108

Human-DNM1L-000429 Rat-DNM1L-035303 Mouse-DNM1L-Q8K1M6 Chick-DNM1L-E1BV15 Darne-DNM1L-E9QF63

<u>109</u>

Human-DNM1L-000429

Rat-DNM1L-035303 Mouse-DNM1L-Q8K1M6 Chick-DNM1L-E1BV15 Darne-DNM1L-E9QF63

<u>110</u>

Human-DNM1L-000429 Rat-DNM1L-035303 Mouse-DNM1L-Q8K1M6 Chick-DNM1L-E1BV15 Darne-DNM1L-E9QF63

111

Human-FIS1-Q9Y3D6 Rat-FIS1-P84817 Mouse-FIS1-Q9CQ92 Darne-FIS1-E90GI1

112

Human-FIS1-Q9Y3D6 Rat-FIS1-P84817 Mouse-FIS1-Q9CQ92 Darne-FIS1-E9QGI1 113 ELLPKGS

Human-FIS1-Q9Y3D6	IRKGIVLLE <mark>ELLPKGS</mark> KEEQRDYVFYLAVG	80
Rat-FIS1-P84817	IRRGIVLLE <mark>ELLPKGS</mark> KEEQRDYVFYLAVG	80
Mouse-FIS1-Q9CQ92	IRRGIVLLE <mark>ELLPKGS</mark> KEEQRDYVFYLAVG	80
Darne-FIS1-E9QGI1	IVKGIQLLE <mark>ELVHTSK</mark> KDDQRDFLFYLAVA	80
	* :** ****: *::***	

Fig S1. Sequence conservation of Drp1/Fis1 peptides. Sequence alignment of all the peptides between Homo sapiens Drp1 (O00429), Rat (O35303), Mouse (Q8K1M6), Chicken (E1BV15) Zebrafish (E9QF63) and Yeast (P54861); and Fis1 Homo sapiens (Q9Y3D6), Rat (P84817), Mouse (Q9CQ92), Zebrafish (E9QGI1) and Yeast (P40515). Amino acids are represented by the one-letter code; star (*) indicate identical amino acids; two points (:) indicate high similarity between amino acids.

STOELLRFPK

ORIIOHCSNYSTOELLRFPKLHDAIVEVVTC 470 QRIIQHCSNYSTQELLRFPKLHDAIVEVVTC 483 QRIIQHCSNYSTQELLRFPKLHDAIVEVVTC 476 INTVRQCT---KKLSQYPHLREEMERIVTT 462 VNTVRQCT----KKLAQYPMLREEMERIVTQ 462 :::*: ::* ::* *:: : :**

KLSAREORD

LLDVPVPVAR <mark>KLSAREORD</mark> CEVIERLI	651
LLDVPVPVAR <mark>KLSAREORD</mark> CEVIERLI	670
LLDVPVPVAR <mark>KLSAREORD</mark> CEVIERLI	657
-ENGSDSFMHSMDPQLE <mark>R</mark> QVETIRNLV	666
DESSSDGFMHSMDPQLE <mark>R</mark> QVETIRNLV	667
: : :*: * * *:	

DLLPRGT

SVLESLVGR <mark>DLLPRGT</mark> GIVTRRPLILQLVH	69
SVLESLVGR <mark>DLLPRGT</mark> GVVTRRPLILQLVH	69
SVLESLVGR <mark>DLLPRGT</mark> GVVTRRPLILQLVH	69
SVLENFVGR <mark>DFLPRGS</mark> GIVTRRPLVLQLVN	76
SVLENFVGK <mark>DFLPRGS</mark> GIVTRRPLVLQLIN	76
**** :**:	

SVEDLLKFEK

MEAVLNELV <mark>SVEDLLKFEK</mark> KFQSEKAAGSV	30
MEAVLNELV <mark>SVEDLKNFER</mark> KFQSEQAAGSV	30
MEAVLNELV <mark>SVEDLKNFER</mark> KFQSEQAAGSV	30
MEAVVSDIV APEDLKKFEK KYNAELVKGPV	30
****: ::* *** :**:** * *	

KGSKEEQRD

IRKGIVLLEELLP <mark>KGSKEEQRD</mark> YVFYLAVG	80
IRRGIVLLEELLP <mark>KGSKEEQRD</mark> YVFYLAVG	80
IRRGIVLLEELLP KGSKEEQRD YVFYLAVG	80
IVKGIQLLEELVH TSKKDDQRD FLFYLAVA	80
* :** ****: *:****:	

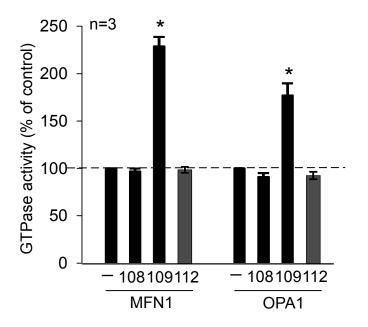


Fig S2. Drp1 P109 increases MFN1 and OPA1 GTPase activity. MFN1 (25ng) and OPA1 (25 ng) recombinant proteins were incubated with peptide 108, 109 or 112. GTPase activity of the proteins was determined. The data are expressed as mean \pm SE of three independent experiments. *, p<0.05 vs. MFN1 or OPA1 protein alone.

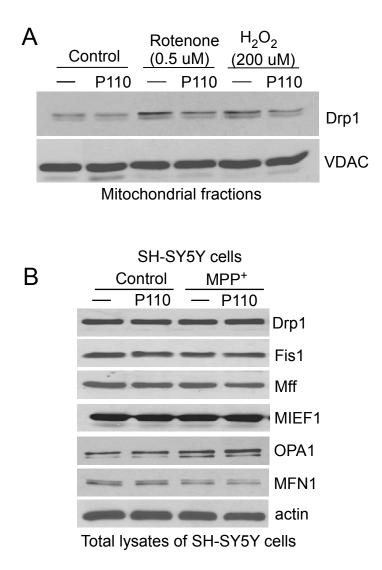


Fig S3. P110 blocked Drp1 translocation to the mitochondria in response to H2O2 or Rotenone. (A) SH-SY5Y cells were treated with P110 (1 μ M) followed by exposure to Rotenone (0.5 μ M, 1 hour) and H2O2 (200 μ M, 2 hours). Mitochondria were isolated and the mitochondrial level of Drp1 was determined by western blot. VDAC was used as a loading control. (B) Total lysates harvested from above cells and mitochondrial fusion/fission related proteins were determined by western blot.

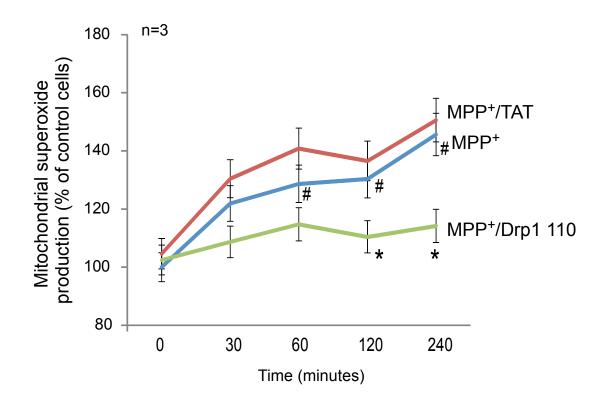
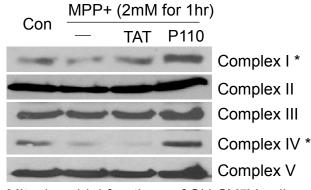


Fig S4. P110 reduced MPP+-induced mitochondrial ROS over time. SH-SY5Y cells were treated with P110 (1 μ M) or control peptide TAT (1 μ M) followed by incubation of MPP+ (2 mM) at the indicated time points. The detection of MitoSOXTM was performed using black 96-well plates in a fluorescence microplate reader at 510 nm excitation and 580 nm emission. All these measurements were normalized to the cell number counted using Hoechst staining. Data are expressed as mean ± S.E. of three independent experiments. *p<0.05 *vs*. MPP+-treated cells; #p<0.05 *vs*. control cells.

N-terminus	TAT	Spacer	Cargo	C-terminus
Peptide 108	H Gly Lys Arg Gln Arg Arg	g Gly Thr Glu Gly Ser Gln Leu	Leu Phe Lys Arg Pro NH ₂	Molecular Weight: 2873.33
Peptide 109	H Gly Lys Arg Gln Arg	g Gly Leu Ala Gly Lys Ser Arg	Glu Arg NH ₂	Molecular Weight: 2757.13
Peptide 110	H Glý Lýs Arg Gln Arg	g Gly Leu Pro Gly Asp Leu Arg	GlyNH ₂ Ser	Molecular Weight: 2411.78
Peptide 111	H Gly Lys Arg Gln Arg	g Gly Val Asp Gly Ser Glu Leu	Leu Phe Lys Glu NH ₂	Molecular Weight: 2862.30
Peptide 112	H ^{Tyr} Gly ^{Arg} Lys ^{Arg} Gln ^{Arg} Arg	g Gly Gly Lys Gly Lys Ser Glu	Glu Arg NH ₂ Gln Asp	Molecular Weight: 2731.05
Peptide 113	H Gly Lys Arg Gln Arg	Gly Leu Pro Gly Glu Leu Lys	Gly NH ₂ Ser	Molecular Weight: 2397.79
Peptide 274	H Gly Lys Arg Gln Arg	Gly Ala Leu Pro	GlyNH ₂ Ser	Molecular Weight: 2367.77
Peptide 275	H Glý Lýs Arg Gln Arg	Gly Ala Pro Gly Asp Leu Arg	GlyNH ₂ Ser	Molecular Weight: 2369.70
Peptide 276	H Gly Lys Arg Gln Arg Arg	Gly Asp Ala Arg	Gly NH ₂ Ser	Molecular Weight: 2369.70
Peptide 277	H ^{Tyr} Gly Arg Lys Arg Gln Arg Arg	g Gly Leu Ala Gly Asp Leu Arg	Gly_NH ₂ Ser	Molecular Weight: 2385.74
Peptide 278	H Gly Lys Arg Gln Arg	Gly Asp Leu Pro Ala	Gly NH ₂ Ser	Molecular Weight: 2326.67
Peptide 279	H Gly Lys Arg Gln Arg Arg			Molecular Weight: 2425.80
Peptide 280	H Gly Lys Arg Gln Arg Arg	Gly Asp Leu Pro Arg	Gly NH ₂	Molecular Weight: 2395.78

Fig S5: Chemical structure of all the peptides that were used in this study. All the peptides are present from the N-terminus to the C-terminus. TAT is on the N-terminus, spacer of two Gly amino acids after that and the cargo sequence which derived from the proteins on the C-terminus. Peptides 108-113 derived from Drp1 and Fis1 proteins. Peptides 274-280 are Ala scan of peptide 110. To peptide P110, two peptides were synthesized (one is DLLPRGT, the other is DLLPRGS). We used DLLPRGS in the current study.



Mitochondrial fractions of SH-SY5Y cells

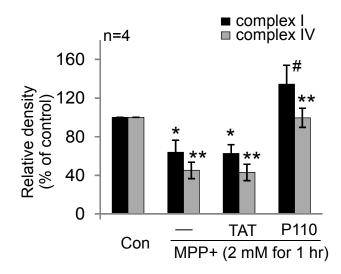


Fig S6. P110 reduced mitochondrial complex I and IV disassembly induced by MPP+. SHSY5Y cells were treated with P110 (1 μ M) or TAT (1 μ M) followed by the addition of MPP+ (2 mM, 1 hour). Mitochondria were isolated and mitochondrial complex I-V assembly was determined by a cocktail of antibodies of complex I-V (Mitosciences/Abcam, MitoProfile Total OXPHOS WB Antibody Cocktail). Upper: representative western blot; Lower histogram: the data are expressed as mean ± S.E. of three independent experiments. The oxidative phosphorylation system in the mitochondria is responsible for generating ATP and consists of five major membrane protein complexes, the mitochondrial complexes I–V. MPP+ is a specific mitochondrial complex I inhibitor. Here, we investigated whether P110 has effects on MPP+-induced defects in mitochondrial complexes. In cultured SH-SY5Y cells, MPP+ treatment disassembled complex I and IV, as evidenced by the reduction of NDUFB8 (component of complex I) and MTCOI (component of complex IV). By contrast, treatment of P110 under the same conditions abolished the reduction of these two proteins, suggesting that P110 treatment recovered the MPP+-induced oxidative phosphorylation defect and mitochondrial integrity.