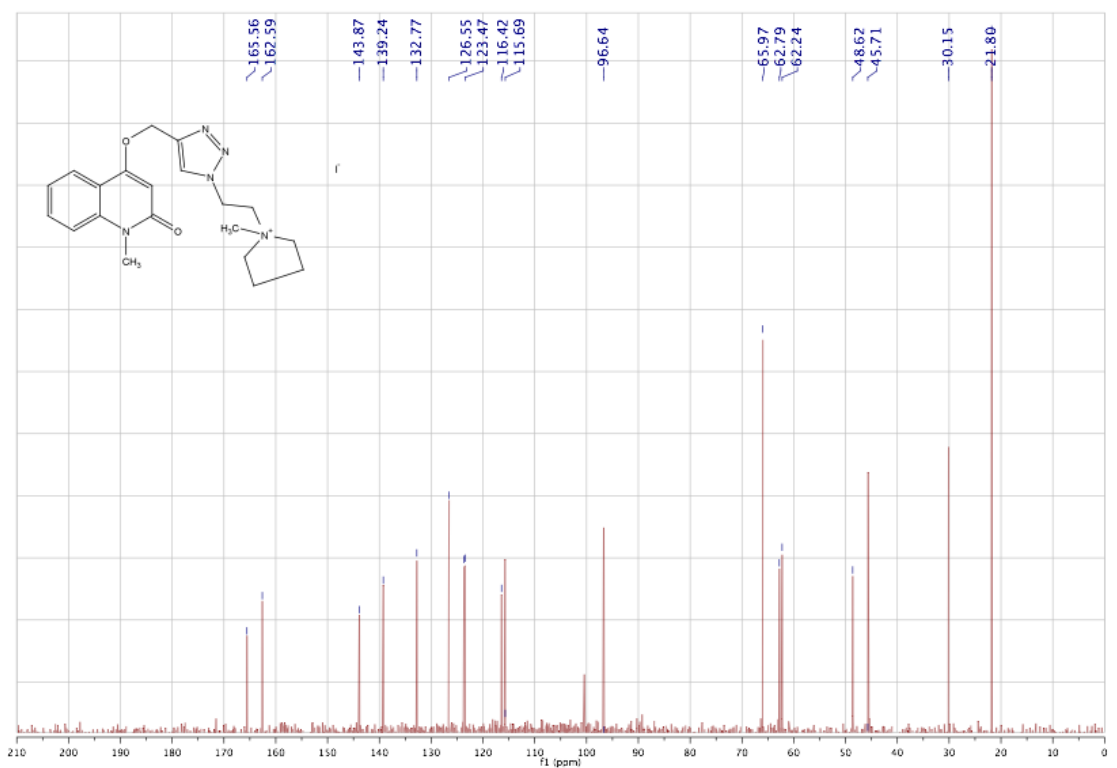
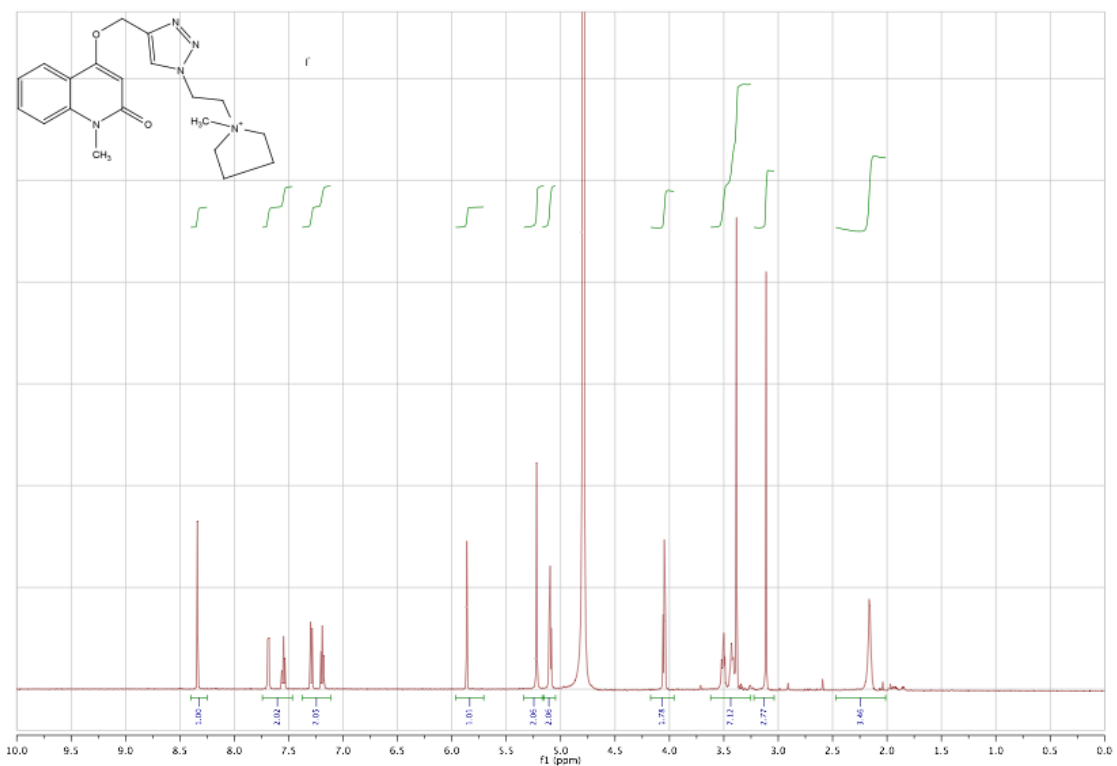
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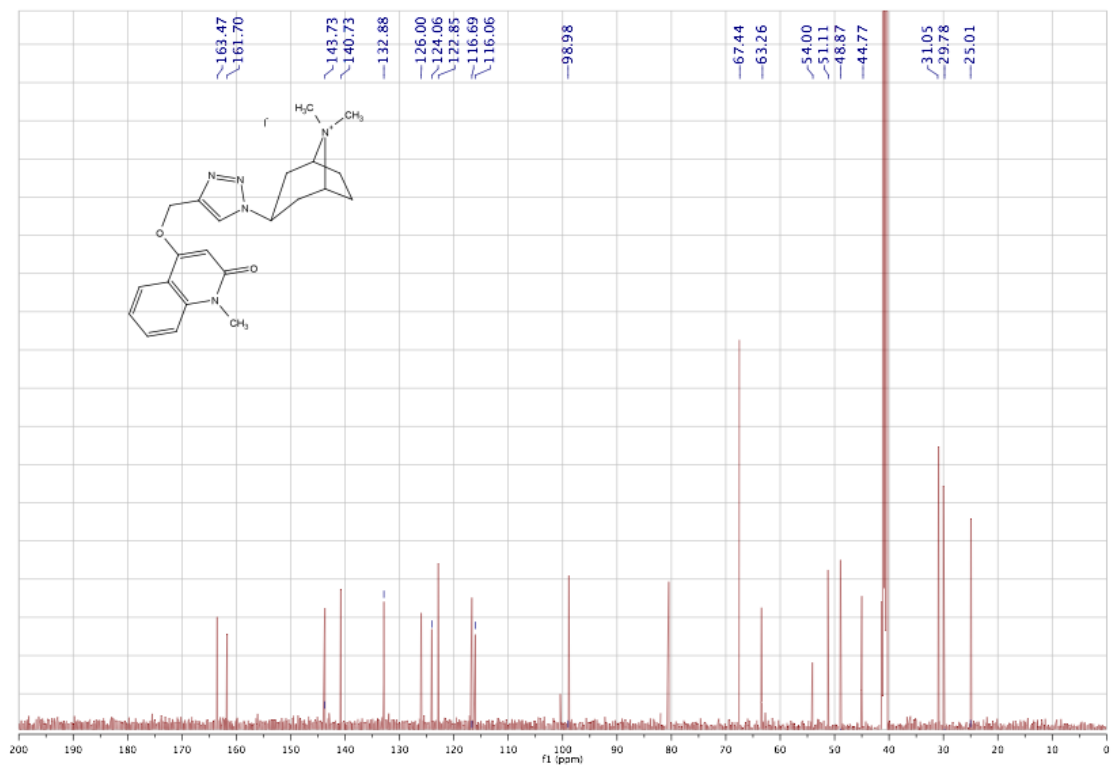
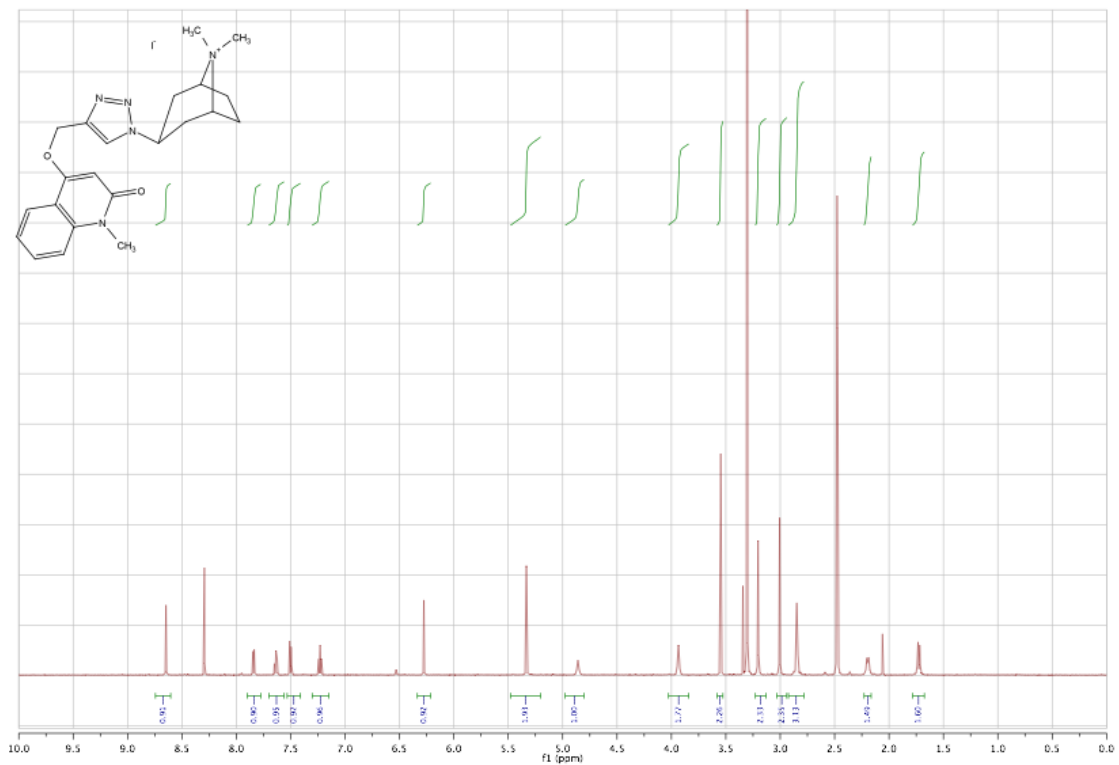
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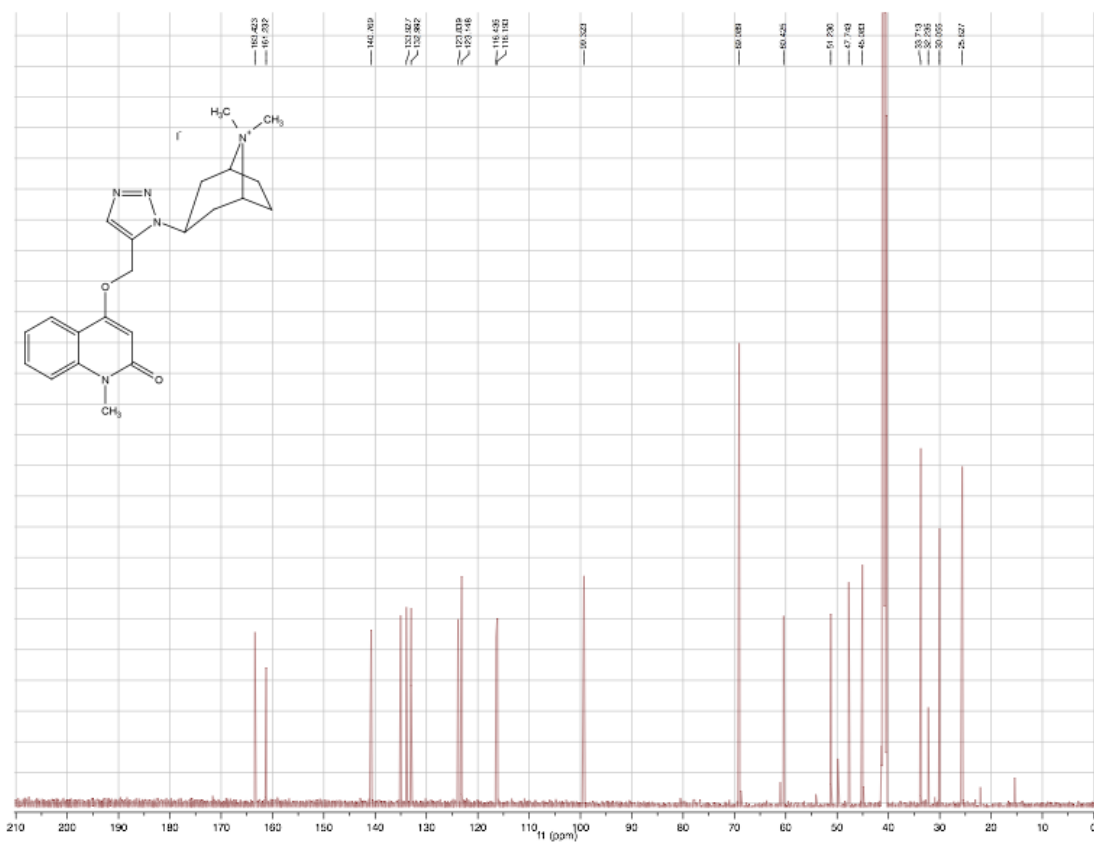
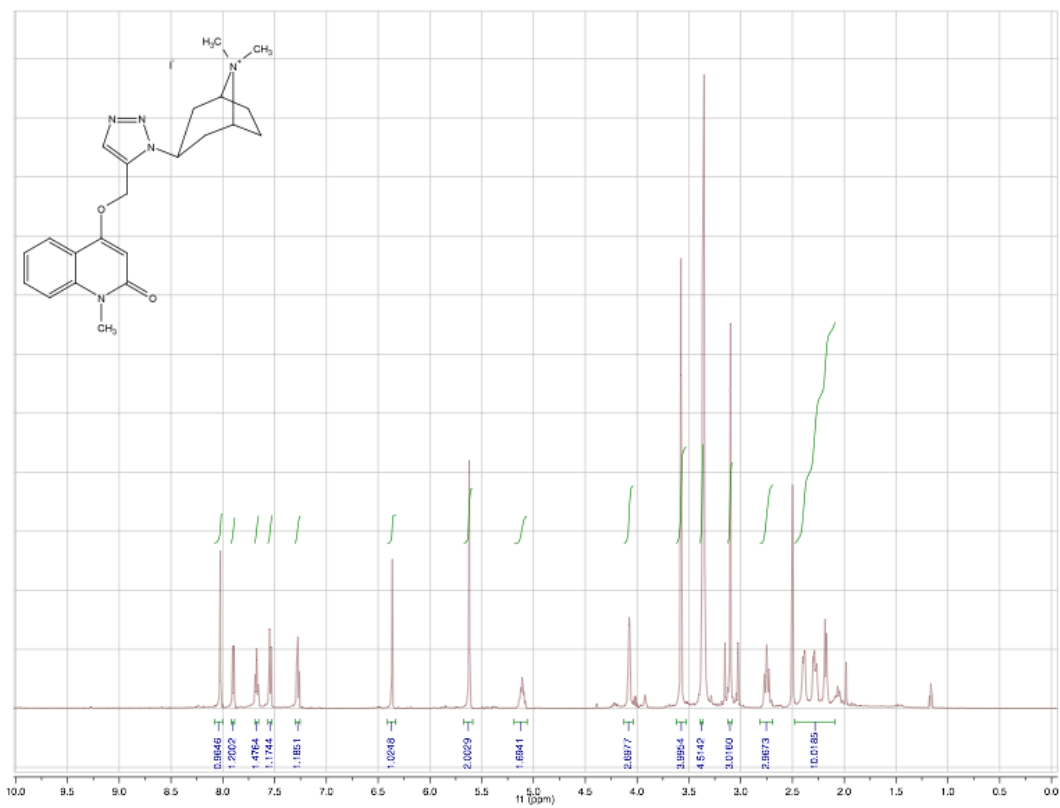
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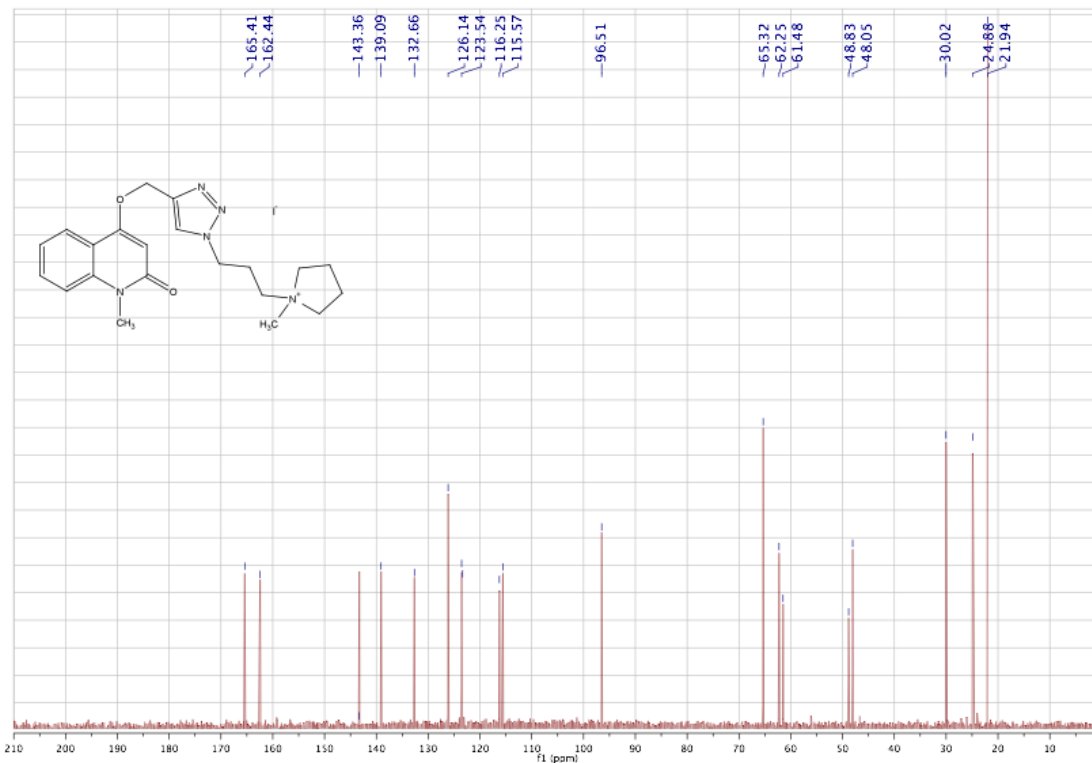
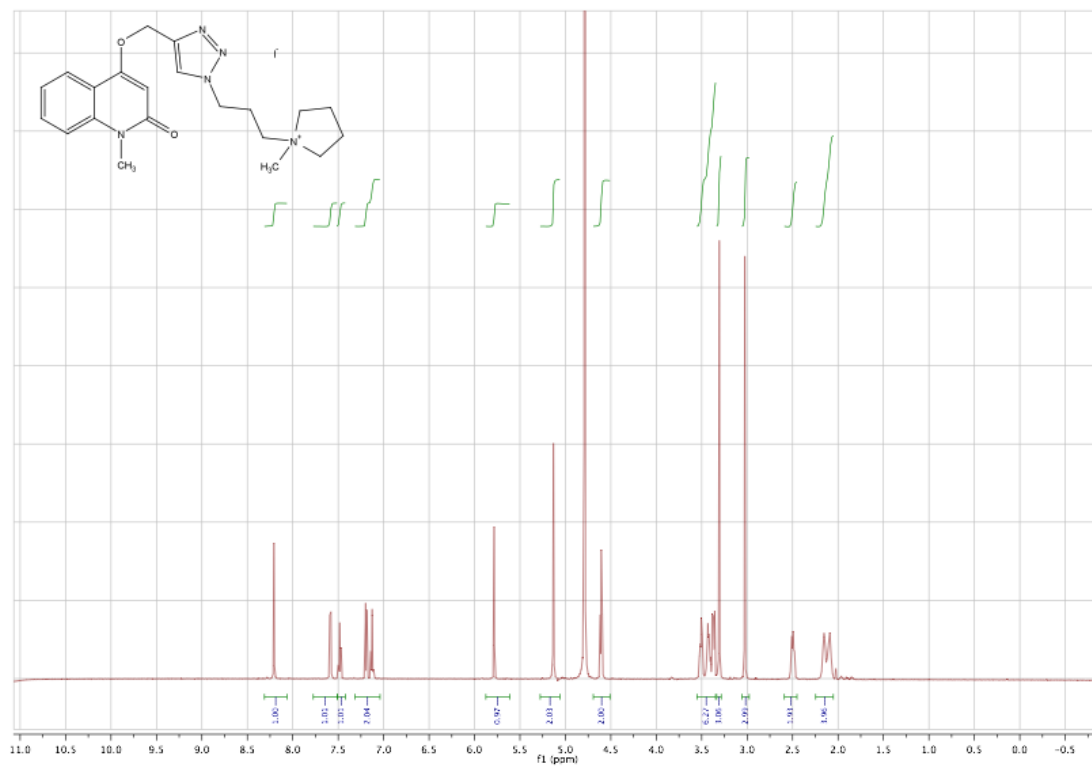
18:



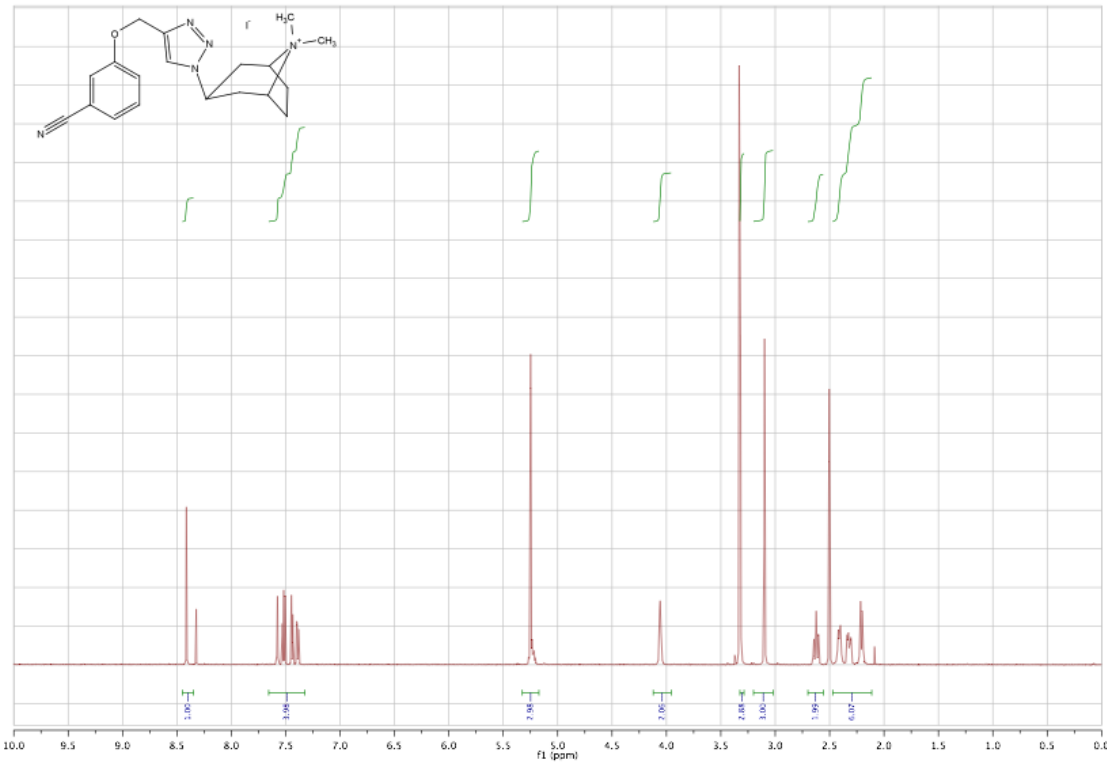
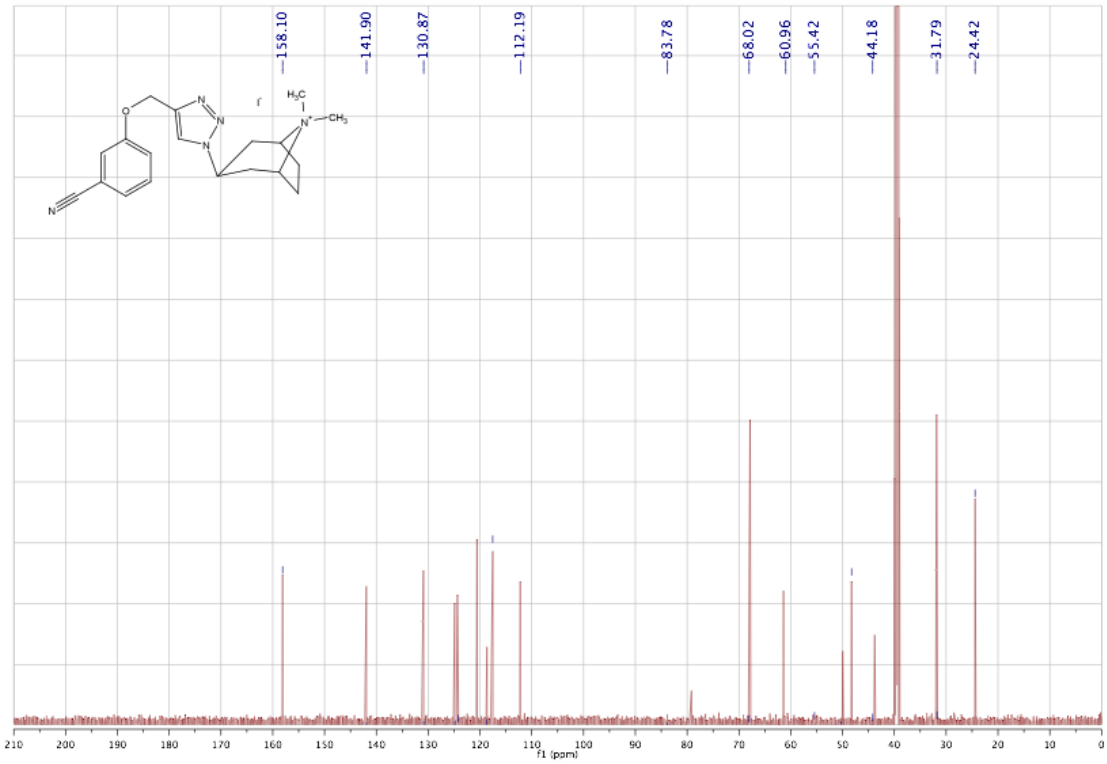
18a:



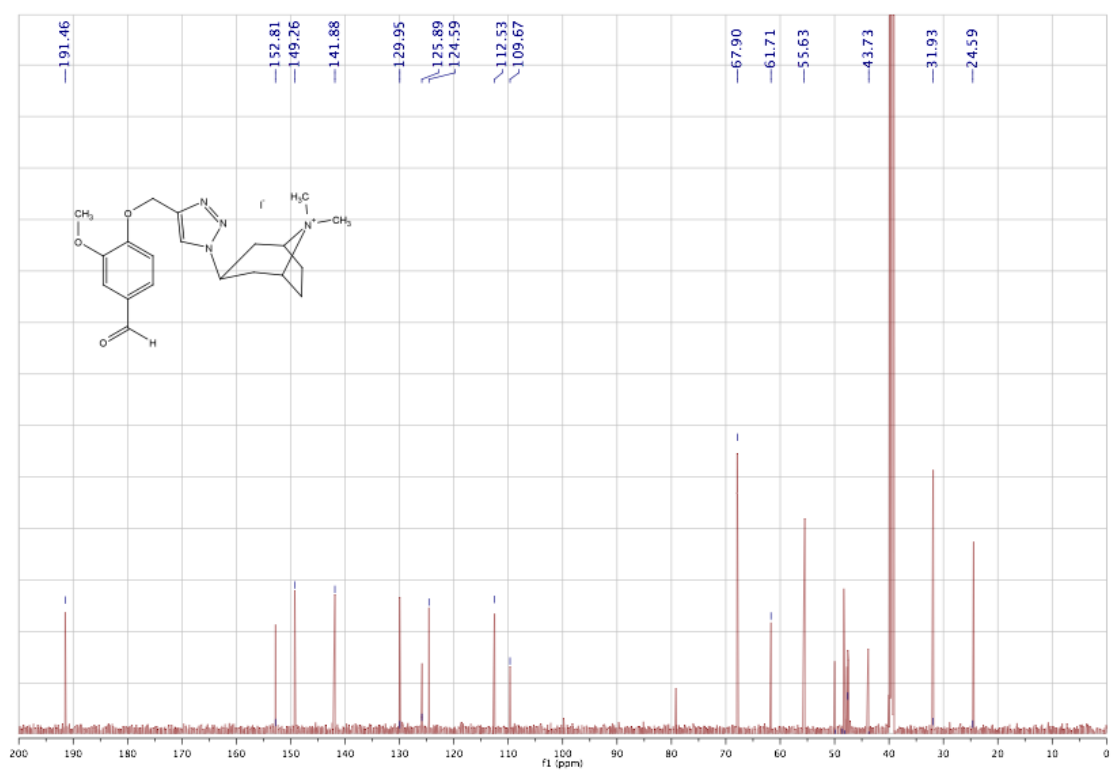
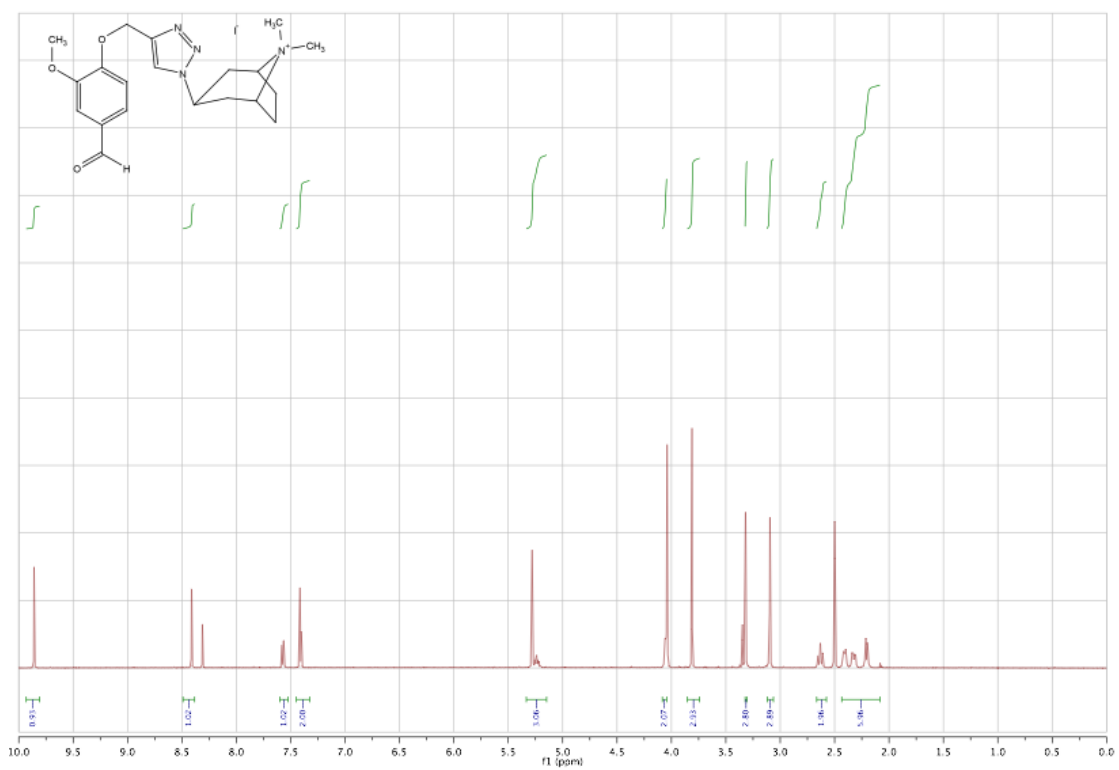
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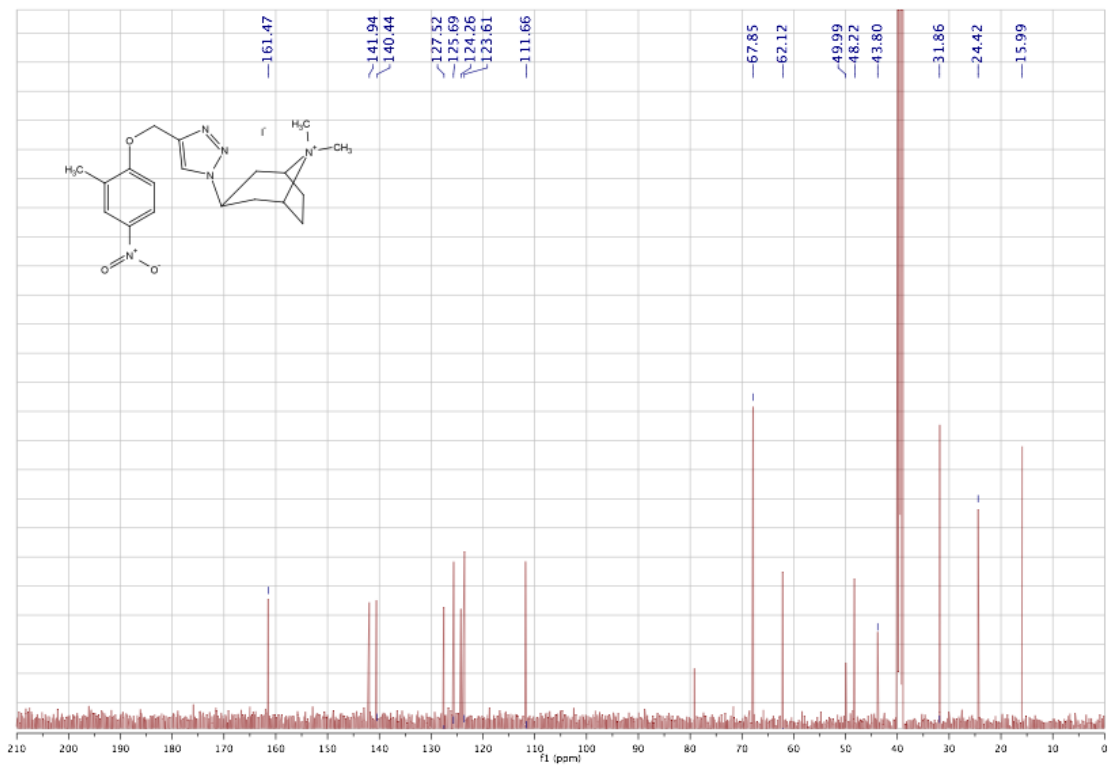
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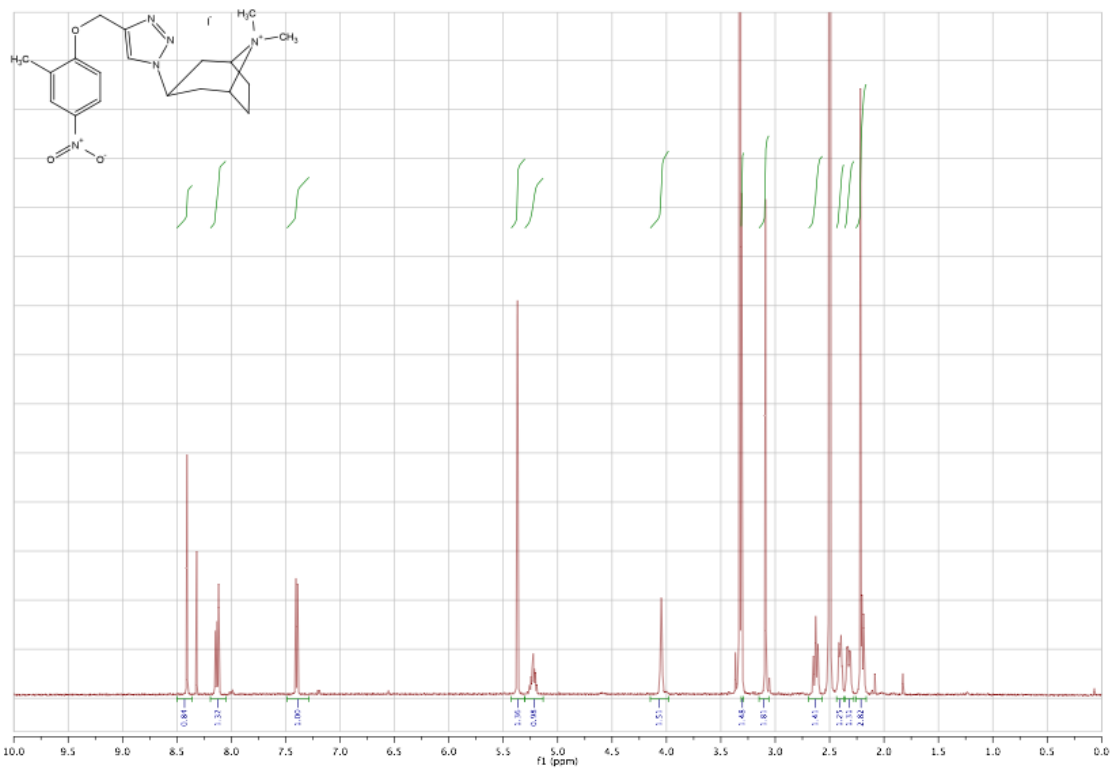


28:

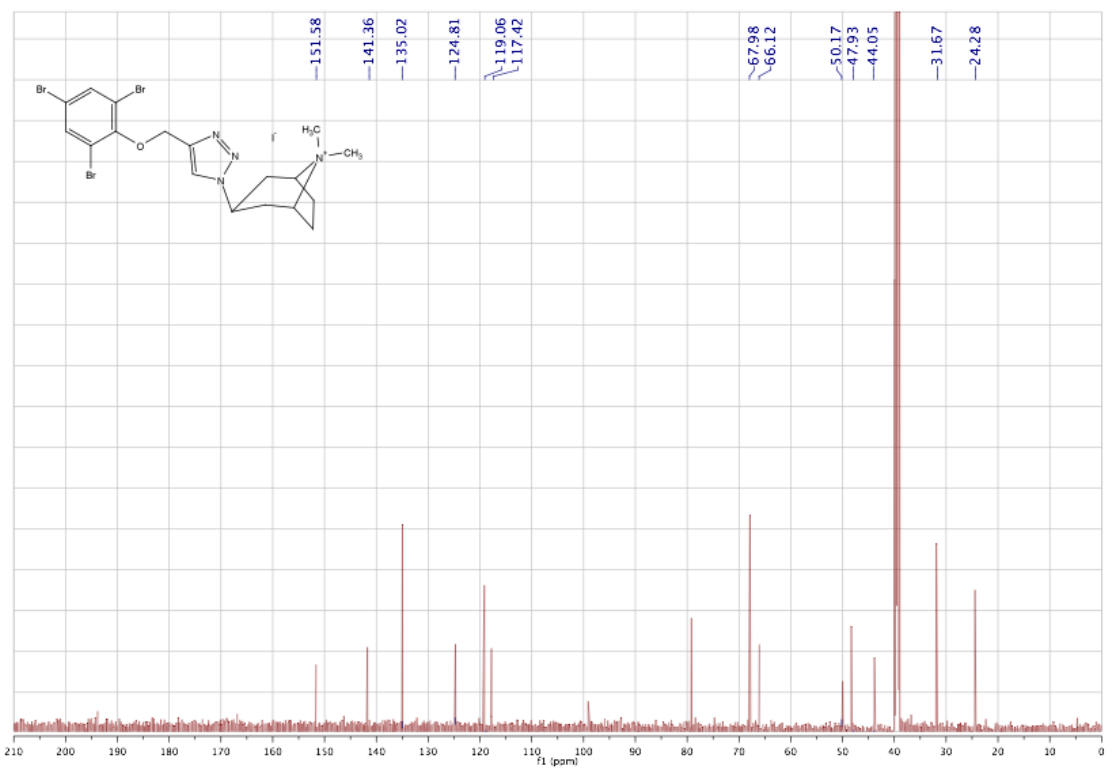


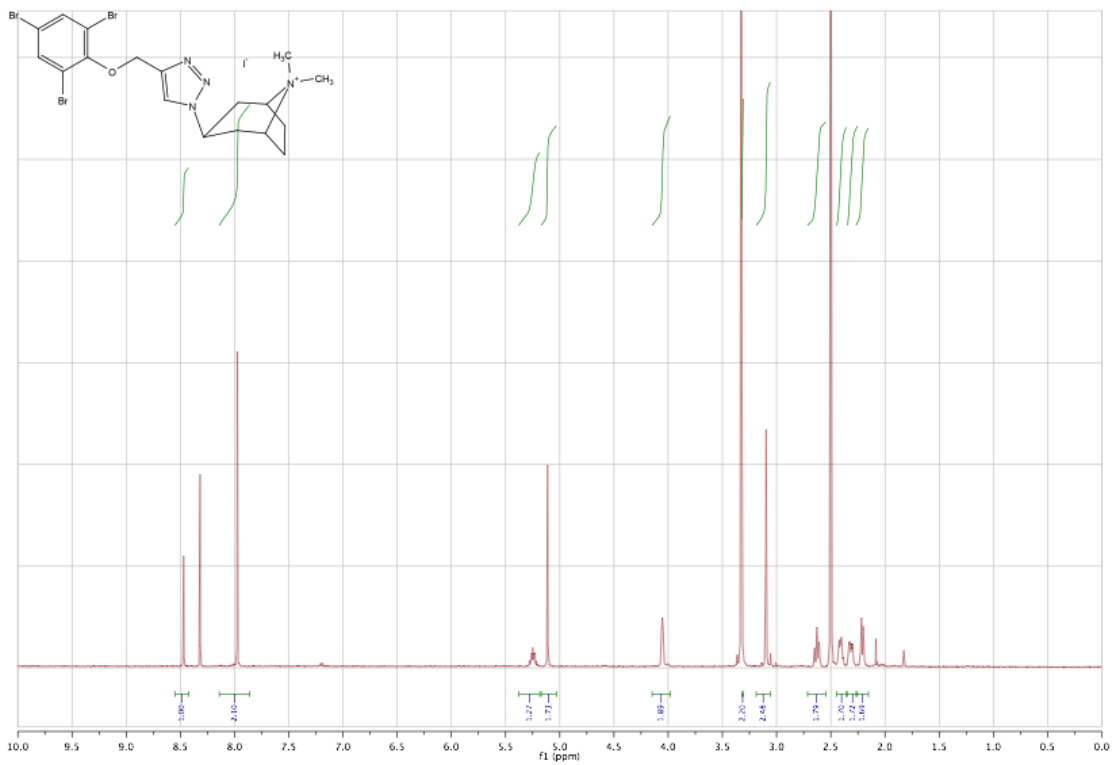
29:





30:





Protein crystallography

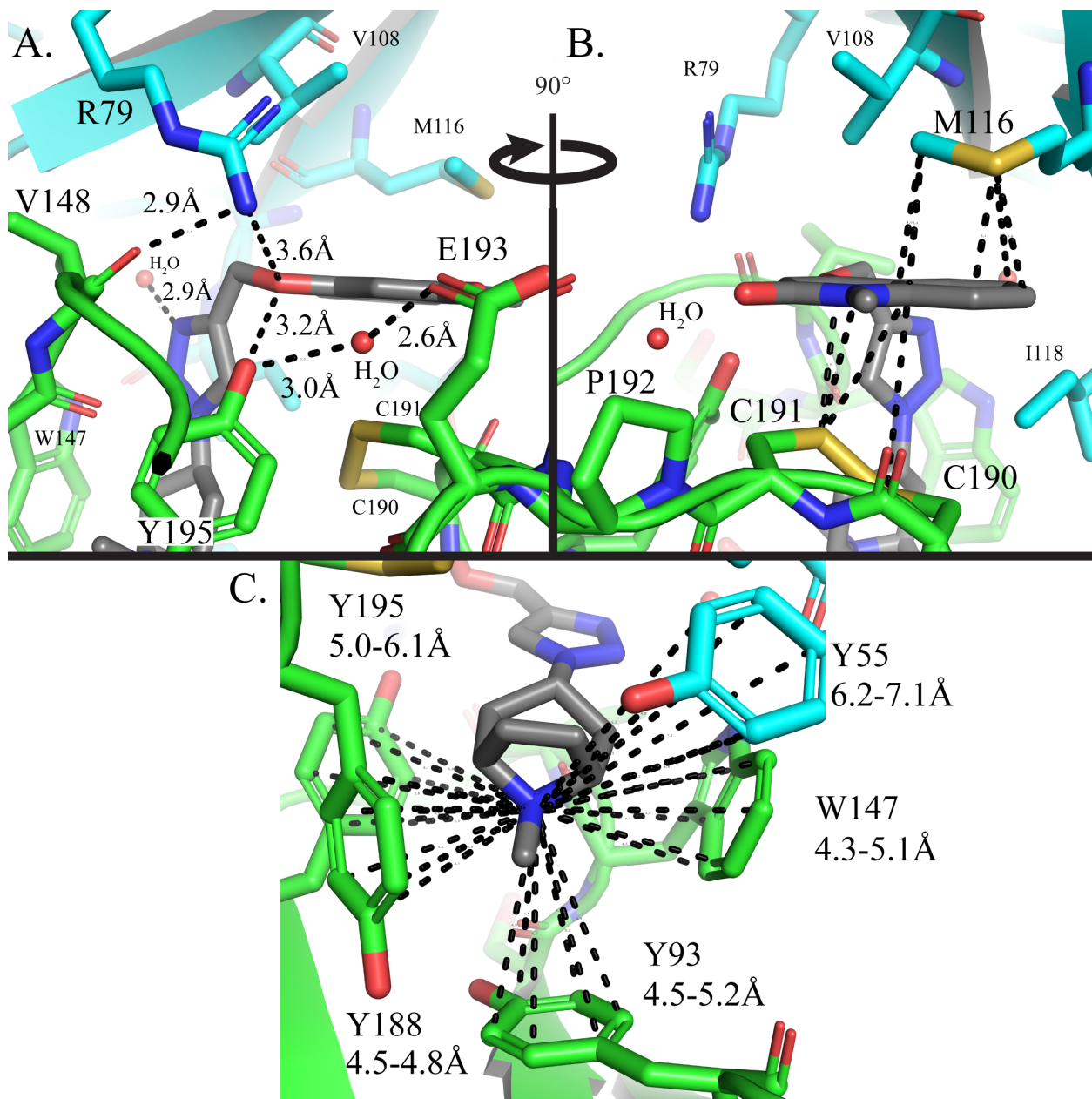
Data set	<i>Ac</i> -AChBP/(18)
Space group	$P2_12_12_1$
Cell constants (Å)	a=88.3 b=115.1, c=131.4 $\alpha=\beta=\gamma=90$
Wavelength (Å)	1.0
Resolution (Å)	43.29 – 2.3
Total/unique reflections	472547/57215
Average redundancy	8.0(6.6)
Completeness (%)	98.5 (94.9)
$\langle I \rangle / \langle \sigma_I \rangle$	28.0 (2.56)
R_{sym}°	0.075 (0.473)
R_{work}° (%)	19.42 (24.68)
R_{free}^{\S} (%)	25.18 (33.95)
Overall B value (Å ²)	36.15
Rmsd bond length (Å)	0.013
Rmsd bond angle (°)	1.185

$R_{\text{sym}} = \sum_h \sum_i |I(h) - I(h)_i| / \sum_h \sum_i I(h)_i$, where $I(h)$ is the mean intensity after rejections.

[§]Numbers in parentheses correspond to the highest resolution shell of data, which were 2.34 to 2.30 for reflections and 2.357 to 2.3 for refinement

^{||} $R_{\text{work}} = \sum_h ||F_{\text{obs}}(h) - F_{\text{calc}}(h)|| / \sum_h |F_{\text{obs}}(h)|$; no I/σ cutoff was used during refinement.

[¶] 5.0% of the observed intensities was excluded from refinement for cross validation purposes.



Supplementary Figure 1: Molecular interactions of complex 18 with *Ac*-AChBP from binding site with best occupancy and electron density.

The primary subunit is colored in green, whereas the complementary subunit is in cyan. Only residues within 4 Å of ligand **18** are shown. A.) Hydrogen bonding network from ether oxygen of **18**. The length of the distances from Arg79 and Tyr195 show a very weak interaction with the ligand, elaborated by the weak density found for the quinolinone in many of the binding pockets (Figure 3A). B.) Select distances shown range from 3.1–5.0 Å, to demonstrate hydrophobic interactions between disulfide of Cys190–191, quinolinone of ligand, and Met116. C.) Aromatic nest of residues interacting with aza-nitrogen in the dimethyl-tropane. Distances are the ranges found from the nitrogen to each carbon on the aromatic ring. Tyr93, Tyr188, and Trp147 are within cation- π distances (4.0–5.0 Å nitrogen to center of aromatic ring), whereas Tyr55 and Tyr195 are more distant.