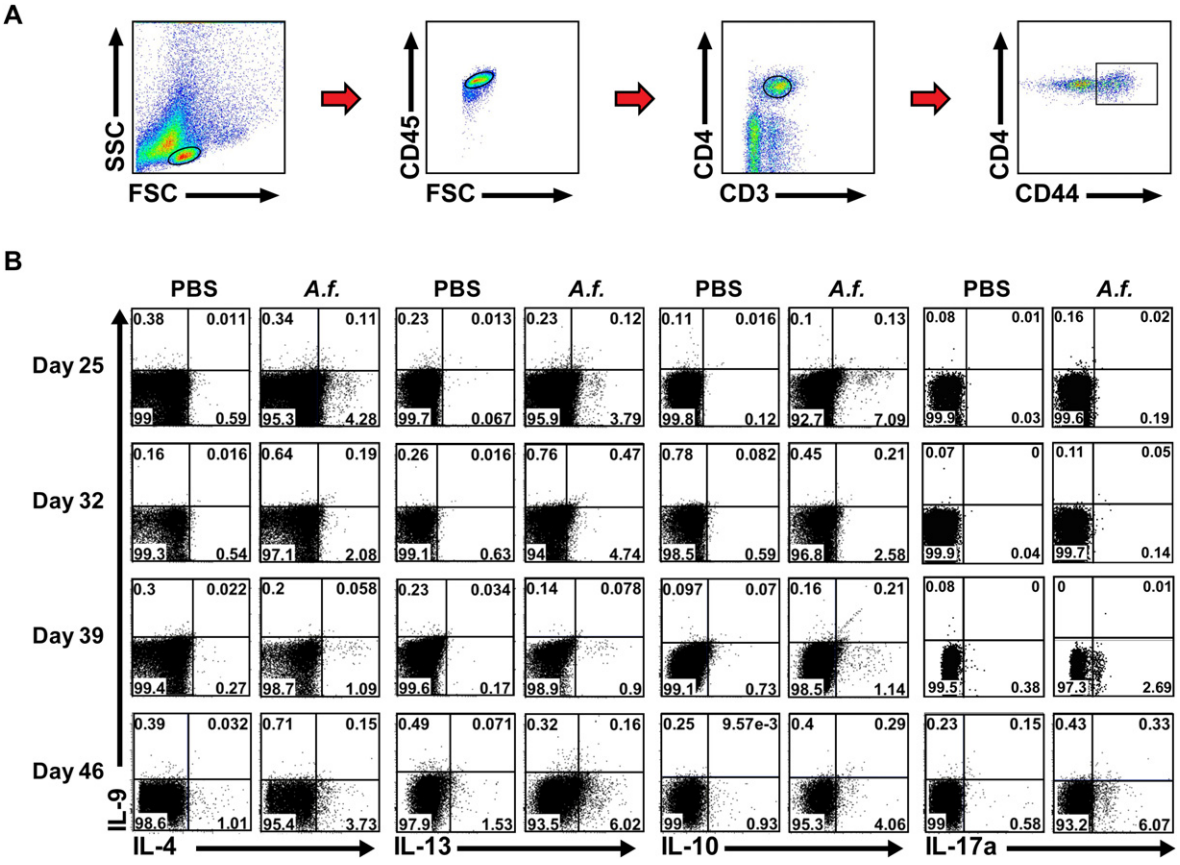

FIG E1. A, Gating strategy for analysis of cytokine production by effector $CD4^+$ T cells in the lungs. $CD45^+CD4^+CD3^+CD44^+$ cells were considered $CD4^+$ Teff cells and analyzed for intracellular cytokine production in the lungs. The lymphocytes were initially selected based on forward scatter *(FSC)* and side scatter *(SSC)*, followed by CD45, CD4, and CD3 gating. Finally, $CD44^+$ cells were selected as $CD4^+$ Teff cells and analyzed for the expression of cytokines. **B**, T_H9 cells do not develop in the LNs after chronic exposure to *A fumigatus (A.f.)*. A group of naive BALB/c mice were immunized intranasally for 46 days with *A fumigatus* (50 μg on weeks 1 and 2 and 20 μg on weeks 3-8 in 50 μL of saline solution) or PBS (negative control), as described in the protocol of immunization (Fig 1, *A*). On days 25, 32, 39, and 46, the percentage of cytokine (IL-9, IL-4, IL-13, and IL-10)–secreting Teff cells in the LNs was then assessed by means of intracellular staining gated on the $CD45^+CD44^+CD4^+$ population.

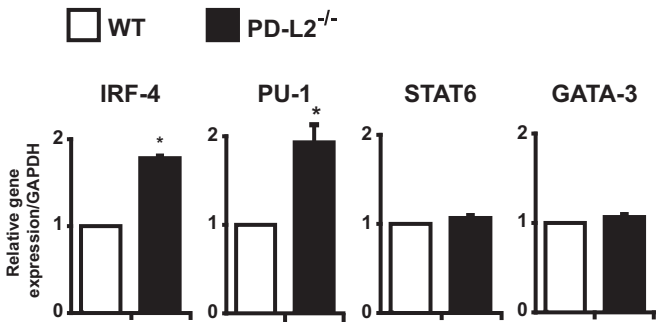

FIG E2. Relative gene expression of the transcription factors IRF4, PU-1, STAT6, and GATA-3 in *in vitro*–primed $CD4^+$ T cells. Purified $CD4^+DO11.10^+$ T cells were put in culture (1×10^5 cells per well) with OVA-loaded bone marrow–derived DCs from WT and PD-$L2^{-/-}$ mice (1:32 ratio) in the presence of IL-4 (10 ng/mL), TGF-β (1 ng/mL), and anti–IFN-γ antibodies (10 μg/mL). After 5 days of culture, relative gene expression of IRF4, PU-1, STAT6, and GATA-3 was analyzed by using quantitative real-time PCR. Data are means ± SEMs of 3 experiments. $*P < .05$, WT versus PD-$L2^{-/-}$ mice.