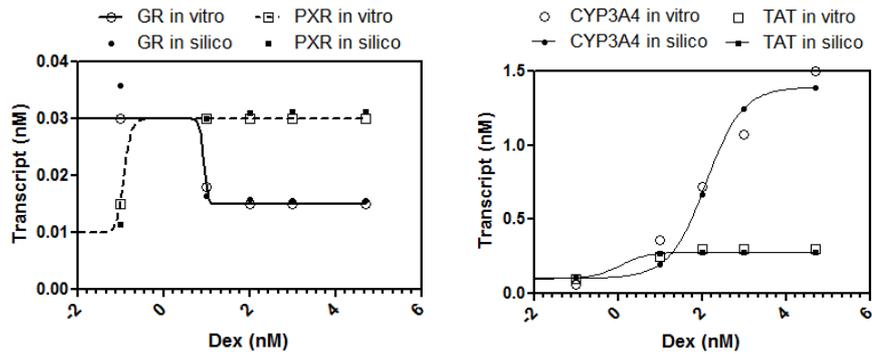
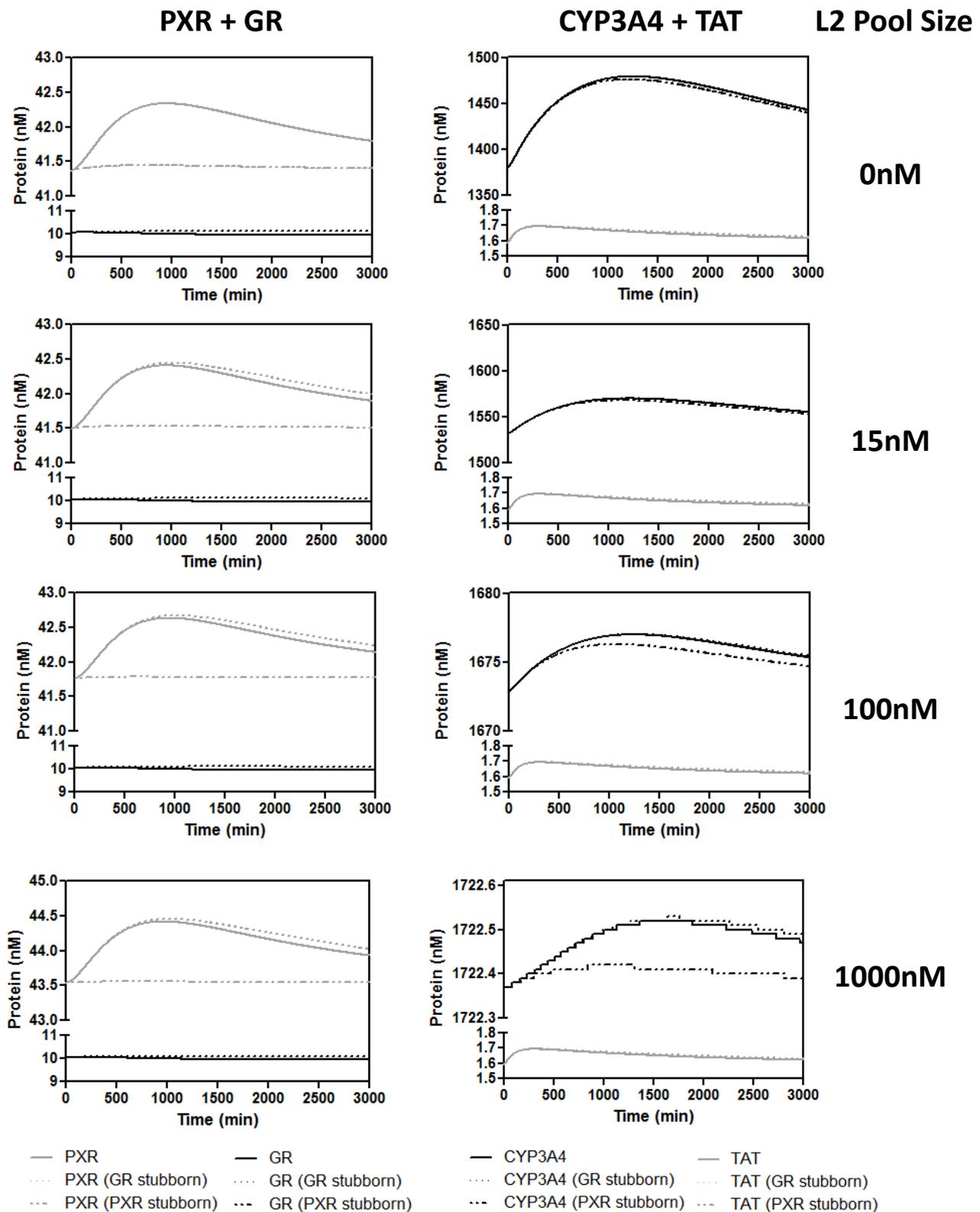


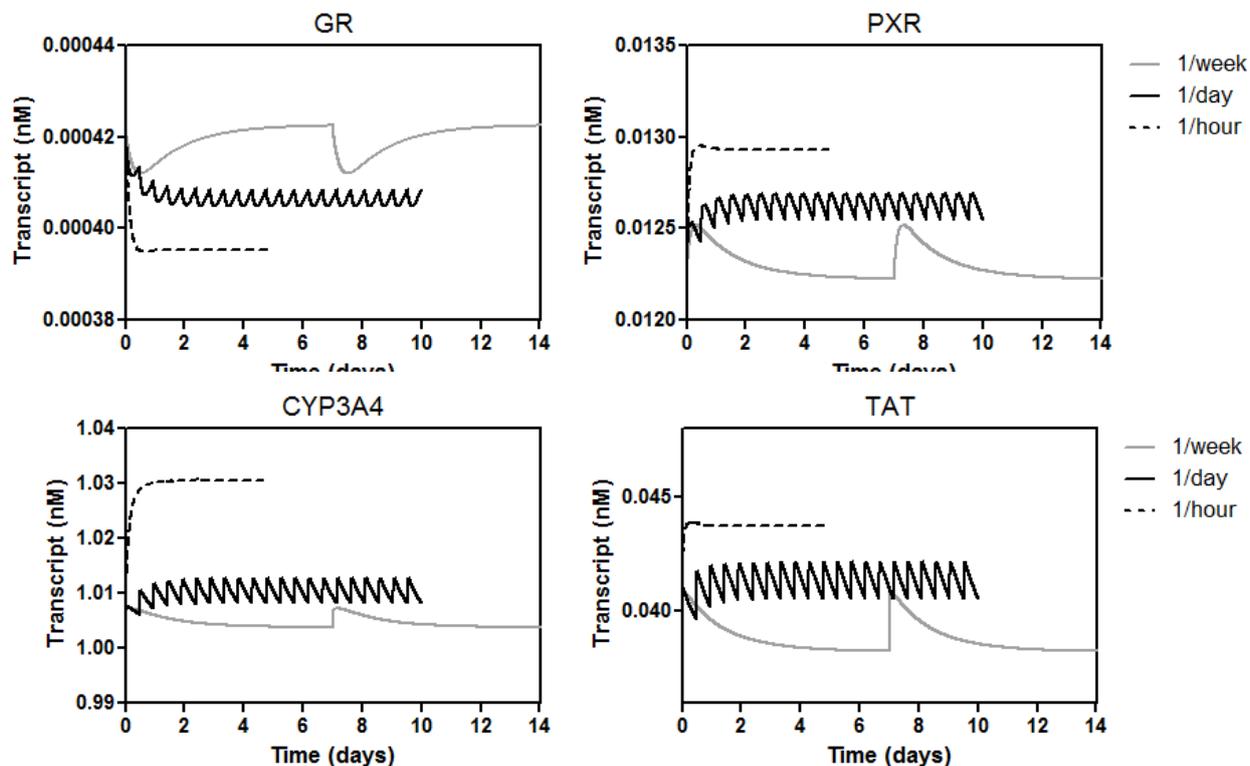
Supplementary Figure S1: GR-PXR signal regulatory network: GR-PXR signal regulatory network was constructed in using CellDesigner (v4.0.1; Systems Biology Institute, <http://celldesigner.org/index.html>), a graphical front-end for creating process diagrams of biochemical networks in Systems Biology Markup Language (SBML) and utilising Systems Biology Graphical Notation (SBGN). Each individual chemical or protein is identified as a species (Gene = gene; mRNA = mRNA; Protein or NR = protein; chemical = chemical). Interactions between species are identified as reactions (re1....ren) , and correspond to reaction identifiers (v1...vn) in Supplementary table S1.



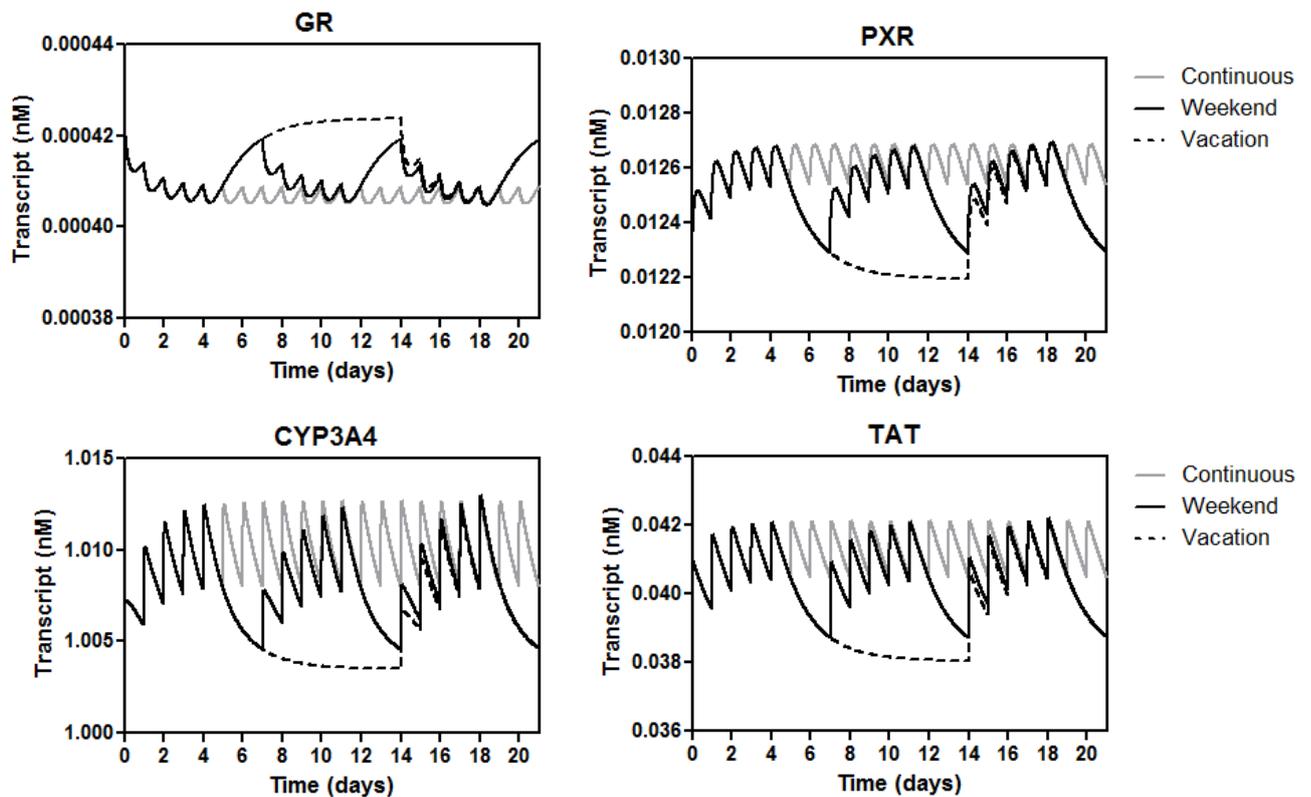
Supplementary Figure S2: In silico model for the response to the artificial glucocorticoid Dexamethasone: Primary human hepatocytes were exposed to vehicle or (0.1-50000 nM) dexamethasone as indicated for 48 hours, RNA extracted and transcript levels of PXR, GR, CYP3A4 and TAT quantified by TaqMan Q-PCR. Average transcript levels (\pm SEM) from triplicate cultures are indicated by open symbols, and the concentration-response curve shown by the full line was fitted to these data. Transcript levels of PXR, GR, CYP3A4 and TAT following 48 hours exposure to indicated concentrations of cortisol were also simulated by the model, and are presented as closed symbols.



Supplementary Figure S3: Impact of L2 pool size on response of the gene regulatory network to an acute stress episode: Protein levels for GR, PXR, CYP3A4 and TAT were simulated over 3000 minutes following addition of a single spike of cortisol, doubling free cortisol concentration from 15-30nM. Simulations were undertaken in the presence of a second PXR ligand pool (L2), at the indicated concentration. Solid lines represent levels under the complete single-ligand model, whereas dashed lines represent levels GRstubborn and PXRstubborn scenarios as indicated.



Supplementary Figure S4: Response of the gene regulatory network to repeat stress episodes: Transcript levels for GR, PXR, CYP3A4 and TAT were simulated over 21 days following addition of repeat spikes of cortisol, such that each spike doubled the free blood cortisol concentration. Frequency of cortisol spikes were 1/week (grey line), 1/day (black line) or 1/hour (dashed line).



Supplementary Figure S5: A low stress vacation required to allow adaptation to a stressful week: Transcript levels for GR, PXR, CYP3A4 and TAT, as well as free blood cortisol, were simulated over 21 days following addition of repeat spikes of cortisol, such that each spike doubled the free blood cortisol concentration. Cortisol spikes were applied 1/day for either continuous throughout the period (grey line), with a weekend break (black line) or a 9-day vacation (dashed line).

Supplementary Table S1: Model Parameters for cortisol (L1) model

Reactions		Parameters
v₁	CYPmRNA synthesis: $k_1 \cdot CYPgene(t)$ [nM/min]	* $k_1 = 0.00321 \text{ min}^{-1}$
v₂	CYPmRNA degradation: $k_2 \cdot CYPmRNA(t)$ [nM/min]	$k_2 = 0.04 \text{ min}^{-1}$ 41,42
v₃	CYPprotein synthesis: $k_3 \cdot CYPmRNA(t)$ [nM/min]	$k_3 = 2.5 \text{ min}^{-1}$ ⁴¹
v₄	CYPprotein degradation: $k_4 \cdot CYPptotein(t)$ [nM/min]	$k_4 = 0.0005 \text{ min}^{-1}$ 43,44
v₅	PXRmRNA synthesis on inactive PXRgene: $k_5 \cdot PXRgene(t)$ [nM/min]	* $k_5 = 5.52e-05 \text{ min}^{-1}$ 1
v₆	PXRmRNA degradation: $k_6 \cdot PXRmRNA(t)$ [nM/min]	** $k_6 = 0.006 \text{ min}^{-1}$ 45
v₇	PXRprotein synthesis: $k_7 \cdot PXRmRNA(t)$ [nM/min]	$k_7 = 10 \text{ min}^{-1}$ ⁴⁶
v₈	PXRprotein degradation: $k_8 \cdot PXRptotein(t)$ [nM/min]	$k_8 = 0.003 \text{ min}^{-1}$ ⁴⁷
v₉	GRmRNA synthesis on inactive GRgene: $k_9 \cdot GRgene(t)$ [nM/min]	* $k_9 = 3.2e-06 \text{ min}^{-1}$
v₁₀	GRmRNA degradation: $k_{10} \cdot GRmRNA(t)$ [nM/min]	** $k_{10} = 0.003 \text{ min}^{-1}$ 45
v₁₁	GRprotein synthesis: $k_{11} \cdot GRmRNA(t)$ [nM/min]	$k_{11} = 19.98 \text{ min}^{-1}$ ⁴⁸
v₁₂	GRprotein degradation: $k_{12} \cdot GRptotein(t)$ [nM/min]	$k_{12} = 0.001 \text{ min}^{-1}$ 48,49
v₁₃	TATmRNA synthesis on inactive TATgene: $k_{13} \cdot TATgene(t)$ [nM/min]	* $k_{13} = 0.000855 \text{ min}^{-1}$
v₁₄	TATmRNA degradation: $k_{14} \cdot TATmRNA(t)$ [nM/min]	$k_{14} = 0.064 \text{ min}^{-1}$ ⁵⁰
v₁₅	Cortisol binding GRprotein: $k_{15f} \cdot Cort(t) \cdot GRprotein - k_{15b} \cdot GRproteinCort(t)$ [nM/min]	$k_{15f} = 60 \text{ nM}^{-1} \text{ min}^{-1}$ (diffusion limited); $k_{15b} = 600 \text{ min}^{-1}$ (Kd = 10 nM ⁵¹)
v₁₆	GRproteinCort degradation: $k_{16} \cdot GRptoteinCort(t)$ [nmoles/min]	$k_{16} = 0.001 \text{ min}^{-1}$ 48,49

v17	Cortisol binding PXRprotein: $k_{17f} \cdot Cort(t) \cdot PXRprotein - k_{17b} \cdot PXRproteinCort(t)$ [nM/min]	$k_{17f} = 60 \text{ nM}^{-1}\text{min}^{-1}$ (diffusion limited); $k_{17b} = 600000 \text{ min}^{-1}$ 52
v18	PXRproteinCort degradation: $k_{18} \cdot PXRptotein Cort(t)$ [nM/min]	$k_{18} = 0.0015 \text{ min}^{-1}$ 47
v19	Cort degradation: $CYPptotein \cdot \frac{V_{mC} \cdot \frac{Cort(t)}{K_{MC}}}{1 + \frac{Cort(t)}{K_{MC}} + \frac{L2(t)}{K_{ML2}}}$ [nM/min]	$V_{mC} = 10 \text{ min}^{-1}$ ($k_{cat} = V_{mC} / 1200(\text{nM max CYP protein conc in cell}) \text{ nM}^{-1} = 0.083$); $K_{MC} = 15000 \text{ nM}$ Since $L2=0$, Competition between cortisol and L2 for CYP is neglected ^{53,54}
v20	PXRpoteinCortisol binding CYPgene: $k_{20f} \cdot PXRproteinCort(t) \cdot CYPgene(t) - k_{20b} \cdot PXRproteinCortCYPgen e(t)$ [nM/min]	$k_{20f} = 60 \text{ nM}^{-1}\text{min}^{-1}$ (diffusion limited); * $k_{20b} = 1 \text{ min}^{-1}$
v21	CYPmRNA synthesis on active PXRproteinCortCYPgene: $k_{21} \cdot PXRproteinCortCYPgene(t)$ [nM/min]	* $k_{21b} = 0.05 \text{ min}^{-1}$
v22	GRproteinCortisol binding PXRgene: $k_{22f} \cdot GRproteinCort(t) \cdot PXRgene(t) - k_{22b} \cdot GRproteinCortPXRgene(t)$ [nM/min]	$k_{22f} = 60 \text{ nM}^{-1}\text{min}^{-1}$ (diffusion limited); * $k_{22b} = 200 \text{ min}^{-1}$
v23	PXRmRNA synthesis on active GRproteinCortPXRgene: $k_{23} \cdot GRproteinC ortPXRgene(t)$ [nM/min]	* $k_{23} = 0.00011 \text{ min}^{-1}$ 1
v24	GRpoteinCortisol binding GRgene: $k_{24f} \cdot GRproteinCort(t) \cdot GRgene(t) - k_{24b} \cdot GRproteinC ortGRgene(t)$ [nM/min]	$k_{24f} = 60 \text{ nM}^{-1}\text{min}^{-1}$ (diffusion limited); * $k_{24b} = 60 \text{ min}^{-1}$
v25	GRmRNA synthesis on active GRproteinCortGRgene: $k_{25} \cdot GRproteinC ortGRgene(t)$ [nM/min]	* $k_{25} = 1.2\text{e-}06 \text{ min}^{-1}$
v26	GRproteinCortisol binding TATgene: $k_{26f} \cdot GRproteinCort(t) \cdot TATgene(t) - k_{26b} \cdot GRproteinC ortTATgene(t)$ [nM/min]	$k_{26f} = 60 \text{ nM}^{-1}\text{min}^{-1}$ (diffusion limited); * $k_{26b} = 300 \text{ min}^{-1}$

v27	TATmRNA synthesis on active GRproteinCortTATgene: $k_{27} \cdot GRproteinCortTATgene(t)$ [nM/min]	* $k_{27} = 0.005 \text{ min}^{-1}$
v28	DEX binding GRprotein: $k_{28f} \cdot DEX(t) \cdot GRprotein - k_{28b} \cdot GRproteinDEX(t)$ [nM/min]	$k_{28f} = 60 \text{ nM}^{-1} \text{ min}^{-1}$ (diffusion limited); $k_{28b} = 60 \text{ min}^{-1}$ (calculated from Kd that is in the order of magnitude of 1 nM ⁵⁵)
v29	GRproteinDEX degradation: $k_{29} \cdot GRproteinDEX(t)$ [nmoles/min]	$k_{29} = 0.001 \text{ min}^{-1}$ 48,49
v30	DEX binding PXRprotein: $k_{30f} \cdot DEX(t) \cdot PXRprotein - k_{30b} \cdot PXRproteinDEX(t)$ [nM/min]	$k_{30f} = 60 \text{ nM}^{-1} \text{ min}^{-1}$ (diffusion limited);; $k_{30b} = 60000 \text{ min}^{-1}$ (calculated from Kd that is in the order of magnitude of 1000 nM ⁵⁶)
v31	PXRproteinDEX degradation: $k_{31} \cdot PXRproteinDEX(t)$ [nmoles/min]	** $k_{31} = 0.0015 \text{ min}^{-147}$
v32	DEX degradation: $CYPprotein \cdot \frac{V_{mDEX} \cdot DEX(t)}{1 + \frac{DEX(t)}{K_{MDEX}} + \frac{Cort(t)}{K_{MC}} + \frac{L2(t)}{K_{M:2}}}$ [nM/min]	$V_{mDEX} = 5.1 \text{ min}^{-1}$; ($k_{cat} = V_{mC} / 1200$ (max CYP protein conc in cell) nM = 0.00425);) $K_{MDEX} = 23000 \text{ nM}$ 57
v33	PXRproteinDEX binding CYPgene: $k_{33f} \cdot PXRproteinDEX(t) \cdot CYPgene(t) - k_{33b} \cdot PXRproteinDEXCYPgene(t)$ [nM/min]	$k_{33f} = 60 \text{ nM}^{-1} \text{ min}^{-1}$ (diffusion limited); * $k_{33b} = 1 \text{ min}^{-1}$
v34	CYPmRNA synthesis on active PXRproteinDEXCYPgene: $k_{34} \cdot PXRproteinDEXCYPgene(t)$ [nM/min]	* $k_{34} = 0.05 \text{ min}^{-1}$
v35	GRproteinDEX binding PXRgene:	$k_{35f} = 60 \text{ nM}^{-1} \text{ min}^{-1}$

	$k_{35f} \cdot GRproteinDEX(t) \cdot PXRgene(t)$ $- k_{35b} \cdot GRproteinDEXPXRgene(t)$ [nM/min]	(diffusion limited); * $k_{35b} = 200 \text{ min}^{-1}$
v36	PXRmRNA synthesis on active GRproteinDEXPXRgene: $k_{36} \cdot GRproteinDEXPXRgene(t)$ [nM/min]	* $k_{36} = 0.00011 \text{ min}^{-1}$
v37	GRproteinDEX binding GRgene: $k_{37f} \cdot GRproteinDEX(t) \cdot GRgene(t)$ $- k_{37b} \cdot GRproteinDEXGRgene(t)$ [nM/min]	$k_{37f} = 60 \text{ nM}^{-1} \text{ min}^{-1}$ (diffusion limited); * $k_{37b} = 60 \text{ min}^{-1}$
v38	GRmRNA synthesis on active GRproteinDEXGRgene: $k_{38} \cdot GRproteinDEXGRgene(t)$ [nM/min]	* $k_{38} = 1.2e-06 \text{ min}^{-1}$
v39	GRproteinDEX binding TATgene: $k_{39f} \cdot GRproteinDEX(t) \cdot TATgene(t)$ $- k_{39b} \cdot GRproteinDEXTATgene(t)$ [nM/min]	$k_{39f} = 60 \text{ nM}^{-1} \text{ min}^{-1}$ (diffusion limited); * $k_{39b} = 300 \text{ min}^{-1}$
v40	TATmRNA synthesis on active GRproteinDEXTATgene: $k_{40} \cdot GRproteinDEXTATgene(t)$ [nM/min]	* $k_{40} = 0.005 \text{ min}^{-1}$
v41	DEX transport through cellular membrane: $k_{41f} \cdot DEXout(t) - k_{41b} \cdot DEXin(t)$ [nM/min]	$k_{41f} = 100 \text{ min}^{-1}$; $k_{41b} = 100 \text{ min}^{-1}$ (taken at high value not to limit DEX uptake)
v42	Endogenous cortisol synthesis: $k_{42f} \cdot Cortisone(t) - k_{42b} \cdot Cort(t)$ [nM/min]	$k_{42f} = 0.016 \text{ min}^{-1}$; $k_{42b} = 0.016 \text{ min}^{-1}$ (fitted to have realistic cortisol concentration)
v43	TATprotein synthesis: $k_{43} \cdot TATmRNA(t)$ [nM/min]	* $k_{43} = 0.5 \text{ min}^{-1}$
v44	TATprotein degradation: $k_{44} \cdot TATprotein(t)$ [nM/min]	$k_{44} = 0.012 \text{ min}^{-1}$ ⁵⁰
V_{cortisolT} ransport	Transport of cortisol through cellular membrane: $k_{ctf} \cdot CortOUT(t) - k_{ctb} \cdot Cort(t)$ [nM/min]	$k_{ctf} = 1000 \text{ min}^{-1}$; $k_{ctb} = 1000 \text{ min}^{-1}$
V_{L2_PXR} _binding	Ligand2 binding PXRprotein: $k_{l2pxrf} \cdot L2(t) \cdot PXRprotein - k_{l2pxrb} \cdot PXRproteinLigand2(t)$ [nM/min]	$k_{l2pxrf} = 0 \text{ nM}^{-1} \text{ min}^{-1}$ ¹ (diffusion limited); $k_{l2pxrb} = 0 \text{ min}^{-1}$ ⁵²

$V_{L2_PXR_deg}$	PXRproteinLigand2 degradation: $k_{l2_pxr} \cdot PXRproteinLigand2(t)$ [nM/min]	$k_{l2_pxr} = 0 \text{ min}^{-1} 47$
$V_{CYPmR_NA_synt_PXR_L2}$	CYPmRNA synthesis on active PXRproteinLigand2CYPgene: $k_{synt_cyp_b} \cdot PXRproteinLigand2CYPgene(t)$ [nM/min]	* $k_{synt_cyp_b} = 0 \text{ min}^{-1}$
$V_{CYPmR_NA_PXR_L2_binding}$	PXRproteinLigand2 binding CYPgene: $k_{bind_cyp_f} \cdot PXRproteinLigand2(t) \cdot CYPgene(t)$ $- k_{bind_cyp_b} \cdot PXRproteinLigand2CYPgene(t)$ [nM/min]	$k_{bind_cyp_f} = 0 \text{ nM}^{-1} \text{ min}^{-1}$ (diffusion limited); * $k_{bind_cyp_b} = 0 \text{ min}^{-1}$
$V_{cort_distrib}$	Cortisol through the blood $k_{distribution} \cdot CortOUT(t)$ [nM/min]	$k_{distribution} = 1000 \text{ min}^{-1}$
V_{cort_CBG}	Transport of cortisol through cellular membrane: $k_{cortCBG_1} \cdot CortOUT(t) - k_{cortCBG_2} \cdot CBG_CortOUT(t)$ [nM/min]	$k_{cortCBG_1} = 60 \text{ nM}^{-1} \text{ min}^{-1}$ (diffusion limited); $k_{cortCBG_2} = 270 \text{ min}^{-1}$
$V_{cort_albumin}$	$k_{cortALB_1} \cdot CortOUT(t) - k_{cortALB_2} \cdot Alb_CortOUT(t)$	$k_{cortALB_1} = 60 \text{ nM}^{-1} \text{ min}^{-1}$ (diffusion limited); $k_{cortALB_2} = 900000 \text{ min}^{-1}$

Supplementary Table S2: Balance Equations for cortisol (L1) model

Balance equations	
dCort(t)/dt	$-V_{15}+V_{16} -V_{17}+V_{18}-V_{19}+V_{42}+ V_{cortisolTransport}$ [nM/min]
dCortAdded(t)/dt	$- V_{cort_distribution}$ [nM/min]
dCortOUT(t)/dt	$V_{cort_distribution} -V_{cort_albumin} - V_{cort_CBG} - V_{cortisolTransport}$ [nM/min]
dCBG_CortOUT(t)/dt	V_{cort_CBG} [nM/min]
dAlb_CortOUT(t)/dt	$V_{cort_albumin}$ [nM/min]
dCBG(t)/dt	$-V_{cort_CBG}$ [nM/min]
dAlb(t)/dt	$-V_{cort_albumin}$ [nM/min]
dCortisone(t)/dt	0 (considered an infinite reservoir)
dCYPgene(t)/dt	$-V_{20}-V_{33}$ [nM/min]
dCYPprotein(t)/dt	$+V_3-V_4$ [nM/min]
dCYPmRNA(t) /dt	$+V_1-V_2 +V_{21}+V_{34}+V_{CYPmRNA_PXR_L2_binding}$ [nM/min]

$dPXR_{proteinCortCYPgene}(t) / dt$	$+V_{20}$ [nM/min]
$dPXR_{proteinLigand2CYPgene}(t) / dt$	$+ V_{CYPmRNA_PXR_L2_binding} - V_{CYPmRNA_synt_PXR_L2}$ [nM/min]
$dPXR_{proteinDEXCYPgene}(t) / dt$	$+V_{33}$ [nM/min]
$dDEX(t)/dt$	$-V_{28}+V_{29} -V_{30}+V_{31}-V_{32}+V_{41}$ [nM/min]
$dLigand2(t)/dt$	$- V_{L2_PXR_binding} + V_{L2_PXR_deg}$ [nM/min]
$dDEXdeg(t)/dt$	$+V_{32}$ [nM/min]
$dDEXout(t)/dt$	$-V_{41}$ [nM/min]
$dGRgene(t)/dt$	$-V_{24} -V_{37}$ [nM/min]
$dGRprotein(t)/dt$	$+V_{11}-V_{12}-V_{15}-V_{12}-V_{28}$ [nM/min]
$dGRmRNA(t)/dt$	$+V_9-V_{10}+V_{25}+V_{38}$ [nM/min]
$dGRproteinCortGRgene(t)/dt$	$+V_{24}$ [nM/min]
$dGRproteinDEXGRgene(t)/dt$	$+V_{37}$ [nM/min]
$dGRproteinCort(t)/dt$	$+V_{15}-V_{16}-V_{22}-V_{24}-V_{26}$ [nM/min]
$dGRproteinDEX(t)/dt$	$+V_{28}-V_{29}-V_{35}-V_{37}-V_{39}$ [nM/min]
$dPXRgene(t)/dt$	$-V_{35} -V_{22}$ [nM/min]
$dPXR_{protein}(t)/dt$	$+V_7-V_8-V_{17}-V_{30}$ [nM/min]
$dPXRmRNA(t)/dt$	$+V_5-V_6+V_{23}+V_{36}$ [nM/min]
$dGRproteinCortPXRgene(t)/dt$	$+V_{22}$ [nM/min]
$dGRproteinDEXPXRgene(t)/dt$	$+V_{35}$ [nM/min]
$dPXR_{proteinCort}(t)/dt$	$+V_{17}-V_{18}-V_{20}$ [nM/min]
$dPXR_{proteinLigand2}(t)/dt$	$+V_{L2_PXR_binding} - V_{L2_PXR_deg} - V_{CYPmRNA_PXR_L2_binding}$ [nM/min]
$dPXR_{proteinDEX}(t)/dt$	$+V_{30}-V_{31}-V_{33}$ [nM/min]
$dTATgene(t)/dt$	$-V_{26}-V_{39}$ [nM/min]
$dTATmRNA(t)/dt$	$+V_{13}-V_{14}+V_{27}+V_{40}$ [nM/min]
$dGRproteinCortTATgene(t)/dt$	$+V_{26}$ [nM/min]
$dGRproteinDEXTATgene(t)/dt$	$+V_{39}$ [nM/min]
$dTATprotein(t)/dt$	$+V_{43}-V_{44}$ [nM/min]

Supplementary Table S3: Conserved Moieties for cortisol (L1) model

Conserved Moieties		
$GR_{gene_{total}}$	$GRgene(t) + GR_{proteinCortGRgene}(t) + GR_{proteinDEXGRgene}(t)$	****0.83 [nM]
$PXR_{gene_{total}}$	$PXRgene(t) + GR_{proteinCortPXRgene}(t) + GR_{proteinDEXPXRgene}(t)$	****0.83 [nM]
$CYP_{gene_{total}}$	$CYPgene(t) + PXR_{proteinCortCYPgene}(t) + PXR_{proteinLigand2CYPgene}(t) + PXR_{proteinDEXCYPgene}(t)$	****0.83 [nM]
DEX_{total}	$DEX(t) + DEXdeg(t) + DEXout(t) +$	1000 [nM]

	GRproteinDEXGRgene(t) + GRproteinDEX(t) + GRproteinDEXPXRgene(t) + PXRproteinDEX(t) + GRproteinDEXTATgene(t) + PXRproteinDEXCYPgene(t)	
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Supplementary Table S4: Initial Conditions for cortisol (L1) model

Initial conditions	
Cort(0)	1.14 [nM]
Cortisone(0)	23 [nM]
CortOUT(0)	0[nM]
CortAdded(0)	0[nM] at t=0 for the initial state, then ~130 [nM] added into the blood as an exposure of cortisol.
CBG_CortOUT(0)	0[nM]
Alb_CortOUT(0)	0[nM]
CBG(0)	550[nM]
Alb(0)	60000[nM]
Ligand2(0)	0[nM]
CYPgene(0)	0.82976 [nM]
CYPprotein(0)	104 [nM]
CYPmRNA(0)	75e-3 [nM] 94x levels of GR mRNA. Experimentally derived and consistent with ^{58,59}
PXRproteinCortCYPgene(0)	0.00024 [nM]
PXRproteinLigand2CYPgene(0)	0 [nM]
PXRproteinDEXCYPgene(0)	0 [nM]
DEX(0)	0 [nM]
DEXdeg(0)	0 [nM]
DEXout(0)	1000 [nM]
GRgene(0)	0.5 [nM]
GRprotein(0)	GRtotal=80.0 nM =GR (47.24) +GRprotCort(32.36) + GRprotCortGRgene (0.33) + GRproteinCortTATgene(0.02) + GRproteinCortPXRgene(0.05) ^{60,61}
GRmRNA/dt	0.0008 [nM] Experimentally derived and consistent with ^{58,62}
GRproteinCortGRgene(0)	0.33 [nM]
GRproteinDEXGRgene(0)	0 [nM]

GRproteinCort(0)	32.36 [nM]
GRproteinDEX(0)	0 [nM]
PXRgene(0)	0.78 [nM]
PXRprotein(0)	*****99.9 [nM]
PXRmRNA(0)	0.007 [nM] 9x levels of GR mRNA. Experimentally derived and consistent with ^{58,59}
GRproteinCortPXRgene(0)	0.05 [nM]
GRproteinDEXPXRgene(0)	0 [nM]
PXRproteinCort(0)	0.01 [nM]
PXRproteinLigand2(0)	0 [nM]
PXRproteinDEX(0)	0 [nM]
TATgene(0)	0.81 [nM]
TATmRNA(0)	0.4[nM] Experimentally derived and consistent with http://www.genecards.org/
TATprotein(0)	0.462028 [nM]
GRproteinCortTATgene(0)	0.02 [nM]
GRproteinDEXTATgene(0)	0 [nM]

Supplementary Table S5: Model Parameters for cortisol (L1) and second ligand (L2) model

Reactions		Parameters
v₁	CYPmRNA synthesis: $k_1 \cdot CYPgene(t)$ [nM/min]	* $k_1 = 0.00321 \text{ min}^{-1}$
v₂	CYPmRNA degradation: $k_2 \cdot CYPmRNA(t)$ [nM/min]	$k_2 = 0.04 \text{ min}^{-1}$ 41,42
v₃	CYPprotein synthesis: $k_3 \cdot CYPmRNA(t)$ [nM/min]	$k_3 = 2.5 \text{ min}^{-1}$ ⁴¹
v₄	CYPprotein degradation: $k_4 \cdot CYPptotein(t)$ [nM/min]	$k_4 = 0.0005 \text{ min}^{-1}$ 43,44
v₅	PXRmRNA synthesis on inactive PXRgene: $k_5 \cdot PXRgene(t)$ [nM/min]	* $k_5 = 5.52e-05 \text{ min}^{-1}$ 1
v₆	PXRmRNA degradation: $k_6 \cdot PXRmRNA(t)$ [nM/min]	** $k_6 = 0.006 \text{ min}^{-1}$ 45
v₇	PXRprotein synthesis: $k_7 \cdot PXRmRNA(t)$ [nM/min]	$k_7 = 10 \text{ min}^{-1}$ ⁴⁶
v₈	PXRprotein degradation: $k_8 \cdot PXRptotein(t)$ [nM/min]	$k_8 = 0.003 \text{ min}^{-1}$ ⁴⁷

v₉	GRmRNA synthesis on inactive GRgene: $k_9 \cdot GRgene(t)$ [nM/min]	* $k_9 = 3.2e-06 \text{ min}^{-1}$
v₁₀	GRmRNA degradation: $k_{10} \cdot GRmRNA(t)$ [nM/min]	** $k_{10} = 0.003 \text{ min}^{-1}$ 45
v₁₁	GRprotein synthesis: $k_{11} \cdot GRmRNA(t)$ [nM/min]	$k_{11} = 19.98 \text{ min}^{-1}$ 48
v₁₂	GRprotein degradation: $k_{12} \cdot GRptotein(t)$ [nM/min]	$k_{12} = 0.001 \text{ min}^{-1}$ 48,49
v₁₃	TATmRNA synthesis on inactive TATgene: $k_{13} \cdot TATgene(t)$ [nM/min]	* $k_{13} = 0.000855 \text{ min}^{-1}$
v₁₄	TATmRNA degradation: $k_{14} \cdot TATmRNA(t)$ [nM/min]	$k_{14} = 0.064 \text{ min}^{-1}$ 50
v₁₅	Cortisol binding GRprotein: $k_{15f} \cdot Cort(t) \cdot GRprotein - k_{15b} \cdot GRproteinCort(t)$ [nM/min]	$k_{15f} = 60 \text{ nM}^{-1} \text{ min}^{-1}$ (diffusion limited); $k_{15b} = 600 \text{ min}^{-1}$ ($K_d = 10 \text{ nM}$ 51)
v₁₆	GRproteinCort degradation: $k_{16} \cdot GRptoteinCort(t)$ [nmoles/min]	$k_{16} = 0.001 \text{ min}^{-1}$ 48,49
v₁₇	Cortisol binding PXRprotein: $k_{17f} \cdot Cort(t) \cdot PXRprotein - k_{17b} \cdot PXRproteinCort(t)$ [nM/min]	$k_{17f} = 60 \text{ nM}^{-1} \text{ min}^{-1}$ (diffusion limited); $k_{17b} = 600000 \text{ min}^{-1}$ 52
v₁₈	PXRproteinCort degradation: $k_{18} \cdot PXRptoteinCort(t)$ [nM/min]	$k_{18} = 0.0015 \text{ min}^{-1}$ 47
v₁₉	Cort degradation: $CYPptotein \cdot \frac{V_{mc} \cdot \frac{Cort(t)}{K_{MC}}}{1 + \frac{Cort(t)}{K_{MC}} + \frac{L2(t)}{K_{ML2}}} \text{ [nM/min]}$	$V_{mc} = 10 \text{ min}^{-1}$ ($k_{cat} = V_{mc} / 1200$ (nM max CYP protein conc in cell) nM = 0.083); $K_{MC} = 15000 \text{ nM}$ Since $L2=0$, Competition between cortisol and L2 for CYP is neglected 53,54
v₂₀	PXRpoteinCortisol binding CYPgene:	$k_{20f} = 60 \text{ nM}^{-1} \text{ min}^{-1}$

	$k_{20f} \cdot PXRproteinCort(t) \cdot CYPgene(t)$ $- k_{20b} \cdot PXRproteinCortCYPgene(t)$ [nM/min]	(diffusion limited); * $k_{20b} = 1 \text{ min}^{-1}$
v21	CYPmRNA synthesis on active PXRproteinCortCYPgene: $k_{21} \cdot PXRproteinCortCYPgene(t)$ [nM/min]	* $k_{21b} = 0.05 \text{ min}^{-1}$
v22	GRproteinCortisol binding PXRgene: $k_{22f} \cdot GRproteinCort(t) \cdot PXRgene(t)$ $- k_{22b} \cdot GRproteinCortPXRgene(t)$ [nM/min]	$k_{22f} = 60 \text{ nM}^{-1} \text{ min}^{-1}$ (diffusion limited); * $k_{22b} = 200 \text{ min}^{-1}$
v23	PXRmRNA synthesis on active GRproteinCortPXRgene: $k_{23} \cdot GRproteinCortPXRgene(t)$ [nM/min]	* $k_{23} = 0.00011 \text{ min}^{-1}$
v24	GRproteinCortisol binding GRgene: $k_{24f} \cdot GRproteinCort(t) \cdot GRgene(t)$ $- k_{24b} \cdot GRproteinCortGRgene(t)$ [nM/min]	$k_{24f} = 60 \text{ nM}^{-1} \text{ min}^{-1}$ (diffusion limited); * $k_{24b} = 60 \text{ min}^{-1}$
v25	GRmRNA synthesis on active GRproteinCortGRgene: $k_{25} \cdot GRproteinCortGRgene(t)$ [nM/min]	* $k_{25} = 1.2 \cdot 10^{-6} \text{ min}^{-1}$
v26	GRproteinCortisol binding TATgene: $k_{26f} \cdot GRproteinCort(t) \cdot TATgene(t)$ $- k_{26b} \cdot GRproteinCortTATgene(t)$ [nM/min]	$k_{26f} = 60 \text{ nM}^{-1} \text{ min}^{-1}$ (diffusion limited); * $k_{26b} = 300 \text{ min}^{-1}$
v27	TATmRNA synthesis on active GRproteinCortTATgene: $k_{27} \cdot GRproteinCortTATgene(t)$ [nM/min]	* $k_{27} = 0.005 \text{ min}^{-1}$
v28	DEX binding GRprotein: $k_{28f} \cdot DEX(t) \cdot GRprotein - k_{28b} \cdot GRproteinDEX(t)$ [nM/min]	$k_{28f} = 60 \text{ nM}^{-1} \text{ min}^{-1}$ (diffusion limited); $k_{28b} = 60 \text{ min}^{-1}$ (calculated from Kd that is in the order of magnitude of 1 nM ⁵⁵)
v29	GRproteinDEX degradation: $k_{29} \cdot GRproteinDEX(t)$ [nmoles/min]	$k_{29} = 0.001 \text{ min}^{-1}$ 48,49
v30	DEX binding PXRprotein: $k_{30f} \cdot DEX(t) \cdot PXRprotein - k_{30b} \cdot PXRproteinDEX(t)$ [nM/min]	$k_{30f} = 60 \text{ nM}^{-1} \text{ min}^{-1}$ (diffusion limited); $k_{30b} = 60000 \text{ min}^{-1}$ (calculated from Kd that is in the order of magnitude of 1000 nM ⁵⁶)

V31	PXRproteinDEX degradation: $k_{31} \cdot PXRptotein DEX (t)$ [nmoles/min]	** $k_{31} = 0.0015$ min^{-147}
V32	DEX degradation: $CYPptotein \cdot \frac{V_{mDEX} \cdot \frac{DEX(t)}{K_{MDEX}}}{1 + \frac{DEX(t)}{K_{MDEX}} + \frac{Cort(t)}{K_{MC}} + \frac{L2(t)}{K_{M:2}}}$ [nM/min]	$V_{mDEX} = 5.1 \text{ min}^{-1}$; ($k_{cat}=V_{mC} / 1200$ (max CYP protein conc in cell) nM = 0.00425); $K_{MDEX} = 23000 \text{ nM}$ 57
V33	PXRpoteinDEX binding CYPgene: $k_{33f} \cdot PXRproteinDEX (t) \cdot CYPgene (t)$ $- k_{33b} \cdot PXRprotein DEXCYPgene (t)$ [nM/min]	$k_{33f} = 60 \text{ nM}^{-1}\text{min}^{-1}$ (diffusion limited); * $k_{33b} = 1 \text{ min}^{-1}$
V34	CYPmRNA synthesis on active PXRproteinDEXCYPgene: $k_{34} \cdot PXRprotein DEXCYPgene (t)$ [nM/min]	* $k_{34} = 0.05 \text{ min}^{-1}$
V35	GRproteinDEX binding PXRgene: $k_{35f} \cdot GRproteinDEX (t) \cdot PXRgene (t)$ $- k_{35b} \cdot GRproteinD EXPXRgene (t)$ [nM/min]	$k_{35f} = 60 \text{ nM}^{-1}\text{min}^{-1}$ (diffusion limited); * $k_{35b} = 200 \text{ min}^{-1}$
V36	PXRmRNA synthesis on active GRproteinDEXPXRgene: $k_{36} \cdot GRproteinD EXPXRgene (t)$ [nM/min]	* $k_{36} = 0.00011 \text{ min}^{-1}$
V37	GRproteinDEX binding GRgene: $k_{37f} \cdot GRproteinDEX (t) \cdot GRgene (t)$ $- k_{37b} \cdot GRproteinD EXGRgene (t)$ [nM/min]	$k_{37f} = 60 \text{ nM}^{-1}\text{min}^{-1}$ (diffusion limited); * $k_{37b} = 60 \text{ min}^{-1}$
V38	GRmRNA synthesis on active GRproteinDEXGRgene: $k_{38} \cdot GRproteinD EXGRgene (t)$ [nM/min]	* $k_{38} = 1.2\text{e-}06 \text{ min}^{-1}$
V39	GRproteinDEX binding TATgene: $k_{39f} \cdot GRproteinDEX (t) \cdot TATgene (t)$ $- k_{39b} \cdot GRproteinD EXTATgene (t)$ [nM/min]	$k_{39f} = 60 \text{ nM}^{-1}\text{min}^{-1}$ (diffusion limited); * $k_{39b} = 300 \text{ min}^{-1}$
V40	TATmRNA synthesis on active GRproteinDEXTATgene: $k_{40} \cdot GRproteinD EXTATgene (t)$ [nM/min]	* $k_{40} = 0.005 \text{ min}^{-1}$
V41	DEX transport through cellular membrane: $k_{41f} \cdot DEXout(t) - k_{41b} \cdot DEXin(t)$ [nM/min]	$k_{41f} = 100 \text{ min}^{-1}$; $k_{41b} = 100 \text{ min}^{-1}$ (taken at high value not to limit DEX uptake)
V42	Endogenous cortisol synthesis:	$k_{42f} = 0.016 \text{ min}^{-1}$;

	$k_{42f} \cdot Cortisone(t) - k_{42b} \cdot Cort(t)$ [nM/min]	$k_{42b} = 0.016 \text{ min}^{-1}$ (fitted to have realistic cortisol concentration)
V₄₃	TATprotein synthesis: $k_{43} \cdot TATmRNA(t)$ [nM/min]	* $k_{43} = 0.5 \text{ min}^{-1}$
v₄₄	TATprotein degradation: $k_{44} \cdot TATptotein(t)$ [nM/min]	$k_{44} = 0.012 \text{ min}^{-1 50}$
V_{cortisolT} ransport	Transport of cortisol through cellular membrane: $k_{ctf} \cdot CortOUT(t) - k_{ctb} \cdot Cort(t)$ [nM/min]	$k_{ctf} = 1000 \text{ min}^{-1}$; $k_{ctb} = 1000 \text{ min}^{-1}$
V_{L2_PXR} _binding	Ligand2 binding PXRprotein: $k_{l2pxrf} \cdot L2(t) \cdot PXRprotein - k_{l2pxrb} \cdot PXRproteinLigand 2(t)$ [nM/min]	$k_{l2pxrf} = 60 \text{ nM}^{-1} \text{ min}^{-1}$ (diffusion limited); $k_{l2pxrb} = 600000 \text{ min}^{-1 52}$
V_{L2_PXR} _deg	PXRproteinLigand2 degradation: $k_{l2_pxr} \cdot PXRptotein Ligand 2(t)$ [nM/min]	$k_{l2_pxr} = 0.0015 \text{ min}^{-1 47}$
V_{CYPmR} NA_synt_P XR_L2	CYPmRNA synthesis on active PXRproteinLigand2CYPgene: $k_{synt_cyp_b} \cdot PXRproteinLigand 2CYPgene(t)$ [nM/min]	* $k_{synt_cyp_b} = 0.05 \text{ min}^{-1}$
V_{CYPmR} NA_PXR_ L2_binding	PXRproteinLigand2 binding CYPgene: $k_{bind_cyp_f} \cdot PXRproteinLigand 2(t) \cdot CYPgene(t)$ $- k_{bind_cyp_b} \cdot PXRproteinLigand 2CYPgene(t)$ [nM/min]	$k_{bind_cyp_f} = 60 \text{ nM}^{-1} \text{ min}^{-1}$ (diffusion limited); * $k_{bind_cyp_b} = 1 \text{ min}^{-1}$
V_{cort_distr} ibution	Cortisol through the blood $k_{distribution} \cdot CortOUT(t)$ [nM/min]	$k_{distribution} = 1000 \text{ min}^{-1}$
V_{cort_CB} G	Transport of cortisol through cellular membrane: $k_{cortCBG_1} \cdot CortOUT(t) - k_{cortCBG_2} \cdot CBG_CortOUT(t)$ [nM/min]	$k_{cortCBG_1} = 60 \text{ nM}^{-1} \text{ min}^{-1}$ (diffusion limited); $k_{cortCBG_2} = 270 \text{ min}^{-1}$
V_{cort_albu} min	$k_{cortALB_1} \cdot CortOUT(t) - k_{cortALB_2} \cdot Alb_CortOUT(t)$	$k_{cortALB_1} = 60 \text{ nM}^{-1} \text{ min}^{-1}$ (diffusion limited);

		$k_{cortALB_2} = 900000$ min^{-1}
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Supplementary Table S6: Balance Equations for cortisol (L1) and second ligand (L2) model

Balance equations	
dCort(t)/dt	$-V_{15}+V_{16} -V_{17}+V_{18}-V_{19}+V_{42}+ V_{cortisolTransport}$ [nM/min]
dCortAdded(t)/dt	$- V_{cort_distribution}$ [nM/min]
dCortOUT(t)/dt	$V_{cort_distribution} -V_{cort_albumin} - V_{cort_CBG} - V_{cortisolTransport}$ [nM/min]
dCBG_CortOUT(t)/dt	V_{cort_CBG} [nM/min]
dAlb_CortOUT(t)/dt	$V_{cort_albumin}$ [nM/min]
dCBG(t)/dt	$-V_{cort_CBG}$ [nM/min]
dAlb(t)/dt	$-V_{cort_albumin}$ [nM/min]
dCortisone(t)/dt	0 (considered an infinite reservoir)
dCYPgene(t)/dt	$-V_{20}-V_{33}$ [nM/min]
dCYPprotein(t)/dt	$+V_3-V_4$ [nM/min]
dCYPmRNA(t) /dt	$+V_1-V_2 +V_{21}+V_{34}+V_{CYPmRNA_PXR_L2_binding}$ [nM/min]
dPXRproteinCortCYPgene(t) /dt	$+V_{20}$ [nM/min]
dPXRproteinLigand2CYPgene(t) /dt	$+ V_{CYPmRNA_PXR_L2_binding}- V_{CYPmRNA_synt_PXR_L2}$ [nM/min]
dPXRproteinDEXCYPgene(t) /dt	$+V_{33}$ [nM/min]
dDEX(t)/dt	$-V_{28}+V_{29} -V_{30}+V_{31}-V_{32}+V_{41}$ [nM/min]
dLigand2(t)/dt	$- V_{L2_PXR_binding}+ V_{L2_PXR_deg}$ [nM/min]
dDEXdeg(t)/dt	$+V_{32}$ [nM/min]
dDEXout(t)/dt	$-V_{41}$ [nM/min]
dGRgene(t)/dt	$-V_{24} -V_{37}$ [nM/min]
dGRprotein(t)/dt	$+V_{11}-V_{12}-V_{15}-V_{12}-V_{28}$ [nM/min]
dGRmRNA(t)/dt	$+V_9-V_{10}+V_{25}+V_{38}$ [nM/min]
dGRproteinCortGRgene(t)/dt	$+V_{24}$ [nM/min]
dGRproteinDEXGRgene(t)/dt	$+V_{37}$ [nM/min]
dGRproteinCort(t)/dt	$+V_{15}-V_{16}-V_{22}-V_{24}-V_{26}$ [nM/min]
dGRproteinDEX(t)/dt	$+V_{28}-V_{29}-V_{35}-V_{37}-V_{39}$ [nM/min]
dPXRgene(t)/dt	$-V_{35} -V_{22}$ [nM/min]
dPXRprotein(t)/dt	$+V_7-V_8-V_{17}-V_{30}$ [nM/min]
dPXRmRNA(t)/dt	$+V_5-V_6+V_{23}+V_{36}$ [nM/min]
dGRproteinCortPXRgene(t)/dt	$+V_{22}$ [nM/min]
dGRproteinDEXPXRgene(t)/dt	$+V_{35}$ [nM/min]
dPXRproteinCort(t)/dt	$+V_{17}-V_{18}-V_{20}$ [nM/min]
dPXRproteinLigand2(t)/dt	$+V_{L2_PXR_binding}-V_{L2_PXR_deg}-V_{CYPmRNA_PXR_L2_binding}$ [nM/min]

dPXRproteinDEX(t)/dt	+V ₃₀ -V ₃₁ -V ₃₃ [nM/min]
dTATgene(t)/dt	-V ₂₆ -V ₃₉ [nM/min]
dTATmRNA(t)/dt	+V ₁₃ -V ₁₄ +V ₂₇ +V ₄₀ [nM/min]
dGRproteinCortTATgene(t)/dt	+V ₂₆ [nM/min]
dGRproteinDEXTATgene(t)/dt	+V ₃₉ [nM/min]
dTATprotein(t)/dt	+V ₄₃ -V ₄₄ [nM/min]

Supplementary Table S7: Conserved Moieties for cortisol (L1) and second ligand (L2) model

Conserved Moieties		
GRgene _{total}	GRgene(t) + GRproteinCortGRgene(t) + GRproteinDEXGRgene(t)	****0.83 [nM]
PXRgene _{total}	PXRgene(t) + GRproteinCortPXRgene(t) + GRproteinDEXPXRgene(t)	****0.83 [nM]
CYPgene _{total}	CYPgene(t) + PXRproteinCortCYPgene(t) + PXRproteinLigand2CYPgene(t)+ PXRproteinDEXCYPgene(t)	****0.83 [nM]
DEX _{total}	DEX(t) + DEXdeg(t) + DEXout(t) + GRproteinDEXGRgene(t) + GRproteinDEX(t) + GRproteinDEXPXRgene(t) + PXRproteinDEX(t) + GRproteinDEXTATgene(t) + PXRproteinDEXCYPgene(t)	1000 [nM]

Supplementary Table S8: Initial Conditions for cortisol (L1) and second ligand (L2) model

Initial conditions	
Cort(0)	1.14 [nM]
Cortisone(0)	23 [nM]
CortOUT(0)	0[nM]
CortAdded(0)	0[nM] at t=0 for the initial state, then ~130 [nM] added into the blood as an exposure of cortisol.
CBG_CortOUT(0)	0[nM]
Alb_CortOUT(0)	0[nM]
CBG(0)	550[nM]
Alb(0)	60000[nM]
Ligand2(0)	100[nM]
CYPgene(0)	0.82976 [nM]
CYPprotein(0)	104 [nM]
CYPmRNA(0)	75e-3 [nM] 94x levels of GR mRNA. Experimentally derived and

	consistent with ^{58,59}
PXRproteinCortCYPgene(0)	0.00024 [nM]
PXRproteinLigand2CYPgene(0)	0 [nM]
PXRproteinDEXCYPgene(0)	0 [nM]
DEX(0)	0 [nM]
DEXdeg(0)	0 [nM]
DEXout(0)	1000 [nM]
GRgene(0)	0.5 [nM]
GRprotein(0)	GRtotal=80.0 nM =GR (47.24) +GRprotCort(32.36) + GRprotCortGRgene (0.33) + GRproteinCortTATgene(0.02) + GRproteinCortPXRgene(0.05) _{60,61}
GRmRNA/dt	0.0008 [nM] Experimentally derived and consistent with ^{58,62}
GRproteinCortGRgene(0)	0.33 [nM]
GRproteinDEXGRgene(0)	0 [nM]
GRproteinCort(0)	32.36 [nM]
GRproteinDEX(0)	0 [nM]
PXRgene(0)	0.78 [nM]
PXRprotein(0)	****99.9 [nM]
PXRmRNA(0)	0.007 [nM] 9x level of GR mRNA. Experimentally derived and consistent with ^{58,59}
GRproteinCortPXRgene(0)	0.05 [nM]
GRproteinDEXPXRgene(0)	0 [nM]
PXRproteinCort(0)	0.01 [nM]
PXRproteinLigand2(0)	0 [nM]
PXRproteinDEX(0)	0 [nM]
TATgene(0)	0.81 [nM]
TATmRNA(0)	0.4[nM] Experimentally derived and consistent with http://www.genecards.org/
TATprotein(0)	0.462028 [nM]
GRproteinCortTATgene(0)	0.02 [nM]
GRproteinDEXTATgene(0)	0 [nM]

*mRNA and protein synthesis rates are fitted according to the experimental data obtained.

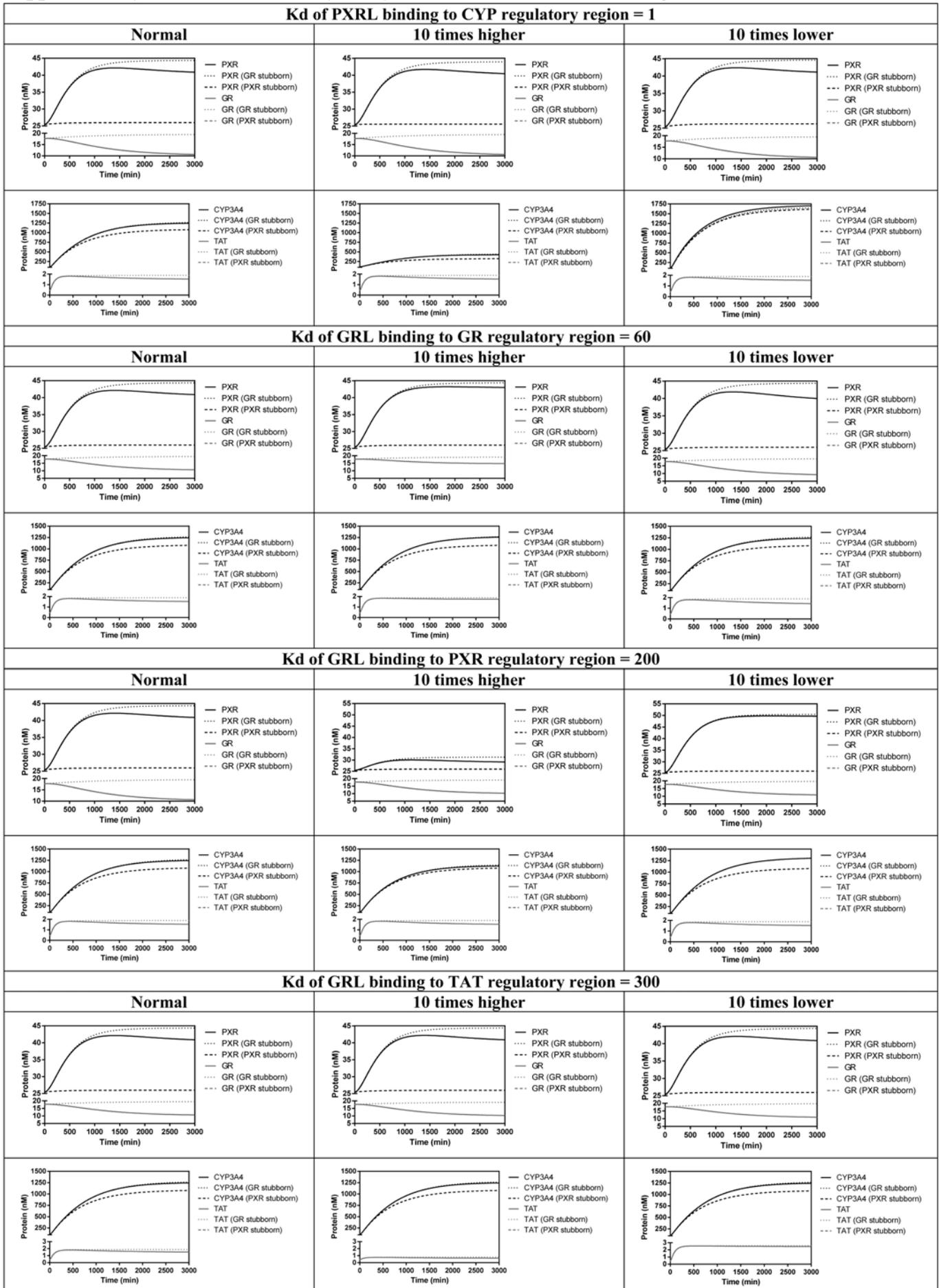
**mRNA and protein degradation are fitted to concur with ⁶³

***Kd of binding of liganded NRs to REs is fitted in the realistic order of magnitude ⁶⁴

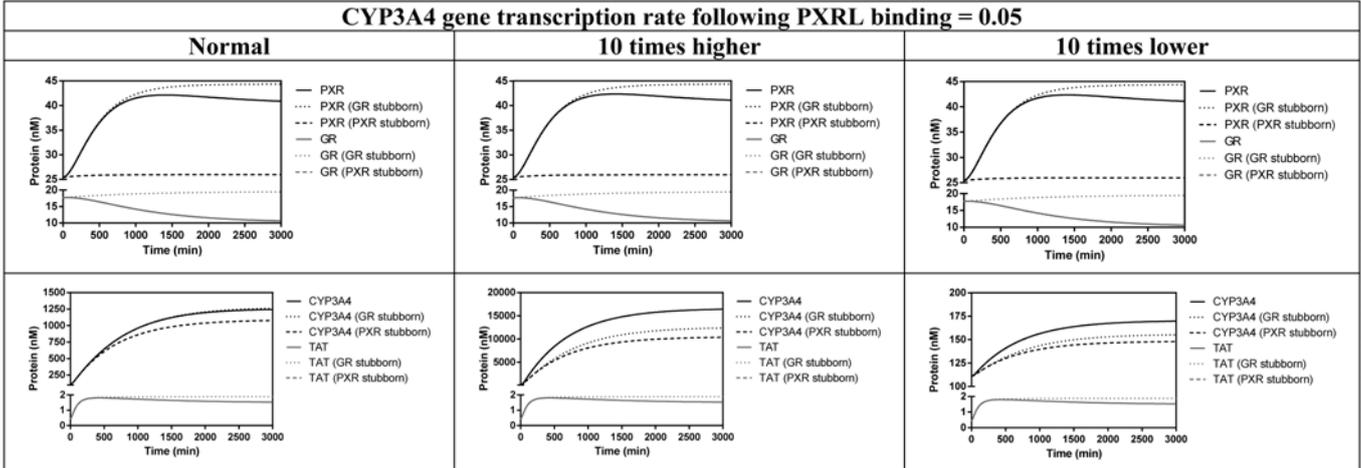
**** Taken in the realistic order of magnitude that is calculated from ($\sim 1.7 \times 10^{-12}$ nmoles of REs (1,000 molecules) per cell (de Kloet et al, 2000), consequently per volume of nucleus. Volume of nucleus is $\sim 0.45 \times 10^{-12}$ L ⁶¹)

*****Taken in the realistic order of magnitude that is calculated from (NR total=NR + NRDEX +NRCortisol is 1.7×10^{-9} nmoles (100,000 molecules) per cell ^{60,65}; volume of cell is $\sim 2 \times 10^{-12}$ L ⁶¹)

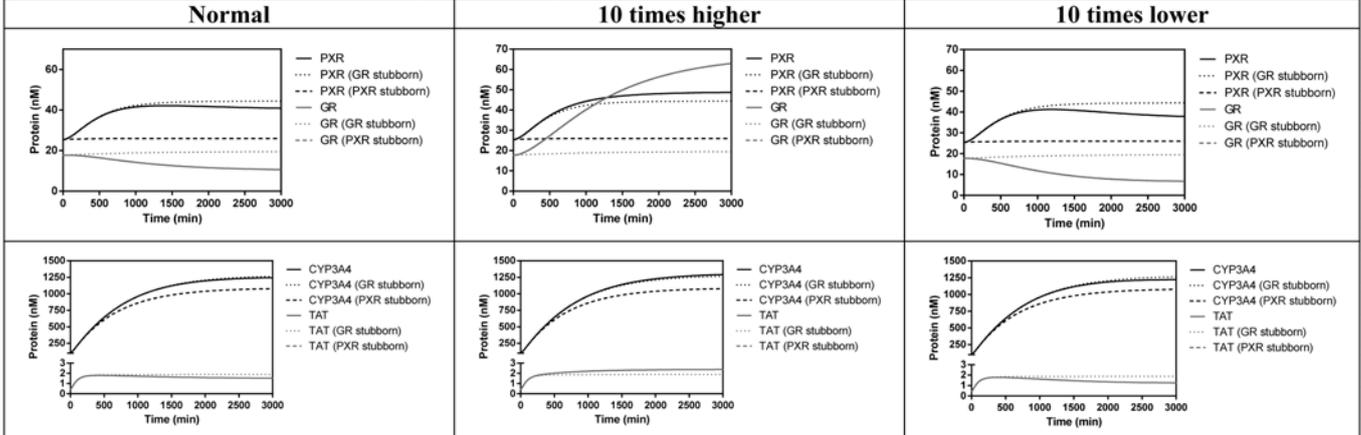
Supplementary Table S9: Sensitivity analysis of main conclusions for Figure 2



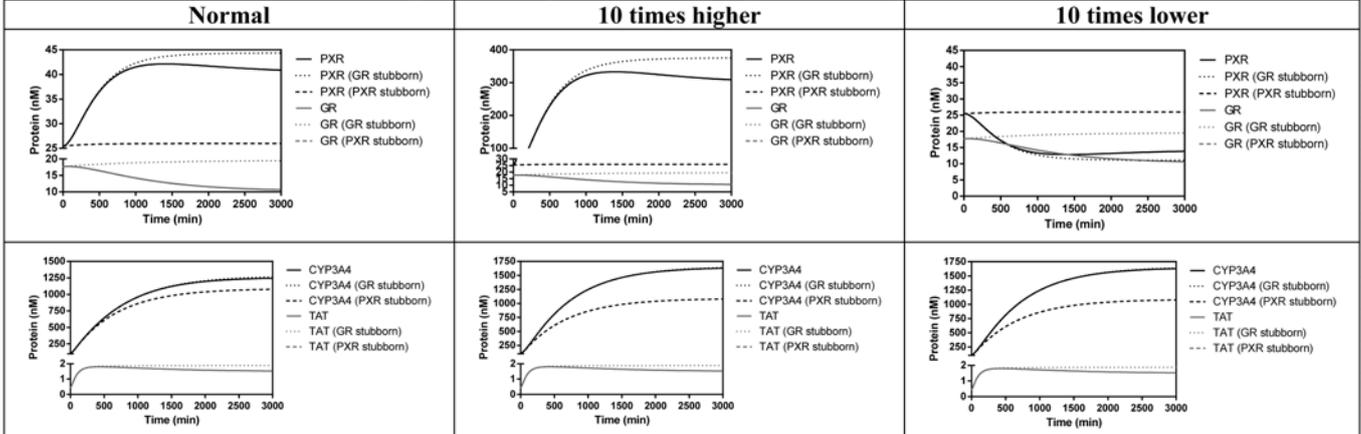
CYP3A4 gene transcription rate following PXRL binding = 0.05



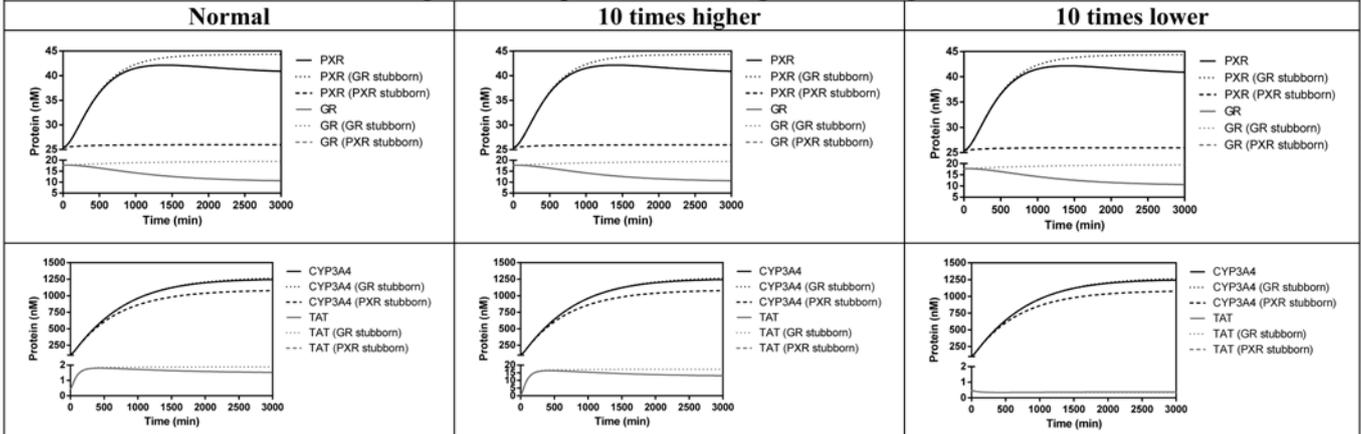
GR gene transcription rate following GRL binding = 1.2e-6



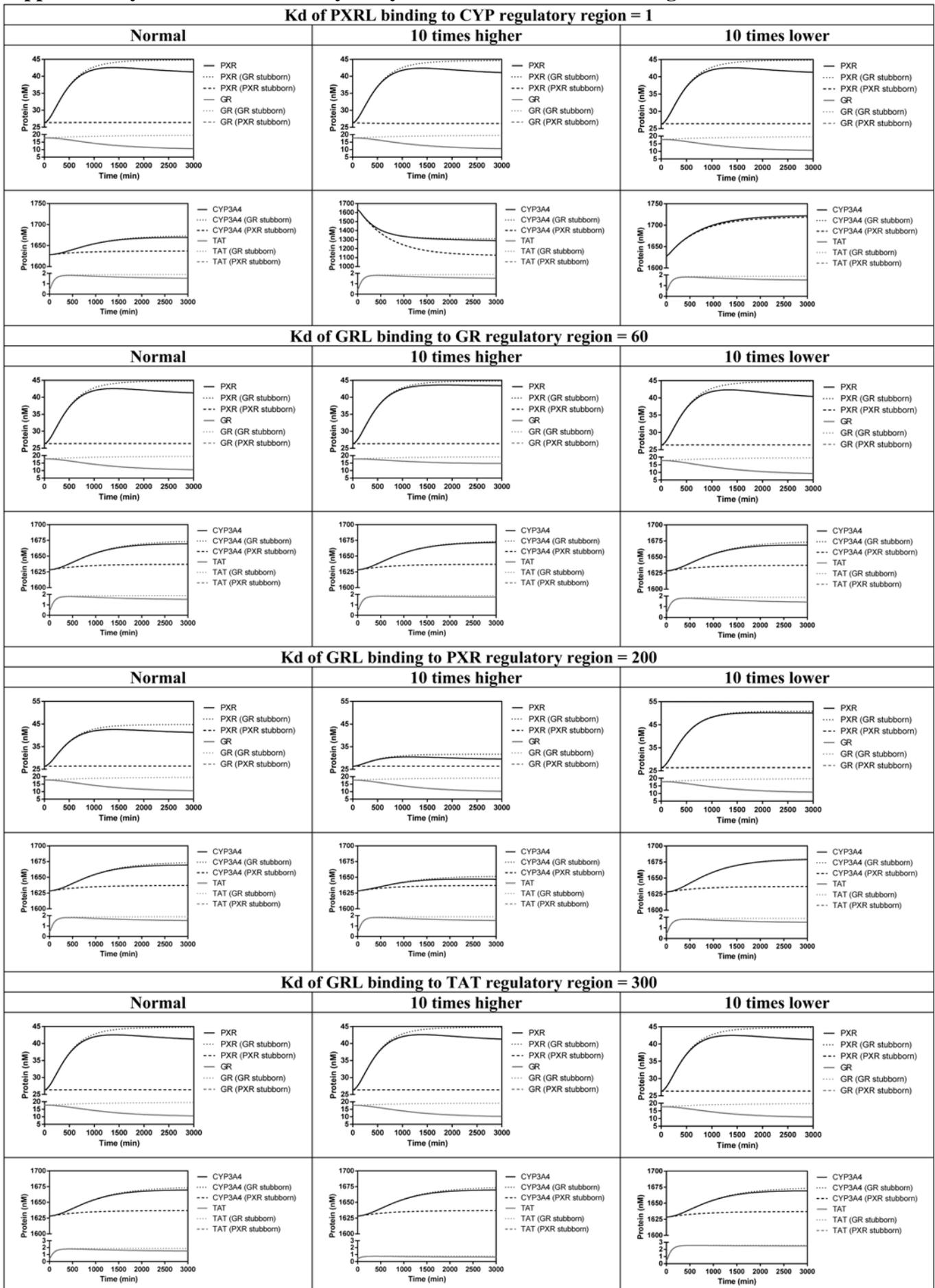
PXR gene transcription rate following GRL binding = 0.00011



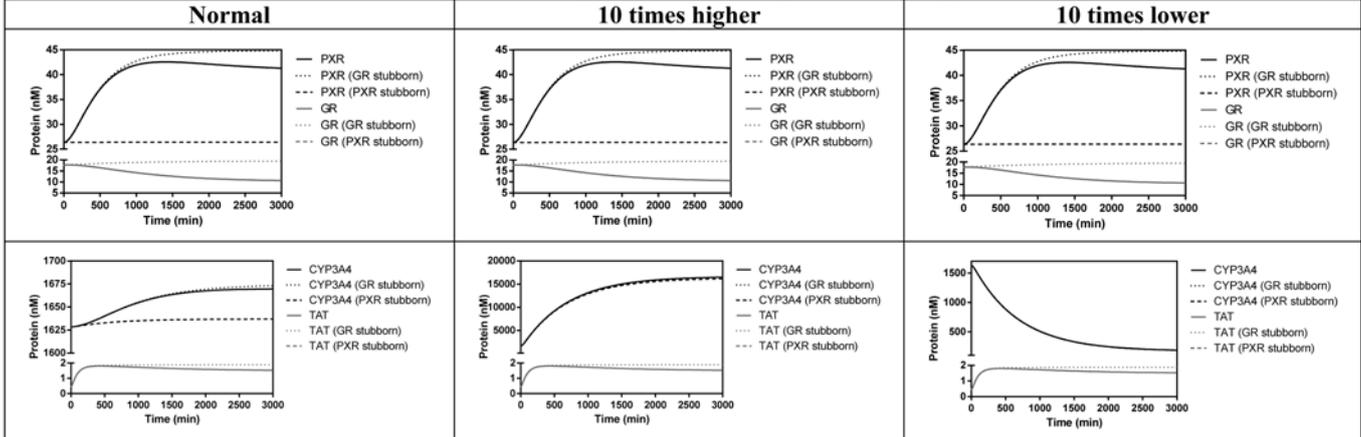
TAT gene transcription rate following GRL binding = 0.005



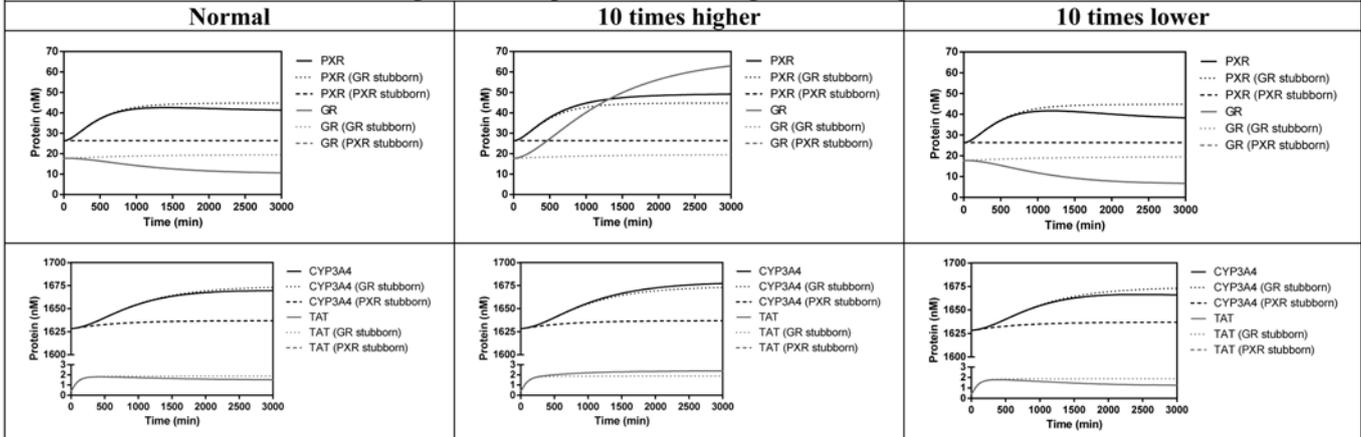
Supplementary Table S10: Sensitivity analysis of main conclusions for Figure 3



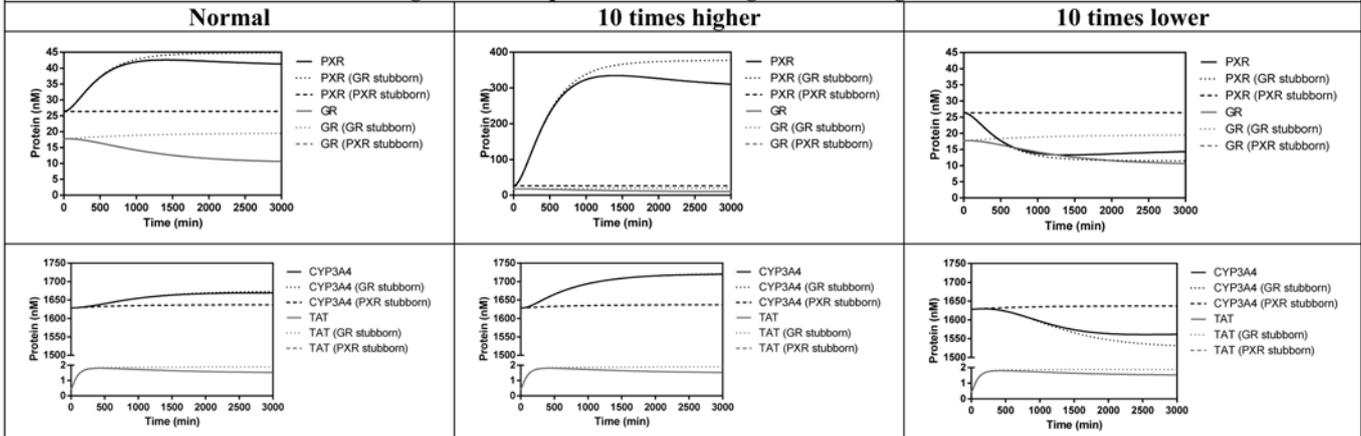
CYP3A4 gene transcription rate following PXRL binding = 0.05



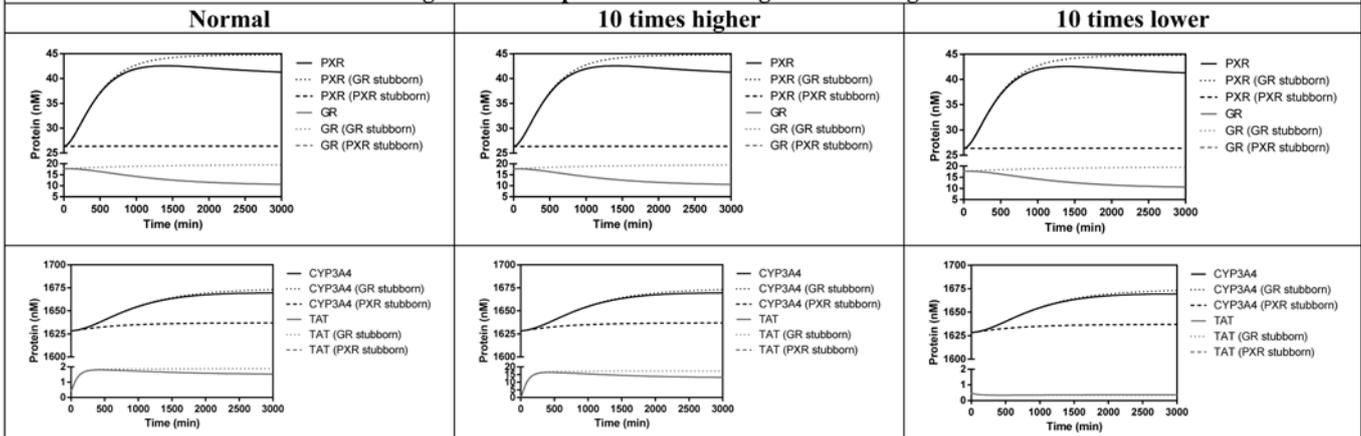
GR gene transcription rate following GRL binding = 1.2e-6



PXR gene transcription rate following GRL binding = 0.00011



TAT gene transcription rate following GRL binding = 0.005



Supplementary References

- 41 Luke, N. S., DeVito, M. J., Shah, I. & El-Masri, H. A. Development of a quantitative model of pregnane X receptor (PXR) mediated xenobiotic metabolizing enzyme induction. *Bull Math Biol* **72**, 1799-1819 (2010).
- 42 Hargrove, J. L. Microcomputer-assisted kinetic modeling of mammalian gene expression. *FASEB J* **7**, 1163-1170 (1993).
- 43 Wang, Y. *et al.* A role for protein phosphorylation in cytochrome P450 3A4 ubiquitin-dependent proteasomal degradation. *J Biol Chem* **284**, 5671-5684 (2009).
- 44 Watkins, P. B., Wrighton, S. A., Schuetz, E. G., Maurel, P. & Guzelian, P. S. Macrolide antibiotics inhibit the degradation of the glucocorticoid-responsive cytochrome P-450p in rat hepatocytes in vivo and in primary monolayer culture. *J Biol Chem* **261**, 6264-6271 (1986).
- 45 Legewie, S., Herzel, H., Westerhoff, H. V. & Bluthgen, N. Recurrent design patterns in the feedback regulation of the mammalian signalling network. *Mol Syst Biol* **4**, 190 (2008).
- 46 Bailey, I., Gibson, G. G., Plant, K., Graham, M. & Plant, N. A PXR-mediated negative feedback loop attenuates the expression of CYP3A in response to the PXR agonist pregnenolone-16alpha-carbonitrile. *PLoS One* **6**, e16703 (2011).
- 47 Masuyama, H., Inoshita, H., Hiramatsu, Y. & Kudo, T. Ligands have various potential effects on the degradation of pregnane X receptor by proteasome. *Endocrinology* **143**, 55-61 (2002).
- 48 Jin, J. Y., DuBois, D. C., Almon, R. R. & Jusko, W. J. Receptor/gene-mediated pharmacodynamic effects of methylprednisolone on phosphoenolpyruvate carboxykinase regulation in rat liver. *J Pharmacol Exp Ther* **309**, 328-339 (2004).
- 49 Hoeck, W., Rusconi, S. & Groner, B. Down-regulation and phosphorylation of glucocorticoid receptors in cultured cells. Investigations with a monospecific antiserum against a bacterially expressed receptor fragment. *J Biol Chem* **264**, 14396-14402 (1989).
- 50 Ramakrishnan, R., DuBois, D. C., Almon, R. R., Pyszczynski, N. A. & Jusko, W. J. Fifth-generation model for corticosteroid pharmacodynamics: application to steady-state receptor down-regulation and enzyme induction patterns during seven-day continuous infusion of methylprednisolone in rats. *J Pharmacokinetic Pharmacodyn* **29**, 1-24 (2002).
- 51 Huizenga, N. *et al.* Decreased ligand affinity rather than glucocorticoid receptor down-regulation in patients with endogenous Cushing's syndrome. *Eur. J. Endocrinol.* **142**, 472-476 (2000).
- 52 El-Sankary, W., Bombail, V., Gibson, G. G. & Plant, N. Glucocorticoid-Mediated Induction of CYP3A4 is Decreased by Disruption of a Protein: DNA Interaction Distinct from the Pregnane X Receptor Response Element. *Drug Metab. Dispos.* **30**, 1029-1034. (2002).
- 53 Ged, C. *et al.* The increase in urinary excretion of 6 beta-hydroxycortisol as a marker of human hepatic cytochrome P450III_A induction. *Br J Clin Pharmacol* **28**, 373-387 (1989).
- 54 Bu, H. Z. A literature review of enzyme kinetic parameters for CYP3A4-mediated metabolic reactions of 113 drugs in human liver microsomes: structure-kinetics relationship assessment. *Curr Drug Metab* **7**, 231-249 (2006).
- 55 Marissal-Arvy, N., Mormede, P. & Sarrieau, A. Strain differences in corticosteroid receptor efficiencies and regulation in Brown Norway and Fischer 344 rats. *J Neuroendocrinol* **11**, 267-273 (1999).
- 56 El-Sankary, W., Gibson, G. G., Ayrton, A. & Plant, N. Use of a reporter gene assay to predict and rank the potency and efficacy of CYP3A4 inducers. *Drug Metab Dispos* **29**, 1499-1504 (2001).
- 57 Emoto, C. *et al.* Non-invasive method to detect induction of CYP3A4 in chimeric mice with a humanized liver. *Xenobiotica* **38**, 239-248 (2008).
- 58 Phillips, A., Hood, S. R., Gibson, G. G. & Plant, N. J. Impact of transcription factor profile and chromatin conformation on human hepatocyte CYP3A gene expression. *Drug Metabolism and Disposition* **33**, 233-242 (2005).
- 59 Noreault, T. L. *et al.* Arsenite decreases CYP3A4 and RXRalpha in primary human hepatocytes. *Drug Metab Dispos* **33**, 993-1003 (2005).

- 60 van Steensel, B. *et al.* Localization of the glucocorticoid receptor in discrete clusters in the cell
nucleus. *J Cell Sci* **108 (Pt 9)**, 3003-3011 (1995).
- 61 Riddick, G. & Macara, I. G. The adapter importin-alpha provides flexible control of nuclear
import at the expense of efficiency. *Mol. Sys. Biol.* **3** (2007).
- 62 Pujols, L. *et al.* Expression of glucocorticoid receptor alpha- and beta-isoforms in human cells
and tissues. *Am J Physiol Cell Physiol* **283**, C1324-1331 (2002).
- 63 Legewie, S., Herzel, H., Westerhoff, H. V. & Bluthgen, N. Recurrent design patterns in the
feedback regulation of the mammalian signalling network. *Mol. Sys. Biol.* **4** (2008).
- 64 Rundlett, K. L. & Armstrong, D. W. Evaluation of free D-glutamate in processed foods.
Chirality **6**, 277-282 (1994).
- 65 Nordeen, S. K., Kuhnel, B., Lawlerheavner, J., Barber, D. A. & Edwards, D. P. A quantitative
comparison of dual control of a hormone response element by progestins and glucocorticoids in
the same cell-line. *Mol. Endocrinol.* **3**, 1270-1278 (1989).