

Table 1: Overview of strains used in this study

Organism	Mutation(s)/Plasmid	Tags	Selection marker(s)	Reference
<i>L. pn.</i> JR32	-	-	-	(47)
<i>L. pn.</i> Corby	-	-	-	(48)
<i>L. pn.</i> JR32	<i>dotA</i> knock-out	-	KmR	(23)
<i>L. pn.</i> JR32	<i>dotB</i> knock-out	-	KmR	(47)
<i>L. pn.</i> Corby	<i>IspDE</i> knock-out	-	KmR	(93)
<i>L. pn.</i> JR32	<i>plcA</i> knock-out	-	HygR	this study
<i>L. pn.</i> JR32	<i>plcB</i> knock-out	-	KmR	this study
<i>L. pn.</i> JR32	<i>plcC</i> knock-out	-	GmR	this study
<i>L. pn.</i> JR32	<i>plcAB</i> knock-out	-	KmR, HygR	this study
<i>L. pn.</i> JR32	<i>plcAC</i> knock-out	-	HygR, GmR	this study
<i>L. pn.</i> JR32	<i>plcBC</i> knock-out	-	KmR, GmR	this study
<i>L. pn.</i> JR32	<i>plcABC</i> knock-out	-	KmR, HygR, GmR	this study
<i>E. coli</i> DH5α	pMMB2002	-	CmR	(21)
<i>E. coli</i> DH5α	pMS12 = pMMB2002 <i>plcC</i> _{JR32}	-	CmR	this study
<i>E. coli</i> BL21	pMS14 = pET160/GW/D-TOPO <i>plcC</i> _{JR32} H179N	N-term. 6×His-Lumio-TEV	AmpR	this study
<i>E. coli</i> BL21	pMS17 = pET160/GW/D-TOPO <i>plcC</i> _{JR32} S336A	N-term. 6×His-Lumio-TEV	AmpR	this study
<i>E. coli</i> DH5α	pMS18 = pMMB2002 <i>plcA</i> _{JR32}	-	CmR	this study
<i>E. coli</i> DH5α	pMS19 = pMMB2002 <i>plcB</i> _{JR32}	-	CmR	this study
<i>E. coli</i> BL21	pMS2 = pET160/GW/D-TOPO <i>plcC</i> _{JR32}	N-term. 6×His-Lumio-TEV	AmpR	this study
<i>E. coli</i> DH5α	pMS29 = pGEMT-EZ 3.84kb <i>plcB</i> _{JR32}	-	AmpR	this study
<i>E. coli</i> BL21	pMS36 = pGP172 <i>plcA</i> _{JR32} (-SP*)	N-term. Strep	AmpR	this study
<i>E. coli</i> BL21	pMS37 = pGP172 <i>plcB</i> _{JR32} (-SP*)	N-term. Strep	AmpR	this study
<i>E. coli</i> BL21	pMS38 = pGP172 <i>plcC</i> _{JR32}	N-term. Strep	AmpR	this study
<i>E. coli</i> DH5α	pPA111= pGEMT-EZ 3.35kb <i>plcC</i> _{JR32}	-	AmpR	this study
<i>E. coli</i> DH5α	pPA114= pGEMT-EZ <i>plcC</i> _{JR32::gmR}	-	AmpR, GmR	this study
<i>E. coli</i> DH5α	pPA127 = pLAW344 <i>plcC::GmR</i>	-	CmR, GmR	this study
<i>E. coli</i> DH5α	pPA151 = pGEMT-EZ <i>plcB</i> _{JR32::kmR}	-	AmpR, KmR	this study
<i>E. coli</i> DH5α	pPA155 = pLAW344 <i>plcB::KmR</i>	-	CmR, KmR	this study
<i>E. coli</i> DH5α	pPA158 = pEGMT-EZ 2.65kb <i>plcA</i> _{JR32}	-	AmpR	this study
<i>E. coli</i> DH5α	pPA159 = pEGMT-EZ <i>plcA</i> _{JR32::hygR}	-	AmpR, HygR	this study
<i>E. coli</i> DH5α	pPA160 = pLAW344 <i>plcA::hygR</i>	-	CmR, HygR	this study
<i>E. coli</i> BL21	pPA163 = pET160/GW/D-TOPO <i>plcA</i> _{JR32} (-SP*)	N-term. 6×His-Lumio-TEV	AmpR	this study
<i>E. coli</i> BL21	pPA164 = pET160/GW/D-TOPO <i>plcB</i> _{JR32} (-SP*)	N-term. 6×His-Lumio-TEV	AmpR	this study
<i>E. coli</i> BL21	pPA175 = pGP172 <i>plcC</i> _{JR32} D251V	N-term. Strep	AmpR	this study
<i>E. coli</i> BL21	pPA176 = pGP172 <i>plcC</i> _{JR32} F253A	N-term. Strep	AmpR	this study
<i>E. coli</i> BL21	pPA177 = pGP172 <i>plcC</i> _{JR32} F244A	N-term. Strep	AmpR	this study
<i>E. coli</i> BL21	pPA178 = pGP172 <i>plcC</i> _{JR32} Y156A	N-term. Strep	AmpR	this study
<i>E. coli</i> BL21	pPA179 = pGP172 <i>plcC</i> _{JR32} F167A	N-term. Strep	AmpR	this study
<i>E. coli</i> BL21	pPA180 = pGP172 <i>plcC</i> _{JR32} H166N	N-term. Strep	AmpR	this study
<i>E. coli</i> BL21	pPA181 = pGP172 <i>plcC</i> _{JR32} D314V	N-term. Strep	AmpR	this study
<i>E. coli</i> BL21	pPA182 = pGP172 <i>plcC</i> _{JR32} R265Q	N-term. Strep	AmpR	this study
<i>E. coli</i> BL21	pPA184 = pGP172 <i>plcC</i> _{JR32} H257N	N-term. Strep	AmpR	this study
<i>E. coli</i> BL21	pPA185 = pGP172 <i>plcC</i> _{JR32} H284N	N-term. Strep	AmpR	this study
<i>E. coli</i> BL21	pPA186 = pGP172 <i>plcC</i> _{JR32} H247N	N-term. Strep	AmpR	this study
<i>E. coli</i> BL21	pPA187 = pGP172 <i>plcC</i> _{JR32} R385Q	N-term. Strep	AmpR	this study
<i>E. coli</i> BL21	pPA189 = pGP172 <i>plcC</i> _{JR32} E286A	N-term. Strep	AmpR	this study
<i>E. coli</i> BL21	pPA190 = pGP172 <i>plcC</i> _{JR32} D63V	N-term. Strep	AmpR	this study
<i>E. coli</i> BL21	pPA191 = pGP172 <i>plcC</i> _{JR32} R326Q	N-term. Strep	AmpR	this study
<i>E. coli</i> BL21	pPA192 = pGP172 <i>plcC</i> _{JR32} H409Q	N-term. Strep	AmpR	this study

*SP=signal peptide

Table 2: Overview of primers used in this study

Plasmid	Gene	Primer name	Primer sequence
pMS2	<i>plcC</i> _{JR32}	lpg0012To_fw lpg0012_rv	5'-CACCATGAACACAACCGAACATAC-3' 5'-AAGACAGCTCAAATGGGTTACT-3'
pMS12	<i>plcC</i> _{JR32}	cegC1_Xba1_fw cegC1_Kpn1_rv3	5'-GATTAACTGCTCTAGATCCATTGCTTCAT-3' 5'-GCCACTAGTTTATCAGGTACCAATCAGACTCCC-3'
pMS18	<i>plcA</i> _{JR32}	plcA_Xba1_fw plcA_Kpn1_rv	5'-TTAACTGAGTCTAGACCCGCTTGTG-3' 5'-ACCATAAGGTACCCATGACTGGATCG-3'
pMS19	<i>plcB</i> _{JR32}	plcB_Xba1_fw plcB_Kpn1_rv	5'-AACTCAACCCTAGACGGTAATGC-3' 5'-CCGGGATTGCGGTACCGATAATAG-3'
pMS36	<i>plcA</i> _{JR32} (-SP)	NStrA_SacII_f2 StrepPlcA_KpnI_r	5'-AATCCGCGGTAATGTTTCCTCTATAAC-3' 5'-GCGGTACCCCTACCAAAAGGATTATC-3'
pMS37	<i>plcB</i> _{JR32} (-SP)	NStrB_SacII_f2 StrepPlcB_KpnI_r	5'-AATCCGCGGTAATGTTGAAGAATATCAC-3' 5'-GCGGTACCTTAAATTATTAACTG-3'
pPA163	<i>plcA</i> _{JR32} (-SP)	pa_topo2_fw plcA_Kpn1_rv	5'-CACCTTTCCCTCTATAACTTATGGTG-3' 5'-ACCATAAGGTACCCATGACTGGATCG-3'
pPA164	<i>plcB</i> _{JR32} (-SP)	pb_topo2_fw plcB_Kpn1_rv	5'-CACCAATGAAGCATCTTCTATCGAGAC-3' 5'-CCGGGATTGCGGTACCGATAATAG-3'
pPA158	2.65kb <i>plcA</i> fragment	plcA_Lokus_fw plcA_Lokus_rv	5'-GCAGGTTTGCCACCATTG-3' 5'-CTAGCCGCCGTCACTATATC-3'
pMS29	3.84kb <i>plcB</i> fragment	plcB_Lokus_fw plcB_Lokus_rv	5'-TCGACATGCGCTGTATGAG-3' 5'-CGCATAACGCAAGGAAATTATC-3'
pPA111	3.35kb <i>plcC</i> fragment	cegC1_EcoRV_fw cegC1_EcoRV_rv	5'-GATATCGTCAGGCTGGCTAAAGTC-3' 5'-GATATCCGCTGGCTGGATCTTC-3'
pMS38	<i>plcC</i> _{JR32}	NStrC_SacII_f2 StrepPlcC_KpnI_r	5'-AATACCGCGGTAATGAACACAACGGAAC-3' 5'-GCGGTACCTCAGACTCCAAATTCAACC-3'
pPA159	inverse PCR for removal of <i>plcA</i> ORF and insertion of XbaI sites	plcAMutXba_fw plcAMutXba_rv	5'-TCTCTAGACTTATGATATGCCGGCT-3' 5'-TCTCTGGACTTACGACAGGCATTGGA-3'
pPA151	inverse PCR for removal of <i>plcB</i> ORF and insertion of XbaI sites	plcBMut_Xba_f plcBMut_Xba_r	5'-TCTCTAGAAAATACTGGAGCGGGCT-3' 5'-TCTCTAGATTGGCCCTTATTGGG-3'
pPA114	insertion of SmaI sites by site directed mutagenesis	cegC1Mut_Sma_r cegC1Mut_SmaI_f cegC1_5'Sma_fw cegC1_5'Sma_rv	5'-CCATTGCCCGGGCTGTATGTTCCGTTGTCA-3' 5'-TGAACACAACGGAACATACAGACCCGGGCAATGG-3' 5'-GAGGAAGAAGAGGCCGGGTGAATTGGGA-3' 5'-TCCCAAATTACCCGGGCCTTCTCCTC-3'
pMS14	<i>plcC</i> _{JR32} H179N	cegC1_H179N_fw/rv	5'-ATATTGGGTAACGCTATCGCATTGAG-3'
pMS17	<i>plcC</i> _{JR32} S336A	cegC1_S336A_fw/rv	5'-CCTAAACGGTATGACCGCA <u>G</u> CACTAA-3'
pPA175	<i>plcC</i> _{JR32} D251V	cegC1_D251V_fw/rv	5'-GCTGTTTACTTTCACTACTATTCTG <u>TG</u> CATTTT-3'
pPA176	<i>plcC</i> _{JR32} F253A	plcC_F253A_f/r	5'-CACTACTATTCTGATCAT <u>G</u> CTGCTACAGGA-3'
pPA177	<i>plcC</i> _{JR32} F244A	plcC_F244A_f/r	5'-CCAAGCTTATTCTATAGAGCTG <u>G</u> CTACTTT-3'
pPA178	<i>plcC</i> _{JR32} Y156A	plcC_Y156A_f/r	5'-TATTTCGCGTAAAGAC <u>G</u> CTGGGAAATG-3'
pPA179	<i>plcC</i> _{JR32} F167A	plcC_F167A_f/r	5'-TCGGAACCAAACCAT <u>G</u> CTACGCCATGGTC-3'
pPA180	<i>plcC</i> _{JR32} H166N	cegC1_H166_qc_fw/rv	5'-GAACCAAAC <u>A</u> TTTTACGCCATG-3'
pPA181	<i>plcC</i> _{JR32} D314V	cegC1_D314V_f/r	5'-GCACGTGGGG <u>T</u> GGGAAATTG-3'
pPA182	<i>plcC</i> _{JR32} R265Q	cegC1_R265Q_fw/rv	5'-GGCGATTAC <u>A</u> AGTCGTATAAAAGAGCG-3'
pPA184	<i>plcC</i> _{JR32} H257N	cegC1_H257N_fw/rv	5'-TCATTTGCTACAGGA <u>A</u> ATATGCCATGAT-3'
pPA185	<i>plcC</i> _{JR32} H284N	cegC1_H284N_fw/rv	5'-GCAATATCCTGGCTAATAATCT <u>A</u> ATGATG-3'
pPA186	<i>plcC</i> _{JR32} H247N	cegC1_H247N_fw/rv	5'-GCTTATTCTATAGAGCTGTTACTTT <u>A</u> ACTACT-3'
pPA187	<i>plcC</i> _{JR32} R385Q	cegC1_R385_qc_fw/rv	5'-AGTATATTAC <u>A</u> GAACAATCTCAG-3'
pPA189	<i>plcC</i> _{JR32} E286A	plcC_E286A_f/r	5'-CCTGGCTAAATCTCATGAT <u>G</u> CAGTAAA-3'
pPA190	<i>plcC</i> _{JR32} D63V	cegC1_D63V_fw/rv	5'-GCGGGTGT <u>C</u> ACTTTACACAGG-3'
pPA191	<i>plcC</i> _{JR32} R326Q	cegC1_R326_qc_fw/rv	5'-ATCAATTAA <u>CC</u> AAATTAGCTGCC-3'
pPA192	<i>plcC</i> _{JR32} H409N	plcC_H409N_f/r	5'-CTCAGAGAAAACCCTGAAAAAA <u>A</u> ATGGGTAT-3'