

Figure. S1
ES129.1 cells were subjected to RNAi-mediated knockdown of Mre11 and NBS1. Western-blot analysis with antisera against Mre11, NBS1 and beta-tubulin, showing a reduction in the expression levels of Mre11 and NBS1, respectively.

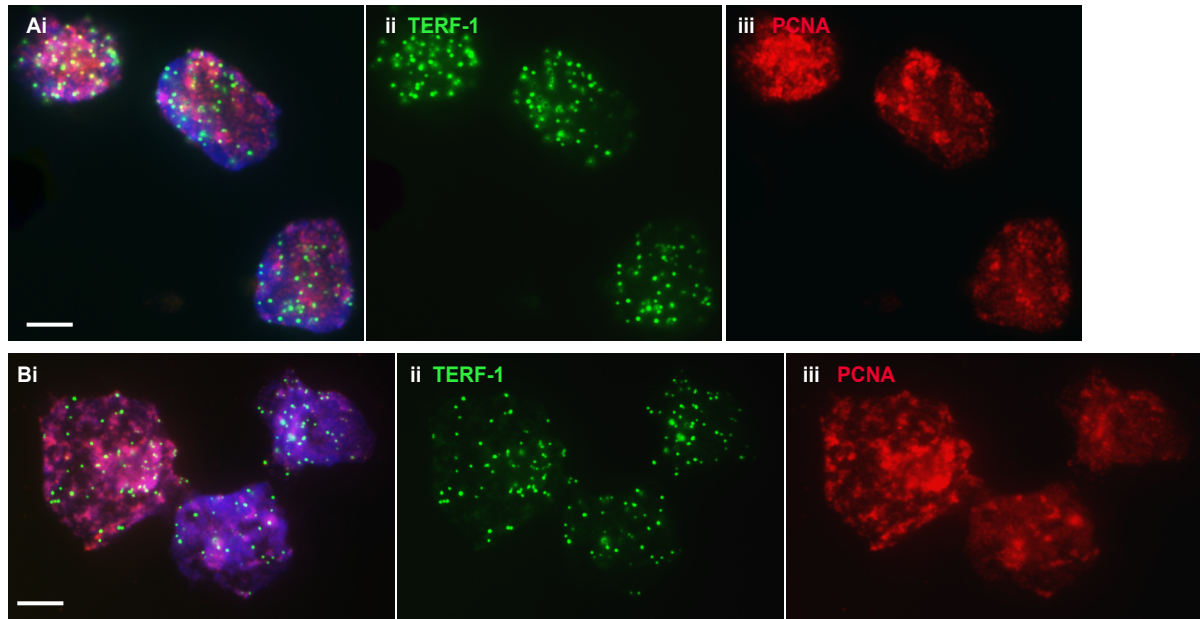


Figure S2. PCNA staining in mouse ES cells depleted of PML bodies.

RNAi-mediated knockdown of PML was performed on ES129.1 cells with specific siRNA oligonucleotides for 48hr, followed by immunofluorescence analysis. Compared to cells transfected with scramble siRNA (A), depletion of PML bodies (B) did not affect the progression of cell cycle or progression of cells through S phase as indicated by the presence of cells with positive PCNA staining (refer to Supplementary material, Table S2c quantitative analysis).

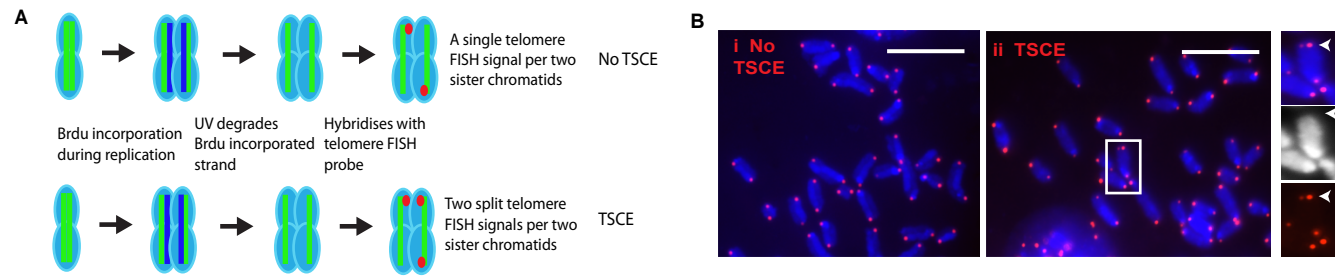


Figure S3. (A) Principle of CO-FISH. Newly replicating DNA strands incorporate BrdU during replication. Treatments with Hoechst and UV specifically degrade these strands and leave each sister-chromatid single-stranded. Consequently a strand-specific telomeric probe will hybridise with only one of the two sister-chromatids at the telomere and show a single signal. When TSCE occurs after replication, splitting of the signal between both chromatids occurs and a double-signals will be detected. The intensities of the two signals may not be symmetric, depending on the site of exchange within the telomere. (B) CO-FISH analysis was performed with telomere probe on ES129.1 cells depleted of PML expression. Examples of metaphase cells showing either absence (left cell) or presence (right cell) of TSCE are shown. Data collected for CO-FISH analysis was presented in Supplementary Table S3b.

Supplementary data

Table S1a. RNAi-mediated knockdown of NBS1 and Mre11. RNAi-mediated knockdown was performed on ES129.1 cells with specific siRNA oligonucleotides against NBS1 and Mre11, and a set of scramble control siRNA oligonucleotides for 48hr. Cells were harvested for reverse-transcription and real-time PCR analysis. Expression levels of NBS1 and Mre11 were measured as Ct values and normalized against Ct values of actin to give Δ Ct values (=NBS1 or Mre11 Ct value- Actin Ct value). Average Δ Ct values were calculated from 3 sets of Δ Ct values. $\Delta\Delta$ Ct values were calculated as differences in Δ Ct values (=NBS1 or Mre11 RNAi Δ Ct - control RNAi Δ Ct) of samples subjected to knockdown with specific siRNA oligonucleotides and those with control scramble siRNA oligonucleotides. Expression ratios and percentages of knockdown were calculated as $2^{-\Delta\Delta$ Ct and $[1 - \text{Expression Ratio}] \times 100\%$, respectively. The data showed a reduction of 74.9% to 78.8% in NBS1 level and 77.46% to 84.69% in Mre11 level, respectively.

NBS1 knockdown N=1

Samples	Primer set	Average Ct	Δ Ct	$\Delta\Delta$ Ct	Fold of expression	% knockdown
Scramble control	NBS1 primer set 1	25.26377133	7.256732667			
NBS1 RNAi set 1		27.67566567	8.804118	1.547385333	0.342129561	65.78704392
NBS1 RNAi set 2		27.01070667	9.622971667	2.366239	0.193950581	80.60494194
Scramble control	NBS1 primer set 2	25.42374733	7.416708667			
NBS1 RNAi set 1		28.00570467	9.134157	1.717448333	0.304086077	69.59139231
NBS1 RNAi set 2		26.91123933	9.523504333	2.106795667	0.232162093	76.78379074
Scramble control	NBS1 primer set 3	25.41877167	7.411733			
NBS1 RNAi set 1		28.07520367	9.203656	1.791923	0.288786859	71.1213141
NBS1 RNAi set 2		27.12061967	9.732884667	2.321151667	0.200107665	79.98923352
Scramble control	Actin primer	18.00703867				
NBS1 RNAi set 1		18.87154767				
NBS1 RNAi set 2		17.387735				

NBS1 knockdown N=2

Samples	Primer set	Average Ct	Δ Ct	$\Delta\Delta$ Ct	Fold of expression	% knockdown
Scramble control	NBS1 primer set 1	26.837439	7.163034333			
NBS1 RNAi set 1		29.098551	9.670301667	2.507267333	0.175888452	82.41115485
NBS1 RNAi set 2		28.84305767	9.583289667	2.420255333	0.186823089	81.31769115
Scramble control	NBS1 primer set 2	27.27044767	7.596043			
NBS1 RNAi set 1		29.25820733	9.829958	2.233915	0.212581064	78.74189356
NBS1 RNAi set 2		28.4302395	9.1704715	1.5744285	0.335776112	66.42238884
Scramble control	NBS1 primer set 3	27.23280633	7.558401667			
NBS1 RNAi set 1		29.67916867	10.25091933	2.692517667	0.154693269	84.53067305
NBS1 RNAi set 2		28.71013367	9.450365667	1.891964	0.26944001	73.05599904
Scramble control	Actin primer	19.67440467				
NBS1 RNAi set 1		19.42824933				
NBS1 RNAi set 2		19.259768				

NBS1 knockdown N=3

Samples	Primer set	Average Ct	Δ Ct	$\Delta\Delta$ Ct	Fold of expression	% knockdown
Scramble control	NBS1 primer set 1	26.09737533	6.532164			
NBS1 RNAi set 1		28.05081367	9.129084	2.59692	0.165290991	83.47090089
NBS1 RNAi set 2		27.54149033	8.097568333	1.565404333	0.337882999	66.2117001
Scramble control	NBS1 primer set 2	26.304097	6.738885667			
NBS1 RNAi set 1		28.430359	9.508629333	2.769743667	0.146630419	85.33695808
NBS1 RNAi set 2		28.24687867	8.802956667	2.064071	0.23914027	76.08597297
Scramble control	NBS1 primer set 3	26.224467	6.659255667			
NBS1 RNAi set 1		28.69802267	9.776293	3.117037333	0.115259907	88.47400929
NBS1 RNAi set 2		28.043052	8.59913	1.939874333	0.260639142	73.93608577
Scramble control	Actin primer	19.56521133				
NBS1 RNAi set 1		18.92172967				
NBS1 RNAi set 2		19.443922				

NBS1 RNAi sets Average % knockdown of NBS 1 from 3 experiments

NBS1 RNAi set 1 **78.82948223**

NBS1 RNAi set 2 **74.93420045**

Mre11 knockdown N=1

Primer set	Samples	Average Ct	Δ Ct	$\Delta\Delta$ Ct	Fold of expression	% knockdown
	Scramble control	24.65532633	6.132106333			
Mre11 primer set 1	Mre11 RNAi set 1	27.971309	9.047531333	2.915425	0.132546915	86.74530848
	Mre11 RNAi set 2	26.9752635	8.341522167	2.209415833	0.216221841	78.37781588
	Scramble control	24.92914567	6.405925667			
Mre11 primer set 2	Mre11 RNAi set 1	28.87453067	9.950753	3.544827333	0.08568418	91.43158203
	Mre11 RNAi set 2	27.30910167	8.675360333	2.269434667	0.207411146	79.25888535
Actin Primer	Scramble control	18.52322				
	Mre11 RNAi set 1	18.92377767				
	Mre11 RNAi set 2	18.63374133				

Mre11 knockdown N=2

Primer set	Samples	Average Ct	Δ Ct	$\Delta\Delta$ Ct	Fold of expression	% knockdown
	Scramble control	24.78264233	6.064400333			
Mre11 primer set 1	Mre11 RNAi set 1	26.679978	8.898928333	2.834528	0.140191618	85.98083819
	Mre11 RNAi set 2	25.84722033	7.890400667	1.826000333	0.282045469	71.79545308
	Scramble control	24.82210433	6.103862333			
Mre11 primer set 2	Mre11 RNAi set 1	26.53305967	8.75201	2.648147667	0.159524767	84.04752328
	Mre11 RNAi set 2	25.675893	7.719073333	1.615211	0.326417203	67.35827966
Actin Primer	Scramble control	18.718242				
	Mre11 RNAi set 1	17.78104967				
	Mre11 RNAi set 2	17.95681967				

Mre11 knockdown N=3

Primer set	Samples	Average Ct	Δ Ct	$\Delta\Delta$ Ct	Fold of expression	% knockdown
	Scramble control	23.50240933	5.053729667			
Mre11 primer set 1	Mre11 RNAi set 1	26.01550333	6.979964333	1.926234667	0.263114987	73.68850127
	Mre11 RNAi set 2	25.770029	7.124001	2.070271333	0.238114712	76.18852879
	Scramble control	23.869041	5.420361333			
Mre11 primer set 2	Mre11 RNAi set 1	26.44168967	7.406150667	1.985789333	0.252474689	74.75253112
	Mre11 RNAi set 2	26.362578	7.71655	2.296188667	0.203600263	79.63997365
Actin Primer	Scramble control	18.44867967				
	Mre11 RNAi set 1	19.035539				
	Mre11 RNAi set 2	18.646028				

Mre11 knockdown N=4

Primer set	Samples	Average Ct	Δ Ct	$\Delta\Delta$ Ct	Fold of expression	% knockdown
	Scramble control	25.50081233	6.463228667			
Mre11 primer set 1	Mre11 RNAi set 1	28.00497467	9.645509667	3.182281	0.110163561	88.98364394
	Mre11 RNAi set 2	26.8625135	9.084040167	2.6208115	0.162576259	83.74237411
	Scramble control	25.45127733	6.413693667			
Mre11 primer set 2	Mre11 RNAi set 1	28.394948	10.035483	3.621789333	0.081233052	91.87669476
	Mre11 RNAi set 2	26.773127	8.994653667	2.58096	0.167129696	83.28703043
Actin Primer	Scramble control	19.03758367				
	Mre11 RNAi set 1	18.359465				
	Mre11 RNAi set 2	17.77847333				

Mre11 RNAi sets Average % knockdown of MRE11 from 4 experiments

Mre11 set 1	84.68832788
Mre11 set 2	77.45604262

Table S1b. FACS analysis of NBS1 and Mre11 knockdown cells. RNAi-mediated knockdown was performed on ES129.1 cells with NBS1 and Mre11 specific siRNA oligonucleotides and a set of scramble control siRNA oligonucleotides for 48hr. Cells were harvested for FACS analysis and percentage of cell population in various stages of cell cycle was shown. RNAi-mediated depletion of NBS1 and Mre11 for 48hr, respectively, did not have a significant impact on the progression of cell cycle.

Cell cycle stages		G1	S	G2/M
Average values (% of cell)	Scramble control RNAi	17.17	68.67	14.34
	NBS1 RNAi set#1	17.99	69.50	12.51
	NBS1 RNAi set#2	16.71	70.87	12.44

Cell cycle stages		G1	S	G2/M
Average values (% of cell)	Scramble control RNAi	18.35	70.18	11.64
	Mre11 RNAi set#1	17.31	73.14	9.74
	Mre11 RNAi set#2	17.94	68.95	13.44

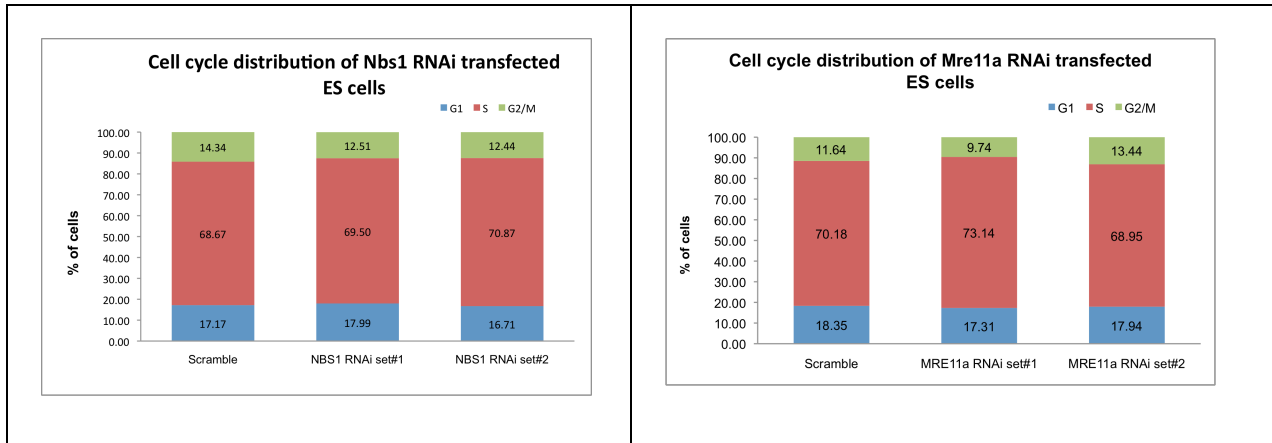


Table S1c. Immunofluorescence analysis of NBS1 and Mre11 knockdown cells. (c) RNAi-mediated knockdown was performed on ES129.1 cells with NBS1 and Mre11 specific siRNA oligonucleotides and a set of scramble control siRNA oligonucleotides for 48hr. Cells were harvested for immunofluorescence analysis and quantification of telomeric signals at PML bodies as indicated by co-staining of telomere FISH and anti-PML antibody staining signals. Percentage of cell population showing positive foci of telomeres and PML bodies remained unchanged between the control cells and those depleted of either NBS1 or Mre11. These data indicate that loss of NBS1 and Mre11 did not have a significant effect on the assembly of telomere-associated PML bodies in mouse ES cells.

		Control RNAi	NBS1 RNAi set#1	NBS1 RNAi set#2
Number of co-localised foci (% of cell population)	<4	38.31	48.21	46.31
	5 to 9	39.22	36.48	29.28
	10 to 14	16.25	10.32	13.97
	>14	6.22	6.84	11.85
% of cell with ≥ 5 positive foci		61.69	51.795	53.69

		Control RNAi	Mre11 RNAi set#1	Mre11 RNAi set#2
Number of co-localised foci (% of cell population)	<4	35.29	32.67	35.29
	5 to 9	40.14	45.81	43.58
	10 to 14	18.35	14.30	13.21
	>14	6.22	7.23	7.92
% of cell with ≥ 5 positive foci		64.71	67.33	64.71

Table S2a. RNAi-mediated knockdown of PML. RNAi-mediated knockdown of PML was performed on ES129.1 cells using 4 sets of specific siRNA oligonucleotides for 48hr. Cells were harvested for reverse-transcription and real-time PCR assays. Expression level of PML was measured as Ct values and normalized against Ct values of actin to give Δ Ct values (=PML Ct value- Actin Ct value). Average Δ Ct values were calculated. $\Delta\Delta$ Ct values were calculated as differences in Δ Ct values (=PML RNAi Δ Ct - control RNAi Δ Ct) of samples subjected to knockdown with PML-specific siRNA oligonucleotides and those with control scramble siRNA oligonucleotides. Expression ratios and percentages of knockdown were calculated as $2^{-\Delta\Delta$ Ct and $[1 - \text{Expression Ratio}] \times 100\%$, respectively. The data showed a reduction of 57.80% to 76.65% in PML level. siRNA oligonucleotides set#3 and #4 were used in further experiments.

PML Primer set 1	Average Ct	ΔCt	$\Delta\Delta$Ct	Fold of expression	% of Knockdown
Scramble control	21.6266455	6.2360745			
PML RNAi set#1	22.989065	7.607481	1.3714065	0.386514247	61.3
PML RNAi set#2	23.33388	7.4574755	1.221401	0.428866044	57.1
PML RNAi set#3	24.012379	8.599505	2.3634305	0.194328513	80.6
PML RNAi set#4	24.0316495	8.440722	2.2046475	0.216937671	78.3
PML Primer set 2	Average Ct	ΔCt	$\Delta\Delta$Ct	Fold of expression	% Knockdown
Scramble control	21.097066	5.706495			
PML RNAi set#1	22.217684	6.8361	1.129605	0.457040843	54.3
PML RNAi set#2	22.439604	6.5631995	0.8567045	0.552212519	44.8
PML RNAi set#3	23.2923735	7.8794995	2.1730045	0.221748385	77.8
PML RNAi set#4	23.299015	7.7080875	2.0015925	0.249724193	75.0
Actin primer	Average Ct				
Scramble control	15.390571				
PML RNAi set#1	15.381584				
PML RNAi set#2	15.8764045				
PML RNAi set#3	15.412874				
PML RNAi set#4	15.5909275				

PML RNAi	Average % of knockdown
PML RNAi set#1	57.80
PML RNAi set#2	50.95
PML RNAi set#3	79.20
PML RNAi set#4	76.65

Table S2b. FACS analysis of PML knockdown cells. RNAi-mediated knockdown of PML was performed on ES129.1 cells using PML specific siRNA oligonucleotides and a set of scramble control oligonucleotides. Cells were harvested for FACS analysis. The depletion of PML expression (after 48hr) has no immediate or significant impact on the cell cycle progression of the cell population.

Cell cycle stages	G1 (%)	S (%)	G2/M (%)
Untransfected	16.84	16.84	16.84
Control RNAi	17.96	64.30	17.76
PML RNAi set#1	16.73	64.26	19.02
PML RNAi set#2	16.15	63.23	20.63

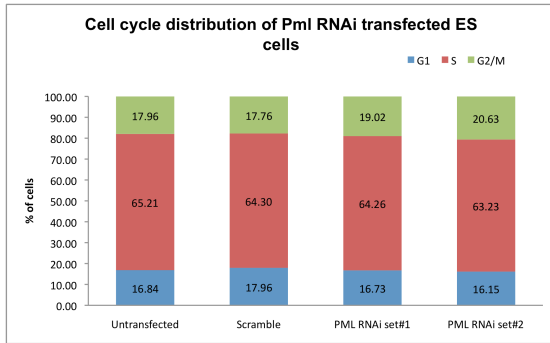


Table S2c. PCNA staining of PML knockdown cells. RNAi-mediated knockdown of PML was performed on ES129.1 cells using PML specific siRNA oligonucleotides and a set of scramble control oligonucleotides. Cells were harvested for immunofluorescence analysis using an antibody against PCNA (see Supplementary Figure S1), an antigen that is expressed in the nuclei of cells during the DNA synthesis or S phase of the cell cycle. RNAi-depletion of PML did not have any impact of percentage of cells showing PCNA staining, suggesting that the progression of cells through S phase was not affected.

Time period after G1/S release (hours) Expt N=1		Telomere-associated PML bodies					
		0	2	4	6	8	10
Scramble control	Cells with positive PCNA foci (35 cells)	4	12	26	32	12	9
	Percentage of cell population with positive PCNA foci (%)	11.00	34.29	74.26	91.43	34.26	25.71
PML RNAi	Number of cells with positive PCNA foci (35 cells)	5	10	23	29	11	10
	Percentage of cell population with positive PCNA foci (%)	14.29	28.57	65.71	82.86	31.42	28.57

Time period after G1/S release (hours) Expt N=2		Telomere-associated PML bodies					
		0	2	4	6	8	10
Scramble control	Cells with positive PCNA foci (35 cells)	6	9	23	33	10	7
	Percentage of cell population with positive PCNA foci (%)	17.14	25.71	65.71	94.28	28.57	20.00
PML RNAi	Number of cells with positive PCNA foci (35 cells)	7	13	29	32	18	12
	Percentage of cell population with positive PCNA foci (%)	20.00	37.14	82.86	91.43	51.43	34.29

Table S3a. Staining with anti-53BP1 antibody in PML knockdown cells. In PML RNAi-mediated knockdown ES129.1 cells, TIFs were determined by staining with an antibody against a DNA damage marker- 53BP1. In ES129.1 cells transfected with control scramble siRNA oligonucleotides, only 10% to 14% of cells showed ≥ 5 TIFs per cell. In contrast, ES129.1 cells subjected to transfection with PML-specific siRNA oligonucleotides showed an increase by 3 to 5 fold (from 10%-14% to 38%-46%) in the population of cells showing ≥ 5 TIFs per cell, indicating that telomere functional integrity was impaired following the loss of PML bodies. Data from the scoring and quantification was presented in Figure 4.

NO of TIFs	Expt n=1		Expt n=2		Expt n=3	
	Control	PML RNAi	Control	PML RNAi	Control	PML RNAi
<5	44	31	43	27	45	27
5 to 9	6	13	5	16	3	15
10 to 14	0	5	2	6	2	5
>14	0	1	0	1	0	3
Total	50	50	50	50	50	50
	Percentage of cell population (%)					
	Expt n=1		Expt n=2		Expt n=3	
	Control	PML RNAi	Control	PML RNAi	Control	PML RNAi
<5	88%	62%	86%	54%	90%	54%
5 to 9	12%	26%	10%	32%	6%	30%
10 to 14	0%	10%	4%	12%	4%	10%
>14	0%	2%	0%	2%	0%	6%
Total	100%	100%	100%	100%	100%	100%

Table S3b CO-FISH analysis in PML knockdown cells. CO-FISH assay (Supplementary Figure S2) was used to determine the rate of TSCE following the loss of PML bodies in ES129.1 cells. Number of metaphase chromosomes showing double-signals out of the total of 600 was determined, in 3 sets of experiments. No significant increase in the rate of TSCE was detected in ES129.1 cells following RNAi-mediated knockdown of PML and subsequently loss of function of PML bodies.

Expt n=1		Expt n=2		Expt n=3	
Control	PML RNAi	Control	PML RNAi	Control	PML RNAi
2/150	4/150	4/150	6/150	5/150	3/150
3/150	6/150	4/150	4/150	4/150	6/150
3/150	1/150	7/150	6/150	2/150	6/150
2/150	2/150	2/150	4/150	3/150	6/150
Average =10/600	Average =13/600	Average =17/600	Average =20/600	Average =14/600	Average =21/600
Average percentage of TSCE (%)					
Control	PML RNAi	Control	PML RNAi	Control	PML RNAi
1.7%	2.2%	2.8%	3.3%	2.3%	3.5%

Table S4. CHIP analysis in PML knockdown cells. ES129.1 cells were transfected with either scramble control or PML-specific siRNA oligonucleotides (set#2). CHIP analysis was performed using antisera against H3K9me3, H4K20me3, ATRX and TERF-1, followed by dot blot analysis by hybridization with a γ -32p ATP end-labelled telomere-specific probe. The membrane was exposed overnight on a Typhoon PhosphoImager screen. The intensities of the signals (average signals from 8 sets of experiments) were quantitated using ImageQuant software. The graph is present is Figure 6. The data showed that there were increases in the levels of H3K9me3 and H4K20me3 at telomeres by ~2.3 and 3.4 folds, respectively. In addition, there was an increase in the level of TERF-1 by 2.5 fold, whereas, ATRX level was reduced by half (0.42 fold), indicating a change in telomere chromatin properties in cells depleted of PML bodies.

		H3K9me3/H4	H4K20me3/H4	ATRX/H4	TERF-1/H4
Set 1	control RNAi	4.312372608	1.589548722	3.657718723	0.457781679
	PML RNAi	10.60924814	6.597930243	1.78199156	0.934848117
Set 2	control RNAi	3.497399774	1.944504368	2.547526039	0.622432459
	PML RNAi	9.099310986	6.519464544	1.686144298	0.564184344
Set 3	control RNAi	3.766466342	0.702050028	3.176433747	0.368899084
	PML RNAi	12.52727107	1.963834465	1.130557592	1.406845001
Set 4	control RNAi	3.999414621	0.186866765	2.715411761	1.079034493
	PML RNAi	9.873186165	1.734839595	0.923926565	1.430664885
Set 5	control RNAi	4.312372608	1.076694409	5.709135979	0.355210816
	PML RNAi	7.583572714	7.134483494	1.412933163	0.676673907
Set 6	control RNAi	3.056582117	1.212031806	4.835579463	0.411449347
	PML RNAi	10.89478202	6.716990087	3.411391508	1.332049875
Set 7	control RNAi	4.166925022	2.337286346	4.203692081	0.064487967
	PML RNAi	5.712851623	4.056942018	0.526188307	1.509585288
Set 8	control RNAi	5.337696703	2.398181595	2.922381585	0.924472095
	PML RNAi	10.45528454	4.557298173	1.604104277	2.951697576

N=8		H3K9me3	H4K20me3	ATRX	TERF-1
Fold enrichment of	Control RNAi	4.056153724	1.430895505	3.720984922	0.535470993
	PML RNAi	9.594438407	4.910222827	1.559654659	1.350818624
	Fold difference	2.37	3.43	0.42	2.52