Supplemental material

JCB

Lechtreck et al., http://www.jcb.org/cgi/content/full/jcb.201207139/DC1

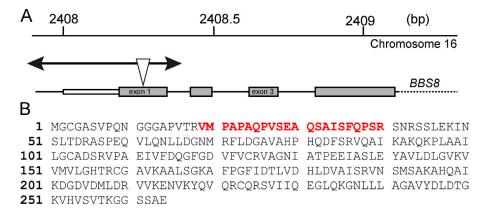


Figure S1. Molecular characterization of an insertional mutation in *C. reinhardtii* BBS8 and sequence of CAH6. (A) Molecular characterization of an insertional mutation in *C. reinhardtii* BBS8. Wild-type *C. reinhardtii* was transformed with a promoter-less fragment of the *AphVII* gene conferring resistance to hygromycin (Berthold et al., 2002). Transformants were screened for lack of phototaxis; the nonphototactic strain LP20.2 was selected and analyzed by restriction enzyme site-directed amplification PCR (González-Ballester et al., 2005; Brown et al., 2012). The strain has an insertion in exon 1 codon 27 between basepairs 82 and 83 (indicated by the triangle; counting from the ATG); no deletion of genomic DNA was detected. This insertion causes an in-frame STOP after 38 residues/codons. The arrows represent the flanking DNA amplified by PCR. The third base of the inserted promoter-less *AphVII* was altered from T to A. We designated this allele *bbs8-1* and here refer to the strain carrying it by the same name. (B) Sequence of CAH6. The peptide identified by mass spectrometry is shown in red.

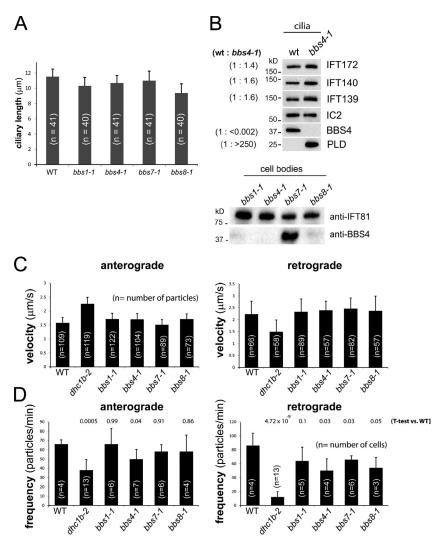


Figure S2. Analysis of ciliary length and IFT in *dhc1b-2* and *bbs* mutants. (A) Ciliary length of wild type and the four *bbs* mutant strains. (B) Western blot analysis of cilia isolated from wild type and *bbs4-1*. For quantification of relative protein amounts, values for band intensities were adjusted for IC2 loading and the ratios (left, parentheses) were then calculated. Velocity (C) and frequency (D) of anterograde and retrograde IFT for wild type and *dhc1b-2* and *bbs* mutants. Standard deviations are indicated in A, C, and D; t test probability data are shown in D.

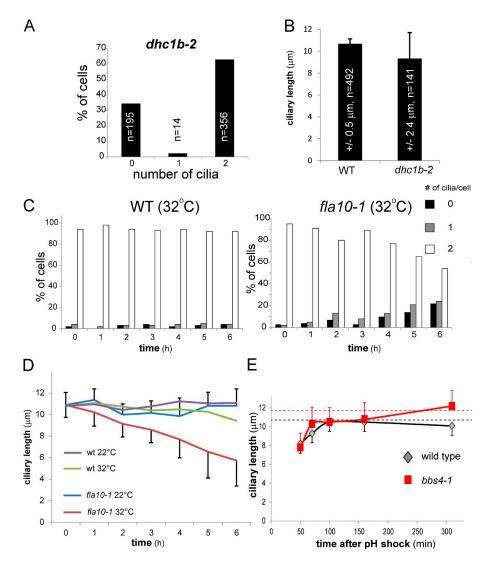


Figure S3. Cilia length and number as a function of time and temperature in *dhc1b-2*, *fla10-1*, and *bbs4-1* mutants. (A and B) Ciliary number and ciliary length, respectively, for the *dhc1b-2* mutant at the permissive temperature (21°C). (C) Ciliary number for wild-type and *fla10-1* cells before (0 h) and at various time points (1–6 h) after shifting the cells to 32°C. The data shown are from a single representative experiment out of two repeats. The number of cilia of wild-type or *fla10-1* cells maintained at 22°C remained at the 0-h level throughout the time course (not depicted). (D) Ciliary length of wild-type and *fla10-1* cells before (0 h) and after various times (1–6 h) of incubation at 22 or 32°C. (E) Ciliary length at various time points after deciliation by pH shock. Measurements are based on isolated cilia viewed by dark-field microscopy. Red, *bbs4-1*; gray, wild type; dashed lines, length of wild-type cilia (black, 10.8 ± 0.91 µm, n = 29) and *bbs4-1* cilia (red, 11.8 ± 1.18 µm, n = 26) before deciliation. At least 24 cilia from different cells were measured for each time point. Standard deviations are indicated in B, D, and E.

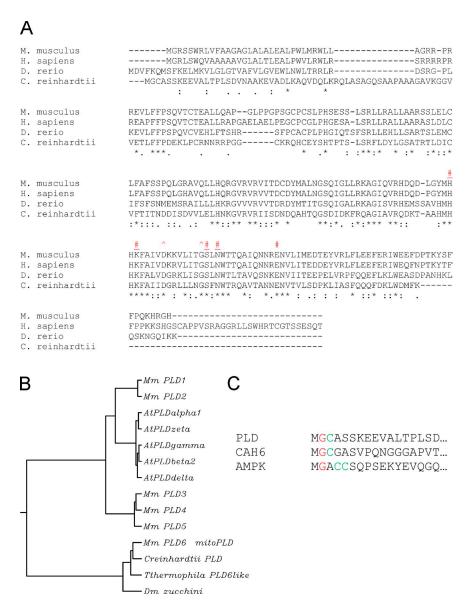


Figure S4. Alignment and phylogenetic analysis of PLDs. (A) Alignment of *C. reinhardtii* and vertebrate PLDs. #, active site; ^, signature sites, <u>#</u>, both. (B) Rooted phylogenetic tree of murine PLDs (Mm_PLD1 [NCBI Protein accession no. NP_001157528.1], Mm_PLD2 [GenBank accession no. AAH68317], Mm_PLD3 [GenBank accession no. AAH76586], Mm_PLD4 [GenBank accession no. AAH58565], Mm_PLD5 [GenBank accession no. AAH00429], and Mm_PLD6 [GenBank accession no. CAl24298.1]), *Arabidopsis thaliana* PLDs (AtPLDalpha1 [GenBank accession no. AEE75720], AtPLDdelta [GenBank accession no. AEE86571], AtPLDgamma [GenBank accession no. AEE83056], AtPLDbeta2 [GenBank accession no. AEE81845], and AtPLDzeta [GenBank accession no. AED93432]), *Drosophila melanogaster* zucchini (GenBank accession no. AAM49862.1), *Tetrahymena thermophila* TTHERM_02188720 (NCBI Protein accession no. XP_001028688), and *C. reinhardtii* PLD (NCBI Protein accession no. XP_001693080). The tree is based on a CLUSTALW multiple sequence alignment using default settings. (C) N-Terminal sequences of PLD, CAH6, and AMPK. Residues predicted to be myristoylated are shown in red and residues predicted to be palmitoylated in green. Online tools (Myristylator, Myr Predictor, and CSS-Palm 2.0) were used for prediction of fatty acid modification.