

## Supplemental Material to:

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## Linkage of autophagy to fungal development, lipid storage and virulence in *Metarhizium robertsii*

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**Figure S1. MrAtg8 is not localized on the lipid droplets (LDs).** The conidia of RFP-MrAtg8 (wild-type strain transformed with an *RFP-MrATG8* fused gene) were stained with the LD-specific fluorescent dye, Bodipy. The stained LDs and RFP-labeled MrAtg8 showed different patterns. Bar, 5 μm.



**Figure S2.** *MrATG8* gene disruption and verification. (A) Construction of the vector pBarATG8 for homologous gene disruption. (B) Transformants were verified by PCR with primers *ATG8*F and *ATG8*R. Pla, plasmid pBarATG8 DNA used as a template; WT, wild type; Lanes 1, 2 and 3 amplified from three independent null mutants; Ect, ectopically integrated transformant with two bands; Comp, complemented mutant acquired by transforming *Mratg8*\Delta with the *MrATG8* gene. (C) RT-PCR verification of *MrATG8* transcripts. Lane 1, wild type (WT); Lane 2, *Mratg8*\Delta; Lane 3, *Mratg8*\Delta complemented transformant with the *MrATG8* gene. *TUB2*, tubulin beta-2 gene used as a reference.



Figure S3. Differences of conidial formation. In contrast to the wild-type and complemented mutant (Comp),  $Mratg8\Delta$  failed to produce conidia either on the nutrient-rich potato dextrose agar (PDA) or the nutrient-poor minimal medium (MM). Bar, 15 µm.



**Figure S4. Appressorium production by the M8-3 mutant on a hydrophobic surface.** When compared to the wild type (Left, a differentiation rate at 88.4%), the mutant could similarly form appressoria (Right, 82.22%) on a hydrophobic surface. CO, conidium; AP, appressorium. Bar, 5 µm.



Figure S5. Phenotyping of  $Mratg1\Delta$ ,  $Mratg4\Delta$  and  $Mratg15\Delta$  complemented with corresponding genes grown on PDA medium. The cultures were grown on the medium for 10 days.





Primer	Sequence (5'- 3')	Purpose of
		use
ATG8UF	CGGGATCCGTGACTACAGAGTAGCAGGCTGTC	For deletion and
ATG8UR	CGGGATCCGATGTCGCTCTTCTCTACCTT	verification of
ATG8LF	GGACTAGTCAGCATCTATGAGGAGCACAAG	MrATG8
ATG8LR	GGACTAGTGCAGTCTGGTAGAGTCCGGTAT	
ATG8F	TGAAAAGCGTAAGGCTGAGG	
ATG8R	CCGAGTAGGTGATGTAGAGGAATC	
TUBF	GATCTTGAACCTGGCACCAT	For RT-PCR
TUBR	CCATGAAGAAGTGCAGACGA	internal control
A8XbU	GCTCTAGACCGAGCATTCAGTCATCATGCGAAG	Mratg8
A8XbD	GCTCTAGATTACGCGGTGCCAAAAGTGTTTTCG	C-terminal
A8XbD1	GCTCTAGATTACGCGGTGGCAAAAGTGTTTTCG	amino acid
A8XbD2	GCTCTAGATTACGCGGTAAAAGTGTTTTCG	mutations
A8XbD3	GCTCTAGATTAGCCAAAAGTGTTTTCGCCC	
A8XbD4	GCTCTAGA TTAGGCAAAAGTGTTTTCGCCC	
A8XbD5	GCTCTAGA TTAAAAAGTGTTTTCGCCCGAG	
RfpXbF	GCTCTAGAGCAGGAATTCATGGCCTCCTCCGAG	For RFP-Mratg8
RfA81	CGTCCTTGAACTTGCTTCGCATGGCGCCGGTGGAGTGGCGGC	fusion
RfA82	GCCGCCACTCCACCGGCGCCATGCGAAGCAAGTTCAAGGACG	
A8Rf1	GACGTCCTCGGAGGAGGCCATCGCGGTGCCAAAAGTGTTTTCG	For Mratg8-RFP
A8Rf2	CGAAAACACTTTTGGCACCGCGATGGCCTCCTCCGAGGACGTC	fusion
RfpXbR	GCTCTAGATTAGGCGCCGGTGGAGTGGCG	
ATG1UF	AACTGCAGACTCATTTCACAGACGCATCC	For deletion,
ATG1UR	AACTGCAGTCAGTACTGGTAACGGTAAGAGG	verification and
ATG1LF	GGACTAGTGAGATCCCTGGATTAGTGGAAGAC	complementatio
ATG1LR	GGACTAGTCTGTCTAGCTCACTGCTAGCTTCT	n of MrATG1
<i>ATG1</i> F	GCGAACTAGGAGACCTGTCG	
ATG1R	GGAGTTCAACGTGGTTTCGT	
ATG1CF	GGACTAGTCCCTTATCACGCTTCCTC	
ATG1CR	GGACTAGTGTTCCCTATCACAAGTCCAA	
ATG4UF	CGGGATCCTGGAATGGACCATGTAGTGAG	For deletion,
ATG4UR	CGGGATCCGACAGAGACAAAGCTGATGCTG	verification and
ATG4LF	GGACTAGTCGCTACTGCTCGTTGTATCCAGTA	complementatio
ATG4LR	GGACTAGTCCTAAGCTAAGCAGCCAGATTC	n of MrATG4
ATG4F	CAGCATCAGCTTTGTCTCTGTC	
ATG4R	ATGCCCAGTCTCGTTCCTACTA	
ATG4CF	GGACTAGTTTCCTGCTGCCCTGTTGG	
ATG4CR	GGACTAGTCCTGAAGGCTGCGATGTT	
ATG15UF	CGGAATTCCGCTCGCTCTTTGAATGT	For deletion,
ATG15UR	CGGGATCCTTGCAGGTGCCAATGTAG	verification and
ATG15LF	GCTCTAGATACAGGAGTCACGGCTACG	complementatio
ATG15LR	CGAGCTCAACCAACATCACAGAGGGA	n ot <i>MrATG15</i>
ATG15F	TTCTGATCCCAAAGCTCCT	
ATG15R	ATCCGAGGTTGTAGTGATA	_
ATG15CF	GCTCTAGAATGAGGGTGTCGTAGAAGAGG	_
ATG15CR	GCTCTAGAAACCAACATCACAGAGGGAGT	
<i>MPL1</i> F	GAACCCTCCCTTCTTCACC	For RT-PCR
MPI IR		analysis of
	CETTOCATTOCATAGACI	MPL1 gene

 Table S1. List of oligonucleotide primers used in the study.

Function*	Gene symbols	S. cerevisiae	M. oryzae	M. robertsii
	ATG1	NP_011335.1	MGG_06393	MAA_03501
	ATG13	NP_015511.1	MGG_00454	MAA_02276
Atg1 kinase and its	ATG17	NP_013527.1	MGG_07667	MAA_07123
regulators	ATG29	NP_015159.1	MGG_02790	MAA_08445
	ATG31	NP_010305.1	None	None
	ATG6	NP_015205.1	MGG_03694	MAA_00746
Ptains 3-kinase complex	ATG14	NP_009686.1	None	None
	ATG5	NP_015176.1	MGG_09262	MAA_02911
	ATG7	NP_012041.1	MGG_07297	MAA_03074
Atg12 conjugation system	ATG10	NP_013058.1	None	None
	ATG12	NP_009776.1	MGG_00598	MAA_02068
	ATG16	NP_013882.1	MGG_05255	MAA_08906
	ATG3	NP_014404.1	MGG_02959	MAA_03709
A 4 - 9	ATG4	NP_014176.2	MGG_03580	MAA_04475
Atg8 conjugation system	ATG7	NP_012041.1	MGG_07297	MAA_03074
	ATG8	NP_009475.1	MGG_01062	MAA_02674
	ATG2	NP_014157.1	MGG_05998	MAA_03175
Atg2-Atg18 complex	ATG18	NP_444297.1	MGG_03139	MAA_05300
	ATG9	NP_010132.1	MGG_09559	MAA_04484
	ATG11	NP_015374.1	MGG_04486	MAA_04312
	ATG19	NP_014559.1	None	None
Cytoplasm to vacuole	ATG20	NP_010170.1	MGG_12832	MAA_05815
targeting (Cvt) pathway	ATG21	NP_015225.1	MGG_03088	MAA_010306
	ATG23	NP_013535.1	None	None
	ATG24	NP_012498.1	MGG_03638	MAA_05264
	ATG27	NP_012357.2	MGG_02386	MAA_01720
	ATG11	NP_015374.1	MGG_04486	MAA_04312
	ATG25	None	None	None
Pexophagy	ATG26	NP_013290.1	MGG_03459	MAA_06719
	ATG28	None	None	None
	ATG30	None	None	None
M:4	ATG32	NP_012120.1	None	None
Mitophagy	ATG33	NP_013460.1	None	None
Autophagic degradation	ATG15	NP_009994.2	MGG_12828	MAA_00506
and recycle	ATG22	NP_009892.1	MGG_09904	MAA_08468
Others	ATG34	NP_014558.1	None	None

**Table S2.** Comparison of autophagy-related genes encoded in the genomes of *S. cerevisiae*, *M. oryzae* and *M. robertsii*.

\*, functional grouping is based on the information acquired from the autophagy database

(<u>http://tp-apg.genes.nig.ac.jp/autophagy/index.html</u>). The genes of *M. robertsii* highlighted in bold were deleted in this study for functional studies.