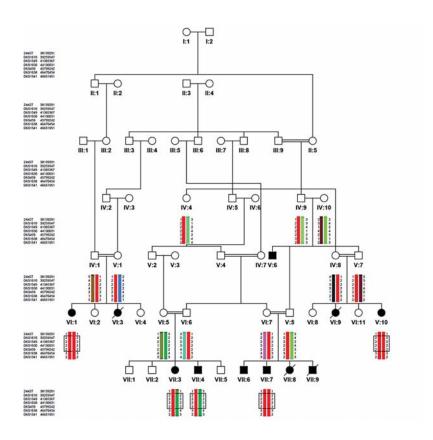
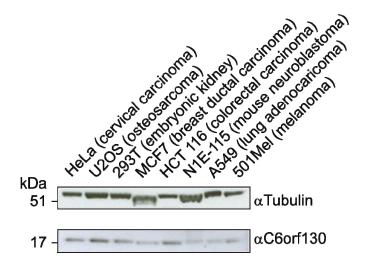
Deficiency of Terminal ADP-Ribose protein Glycohydrolase TARG1/C6orf130 in neurodegenerative disease

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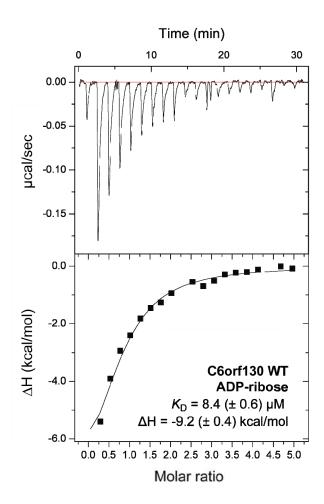
SUPPLEMENTARY FIGURES AND TABLES



Supplementary Figure 1. Refinement mapping of the region on chromosome 6 using microsatellite markers.

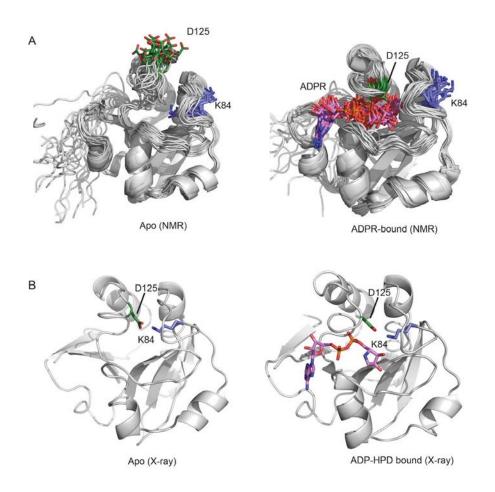


Supplementary Figure 2. Expression profile showing ubiquitous expression of C6orf130 protein across a range of cell lines derived from different human and mouse tissues, as tested by Western blot.

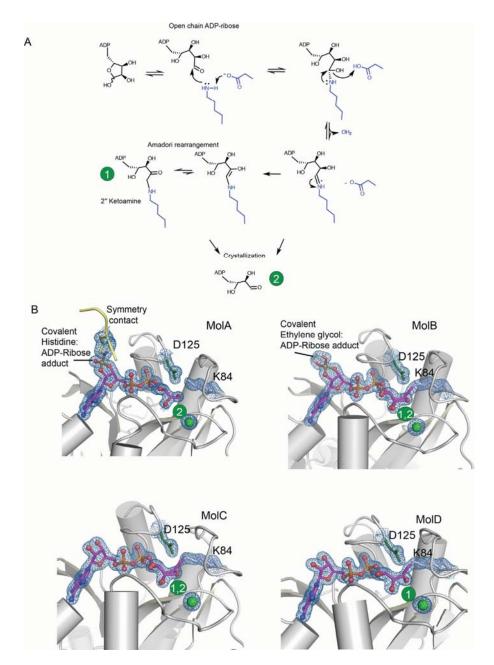


C6orf130	K _d (μM) N	
WT	8.4	0.76
K84A	no binding	
G123E	no binding	
D125A	2.6	0.77

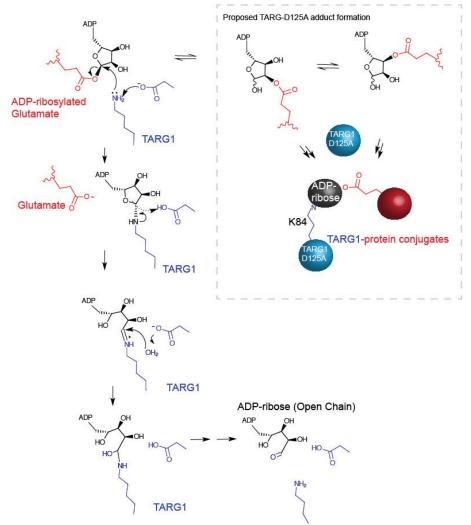
Supplementary Figure 3. Summary of ADP-ribose dissociation constants for C6orf130 protein and its mutant forms as determined by Isothermal Titration Calorimetry. Fitted dissociation constant and number of binding sites are indicated in the table.



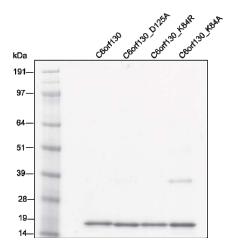
Supplementary Figure 4. Structural comparisons of C6orf130 NMR (Petersen *et al*, 2011) and X-ray structures (This study). (**A**) Apo (RCSB:2LGR) C6orf130 and ADP-ribose bound (RCSB 2L8R) NMR ensembles are displayed, showing an active site excluded conformation of K84. (**B**) X-ray structures of C6orf130/TARG1 show K84 in close proximity to D125 and bound ligand.



Supplementary Figure 5. TARG1/C6orf130 reaction with ADP-ribose. (**A**) Lys 84 reacts with the C1 position of ADP-ribose. Following dehydration, an imine intermediate converts to form a 2"- ketoamine (species "1", green circle) via an Amadori rearrangement (Cervantes-Laurean et al, 1993), and similar to that observed in the X-ray structure of DraG-ADP-ribose complex (Berthold et al, 2009). (**B**) Sigma-A weighted 2Fo-Fc electron density maps (contoured at 1.0 σ for the four C6orf130-ADP-ribose complexes in the crystallographic asymmetric unit. For Molecule D, geometry of the K84 covalent adduct is most consistent for the 2"-ketoamine (species "1"). A second co-crystallized ADP-ribose derived species arising during the three-month crystal growth (MW=528.12 Da, species "2") is apparent for MolA. Molecules B and C, have been modeled with mixed occupancy of species 1 and 2. Molecules A and B additionally are modified by covalent adducts to symmetry related histidine or ethylene glycol on the distal ribose sugars. The nature of these later linkages is uncertain, but the electron density is consistent with a boron bridge, suggesting they arose from contaminating borate in ADP-ribose, crystallization reagents or protein samples.



Supplementary Figure 6. Proposed reaction mechanism for the hydrolysis of a terminal ADP-ribose from PARP. A single ADP-ribose is attached to a glutamate side chain of PARP (red). When the PARP side chain is linked to the 1"-hydroxyl, the Asp-Lys side chains of C6orf130 displace the PARP glutamate, forming a Schiff-base intermediate, which is resolved by water-mediated hydrolysis. Acyl migration (Kasamatsu et al, 2011, Sauve et al, 2001) of the PARP side chain may facilitate 2" or 3"- ADP-ribose linkages. In this case, C6orf130/TARG1 D125A forms adduct without releasing the PARP side chain.



Supplementary Figure 7. Purification of human C6orf130 proteins. Wild-type and mutant proteins were purified as His⁶-tagged proteins by affinity chromatography.

Clinical presentation:	
Onset and course of the disease	4-6 months after the birth, progressive, expiring by age 10-11 years mainly because of aspiration pneumonia
Neurological findings	
Neurodevelopment	Severe delay
Seizure	Generalised tonic-clonic (grand mal)
Motor neuron	Quadriplegia, absence of tendon reflexes
Visual, Hearing	Not assessable
Other neurological findings	Aphasia, partial absence of swallowing reflex
Other systems	No evidence for cardiovascular, renal, gastrointestinal or endocrine abnormalities
Laboratory tests	
Blood, urine, and CSF tests	Normal results for: blood chemistry, blood sugar, complete blood count (CBC), kidney function tests, liver function testes, cerebro-spinal fluid, cellular and biochemistry tests, urine biochemistry, blood gas and amino acids profile
Skin fibroblast cultured analysis Brain and spinal cord CT and MRI scan	Negative for ketolysis defect Normal
EEG	Abnormal indicating general seizure without brain cortex atrophy
EMG & NCV	Abnormal indicating motor- sensorial neuropathy

Supplementary Table 1. Clinical and paraclinical assessment of the 6 affected cases age from 2 to 9 years.

	Heterozygous	Homozygous
Single nucleotide variants (SNVs):		
Coding	10869	7547
Synonymous	5586	3925
Non-synonymous missense	5058	3458
Stopgain	61	20
Stoploss	17	16
Splice site (+/-10bp)	1332	945
Insertion and deletions (indels):		
Coding		
Frameshift deletion	42	18
Frameshift insertion	20	38
In-frame deletion	44	30
In-frame insertion	28	35

Supplementary Table 2. The number of variants for SNVs and indels for the affected case number VII7 identified by new generation sequencing (NGS).

Gene	Description	High expression
DAAM2	dishevelled associated activator of morphogenesis 2	Whole Brain
MOCS1	molybdenum cofactor synthesis 1	Hypothalamous
UBR2	ubiquitin protein ligase E3 component n-recognin 2	Cortex
TBCC	tubulin folding cofactor C	Whole brain
KIAA0240	uncharacterized protein KIAA0240	Cerebellum
KLHDC3	kelch domain containing 3	Whole brain
C6orf108	chromosome 6 open reading frame 108	Whole brain
TTBK1	tau tubulin kinase 1	Cortex and cerebellum
C6orf154	chromosome 6 open reading frame 154	Occipital lobe
AARS2	alanyl-tRNA synthetase 2, mitochondrial (putative)	Whole brain

Supplementary Table 3. The list of genes located on the putative region (Chr6p21) which are highly expressed in the central nervous system and were screened for two affected cases by Sanger sequencing. They did not harbour any pathological variant that could cause the disease phenotype.

	C6orf130 Apo	C6orf130 ADP-HPD	C6orf130 ADP-ribose
Data callection	Сооптоо Аро	COUNTSU ADF-NFD	COULTSO ADE-HBOSE
Data collection	00	D0 0 0	DO.
Space group	C2	P2 ₁ 2 ₁ 2 ₁	P2 ₁
Cell dimensions			
a, b, c (Å)	63.32, 75.37, 35.00	50.80, 72.00, 81.92	73.38, 56.22, 73.79
α, β, γ (°)	90, 100.63, 90	90, 90, 90	90, 94.47, 90
Resolution (Å)	50-1.35 (1.40-1.35)	50-1.25 (1.27-1.25)	50-1.55 (1.58-1.55)
R _{sym} (%)	9.9 (44.5)	7.6 (40.2)	4.3 (53.4)
ΙΙσΙ	11.7 (2.2)	19.3 (2.6)	24.3 (2.5)
Completeness (%)	98.1 (96.2)	94.3 (64.6)	97.9 (96.2)
Redundancy	3.5 (2.9)	5.0 (3.8)	3.8 (3.8)
		()	- ()
Refinement			
Resolution (Å)	50-1.35	50–1.25	50-1.55
No. reflections	32.791	75,062	85,113
R _{work} / R _{free}	12.6 / 16.5	12.0 / 15.4	13.9 / 17.8
No. atoms	12.07 10.0	12.07 10.4	10.07 17.0
Protein	1236	2570	5824
Ligand/ion	n/a	70	272
Water	239	628	658
	239	020	036
B-factors	10.0	40.0	04.0
Protein	19.8	10.2	24.9
Ligand/ion	n/a	19.8	27.7
Water	34.7	26.1	35.2
R.m.s. deviations			
Bond lengths (Å)	0.015	0.015	0.008
Bond angles (°)	1.72	1.91	1.19

Each dataset was collected from a single crystal.
Values in parentheses are for highest-resolution shell.

Supplementary Table 4. Data collection and refinement statistics

Supplementary Table 5. Data collection and refinement statistics

	TARG1/C6orf130 Apo	TARG1/C6orf130 ADP-HPD	TARG1/C6orf130 ADP-ribose
Data collection			
Space group	C2	P2 ₁ 2 ₁ 2 ₁	P2₁
Cell dimensions			·
a, b, c (Å)	63.32, 75.37, 35.00	50.80, 72.00, 81.92	73.38, 56.22, 73.79
α, β, γ (°)	90, 100.63, 90	90, 90, 90	90, 94.47, 90
Resolution (Å)	50-1.35 (1.40-1.35)	50-1.25 (1.27-1.25)	50-1.55 (1.58-1.55)
R _{sym} (%)	9.9 (44.5)	7.6 (40.2)	4.3 (53.4)
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Refinement			
Resolution (Å)	50-1.35	50-1.25	50-1.55
No. reflections	32,791	75,062	85,113
Rwork / Rfree	12.6 / 16.5	12.0 / 15.4	13.7 / 17.5
No. atoms			
Protein	1236	2527	4867
Ligand/ion	n/a	137	248
Water	239	628	668
B-factors			
Protein	19.1	9.8	27.3
Ligand/ion	n/a	10.7	27.7
Water	34.0	26.1	35.0
R.m.s. deviations	0.045	0.045	0.000
Bond lengths (Å)	0.015	0.015	0.008
Bond angles (°)	1.72	1.91	1.19

Each dataset was collected from a single crystal.
Values in parentheses are for highest-resolution shell.