

Supplementary Material

Domain Dissection and Characterization of the Aminoglycoside Resistance Enzyme ANT(3'')-Ii/AAC(6')-IId from *Serratia marcescens*

Keith D. Green^b and Sylvie Garneau-Tsodikova^{a,b,*}

^aDepartment of Medicinal Chemistry, ^bLife Sciences Institute, 210 Washtenaw Ave, University of Michigan, Ann Arbor, MI 48109-2216, U.S.A. (*Corresponding author E-mail: sylviegt@umich.edu; Phone: 734-615-2736)

Table S1: Primers used for the PCR amplification of various constructs (full-length and truncated) of *ant(3'')-Ii/aac(6')-IId*

gene	5' primer	3' primer
<i>ant(3'')-Ii/aac(6')-IId (NHis)</i>	TTAGACC <u>ATATG</u> AGTAACGCAG	TCGATGCTCGAGTTAGGCATCCTGCG
<i>ant(3'')-Ii/aac(6')-IId (CHis)</i>	TTAGACC <u>ATATG</u> AGTAACGCAG	ATGGAA <u>CTCGAGGGC</u> ATCACTGCG
<i>aac(6')-IId(261-463) (NHis)</i>	GCCACTC <u>CATATG</u> CTTGGTGCCCATGCC	TCGATGCTCGAGTTAGGCATCCTGCG
<i>aac(6')-IId(261-463) (CHis)</i>	GCCACTC <u>CATATG</u> CTTGGTGCCCATGCC	TCGATGCTCGAGTTAGGCATCCTGCG
<i>aac(6')-IId(264-463) (NHis)</i>	CTTGGTC <u>CATATG</u> CCAGTGATGTCTAAAAC	TCGATGCTCGAGTTAGGCATCCTGCG
<i>aac(6')-IId(264-463) (CHis)</i>	CTTGGTC <u>CATATG</u> CCAGTGATGTCTAAAAC	TCGATGCTCGAGTTAGGCATCCTGCG
<i>aac(6')-IId(267-463) (NHis)</i>	ATGCCAC <u>ATATG</u> TCTAAAACAAAGTTAG	TCGATGCTCGAGTTAGGCATCCTGCG
<i>aac(6')-IId(267-463) (CHis)</i>	ATGCCAC <u>ATATG</u> TCTAAAACAAAGTTAG	TCGATGCTCGAGTTAGGCATCCTGCG
<i>ant(3'')-Ii(1-260) (CHis)</i>	TTAGACC <u>ATATG</u> AGTAACGCAG	CATGGCCTCGAGCAATTTAGTGGCTTC
<i>ant(3'')-Ii(1-260) (NHis)</i>	TTAGACC <u>ATATG</u> AGTAACGCAG	TGGCATCTCGAGTCACAATTTAGTGGCTTC
<i>ant(3'')-Ii(1-263) (CHis)</i>	TTAGACC <u>ATATG</u> AGTAACGCAG	CATCACCTCGAGGGCACCAAGCAATTTAG
<i>ant(3'')-Ii(1-263) (NHis)</i>	TTAGACC <u>ATATG</u> AGTAACGCAG	AGACATCTCGAGTCAGGCACCAAGCAATTTAG
<i>ant(3'')-Ii(1-266) (CHis)</i>	TTAGACC <u>ATATG</u> AGTAACGCAG	TGTCACCTCGAGCACTGGCATGGCACCAAG
<i>ant(3'')-Ii(1-266) (NHis)</i>	TTAGACC <u>ATATG</u> AGTAACGCAG	CTTTGTCTCGAGTCACACTGGCATGGCACCAAG

The introduced restriction sites are underlined for each primer. All of the 5' primers introduced an *NdeI* restriction site. All of the 3' primers introduced an *XhoI* restriction site.

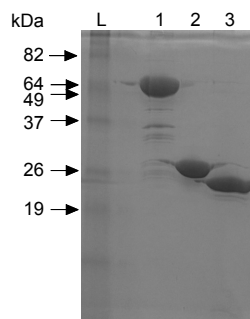


Fig. S1. Coomassie blue-stained 15% Tris-HCl SDS-PAGE gel showing the purified 55.0-kDa NHis₁₀-tagged ANT(3'')-Ii/AAC(6')-IId (Lane 1), 30.3-kDa CHis₆-tagged ANT(3'')-Ii(1-266) (Lane 2), and 24.0-kDa NHis₆-tagged AAC(6')-IId(267-463) (Lane 3). L = BenchMark™ Pre-Stained Ladder from Invitrogen. 6 µg of each protein was loaded on the gel.

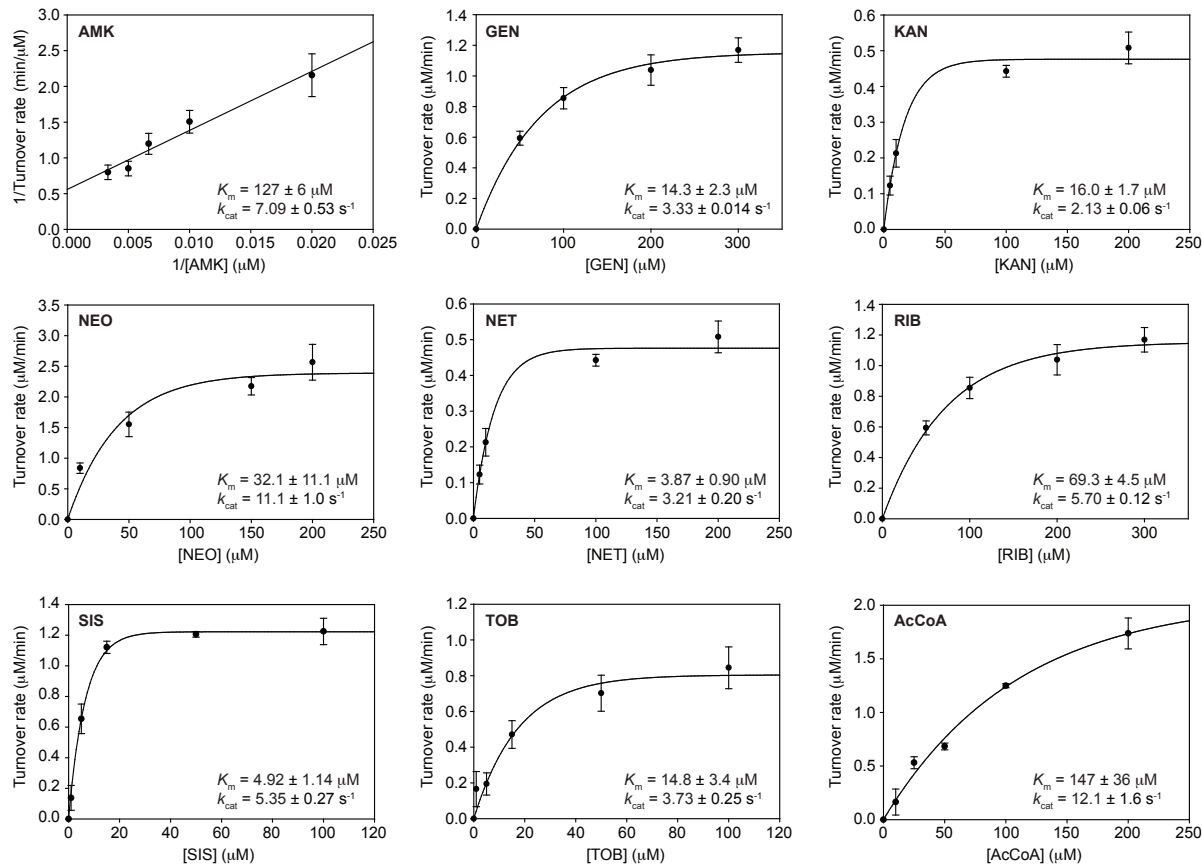


Fig. S2. Michaelis-Menten and Lineweaver-Burk kinetics for a variety of AGs and AcCoA during acetylation by AAC(6')-IId(267-463).

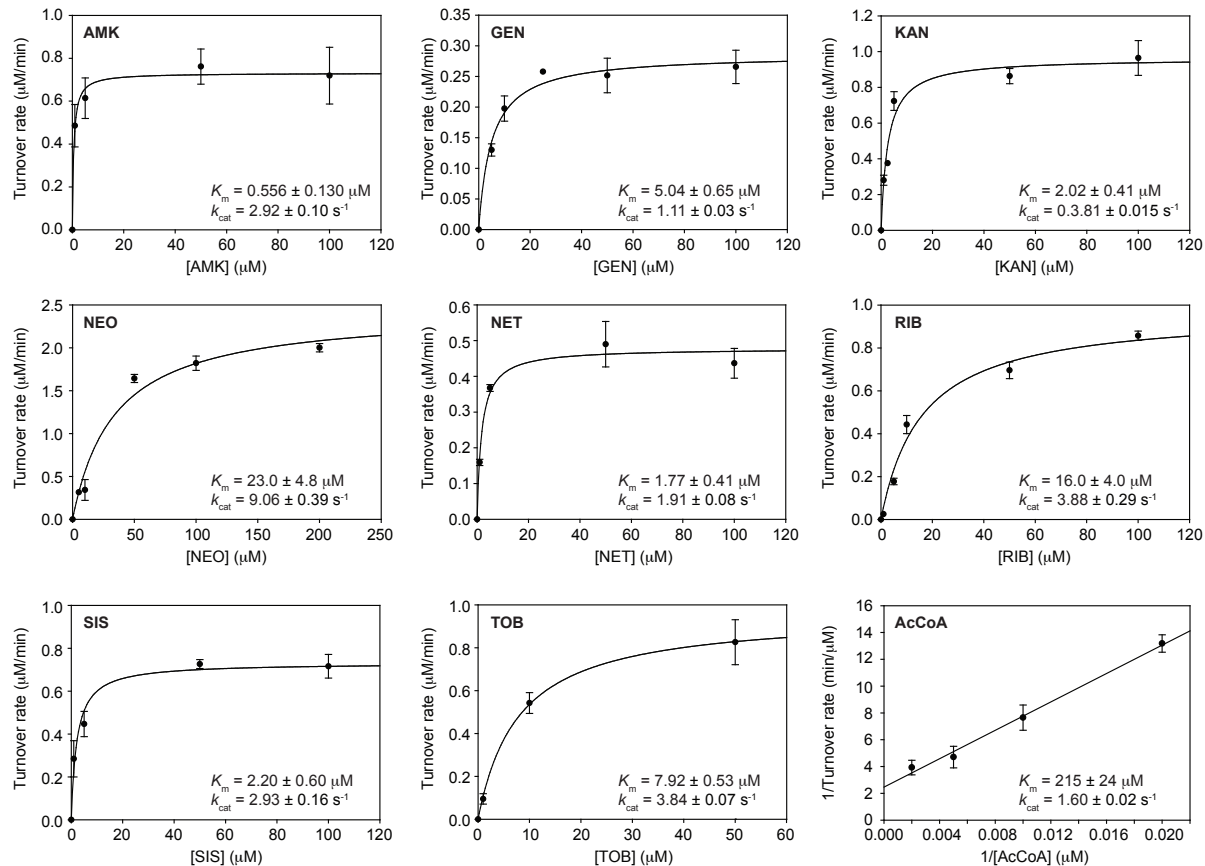


Fig. S3. Michaelis-Menten and Lineweaver-Burk kinetics for a variety of AGs and AcCoA during acetylation by ANT(3'')-Ii/AAC(6')-IId.

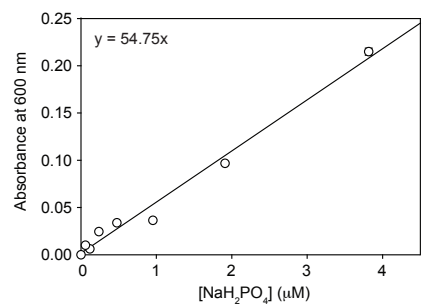


Fig. S4. Standard curve for P_i concentrations.

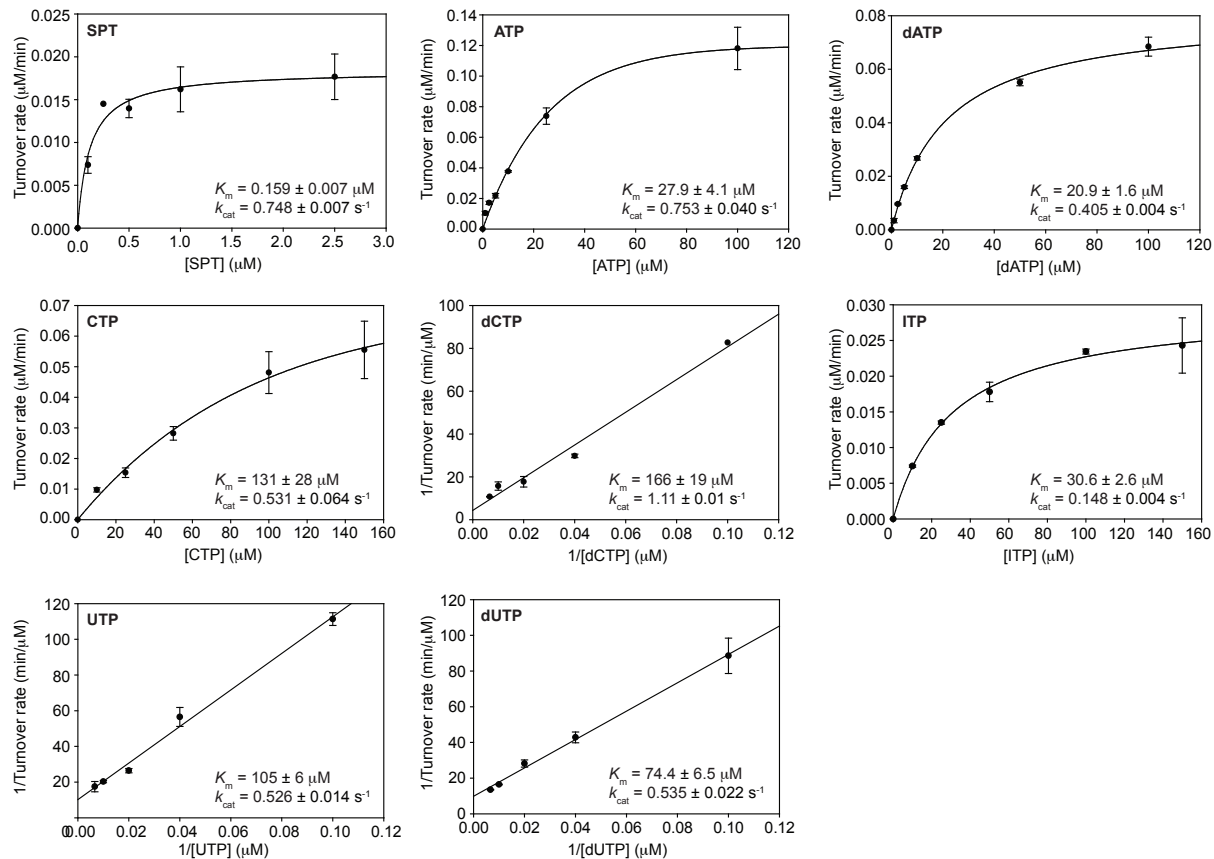


Fig. S5. Michaelis-Menten and Lineweaver-Burk kinetics for SPT as well as for a variety of NTPs and dNTPs during nucleotidylation by ANT(3'')-Ii/AAC(6')-IId.

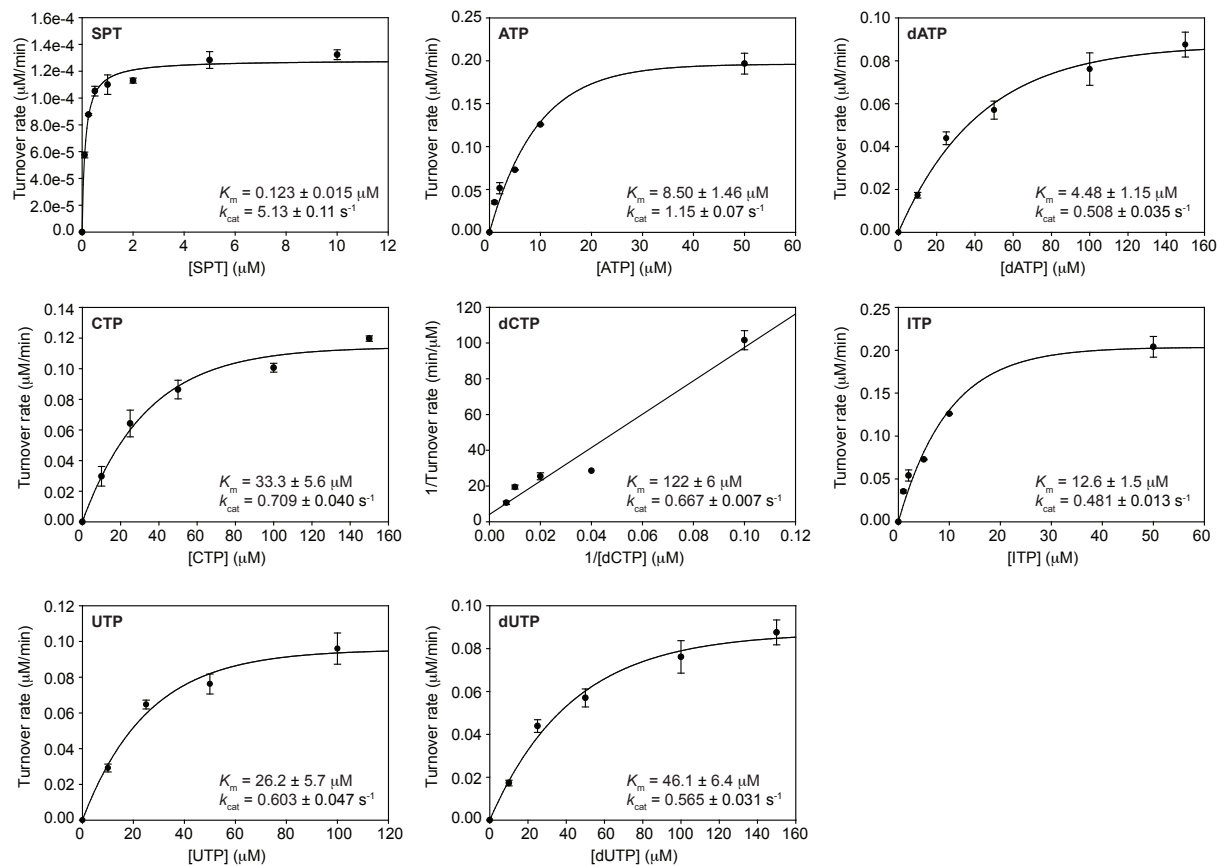


Fig. S6. Michaelis-Menten and Lineweaver-Burk kinetics for SPT as well as for a variety of NTPs and dNTPs during nucleotidylation by ANT(3'')-Ii(1-266).

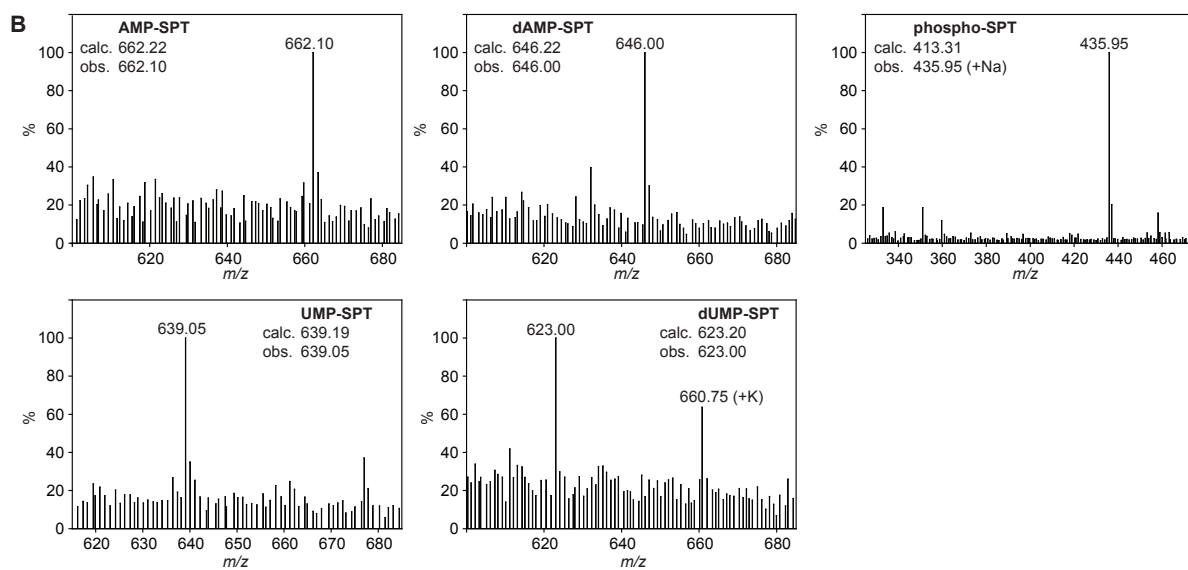
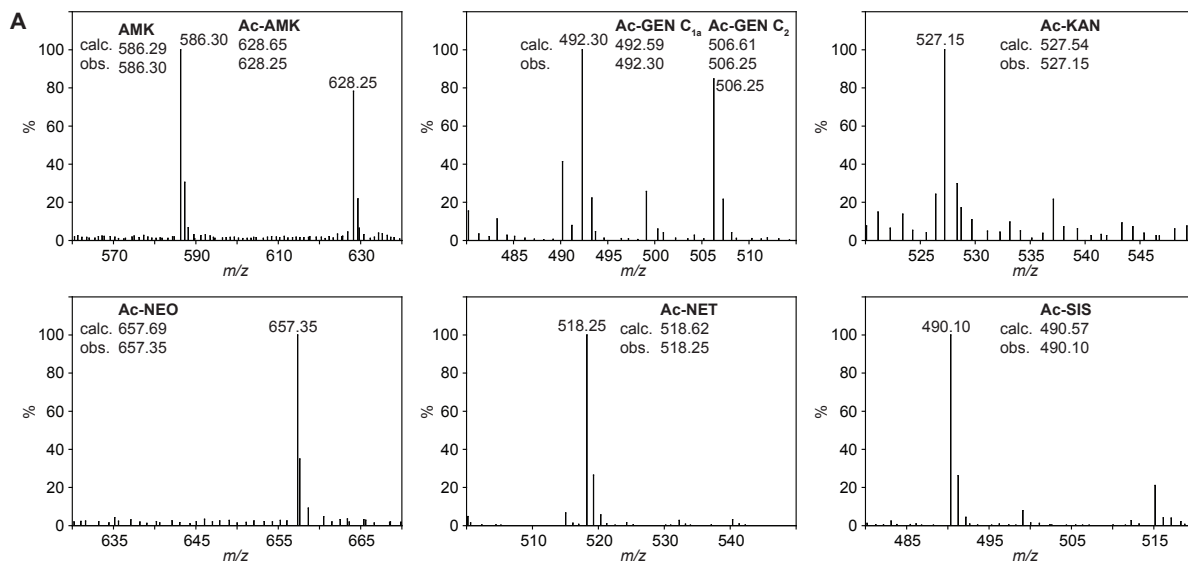


Fig. S7. Representative mass spectra for **A.** the acylation of various AGs, and **B.** nucleotidylation of SPT with various NTPs, dNTPs, and TP by ANT(3'')-Ii/AAC(6')-IId.