

Fig. S1: Electron microscope images of A β ₁₋₄₀ fibrils prepared as described in “Methods” section. a) the batch used to prepare frozen solution A β -VF fibrils; b) batch used to prepare frozen solution A β -VG fibrils.

Fig. S2: DNP build-up curves measured using proton signals and ¹³C-CP signals, monoexponential fitting curves are shown in red. a) A β -VF fibrils in frozen solution, b) A β -VF peptide in frozen solution, c) A β -VG fibrils in frozen solution, d) transverse relaxation curves for samples in frozen solution.

Fig. S3: Schematic representation of relative orientation of CSA tensors in the molecular frame and with respect to each other.

Fig. S4: Series of simulated 2D patterns for a wide range of various (ϕ, ψ) angles. Spectral broadening is set to match the one found in room temperature measurements, exchange time 2.5 s

Fig. S5: $\chi^2_{red}(\phi, \psi)$ plots for the best fits to individual 2D exchange spectra of A β -VF fibrils with various $\tau_{exchange}$. (data are shown in Fig. 6), a) $\tau_{exchange}=0.1$ s, b) $\tau_{exchange}=0.5$ s, c) $\tau_{exchange}=1.0$ s, d) $\tau_{exchange}=2.5$ s, e) $\tau_{exchange}=10$ s.

Fig. S6: RMSD plot for simulated data. The RMSD is calculated as

$$RMSD(\phi, \psi) = \sqrt{\sum_i^N (\lambda S_i(\phi, \psi) - \lambda S_i(\phi_0, \psi_0))^2}$$

, where a spectrum S_i simulated for specific (ϕ_0, ψ_0) angles is compared to all other simulations, the optimal scaling factor λ is calculated

as $\lambda = \frac{\sum_i^N S_i(\phi_0, \psi_0) S_i}{\sum_i^N S_i(\phi_0, \psi_0)^2}$ a) $(\phi_0, \psi_0) = (-50^\circ, -60^\circ)$; b) $(\phi_0, \psi_0) = (-120^\circ, 120^\circ)$; c) $(\phi_0, \psi_0) = (-150^\circ, 150^\circ)$; d) $(\phi_0, \psi_0) = (-75^\circ, -150^\circ)$