

Supporting Information

Valkenburg et al. 10.1073/pnas.1302935110

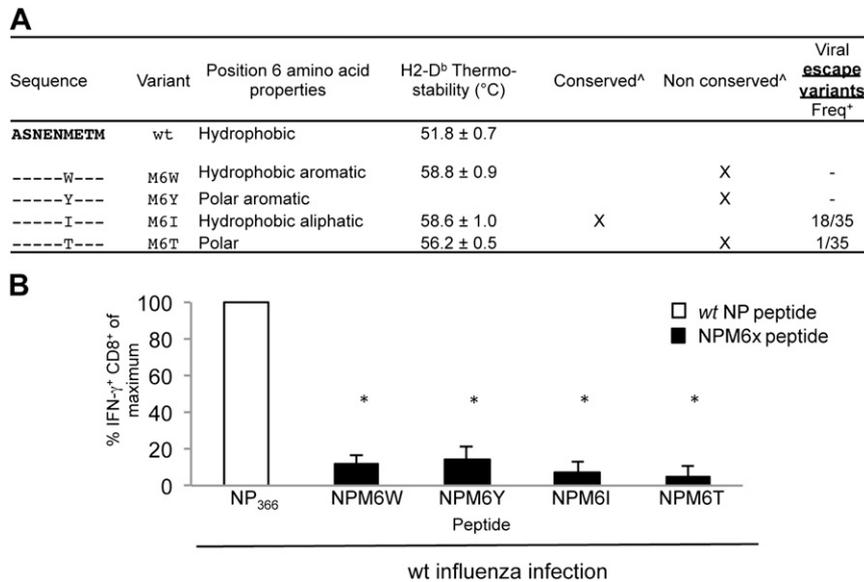
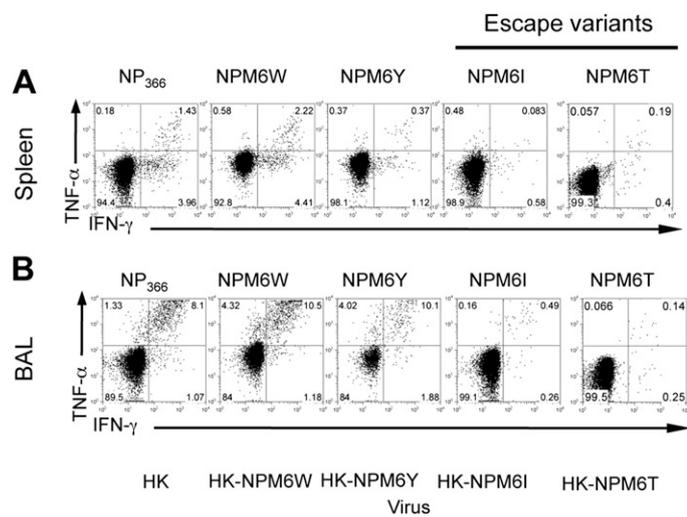


Fig. 51. The wt D^bNP-specific CD8⁺ T cells show little cross-reactivity with the substituted NPM6X peptides. (A) Various amino acid substitutions at PM6 for the NP₃₆₆ peptide caused biochemical changes that in no way diminished the thermostability of NPM6X binding to H-2D^b. Temperatures (°C) represent the point where 50% of the peptide-MHC was unfolded. Amino acid substitutions were either conserved or nonconserved. The NPM6I and NPM6T escape variants were previously isolated from the lungs of infected mice (1). (B) Spleen cytotoxic T lymphocytes (CTLs) from mice primed intraperitoneally with the wt PR8 virus, then challenged intranasally with the wt HK virus 6 wk later, were analyzed for CD8⁺ T-cell specificity on day 8 after secondary infection. Cells were stimulated directly ex vivo for 5 h (in the presence of Brefeldin A) with either the wt NP₃₆₆ or a panel of NPM6X peptides. IFN- γ production was measured by intracellular cytokine staining (ICS), and the background fluorescence was subtracted for the “no peptide” controls. Data represent mean \pm SD ($n = 5$). Experiments were repeated at least twice. * $P \leq 0.01$ relative to wt HK NP₃₆₆-specific response.

1. Wu TD, Brutlag DL (1996) Discovering empirically conserved amino acid substitution groups in databases of protein families. *Proc Int Conf Intell Syst Mol Biol* 4:230–240.
2. French S, Robson B (1983) What is a conservative substitution? *J Mol Evol* 19:171–175.
3. Price GE, Ou R, Jiang H, Huang L, Moskophidis D (2000) Viral escape by selection of cytotoxic T cell-resistant variants in influenza A virus pneumonia. *J Exp Med* 191(11):1853–1867.



C Spleen

Virus	Peptide #	Mean	SD	P values
HK	NP	323908.45	71064.50	
HK-NPM6I	M6I	39238.75	32779.61	>0.0001
HK-NPM6T	M6T	23065.25	15432.12	>0.0001
HK-NPM6W	M6W	472316.95	134165.47	0.06(above wt)
HK-NPM6Y	M6Y	128990.28	70575.50	0.002

D BAL

Virus	Peptide #	Mean	SD	P values
HK	NP	9557.1	1982	
HK-NPM6I	M6I	1671.03	1024.86	>0.0001
HK-NPM6T	M6T	2249.02	782.48	>0.0001
HK-NPM6W	M6W	20150.06	18318.86	NS
HK-NPM6Y	M6Y	6675.18	4401.63	NS

Fig. 52. The NPM6I and NPM6T escape variants generate minimal CD8⁺ T-cell responses. Following intranasal infection with either the WT HK or mutant HK-NPM6X virus, (A and C) splenocytes or (B and D) BAL cells (from the site of infection) were stimulated with the WT NP or NPM6X peptides corresponding to the infecting virus. (A and B) Representative FACS plots are shown. (C and D) Numbers of epitope-specific CD8⁺ T cells in (C) spleen and (D) BAL are presented as mean ± SD. Background “no peptide” values were subtracted. Experiments were repeated at least twice. P values are shown for the comparison between the WT D^bNP₃₆₆ CD8⁺ T-cell responses and NPM6X⁺CD8⁺ populations.

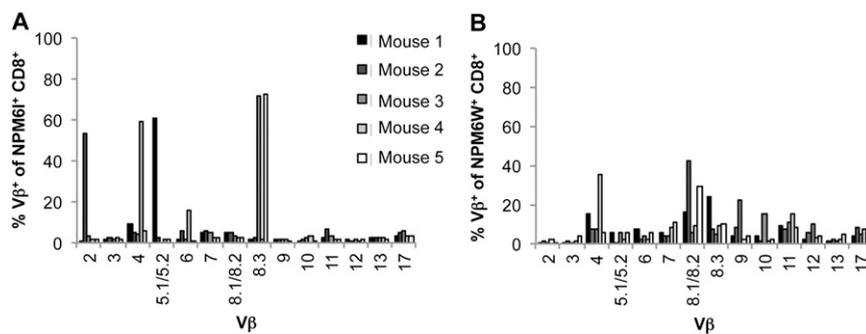


Fig. 53. Limited T-cell receptor (TCR) Vβ use for the low precursor frequency NPM6I⁺ CD8⁺ T cells. Splenocytes were stained with D^bNPM6X-allophycocyanin tetramer and a panel of 14 FITC mAb specific for the TCR Vβ region. Vβ use was assessed for secondary acute responses generated by homologous priming with PR8-NPM6I followed by infection with (A) HK-NPM6I or (B) HK-NPM6W. Data represent individual responses (n = 4–5 mice).

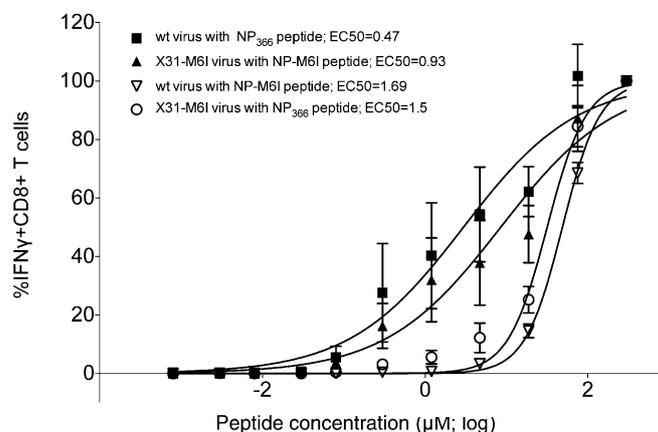


Fig. 54. Optimal stimulation with the homologous peptides. Responsiveness of CD8⁺ T cells to limited peptide concentrations was determined as a measure of functional TCR avidity for the cognate peptide and class I MHC (pMHCI) complex. D^bNP₃₆₆+CD8⁺ and D^bNPM6I CD8⁺ T cells recovered from mice primed and challenged with the wt or NPM6I viruses (PR8 prime, HK boost) were assessed for functional TCR avidity using the IFN- γ ICS assay. The enriched splenocytes analyzed by IFN- γ ICS were stimulated in vitro with fourfold dilutions of the NP₃₆₆, or NPM6I peptide. Data represent mean \pm SD ($n = 4$) and were plotted as sigmoidal curves by transforming peptide concentrations to log values. EC₅₀ values are shown in the figure.

Table S1. Data collection and refinement statistics

Data collection and refinement	Db-NPM6T	Db-NPM6W	Db-NPM6I	Db-H155A-NP	Db-H155AA-NPM6I
Data collection statistics					
Temperature	100K	100K	100K	100K	100K
Space group	P 2 ₁	P 2 ₁ 2 ₁ 2	P2	C2	P2
Cell dimensions (a, b, c), Å	79.72, 84.40, 136.03 $\beta = 90.89^\circ$	86.28, 153.62, 80.50	83.69, 72.27, 86.83 $\beta = 103.28^\circ$	92.33, 111.39, 58.00 $\beta = 122.72^\circ$	83.95, 72.93, 86.35 $\beta = 103.10^\circ$
Resolution, Å	100–2.60 (2.70–2.60)	100–2.50 (2.60–2.50)	50.00–2.00 (2.10–2.00)	100–2.20 (2.30–2.20)	100–2.00 (2.10–2.00)
Total no. of observations	435,398 (42,786)	259,485 (26,613)	284,953 (36,523)	93,158 (11,487)	254,265 (35,169)
No. of unique observations	52,751 (5,093)	35,931 (3,672)	64,509 (8,223)	24,876 (3,089)	68,339 (9,332)
Multiplicity	8.2 (8.4)	7.2 (7.2)	4.4 (4.4)	3.7 (3.7)	3.7 (3.7)
Data completeness, %	94.5 (85.3)	94.0 (88.5)	94.4 (89.1)	98.7 (98.4)	99.3 (99.9)
I/σ_I	15.5 (3.1)	15.9 (3.5)	11.5 (3.2)	12.4 (2.6)	13.5 (2.8)
R _{merge} ,* %	11.7 (68.8)	10.9 (58.9)	10.3 (48.0)	8.2 (58.5)	6.7 (48.3)
Refinement statistics					
Nonhydrogen atoms					
Protein	12,768	6,322	6,453	3,216	6,403
Water	211	181	848	171	830
No. of monomer	4	2	2	1	2
R _{factor} , [†] %	17.8	20.9	19.1	19.4	18.1
R _{free} , [†] %	24.7	24.5	23.1	23.7	23.6
RMS bond lengths, Å	0.010	0.010	0.009	0.010	0.010
RMS bond angles, °	1.17	1.17	1.06	1.12	1.09
Ramachandran plot, %					
Favored/allowed regions	99.2	98.2	99.1	98.8	99.4
Generously allowed regions	0.7	1.1	0.6	1.2	0.6
Disallowed regions	0.1	0.8	0.3	0	0
PDB code	4HUW	4HUV	4HUU	4HUX	4HV8

Values in parentheses are for the highest-resolution shell.

*R_{merge} = $\sum |I_{hkl} - \langle I_{hkl} \rangle| / \sum I_{hkl}$.

[†]R_{factor} = $\sum_{hkl} ||F_o| - |F_c|| / \sum_{hkl} |F_o|$ for all data except ~ 5% which were used for R_{free} calculation.

Table S2. Frequency of TCR β clonotypes in D^bNPM6I+V β 8.3+CD8⁺ cells and D^bNPM6W+ CD8⁺ T cells within multiple V β , after infection with HK-NPM6I or HK-NPM6W viruses

CDR3 β	J β	Amino acid length	Frequency, %				
			M1	M2	M3	M4	M5
D^bNPM6I							
V β 8.3							
SDWRGEQ	2.6	7	95				
SDARGFLAEQ	2.1	10	5				
SDWGTGGKAEQ	2.1	11		100			
V β 1							
SQDLGGIYEQ	2.7	10			100		
V β 4							
SQEAGGYEQ	2.7	9				93	
SQDRTGGRDEQ	2.7	11				3	
SQDEFANTEV	1.1	10				3	
Total sequences			40	35	32	30	
D^bNPM6W							
V β 4							
SQYWGGANEQ	2.1	10				15	
SQDRGLEQ	2.6	8				27	
SQDRRNSYNSPL	1.6	12				29	
SQEGTGYSGNTL	1.3	12				27	
SQYWGGYAEQ	2.1	9				2	
V β 8.1							
GAGLGGRAETL	2.3	11		6			
GDARTGVAETL	2.3	11					6
GDASLYAEQ	2.1	9		3			
GDHRGGIYAEQ	2.1	11					3
GDRDDNYAEQ	2.1	10		6			
GDTGRYAEQ	2.1	9		6			
GEPGLGPYEQ	2.6	10					6
GGGTFVYEQ	2.6	9		10			
GGLGGNYAEQ	2.1	10					54
GVGSNYAEQ	2.1	9		26			
GVRQSSGNTL	1.3	10					20
V β 8.2							
SDSWGGAEQ	2.6	9					
SEHRGRTEV	1.1	9		26			
SPGRGGAE	2.1	9		6			
SPGRGGAEQ	2.1	9		10			
V β 8.3							
SDGTGAQDTQ	2.5	10	89				
SDGTGAKTPSTLGQ		15	3				
SDGTGAQDTSTLGQ		15	3				
SDRDRVYAEQ	2.1	10	3				
SENGVEQ	2.6	7	3				
V β 9							
SRIRGGRDTQ	2.5	10			18		
SRDRAEQ	2.6	7			32		
SHRGLNQAP	1.5	9			47		
RHRGANTEV	1.1	9			3		
Total sequences			38	31	34	41	35

