

SUPPLEMENTARY DATA

FIG. S1. Chromosome locations of *Ta-PHR1-A1*, *B1* and *D1*. *Ta-PHR-A1*, *Ta-PHR-B1* and *Ta-PHR-D1* were located between 7AS8-0.45 and 7AL1-0.39 on chromosome 7A, between 4BS4-0.37 and 4BL-0.71 on chromosome 4B and between 4DL9-0.31-0.56 on chromosome 4D, respectively.

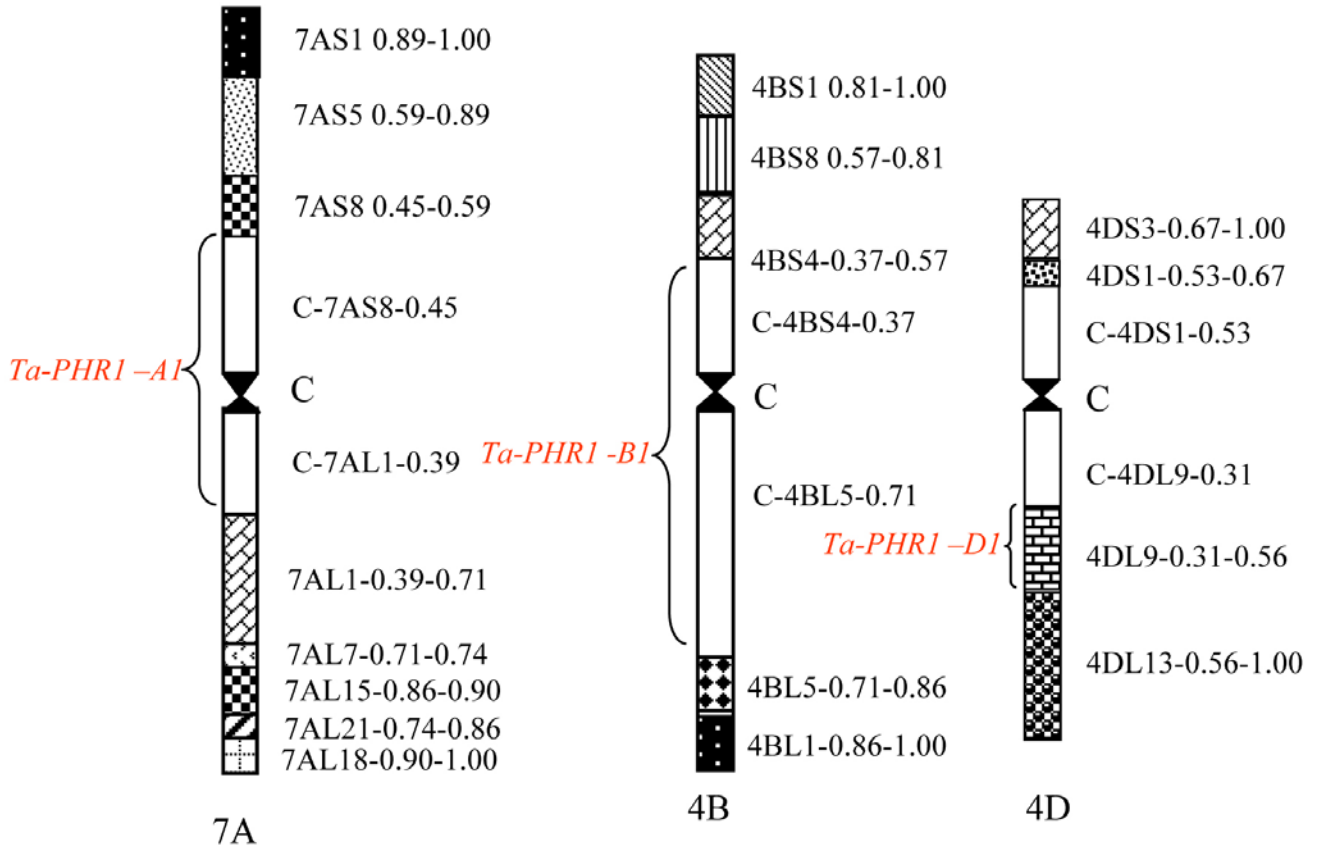


FIG. S2. Alignment of Ta-PHR1 protein sequences. MYB, MYB domain; C-C, CC domain. Multiple sequence alignment was carried out using the ClusTa-IX 1.81 program (Thompson *et al.*, 1997) with default multiple alignment parameters.

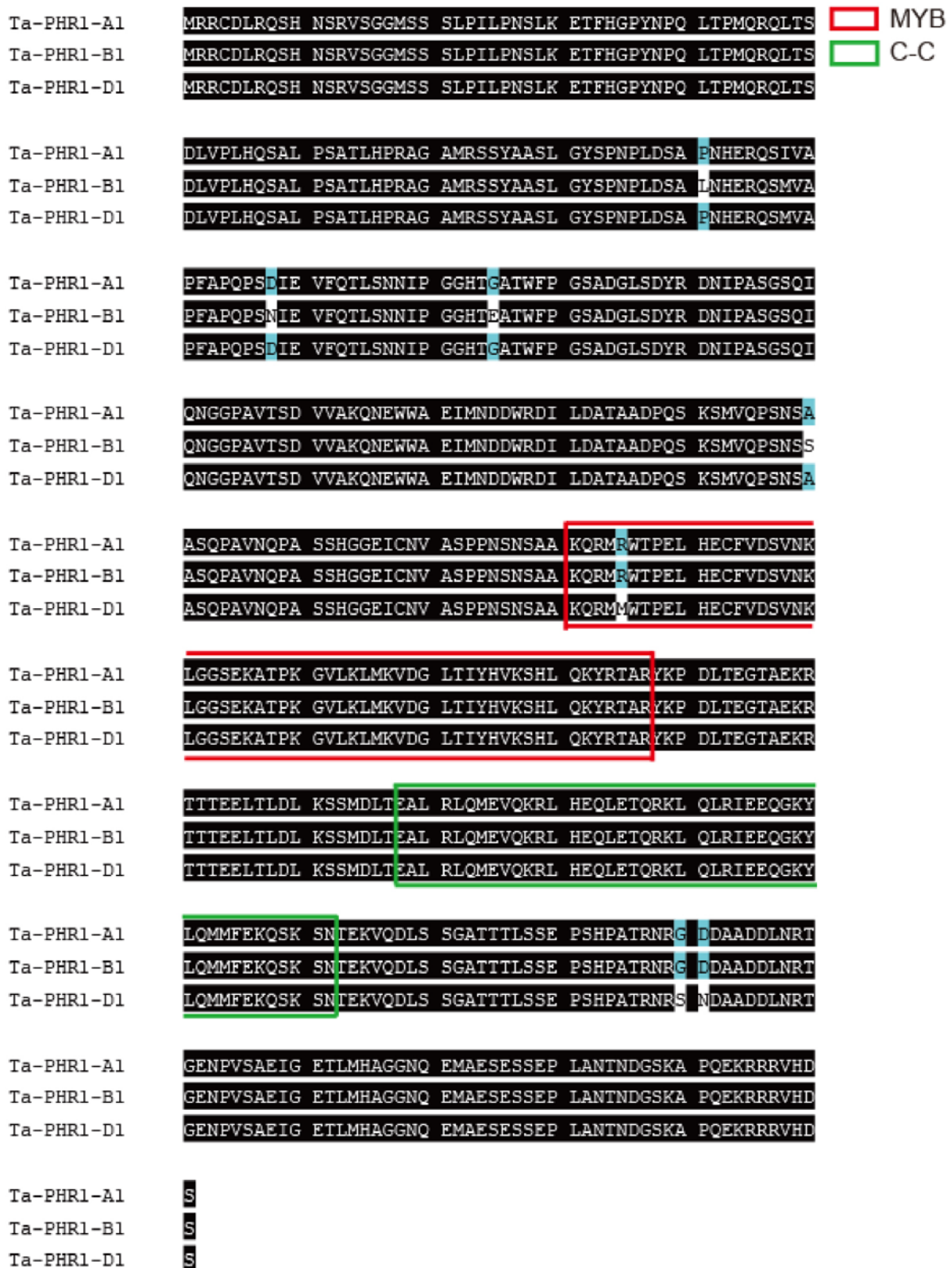


FIG. S3. Expression level of phosphate-starvation-inducible genes in wild-type (WT) and its *Ta-PHR1* transgenic lines under Pi-sufficient (200 μ M Pi) and Pi-deficient (10 μ M Pi) conditions. Seeds were germinated for 7 d, and then the germinated seeds with residual endosperm removed were transferred to nutrient solution containing 10 μ M Pi and 200 μ M Pi, respectively. After the plants were grown in the low and high P nutrient solutions for 7 d, the shoots and roots were collected separately for gene expression analysis. S, shoot; R, root; +P, 200 μ M Pi; -P, 10 μ M Pi. Values are mean \pm s.e. of three replications.

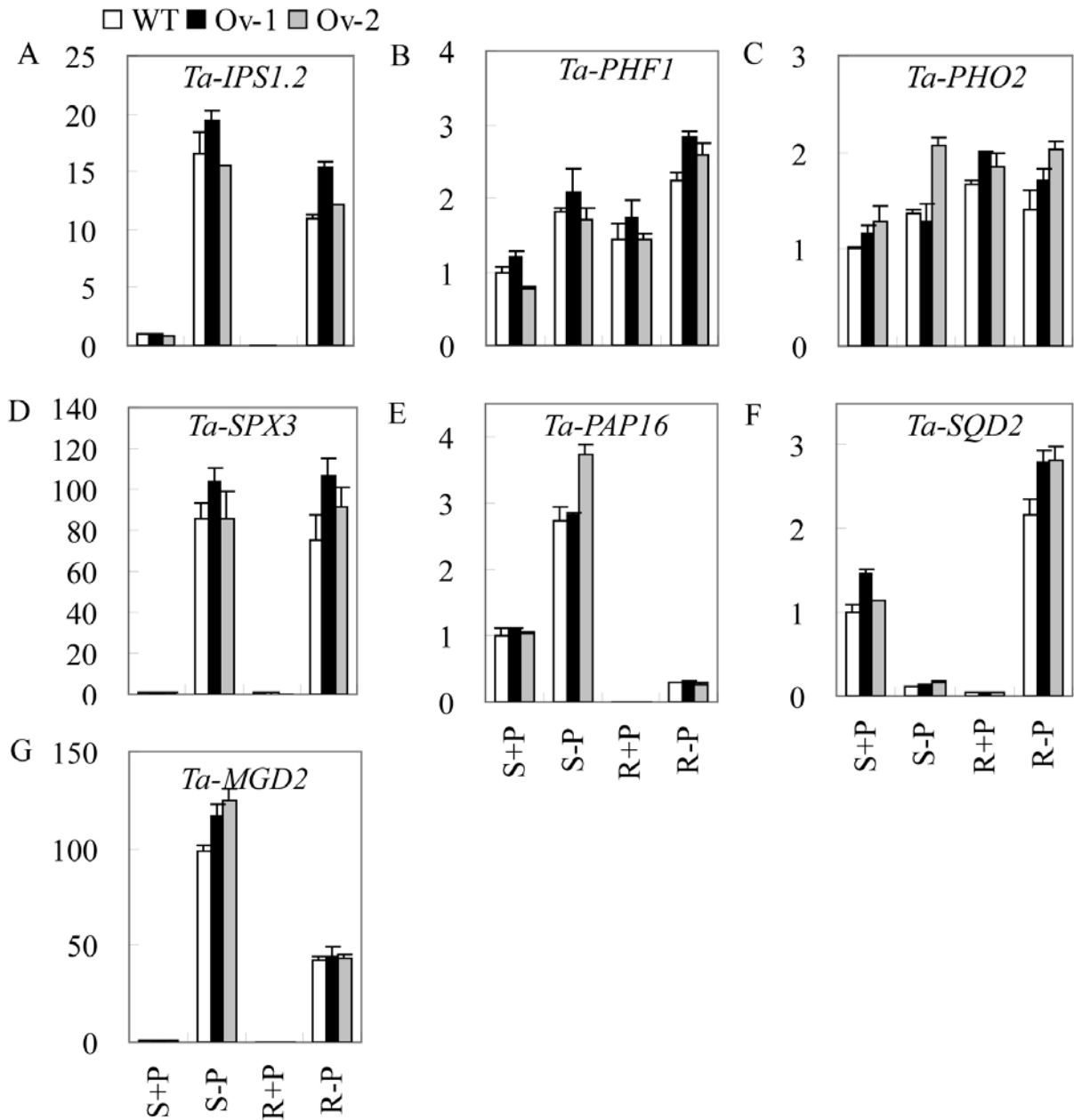


TABLE S1. Primers for RT-PCR or Real-time PCR analysis.

Gene	Forward primer sequence(5'-3')	Reverse primer sequence(5'-3')	Description		
<i>Ta-PHR1</i>	<i>Ta-PHR1CF</i>	GAATGGTGGGCAGAAATa-ATG	<i>Ta-PHR1CR</i>	GGGGTTCTCTCCTGTTCa-TTT	Amplify a fragment from wheat cDNA template
	<i>Ta-PHR1FE</i>	GAATTCATGAGGAGGTGTGATCTGAG	<i>Ta-PHR1RS</i>	GGTCGATTa-ACTa-TCATGCACCCTTCG	Amplify for coding sequences of Ta-PHR1-A1
	<i>Ta-PHR1qF</i>	TTCCCAGGTTCCGGCTGATG	<i>Ta-PHR1qR</i>	CTGCCACCATTTCATTTTGT	Real time PCR primer of Ta-PHT1.2
	<i>Ta-PHR1_BiFC_F</i>	GGGACAAGTTTGTa-CAAAAAAGCAGGCTGCATGGAGTa-TCCATGGCTG	<i>Ta-PHR1_p101-CFP_R</i>	GGGGACCACTTTGTa-CAAGAAAGCTGGGTCTGATTGGCGTAATCCCAA	Amplify for coding sequences of Ta-PHR1-A1 for
<i>BiCF</i>					
<i>Ta-PHT1.2 promoter</i>	<i>proTa-PHT1.2FK</i>	GGTa-CCAGTa-CGGCATA-CGGATTTCAATGC	<i>proTa-PHT1.2RS</i>	GTCGACGGCCGCGGCGATCTCTC	Amplify for coding sequences of Ta-PHT1.2
<i>Ta-Actin</i>	<i>Ta-Actin qF</i>	ACCTTCAGTTGCCAGCAAT	<i>Ta-Actin qR</i>	CAGAGTCGAGCACAATa-CCAGTTG	Real time PCR primer of Ta-Actin
<i>Ta-PHT1.2</i>	<i>Ta-PHT1.2qF</i>	GCATCTa-CTa-CACCGACCCT	<i>Ta-PHT1.2qR</i>	ACGCGATGGAGCAGAGGAC	Real time PCR primer of Ta-PHT1.2
<i>Ta-PHT1.6</i>	<i>Ta-PHT1.6qF</i>	CAGCTCTTCTTCGGCTGGCT	<i>Ta-PHT1.6qR</i>	CCGAGCCAGAAGCGGAAGAA	Real time PCR primer of Ta-PHT1.6
<i>Ta-PHF1</i>	<i>Ta-PHF1qF</i>	GAAAGAATGGCAAATCTGGCTC	<i>Ta-PHF1qR</i>	TGCATGGCTTGGTa-GGTCG	Real time PCR primer of Ta-PHF1
<i>Ta--IPS1.1</i>	<i>Ta-IPS1.1qF</i>	ATGGATCCGGCGTTGGCTa-G	<i>Ta-IPS1.1qR</i>	AGTTGCCCTa-CCTTa-GTa-GAGGTGA	Real time PCR primer of Ta-IPS1.1
<i>Ta-IPS1.2</i>	<i>Ta-IPS1.2qF</i>	ATGGATCCGGCGTTGGCTa-G	<i>Ta-IPS1.2qR</i>	ATa-TTTa-TTGATGATTa-CACTa-GTCA	Real time PCR primer of Ta-IPS1.2
<i>Ta-PHO2</i>	<i>Ta-PHO2 qF</i>	AGTTTa-TGAGGAAAGGATGGACCT	<i>Ta-PHO2 qR</i>	ATa-GTGAACGGAAGGTGGTTCAT	Real time PCR primer of Ta-PHO2
<i>Ta-SPX3</i>	<i>Ta-SPX3 qF</i>	GTGGAAGGACGAGTTCCTGAGC	<i>Ta-SPX3 qR</i>	TCCCGGTGTGTGATGATGAAGAA	Real time PCR primer of Ta-SPX3
<i>Ta-PAP16</i>	<i>Ta-PAP16 qF</i>	CGTa-CGAGAACTCTGGCTGTGT	<i>Ta-PAP16 qR</i>	AGAGAACGGCTTCTCGGCTa-TCT	Real time PCR primer of Ta-PAP16
<i>Ta-SQD2</i>	<i>Ta-SQD2 qF</i>	TGCCAGTGGAGATGTGTTTGTGAT	<i>Ta-SQD2 qR</i>	AAGCTGGTCTTTCCTTCCTGAT	Real time PCR primer of Ta-SQD2
<i>Ta-MGD2</i>	<i>Ta-MGD2 qF</i>	AGAGCTCCTa-CAAGTTCATGGTGAA	<i>Ta-MGD2 qR</i>	AGTGTTGAGGTCGGTGATGACAG	Real time PCR primer of Ta-MGD2
<i>Ta-ASA1</i>	<i>Ta-ASA1 qF</i>	ACGTTGAACGATa-TTCACATGTCA	<i>Ta-ASA1 qR</i>	AACCCCTCAAAGCCACCACTGTa-	Real time PCR primer of Ta-ASA1
<i>Ta-GH3.1</i>	<i>Ta-GH3.1 qF</i>	TGTCCTa-CACCATCATGCCGA	<i>Ta-GH3.1 qR</i>	CTCGTCCGTCTTGTCCGACT	Real time PCR primer of Ta-GH3.1
<i>Ta-CYCD2</i>	<i>Ta-CYCD2 qF</i>	GAGGGACGCCATa-GATTGGATT	<i>Ta-CYCD2 qR</i>	TGAGCACCAAAGCTCCATCCT	Real time PCR primer of Ta-CYCD2