

Supplementary Table 2. Signature DNA pair sequences used for resistance gene search and plasmid replicon typing.

Gene Description	Primer labels	Sequence (5'-3')	Function (Ambler Class)	Reference
Beta-lactamases				
<i>bla</i> CTX-M-1 group	CTX-M Gp1.F CTX-M Gp1.R	GGAATCTGACGCTGGGTAAA GGTTGAGGCTGGGTGAAGTA	ESBL Ambler A	(3)
<i>bla</i> CTX-M-9 group	CTX-M Gp9.F CTX-M Gp9.R	GGTGATGAACGCTTTCCAAT TCAATTTGTTTCATGGCGGTA	ESBL Ambler A	(3)
<i>bla</i> CTX-M group	CTX-M-U1 CTX-M-U2	ATGTGCAGYACCAGTAARGTKATGGC TGGGTRAARTARGTSACCAGAAYCAGCGG	ESBL Ambler A	(1)
<i>bla</i> CTX-M group 1 including CTX-M-1, CTX-M-3 and CTX-M-15	MultiCTXMGp1 F MultiCTXMGp1-2 R	TTAGGAARTGTGCCGCTGYA CGATATCGTTGGTGTRCCAT	ESBL Ambler A	(2)
<i>bla</i> CTX-M group 2 including CTX-M-2	MultiCTXMGp2 F MultiCTXMGp1-2 R	CGTTAACGGCACGATGAC CGATATCGTTGGTGTRCCAT	ESBL Ambler A	(2)
<i>bla</i> CTX-M group 9 including CTX-M-9 and CTX-M-14	MultiCTXMGp9 F MultiCTXMGp9 R	TCAAGCCTGCCGATCTGGT TGATTCTCGCCGCTGAAG	ESBL Ambler A	(2)
<i>bla</i> CTX-M group 8/25 (CTX-M-8, CTX-M-25, CTX-M-26 and CTX-M-39 to CTX-M-41)	CTX-Mg8/25 F CTX-Mg8/25 R	AACRCRCAGACGCTCTAC TCGAGCCGGAASGTGYAT	ESBL Ambler A	(2)
<i>bla</i> TEM	TEM-164.SE TEM-165.AS	TCGCCGCATACACTATTCTCAGAATGA ACGCTCACCGGCTCCAGATTTAT	ESBL Ambler A	(10)
<i>bla</i> TEM variants including TEM-1 and TEM-2	MultiTSO-T F MultiTSO-T R	CATTTCGGTGTCCGCTTATTC CGTTCATCCATAGTTGCTGAC	ESBL Ambler A	(2)
<i>bla</i> SHV	<i>bla</i> SHV.SE <i>bla</i> SHV.AS	ATGCGTTATATTCGCTGTG TGCTTTGTTATTCGGGCCAA	ESBL Ambler A	(11)
<i>bla</i> SHV-5/12	SHV-5/12.F SHV-5/12.R	AGCTGCTGCAGTGGATGGT CAATGCGCTCTGCTTTGTTA	ESBL Ambler A	(3)
<i>bla</i> SHV variants including SHV-1	MultiTSO-S F MultiTSO-S R	AGCCGCTTGAGCAAATTAAC ATCCCCGAGATAAATCACCAC	ESBL Ambler A	(2)
<i>bla</i> VEB1	VEB.F VEB.R	CAAATGCACAAGGATTGGAA ATTCCGGAAGTCCCTGTTTT	ESBL Ambler A	(3)
<i>bla</i> VEB-1 to VEB-6	MultiVEB F MultiVEB R	CATTTCCCGATGCAAAGCGT CGAAGTTCTTTGGACTCTG	ESBL Ambler A	(2)
<i>bla</i> GES-1 to GES-9 and GES-11	MultiGES F MultiGES R	AGTCGGCTAGACCCGAAAG TTTGTCCTGTCTCAGGAT	ESBL Ambler A	(2)
<i>bla</i> GES-1 to GES-9 and GES-11	MultiGES F MultiGES R	AGTCGGCTAGACCCGAAAG TTTGTCCTGTCTCAGGAT	ESBL Ambler A	(2)
<i>bla</i> PER-1 and PER-3	MultiPER F MultiPER R	GCTCCGATAATGAAAGCGT TTCGGCTTGACTCGGCTGA	ESBL Ambler A	(2)
<i>bla</i> KPC-1 to KPC-5	MultiKPC F MultiKPC R	CATTCAAGGGCTTTCTTGCTGC ACGACGGCATAGTCATTTGC	Carbapenemase Ambler A	(2)
<i>bla</i> LAT-1 to LAT-3, BIL-1, CMY-2 to CMY-7, CMY-12 to CMY-18 and CMY-21 to CMY-23	CITMF CITMR	TGGCCAGAAGTACAGGCAAA TTTCTCTGAACGTGGCTGGC	Cephalosporinase Ambler C	(13)
<i>bla</i> LAT-1 to LAT-3, BIL-1, CMY-2 to CMY-7, CMY-12 to CMY-18 and CMY-21 to CMY-23	MultiCaseCIT F MultiCaseCIT R	CGAAGAGGCAATGACCAGAC ACGGACAGGGTTAGGATAGY	Cephalosporinase Ambler C	(2)
<i>bla</i> FOX-1 to FOX-5	MultiCaseFOX F MultiCaseFOX R	CTACAGTGCGGGTGGTTT CTATTTGCGGCCAGGTGA	Cephalosporinase Ambler C	(2)
<i>bla</i> MOX-1, MOX-2, CMY-1, CMY-8 to CMY-11 and CMY-19	MultiCaseMOX F MultiCaseMOX R	GCAACAACGACAATCCATCCT GGGATAGGCGTAACTCCTCCAA	Cephalosporinase Ambler C	(2)
<i>bla</i> DHA	DHAMF DHAMR	AACTTTCACAGGTGTGCTGGGT CCGTACGCATACTGGCTTTGC	Cephalosporinase Ambler C	(13)

<i>bla</i> DHA-1 and DHA-2	MultiCaseDHA F MultiCaseDHA R	TGATGGCACAGCAGGATATTC GCTTTGACTCTTTCCGGTATTCG	Cephalosporinase Ambler C	(2)
<i>bla</i> ACC-1 and ACC-2	MultiCaseACC F MultiCaseACC R	CACCTCCAGCGACTTGTAC GTTAGCCAGCATCCAGATCC	Cephalosporinase Ambler C	(2)
<i>bla</i> ACT-1 and MIR-1	MultiCaseEBC F MultiCaseEBC R	CGGTAAAGCCGATGTTGCG AGCCTAACCCCTGATACA	Cephalosporinase Ambler C	(2)
<i>bla</i> IMP	imp-BE1 imp-BE2	CAYGGTTTGGTGGTTCTTGTA CCTTTAACVGCCTGYCTYMT	Carbapenemase (MBL) Ambler B	(5)
<i>bla</i> IMP variants except IMP-9, IMP-16, IMP-18, IMP-22 and IMP-25	MultiIMP F MultiIMP R	TTGACACTCCATTTACDG GATYGAGAATTAAGCCACYCT	Carbapenemase (MBL) Ambler B	(2)
<i>bla</i> VIM	Vim-F Vim-R	GATGGTGTGGTTCGCATA CGAATGCGCAGCACCAG	Carbapenemase (MBL) Ambler B	(4)
<i>bla</i> VIM variants including VIM-1 and VIM-2	MultiVIM F MultiVIM R	GATGGTGTGGTTCGCATA CGAATGCGCAGCACCAG	Carbapenemase (MBL) Ambler B	(2)
<i>bla</i> NDM1	NDM.F NDM.R	CTCCAACGGTTTGATCGTC ATTGGCATAAGTCGCAATCC	Carbapenemase (MBL) Ambler B	(3)
<i>bla</i> OXA-1, OXA-4 and OXA-30	MultiISO-O F MultiISO-O R	GGCACCAGATTCACCTTCAAG GACCCAAAGTTCTGTAAGTG	OXA beta-lactamase Ambler D	(2)
<i>bla</i> OXA-48-like	MultiOXA-48 F MultiOXA-48 R	GCTTGATCGCCCTGATT GATTTGCTCCGTGGCCGAAA	OXA carbapenemase Ambler D	(2)

Non-beta-lactamases

Gene Description	Primer labels	Sequence (5'-3')	Gene Function	Reference
qnrA	qnrAF qnrAR	ATTTCTCACGCCAGGATTTG GATCGGCAAAGGTTAGGTCA	Quinolone resistance, plasmid	(8)
qnrB	qnrBF qnrBR2	GATCGTGAAAGCCAGAAAGG ATGAGCAACGATGCCTGGTA	Quinolone resistance, plasmid	(8)
qnrC	qnrCF qnrCR	GGGTTGTACATTTATTGAATCG CACCTACCCATTTATTTCA	Quinolone resistance, plasmid	(8)
qnrS	qnrSmF qnrSmR	GCAAGTTCATTGAACAGGGT TCTAAACCGTCGAGTTCGGCG	Quinolone resistance, plasmid	(8)
qep	qepAF qepAR	AACTGCTTAGCCCGTAGAT GTCTACGCCATGGACCTCAC	Quinolone efflux pump, plasmid	(8)
gyrA	gyrAWF gyrAWR	AAATCTGCCGTGTCGTTGGT GCCATACCTACGGCGATACC	Quinolone resistance, chromosomal (topoisomerase II)	(8)
parC	parCWF parCWR	CTGAATGCCAGCGCCAAATT GCGAACGATTTCCGGATCGTC	Quinolone resistance, chromosomal (topoisomerase IV)	(8)
3aac(6')-Ib	aac6'-A aac6'-B	TTGCGATGCTCTATGAGTGGCTA CTCGAATGCCTGGCGTGT	Aminoglycoside acetyltransferase	(9)
aac(6')-Ib-cr	aac(6')-Ib-cr F aac(6')-Ib-cr R	GCAACGCAAAAACAAGTTAGG GTGTTTGAACCATGTACA	Aminoglycoside acetyltransferase plus ciprofloxacin resistance	(12)
aac(6')-Ic	aac6'-Ic (F) aac6'-Ic (R)	CTACGATTACGTCAACGGCTGC TTGCTTCGCCACTCCTGCACC	Aminoglycoside acetyltransferase	(7)
aac(6')-Id	aac6'-Id (F) aac6'-Id (R)	TGGCTGCAACTTCGTTTTCTCC GGTGAAATCTATGGCTTCCG	Aminoglycoside acetyltransferase	(7)
aac(6')-Ie	aac6'-Ie (F) aac6'-Ie (R)	ACATTATACAGAGCCTTGGG CTCGTGTAAATTCATGTTCTGGC	Aminoglycoside acetyltransferase	(7)
aac(6')-Ig	aac6'-Ig (F) aac6'-Ig (R)	CCTATTCCGATCACCAAGC TTGTTTTGCCACACTTCG	Aminoglycoside acetyltransferase	(7)
aac(6')-Ih	aac6'-Ih (F) aac6'-Ih (R)	ATTGGCTTATACCGACACC GGCTGATCTGATTATCCAACGC	Aminoglycoside acetyltransferase	(7)
aac(6')-Ii	aac6'-Ii (F) aac6'-Ii (R)	AAGAATATGGAGACGCTCGGC ACAGGTCGTTTGACTTAACG	Aminoglycoside acetyltransferase	(7)
aac(6')-Ij	aac6'-Ij (F) aac6'-Ij (R)	CATGACGAGGCACATTTACAGG GGCTAATACGATTATCGAGGGC	Aminoglycoside acetyltransferase	(7)
aac(6')-Il	aac6'-Il (F)	TGCTTGGGAATATGTCTGG	Aminoglycoside	(7)

	aac6'-II (R)	TTGTTGGGCTGTTCTTCTAGC	acetyltransferase	
aac(6')-Iib	aac(6')-Iib (F) aac(6')-Iib (R)	CGCTTGTTGATTTGCTGTTGCG TTGAAACGACCTTGACCTTCCG	Aminoglycoside acetyltransferase	(7)
ant(3'')-Ia	ant(3'')-Ia (F) ant(3'')-Ia (R)	TCGACTCAACTATCAGAGG ACAATGGTGACTTCTACAGCG	Aminoglycoside acetyltransferase	(7)
armA	armA F armA R	AGGTTGTTTCCATTTCTGAG TCTCTCCATTCCTTCTCC	Aminoglycoside resistance (16S rRNA methylase)	(15)
rmtA	rmtA F rmtA R	CTAGCGTCCATCCTTCTC TTGCTTCCATGCCCTTGCC	Aminoglycoside resistance (16S rRNA methylase)	(15)
rmtB	rmtB F rmtB R	CCCAAACAGACCGTAGAGGC CTCAAACCTGGCGGCAAGC	Aminoglycoside resistance (16S rRNA methylase)	(15)
rmtC	rmtC F rmtC R	CGAAGAAGTAACAGCCAAAG ATCCCAACATCTCTCCCACT	Aminoglycoside resistance (16S rRNA methylase)	(15)
catA1	CAT F CAT R	AAGTTGGCAGCATTACCCG TCGTGGTATTCACTCCAGAGCG	Chloramphenicol resistance	(12)
tetA	TetA F TetA R	GGTCTTGCTCGTCTCGCTGG AACGCCATCCATCCCGTG	Tetracycline resistance	(12)
tetB	TetB F TetB R	CCTTATCATGCCAGTCTTGC ACTGCCGTTTTTTCGCC	Tetracycline resistance	(12)
tetC	TetC F TetC R	ACTTGGAGCCACTATCGAC CTACAATCCATGCCAACCC	Tetracycline resistance	(12)
tetD	TetD F TetD R	TGGGCAGATGGTCAGATAAG CAGCACACCCTGTAGTTTTTC	Tetracycline resistance	(12)
tetE	TetE F TetE R	TTAATGGCAACAGCCAGC TCCATACCCATCCATTCCAC	Tetracycline resistance	(12)
tetY	TetY F TetY R	ACCGCACTCATTGTTGTC TTCCAAGCAGCAACACAC	Tetracycline resistance	(12)
aphA1-Ia	Aph F Aph R	AACGTCTTGCTCGAGGCCGCG GGCAAGATCCTGGTATCGGTCTGC	Aminoglycoside (neomycin/kanamycin) resistance	(12)
aadB	AadB F AadB R	GGGCGCGTCATGGAGGAGTT TATCGCGACCTGAAAGCGGC	Adenyltransferase Trimethoprim and low- level streptomycin resistance	(12)
strA	StrA F StrA R	CCAATCGCAGATAGAAGGCAAG ATCAACTGGCAGGAGGAACAGG	Streptomycin resistance	(12)
dfrA1	dfr1-F dfr1-R	TGGTAGCTATATCGAAGAATGGAGT TATGTTAGAGGCGAAGTCTTGGGTA	Dihydrofolate reductase Trimethoprim resistance	(6)
dfrA5	dfr5-F dfr5-R	AGTACTCTTTAAAGCCTTGACGTA GTGTTGCTCAAAAACAACCTTCG	Dihydrofolate reductase Trimethoprim resistance	(6)
dfrA7, dfrA17	dfr7/17-F dfr7/17-R dfr7-R dfr17-R	ACATTTGACTCTATGGGTGTTCTTC AAAACGTTCAAAACCAAATTGAA ACCTCAACGTGAACAGTAGACAAAT TCTCTGGCGGGGGTCAAATCTAT	Dihydrofolate reductase Trimethoprim resistance	(6)
dfrA12	dfr12-F dfr12-R	GAGCTGAGATATACTCTGGCACT GTACGGAATTACAGCTTGAATGGT	Dihydrofolate reductase Trimethoprim resistance	(6)
sul1	Sul1-F Sul1-R	TGGTGACGGTGTTCGGCATTTC GCGAGGGTTTCCGAGAAGGTG	Sulfonamide resistance	(14)
sul2	Sul2-F Sul2-R	CGGCATCGTCAACATAACC GTGTGCGGATGAAGTCAG	Sulfonamide resistance	(14)
sul3	Sul3-F Sul3-R	CATTCTAGAAAACAGTCGTAGTTCC CATCTGCAGCTAACCTAGGGCTTTGGA	Sulfonamide resistance	(14)

References

1. **Boyd, D. A., S. Tyler, S. Christianson, A. McGeer, M. P. Muller, B. M. Willey, E. Bryce, M. Gardam, P. Nordmann, and M. R. Mulvey.** 2004. Complete nucleotide sequence of a 92-kilobase plasmid harboring the CTX-M-15 extended-spectrum beta-lactamase involved in an outbreak in long-term-care facilities in Toronto, Canada. *Antimicrobial agents and chemotherapy* **48**:3758-3764.
2. **Dallenne, C., A. Da Costa, D. Decre, C. Favier, and G. Arlet.** 2010. Development of a set of multiplex PCR assays for the detection of genes encoding important beta-lactamases in Enterobacteriaceae. *The Journal of antimicrobial chemotherapy* **65**:490-495.
3. **Ellem, J., S. R. Partridge, and J. R. Iredell.** 2011. Efficient direct extended-spectrum beta-lactamase detection by multiplex real-time PCR: accurate assignment of phenotype by use of a limited set of genetic markers. *Journal of clinical microbiology* **49**:3074-3077.
4. **Ellington, M. J., J. Kistler, D. M. Livermore, and N. Woodford.** 2007. Multiplex PCR for rapid detection of genes encoding acquired metallo-beta-lactamases. *The Journal of antimicrobial chemotherapy* **59**:321-322.
5. **Espedido, B. A., S. R. Partridge, and J. R. Iredell.** 2008. bla(IMP-4) in different genetic contexts in Enterobacteriaceae isolates from Australia. *Antimicrobial agents and chemotherapy* **52**:2984-2987.
6. **Grape, M., A. Motakefi, S. Pavuluri, and G. Kahlmeter.** 2007. Standard and real-time multiplex PCR methods for detection of trimethoprim resistance *df*r genes in large collections of bacteria. *Clinical microbiology and infection : the official publication of the European Society of Clinical Microbiology and Infectious Diseases* **13**:1112-1118.
7. **Hannecart-Pokorni, E., F. Depuydt, L. de wit, E. van Bossuyt, J. Content, and R. Vanhoof.** 1997. Characterization of the 6'-N-aminoglycoside acetyltransferase gene *aac(6')*-Im [corrected] associated with a *sulI*-type integron. *Antimicrobial agents and chemotherapy* **41**:314-318.
8. **Kim, H. B., C. H. Park, C. J. Kim, E. C. Kim, G. A. Jacoby, and D. C. Hooper.** 2009. Prevalence of plasmid-mediated quinolone resistance determinants over a 9-year period. *Antimicrobial agents and chemotherapy* **53**:639-645.
9. **Minarini, L. A., L. Poirel, V. Cattoir, A. L. Darini, and P. Nordmann.** 2008. Plasmid-mediated quinolone resistance determinants among enterobacterial isolates from outpatients in Brazil. *The Journal of antimicrobial chemotherapy* **62**:474-478.
10. **Monstein, H. J., A. Ostholm-Balkhed, M. V. Nilsson, M. Nilsson, K. Dornbusch, and L. E. Nilsson.** 2007. Multiplex PCR amplification assay for the detection of blaSHV, blaTEM and blaCTX-M genes in Enterobacteriaceae. *APMIS : acta pathologica, microbiologica, et immunologica Scandinavica* **115**:1400-1408.
11. **Paterson, D. L., K. M. Hujer, A. M. Hujer, B. Yeiser, M. D. Bonomo, L. B. Rice, R. A. Bonomo, and G. International Klebsiella Study.** 2003. Extended-spectrum beta-lactamases in *Klebsiella pneumoniae* bloodstream isolates from seven countries: dominance and widespread prevalence of SHV- and CTX-M-type beta-lactamases. *Antimicrobial agents and chemotherapy* **47**:3554-3560.

12. **Pazhani, G. P., S. Chakraborty, K. Fujihara, S. Yamasaki, A. Ghosh, G. B. Nair, and T. Ramamurthy.** 2011. QRDR mutations, efflux system & antimicrobial resistance genes in enterotoxigenic *Escherichia coli* isolated from an outbreak of diarrhoea in Ahmedabad, India. *The Indian journal of medical research* **134**:214-223.
13. **Perez-Perez, F. J., and N. D. Hanson.** 2002. Detection of plasmid-mediated AmpC beta-lactamase genes in clinical isolates by using multiplex PCR. *Journal of clinical microbiology* **40**:2153-2162.
14. **Saenz, Y., L. Brinas, E. Dominguez, J. Ruiz, M. Zarazaga, J. Vila, and C. Torres.** 2004. Mechanisms of resistance in multiple-antibiotic-resistant *Escherichia coli* strains of human, animal, and food origins. *Antimicrobial agents and chemotherapy* **48**:3996-4001.
15. **Shin, S. Y., K. C. Kwon, J. W. Park, J. H. Song, Y. H. Ko, J. Y. Sung, H. W. Shin, and S. H. Koo.** 2009. Characteristics of *aac(6')-Ib-cr* gene in extended-spectrum beta-lactamase-producing *Escherichia coli* and *Klebsiella pneumoniae* isolated from Chungnam area. *The Korean journal of laboratory medicine* **29**:541-550.