

SUPPLEMENTAL DATA

Supplementary methods:

Antibodies:

In order to perform the western blot showed in Fig. S3, we used anti-CREB (1/1000, Cell Signaling) and anti-P-CREB (1/1000, Cell Signaling).

Membrane extraction:

Protein extracts from pituitary glands were prepared using 4 mice/genotype; tissues were homogenized in 200 μ l of extraction buffer (10 mM TrisHCl, 5 mM EDTA; pH 7.4). Homogenates were briefly centrifuged at 2000 rpm for ten minutes at 4°C. The supernatant was collected and the pellet was re-extracted in the same way. Supernatants from both extractions were pooled and centrifuged at 45000x g for 45 min at 4°C (Optima TLX Ultracentrifuge, Beckman Coulter). The obtained pellets were resuspended in 750 μ l of extraction buffer followed by a second centrifugation in the same conditions. The resulting pellets were finally resuspended in 30 μ l of 50 mM TrisHCl buffer (pH 7.4). Protein quantifications were performed by Bradford. 30 μ g of proteins/genotype were analyzed by western blot using mouse monoclonal antibodies directed against D2R (1:4000), as previously described (Doi et al, Nat Neuroscience 2006). To control for the amount of proteins loaded, blots were also incubated with a membrane marker (pan-cadherin antibodies; Abcam; 1:1000).

Legends

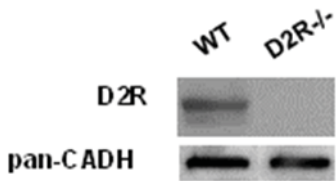
Fig.S1: In vivo deletion of D2S or D2L in the pituitary gland leads to a similar amount of proteins immune-reactive to anti-D2R antibodies in membrane extracts. Western blot analyses of membrane extracts from pituitary glands of WT, D2L^{-/-} and D2S^{-/-} mice show that deletion of either D2L in D2L^{-/-} or D2S in D2S^{-/-} pituitaries results into levels of D2 reactive bands similar to that of WT pituitaries. D2R antibodies detect a D2 specific bands at the approximate size of 90 kD. (A) Representative western blot of WT and D2R^{-/-} pituitary extracts using mouse monoclonal D2R antibodies show the presence of a D2 specific band only in WT but not in D2R^{-/-} extracts. Incubation of western blots with anti-pan-cadherin antibodies, a marker of membrane proteins, was used as internal control of loaded quantities of proteins. (B) Representative western blot of protein extracts from WT, D2L^{-/-} and D2S^{-/-} pituitaries incubated with antibodies directed against D2R and pan-cadherin as internal control. (C) Quantification of D2-specific band intensity in membrane extracts of WT, D2L^{-/-} and D2S^{-/-} pituitaries as shown in B), shows similar levels of D2-specific bands in all genotypes. Bars represent the ratio between values from D2R and pan-Cadherin specific bands quantified using Image J. Statistical analyses were performed by one-way ANOVA.

Fig.S2: Absence of gender differences in the transduction pathways activated by either D2L or D2S. Western blot analyses of pituitary extracts from 4-month-old WT, D2R^{-/-}, D2L^{-/-} and D2S^{-/-} male mice showed that D2L or D2S affect the ERK and AKT pathways in males D2L^{-/-} or D2S^{-/-} pituitaries as they do in females. Bars represent the ratio between phospho-ERK 1/2/ERK 1/2 (panel A) and phospho-AKT/AKT (panel B). At least 4 animals per genotype were used

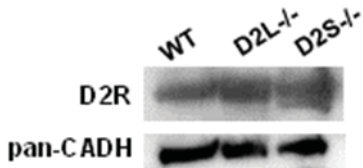
and results analyzed by one-way ANOVA followed by Bonferroni's test (A: Genotype: $F_{(3, 16)}=40.57$, B: Genotype: $F_{(3, 16)}=32.35$).

Fig. S3: The level of CREB phosphorylation is not altered either by D2L or D2S ablation in the pituitary gland. Comparative western blot analyses of CREB phosphorylation levels in 4-month-old WT, D2L^{-/-} and D2S^{-/-} pituitary extracts from mice treated either with saline or 20mg/kg haloperidol. Blots were analyzed using antibodies directed against total CREB and CREB phosphorylated on serine 133 (P-CREB). Bars represent the ratio between P-CREB/CREB levels obtained after quantifications. Statistical analyses were performed on at least 5 animals/genotype analyzed by two-way ANOVA followed by the appropriate post-hoc analysis. (Treatment: $F_{(5, 27)}: 17.77$). * $p < 0.05$ vs respective genotype without HAL.

A



B



C

