

Supplementary figure legends

Figure S1 Identification of the MICAL-L1 region that binds to Syndapin2. **(A)** HeLa cells on cover-slips were mock-treated or treated with either MICAL-L1-, Syndapin2-, or Bin1-SiRNA for 72 h, and transfected with GFP-Myc-EHD1 for the last 24 h. Bar, 10 μ m. **(B)** HeLa cells were mock-treated or treated with Bin1- or Syndapin2-SiRNA for 72 h and subjected to immunoblot analysis with antibodies against Bin1, Syndapin2 and actin (control). **(C-F)** Yeast two-hybrid studies were performed to assess Syndapin2 binding to: **(C)** MICAL-L1, EHD1, EHD2, EHD3 and EHD4. **(D)** MICAL-L1 and a series of truncation mutants, as indicated. **(E)** MICAL-L1 residues 1-368, 368-468, 468-863 and MICAL-L1 residues 368-468 with PXXP-to-AXXA mutations at each PXXP motif. **(F)** MICAL-L1 residues 468-863 and its mutant counterpart with PRAP mutated to ARAA beginning at residue 480. **(G)** Yeast two-hybrid studies were performed with MICAL-L1 as prey and bait. **(H)** Sequence comparison of the PXXP Syndapin-binding motifs of MICAL-L1 and dynamin1. **(I)** HeLa cells were transfected with both GFP-Myc-EHD1 and HA-Syndapin2 for 48 h. Cells were lysed and lysate was subjected to immunoprecipitation using anti-MICAL-L1 antibodies. The cell lysate and the immunoprecipitate were then subjected to immunoblot analysis with antibodies against GFP, the HA-tag and actin. **(J)** HeLa cell lysates were transfected with either GFP or GFP-Myc-EHD3 for 48 h, and subjected to GST pull-down with GST-Syndapin2 as bait. The cell lysate and precipitate were then subjected to immunoblot analysis with antibodies against the GFP-tag.

Figure S2 Exogenously expressed Syndapin2 and MICAL-L1 co-localize on tubular endosomes. **(A)** HeLa cells were mock-treated or treated with Bin1-SiRNA for 72 h. Cells were immunostained for endogenous Syndapin2 and MICAL-L1. Bar, 10 μ m. **(B)** HeLa cells were transfected with Tomato-Syndapin2, GFP-MICAL-L1 or co-transfected with both. Singly transfected cells were stained for endogenous MICAL-L1 or Syndapin2 as indicated. Yellow arrowheads mark co-localization to tubular endosomes. Bar, 10 μ m.

Figure S3 Syndapin2 and MICAL-L1 dynamically associate with tubular recycling endosomes. FRAP experiments were performed on HeLa cells transfected with Tomato-Syndapin2 and GFP-MICAL-L1. Images were taken before and after photobleaching at the indicated times. Bar, 10 μm .

Figure S4 Loss of Syndapin2 affects organization of early endosomes, lysosomes and golgi. (A) HeLa cells were mock-treated or Syndapin2-SiRNA-treated for 72 h and immunostained for either EEA1, Giantin or Lamp1. (B) HeLa cells were mock-treated or Syndapin2-SiRNA-treated for 72 h and immunostained for endogenous SNX1. Insets are shown below each image. (C) HeLa cells were mock-treated or Syndapin2-SiRNA-treated for 72 h and immunostained for endogenous Rabankyrin-5. Insets are shown below the mock images. Bar, 10 μm .

Figure S5 Purification of His-tagged MICAL-L1-CC domain. (A) Samples were collected at various stages of purification of the His-tagged MICAL-L1-CC domain: before and after IPTG induction, after cell lysis, and after cell lysate precipitation to pellet inclusion bodies. Inclusion bodies were then resuspended and precipitated in 1 M urea buffer. Pellets were further solubilized in 8 M urea buffer and both soluble and insoluble fractions were collected. These samples were run on SDS page and stained with Coomassie Blue. (B) Unfolded protein in 8 M urea buffer was bound to Ni-NTA column and eluted. Proteins from the eluted fractions were separated SDS-PAGE and stained with Coomassie Blue. (C) The fractions rich in His-tagged MICAL-L1-CC domain were pooled together (eluted protein) and dissolved in ten times the volume of refolding buffer. The protein was then concentrated using centrifugal filtration devices. Samples were separated on SDS-PAGE and analyzed by Coomassie staining. (D) The refolded and concentrated His-tagged-MICAL-L1-CC domain was subjected to circular dichroism.

Figure S6 Lipid overlay assay of MICAL-L1 and Syndapin2. (A-B) Lipid overlay assays were performed with His-tagged MICAL-L1-CC domain, GST-Syndapin2 (full length) or GST-

Syndapin2-F-BAR domain. The lipid strips were then immunoblotted for the respective tags.

(C) Phosphatidic acid-binding probes localize at the tubular endosomes. HeLa cells were transfected with Spo20p phosphatidic acid-binding domain (PABD) probe or its non-interacting mutant (L67P) as indicated by immunostaining for endogenous MICAL-L1. Bar, 10 μm .

(D) HeLa cells transfected with the HA-MICAL-L1-CC domain for 16 h were co-stained with antibodies against the HA-tag together with either endogenous MICAL-L1 or Syndapin2. Arrows indicate partial co-localization of the MICAL-L1-CC domain and Syndapin2 on tubular membranes.

Figure S7 HeLa cells transfected with HA-MICAL-L1-CC domain for 16 h were incubated with Alexa-Fluor 568-labeled transferrin for 5 min. The cells were either fixed (pulse 5 min.) or fixed after 10 min. (Chase 10 min.) or 20 min. (Chase 20 min.) incubations in complete media. Cells were then immunostained with antibodies against the HA-tag. Bar 10 μm .

Figure S8 *In vitro* LMV tubulation with liposomes containing phosphatidylserine (PS) and PI(4,5)P₂. LMV tubulation was assessed in the absence of protein, GST-Synd2, GST-Synd F-BAR domain or the CC domain of MICAL-L1. LMVs were comprised of a mass ratio of 70% PC, 10% Rhodamine-PE and either 20% PS or PI(4,5)P₂. Insets depict the region in the white box. Bar, 10 μm .

Figure S1

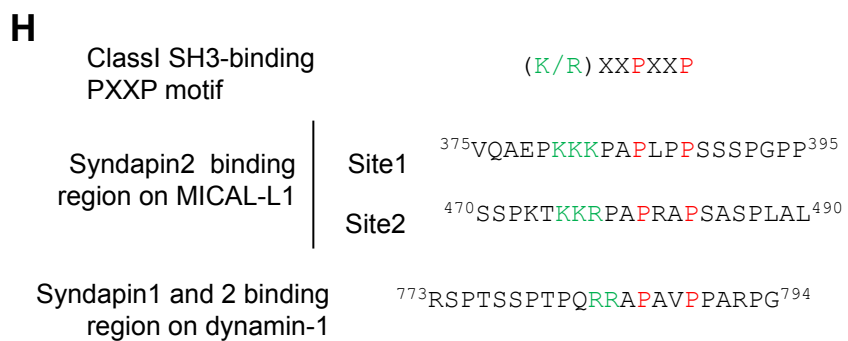
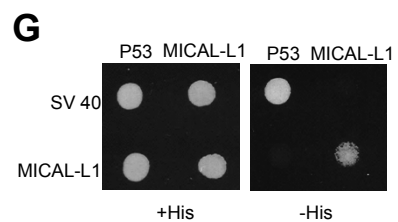
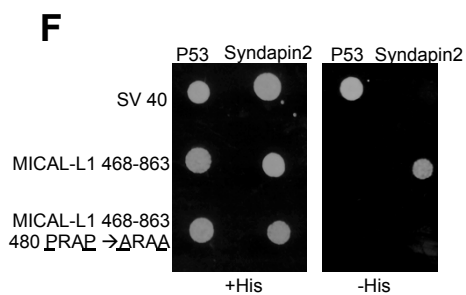
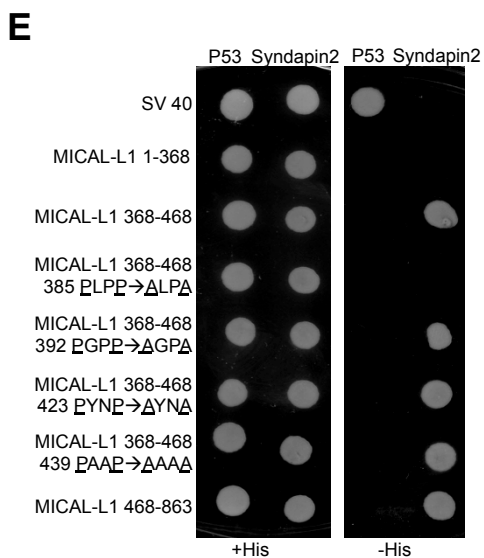
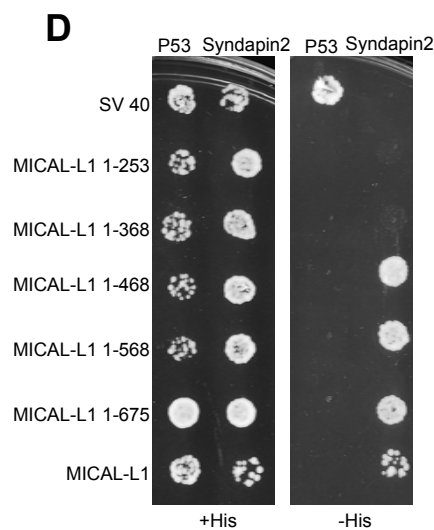
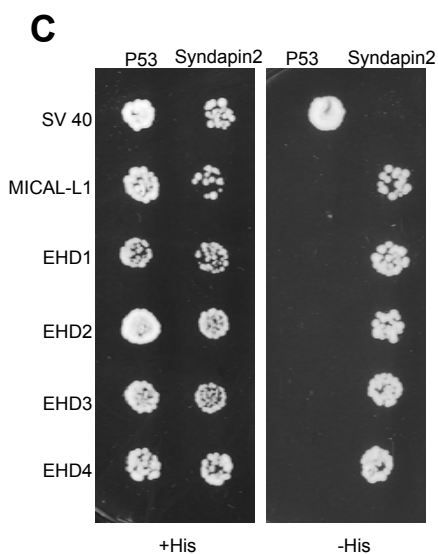
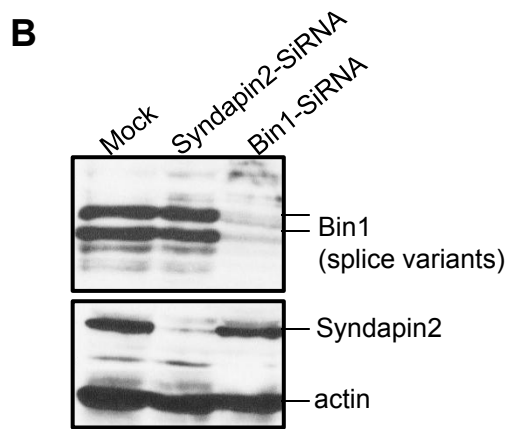
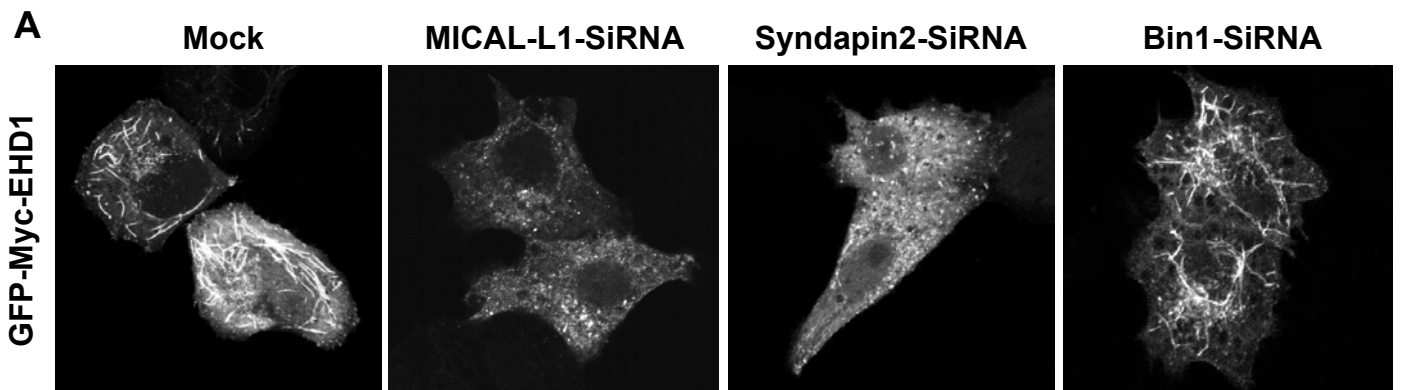
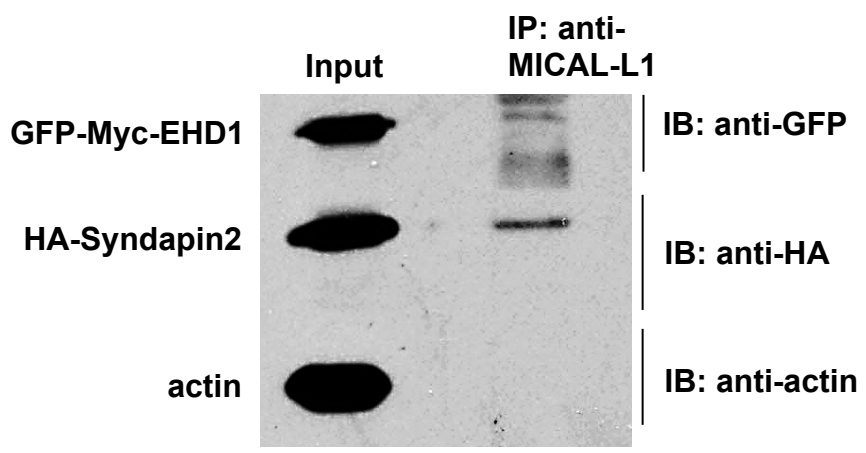


Figure S1

I



J

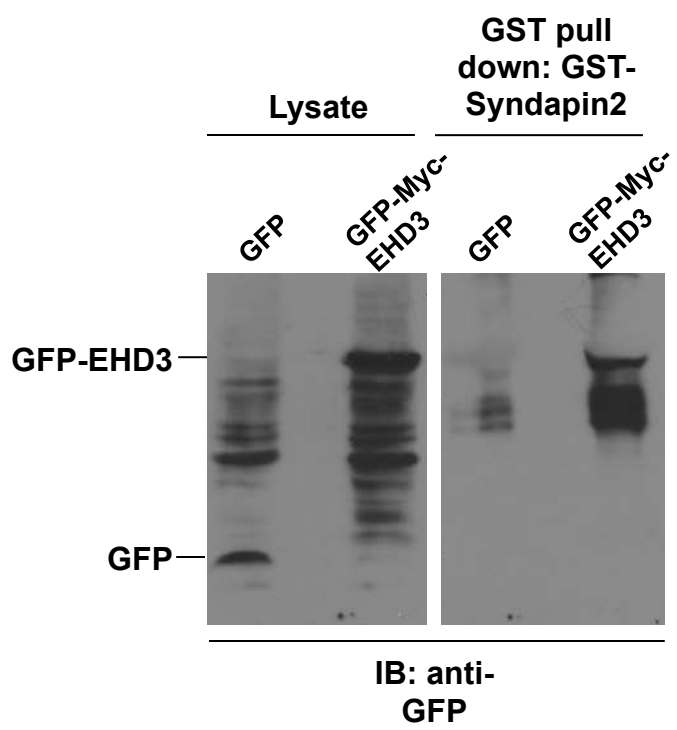


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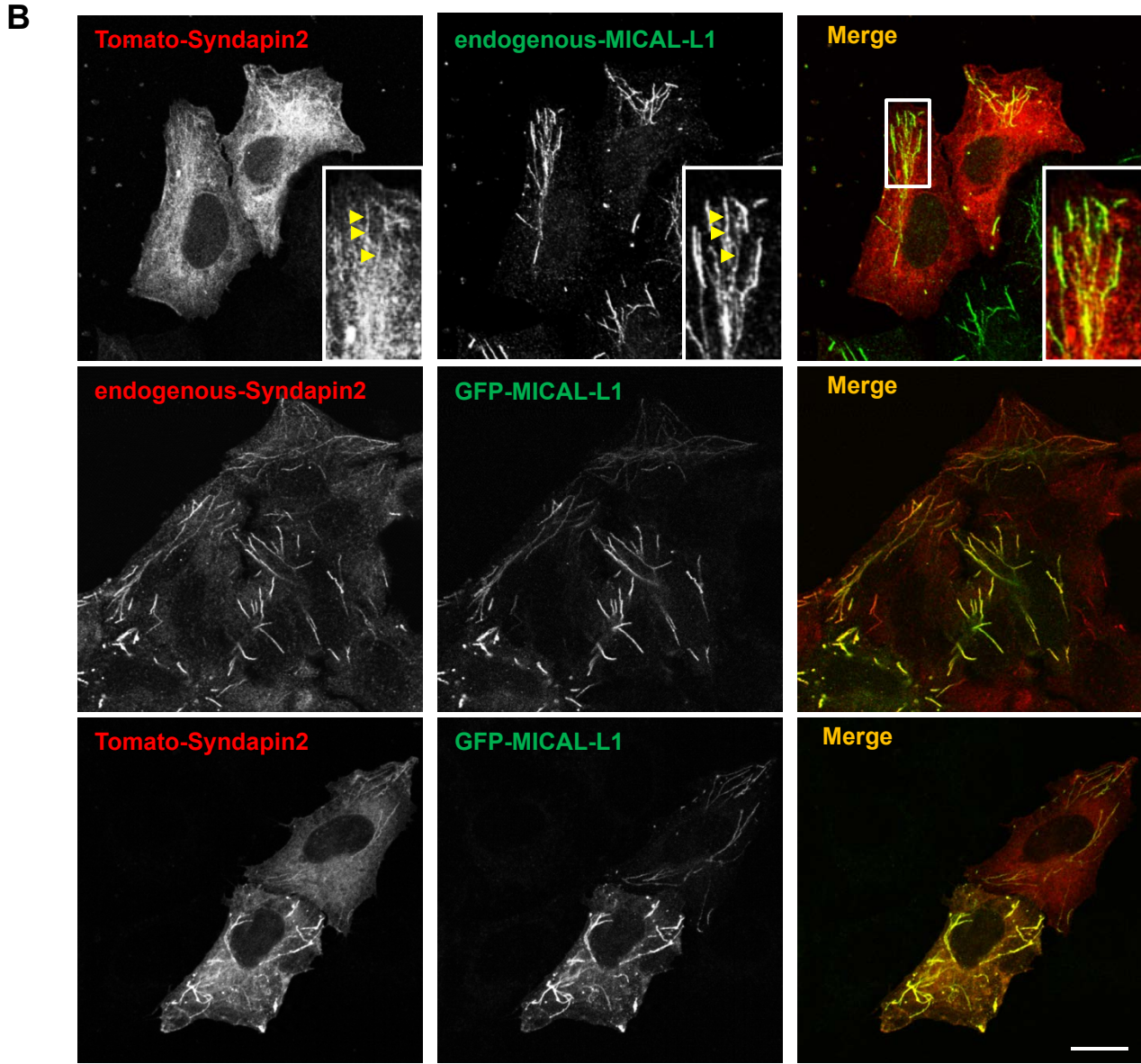
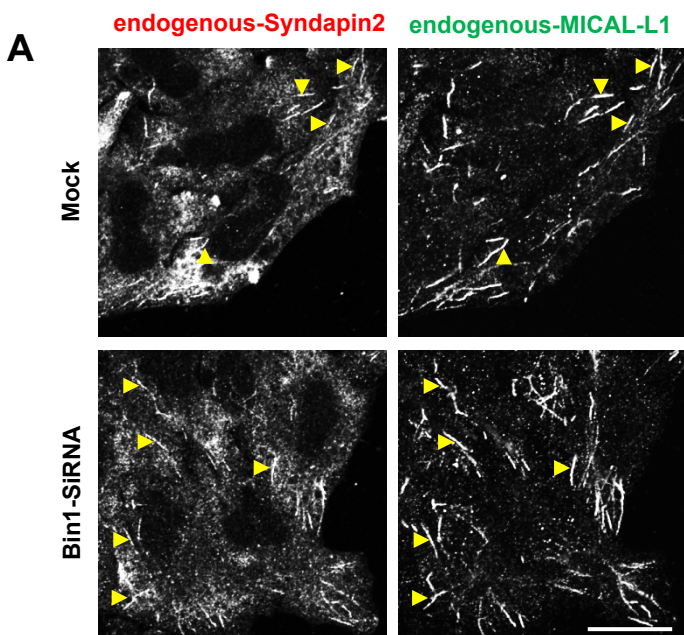


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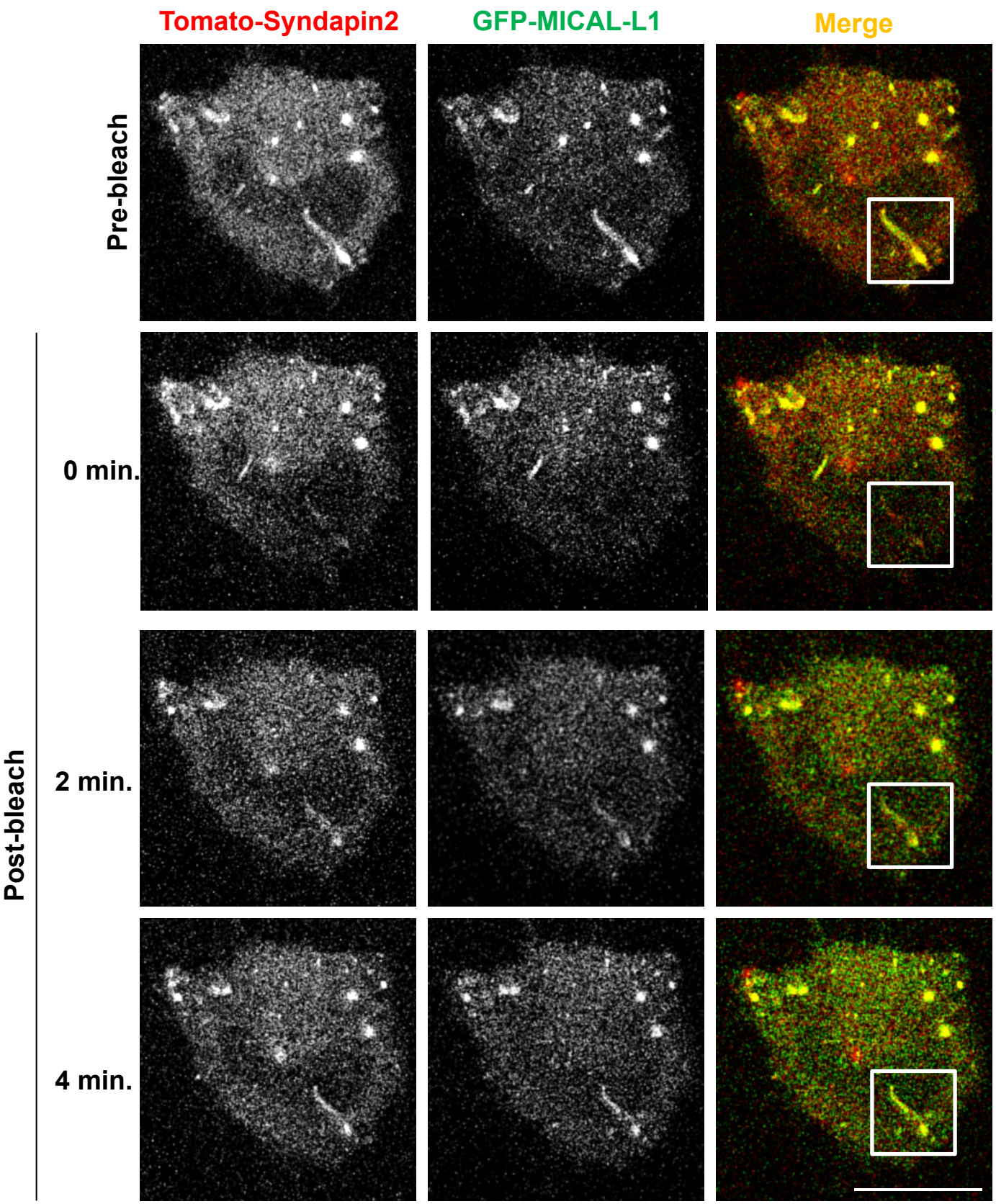


Figure S4A

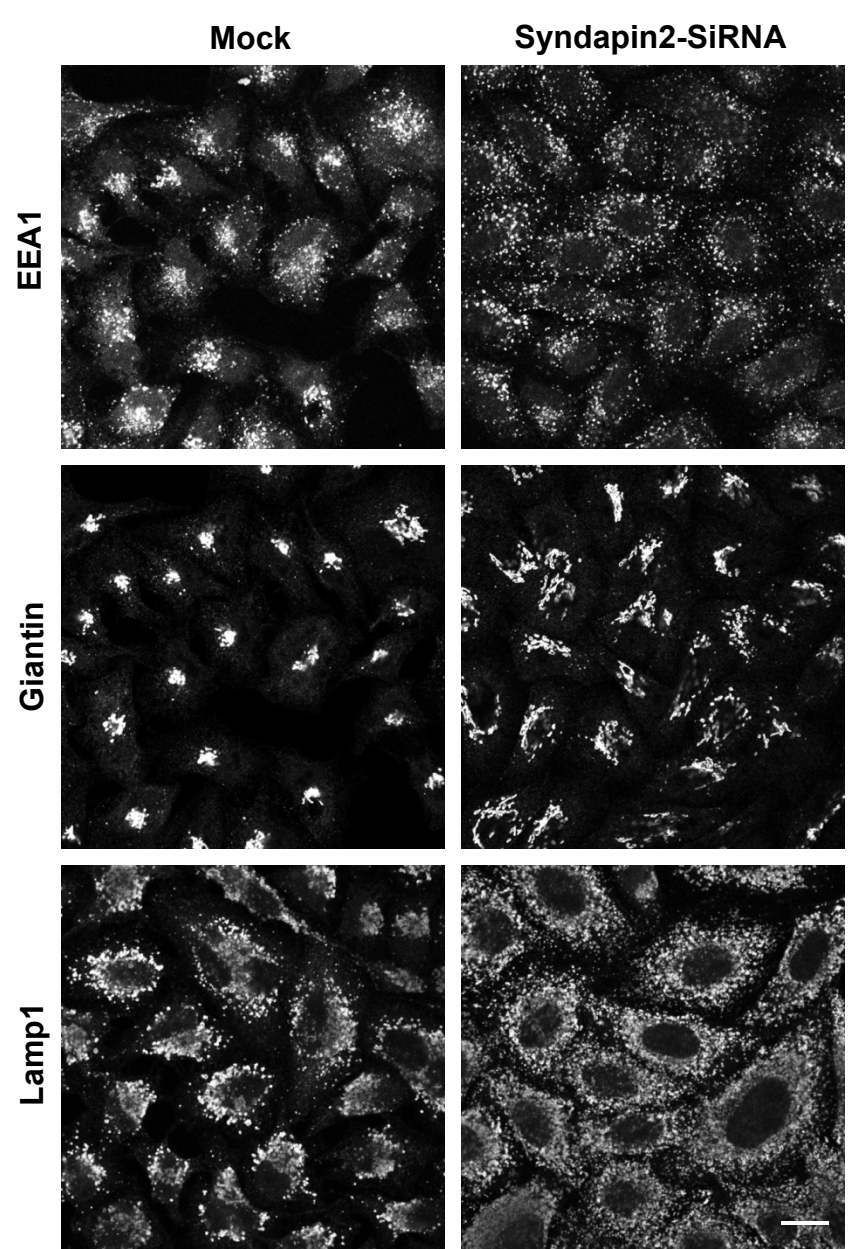


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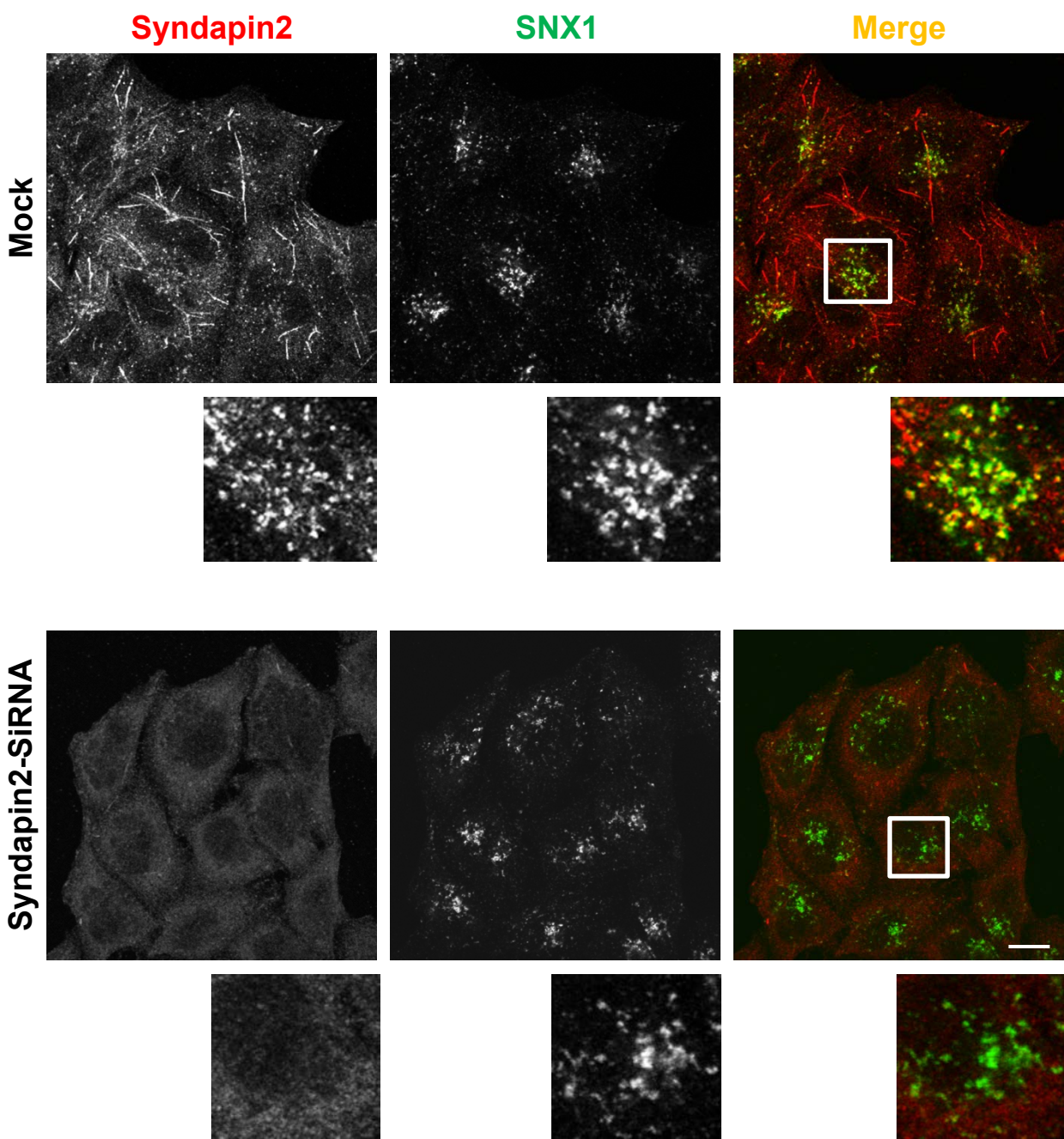


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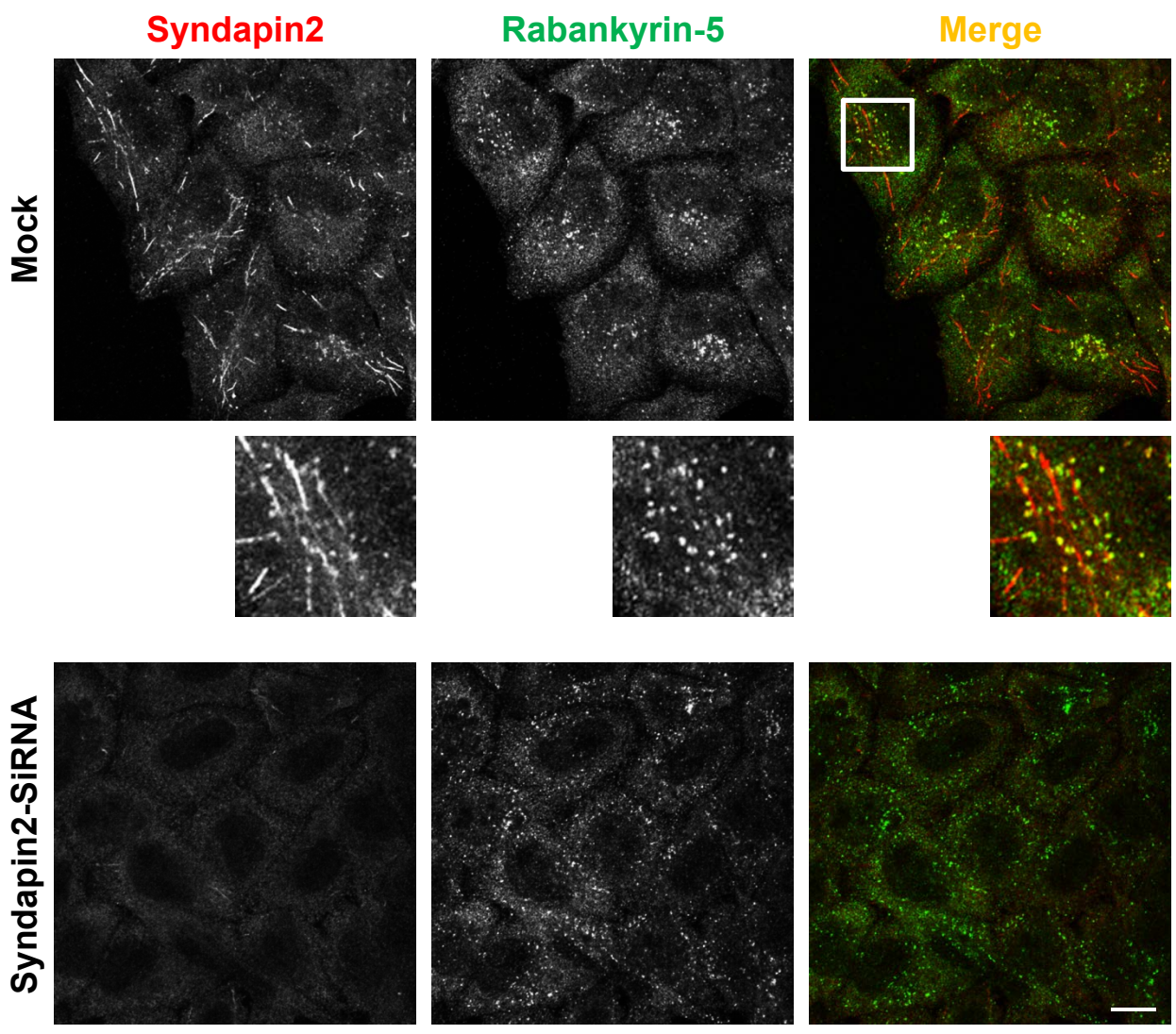


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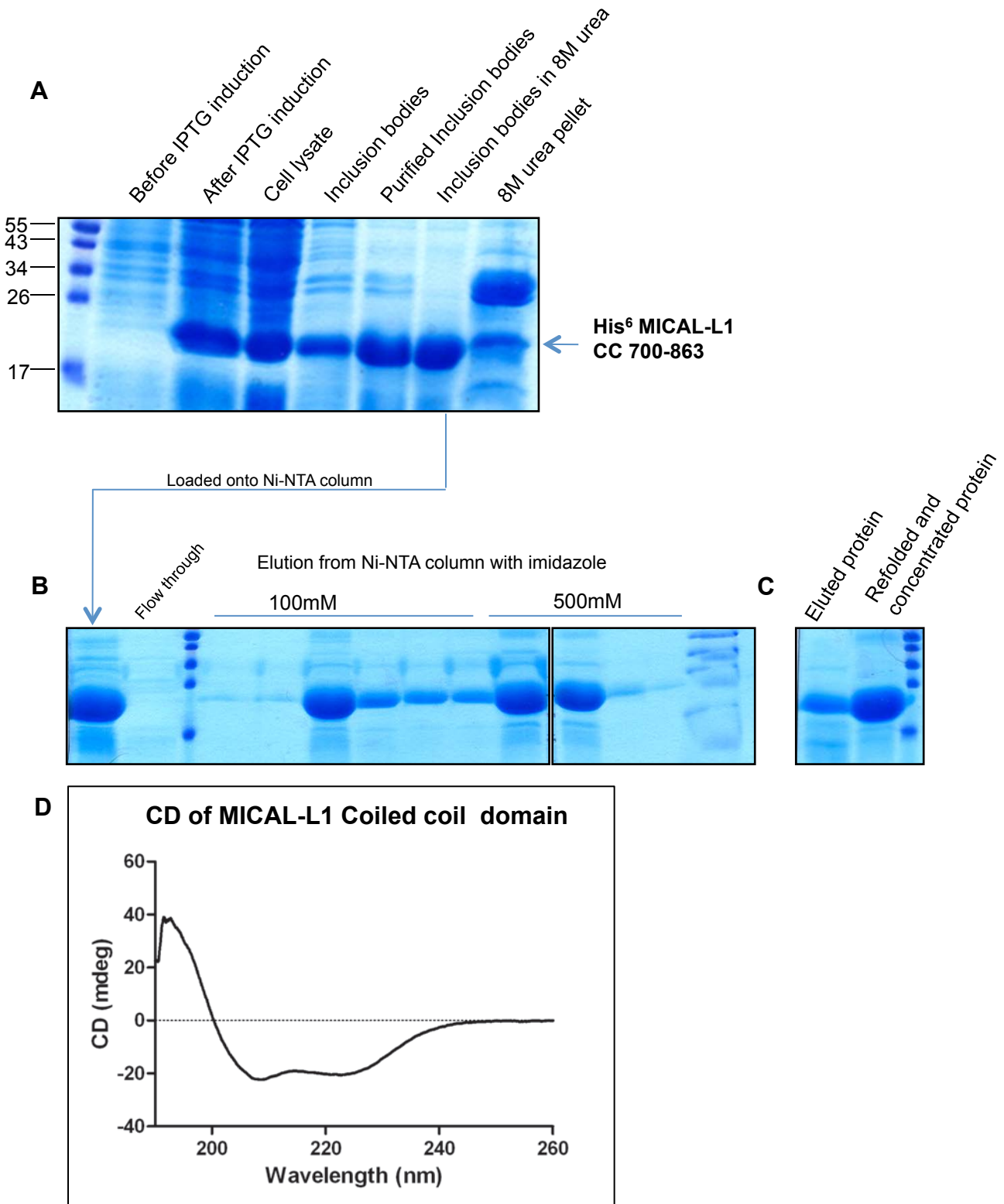
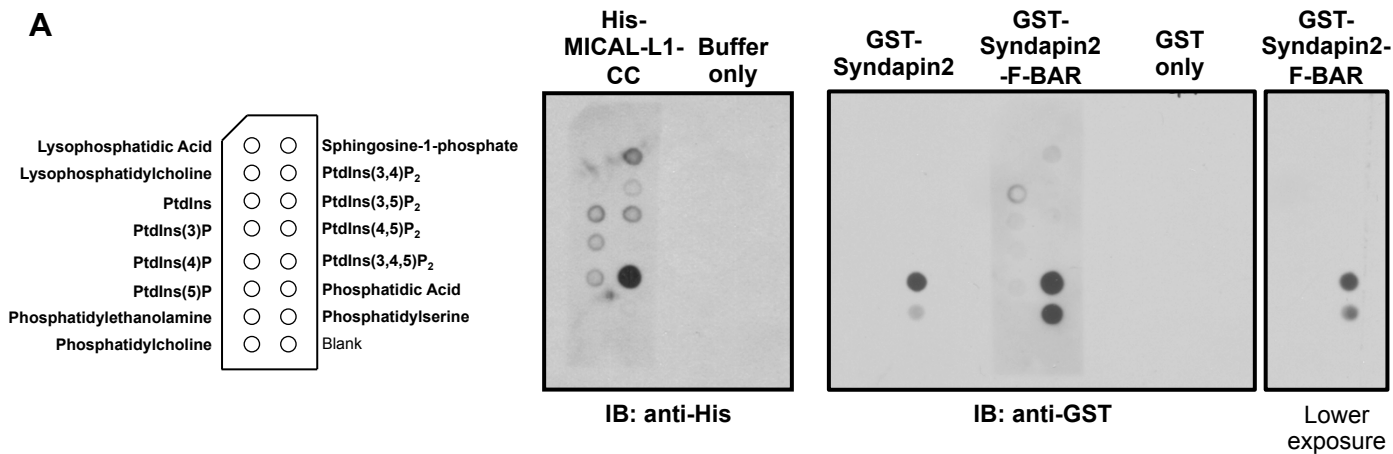
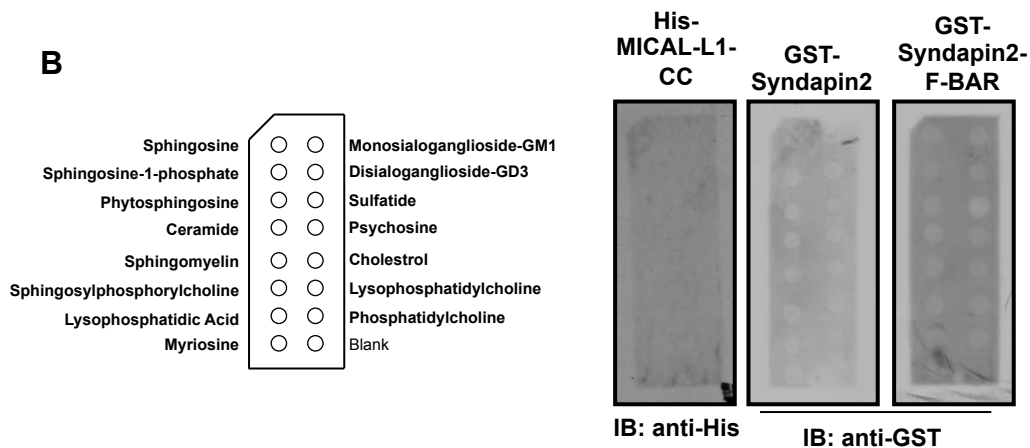


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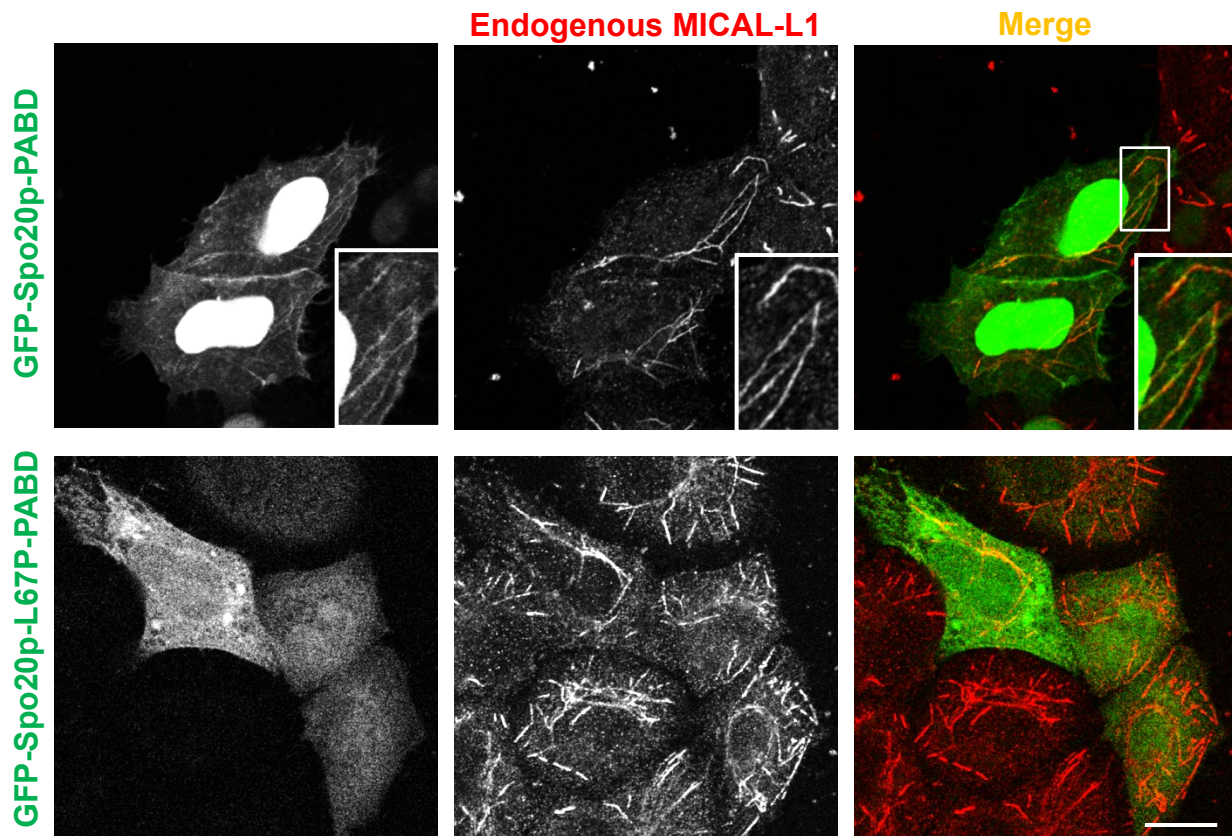
A



B



C



D

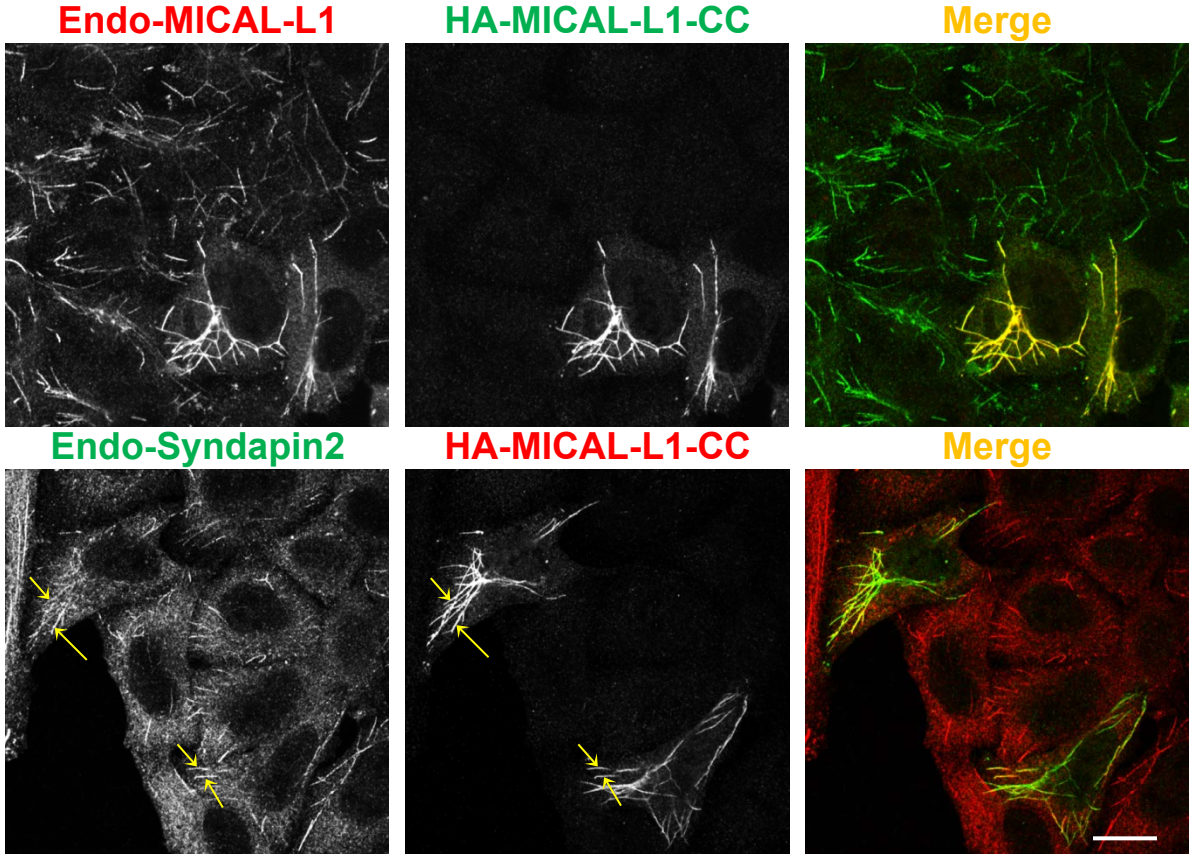


Figure S7

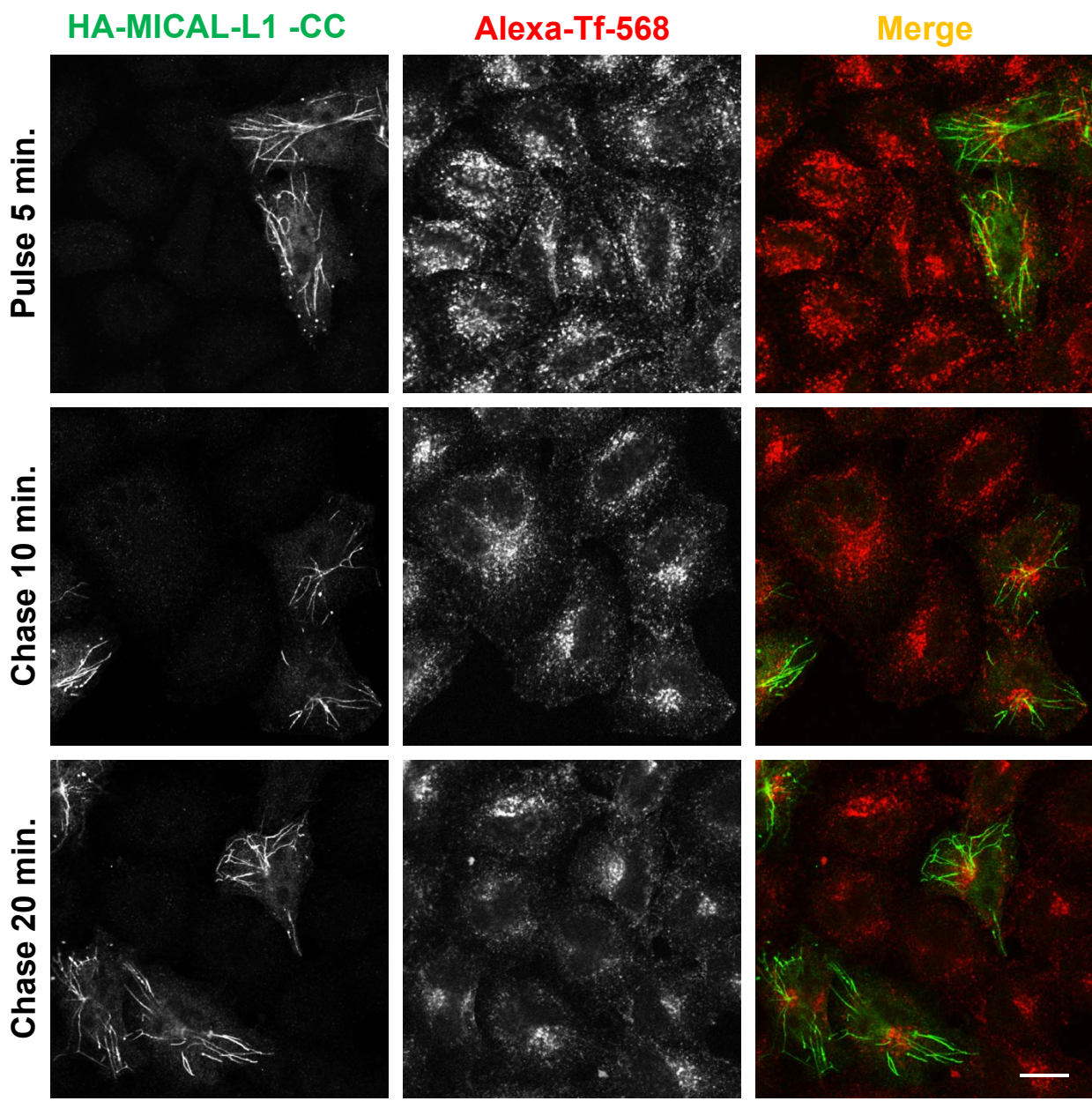


Figure S8

