

**Supplementary Information for**  
**Prostaglandin E<sub>2</sub> promotes Th1 differentiation via synergistic**  
**amplification of IL-12 signaling by cAMP and PI3-Kinase**

Chengcan Yao, Takako Hirata, Kitipong Soontrapa, Xiaojun Ma, Hiroshi Takemori and  
Shuh Narumiya\*

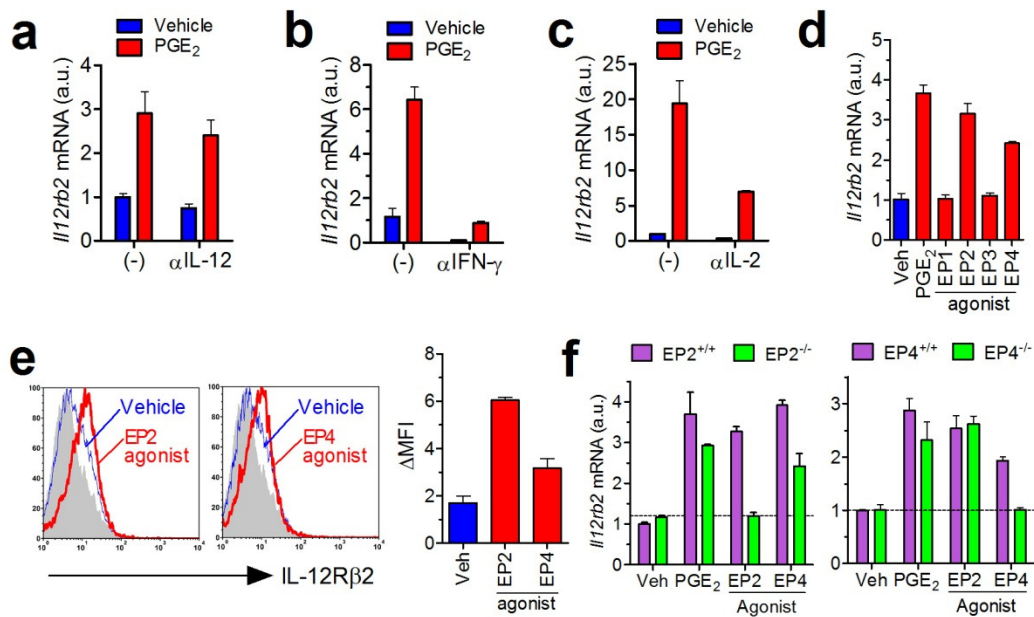
\*To whom correspondence should be addressed. E-mail: snaru@mfour.med.kyoto-u.ac.jp

Supplementary Figure S1. PGE<sub>2</sub>–EP2/EP4 signaling induces IL-12Rβ2 expression in  
TCR-activated CD4<sup>+</sup> T cells.

Supplementary Table S1. Primer sets used for real-time PCR.

Supplementary Table S2. Primer sets used for *Il12rb2* gene locus in ChIP assay.

Supplementary Table S3. Primer sets used for *Ifngr1* gene locus in ChIP assay.



**Supplementary Figure S1. PGE<sub>2</sub>-EP2/EP4 signaling induces IL-12Rβ<sub>2</sub> expression in TCR-activated CD4<sup>+</sup> T cells.** (a,b) Expression of *Il12rb2* mRNA in WT CD4<sup>+</sup> T cells activated for 24 h with anti-CD3 and anti-CD28 in the absence or presence of PGE<sub>2</sub> with or without anti-IL-12 (a) or anti-IFN-γ (b). (c) Expression of *Il12rb2* mRNA in IFN-γR1<sup>-/-</sup> CD4<sup>+</sup> T cells activated for 24 h with anti-CD3 and anti-CD28 in the absence or presence of PGE<sub>2</sub> with or without anti-IL-2. (d) Expression of *Il12rb2* mRNA in T cells activated with αCD3/CD28 in the absence or presence of PGE<sub>2</sub> or agonists selective to EP1 to EP4 for 24 h. (e) IL-12Rβ<sub>2</sub> protein expression in T cells activated for 48 h with αCD3/CD28 in the absence or presence of PGE<sub>2</sub> or selective agonists to EP2 or EP4. (f) Expression of *Il12rb2* mRNA in WT, EP2<sup>-/-</sup> (left) and EP4<sup>-/-</sup> (right) T cells activated for 24 h with anti-CD3 and anti-CD28 in the absence or presence of PGE<sub>2</sub> or selective agonists to EP2 or EP4. Data shown as mean ± SEM are representative of two or more independent experiments with triplicates.

**Supplementary Table S1: Primer sets used for real-time PCR.**

Gene	Universal Probe Library, mouse*	Primer Sequence	
<i>Cd69</i>	#29	Forward	5'-AACGGAAAATAGCTCTTCACATCT-3'
		Reverse	5'-TGATGCTTCTCAAATGTATACTGG-3'
<i>Fosb</i>	#71	Forward	5'-GTTTCGAGAGAGCGGAAC-3'
		Reverse	5'-GCCTTTTCCTCTTCAAGCTG-3'
<i>Gapdh</i>	/	Forward	5'-TGAACGGGAAGCTCACTGG-3'
		Reverse	5'-TCCACCACCCTGTTGCTGTA-3'
<i>Icer</i>	#4	Forward	5'-GCTGAGGCTGATGAAAAACA-3'
		Reverse	5'-GCCACACGATTTTCAAGACA-3'
<i>Ifng</i>	#21	Forward	5'-ATCTGGAGGAACTGGCAAAA-3'
		Reverse	5'-TTCAAGACTTCAAAGAGTCTGAGGTA -3'
<i>Ifngr1</i>	#69	Forward	5'-TCAAAGAGTTCCCTTATGTGCCTA-3'
		Reverse	5'-TACGAGGACGGAGAGCTGTT-3'
<i>Il12rb2</i>	#103	Forward	5'-GTGCCAGGATCCCTCTCTG -3'
		Reverse	5'-GAATGCAACTCTGGTTCTCCA -3'
<i>Il2rb</i>	#91	Forward	5'-AGCATGGGGGAGACCTTC-3'
		Reverse	5'-GGGGCTGAAGAAGGACAAG-3'
<i>Nr4a2</i>	#64	Forward	5'-TCAGAGCCCACGTTCGATT-3'
		Reverse	5'-TAGTCAGGGTTTGCCTGGAA-3'
<i>Ptger4</i>	#77	Forward	5'-CCTAACCCACCCCTACAGGT-3'
		Reverse	5'-AGAAGGACGCGTTGACTCC-3'
<i>Rn18s</i>	#48	Forward	5'-GCAATTATCCCATGAACG-3'
		Reverse	5'-GGGACTTAATCAACGCAAGC-3'
<i>Tbx21</i>	#19	Forward	5'-TCAACCAGCACCAGACAGAG -3'
		Reverse	5'-AAACATCCTGTAATGGCTTGTG -3'

\* Universal ProbeLibrary (Roche) were used together with the LightCycler Taqman Master kit (Roche). Otherwise, FastStart DNA Master<sup>PLUS</sup> SYBR Green I kit (Roche) was used without Universal Probe Library.

**Supplementary Table S2. Primer sets used for *Ili2rb2* gene locus in ChIP assay.**

Primer Set	Universal Probe Library, mouse*	Primer Sequence	
A	/	Forward	5'-TTATGTCCTGCAGACAGGAGTG-3'
		Reverse	5'-CATCTGCAGGTAAATCAAAGCA-3'
B		Forward	5'-AGGGAAATTACTGGAATGATTTATGG-3'
		Reverse	5'-GACTGAGCAATTACTAGATTCTTGGA-3'
		Probe	5'-CTGCAGTCCAATAACCCAACAATAGGTAC-3'
C	#69	Forward	5'-CAGCACGTCTTCTGTATAAACTGG-3'
		Reverse	5'-TGCACTGACTTGCTAACACATCT-3'
D		Forward	5'-GAGACTAGGATGGAAAATATAATCAA-3'
		Reverse	5'-TCTGGATGAAGCCTCTCTGA-3'
		Probe	5'-TTCTGCAATACTCATAGATTGGTGCCTAGC-3'
E		Forward	5'-GGACCTCACAGAGACAGAAACA-3'
		Reverse	5'-ACACCAAGCTAACAACCATAACATA-3'
		Probe	5'-ATGCAGAACTACCACAGACCCAAGCAGTC-3'
F	#110	Forward	5'-CGTCAGAAATGGAACTGAGC-3'
		Reverse	5'-TGCCTGTGATGAAACTCAGC-3'
G	#98	Forward	5'-TGAGTGCCACTGTGGTTATGA-3'
		Reverse	5'-CCAACACGTCATCCAAAATG-3'
H	/	Forward	5'-AAATTTGGGCTTTTAAATATTTTGC-3'
		Reverse	5'-GGCCCACTTGAGCTATGGAG-3'
I	/	Forward	5'-GTCCACTTTCGGTTTTATATGTCC-3'
		Reverse	5'-GAACTTGCCCAGTGCAGGAG-3'
J	#109	Forward	5'-TCTGCCTGGACACTTGTTCAT-3'
		Reverse	5'-GACGCAGAGAGGGATCCTG-3'
K	#82	Forward	5'-TGCCGTCACTGCTTTCAATA-3'
		Reverse	5'-TTCCAACAGCAAAGGAAACC-3'
L	#82	Forward	5'-TGCCGTCACTGCTTTCAATA-3'
		Reverse	5'-CAAATGCTGTTTCCCTGTGC-3'
M	/	Forward	5'-TAGTCCGTGTTACTGCCATCAA-3'
		Reverse	5'-GTAAATGACGCCTGATTAAGTGC-3'
N	#32	Forward	5'-CTCCGTGGGACATCAGAATC-3'
		Reverse	5'-GCCCTCATCTTCCCCT-3'
O	#50	Forward	5'-TCATGACGTCAGGTGACCTACTA-3'
		Reverse	5'-GCAGTTGATTATATACCCGAATTTCT-3'
P	#5	Forward	5'-TGCATTTCAAAAATATAAACCATCA-3'
		Reverse	5'-GATTTTGGCTTTGGGGATTT-3'

\* Universal Probe Library (Roche) or specified probes were used together with the LightCycler Taqman Master kit (Roche). Otherwise, FastStart DNA Master<sup>PLUS</sup> SYBR Green I kit (Roche) was used without Universal Probe Library.

**Supplementary Table S3. Primer sets used for *Ifngr1* gene locus in ChIP assay.**

Primer Set	Universal Probe Library, mouse*	Primer Sequence	
A	/	Forward	5'-CATGCCACTGCAGTAAACTGTC-3'
		Reverse	5'-CCAGTTTGGACTCTGTCTCTTCC-3'
B	#13	Forward	5'-ACGTGTCACTCCCTGATTGG-3'
		Reverse	5'-GCCGAGGGTGGTTCTCTACT-3'
C	/	Forward	5'-TTCTGAAATGTGTGACGTGAGG-3'
		Reverse	5'-GCTCATCGACCCTTAACAAAAA-3'
D	/	Forward	5'-CTATTCCTTCCCCCTTAATCTCC-3'
		Reverse	5'-CTGTGCGCATGCTTCCTAAG-3'
E		Forward	5'-CCCACACGCACACGCATG-3'
		Reverse	5'-CCACTGAGGAAGAGGCACTGTA-3'
		Probe	5'-CGCACGCACGTCTGATACTGTAATCTCATT-3'
F	#16	Forward	5'-CCTCAGTGGATACAAGGTCCA-3'
		Reverse	5'-GGATACCCAAGATATAGCTATGCAG-3'
G	/	Forward	5'-CGGAAACACTTAACTCCCAAGC-3'
		Reverse	5'-CCATTTCTGAGGAAGGATCTGG-3'
H	#64	Forward	5'-CACTTCATAATCTGCTTCCTGGT-3'
		Reverse	5'-ATGCGGCTTGCAGATTTT-3'
I	/	Forward	5'-CATCTGTAGCACCAAGGGTGAG-3'
		Reverse	5'-CCTTGTGCAGGTTCAAGATTGAG-3'
J	#107	Forward	5'-TTTATCCTGACGTCAAAGTCTGAG-3'
		Reverse	5'-CCAGCTAAGGCTTGTATTTGTC-3'
K	/	Forward	5'-TGACGTCAAAGTCTGAGAAATGC-3'
		Reverse	5'-TCTCTGAGTCTTCCAAGCAAAATG-3'
L	/	Forward	5'-TGTAGGAGGGGGTGCAAATG-3'
		Reverse	5'-ACACTCTTGTGCTGCCTCACTC-3'
M		Forward	5'-CCAGGCTGGTCTCTCAAGCTT-3'
		Reverse	5'-CCTGGCCTTGGCGTCA-3'
		Probe	5'-CTTGCGGACTTGGC-3'
N	/	Forward	5'-GATTCTGCTGGTGGTCCTGAT-3'
		Reverse	5'-CAGCGGCTCGGAGAGATTAC-3'
O	/	Forward	5'-GGCATGGGGAGACCTCTTTAAC-3'
		Reverse	5'-AATCAGGTCAAGCAGGATTG-3'
P	#40	Forward	5'-GGCCAAAGCGAGACTGAA-3'
		Reverse	5'-GACGTCAGACATGGAGATGC-3'

\* Universal Probe Library (Roche) or specified probes were used together with the LightCycler Taqman Master kit (Roche). Otherwise, FastStart DNA Master<sup>PLUS</sup> SYBR Green I kit (Roche) was used without Universal Probe Library.