

# A biomathematical model of human erythropoiesis under erythropoietin and chemotherapy administration

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## A Supplement Material

### A.1 Important Model Variables and Mechanisms

Here, we briefly describe all major model variables and mechanisms of the cell kinetic model. We also provide all equations necessary to run the model. Since the presented model is closely related to a former model of granulopoiesis proposed by our group, details of regulation principles can be found in [1].

### A.2 Cell Kinetic Model

#### Amplification Splitting

Influx and efflux of cells of a compartment were amplified so that the product equals the over-all amplification ( $A_X^{\text{in}}(t) \cdot A_X^{\text{out}}(t) = A_X(t)$ ). The effect is a delayed reaction of efflux and compartment size to changes in amplification rates. It applies to all compartments with amplification, namely BE, CE, and MEB. See [1] for details.

#### Self renewal probability $p$

According to [2], this stem cell quantity is regulated by the demand of the hematopoietic bone marrow system.

$$p = F(C_S^{\text{rel}}(t), C_E^{\text{rel}}(t), C_G^{\text{rel}}(t), p_\delta, \theta_E, \theta_G, \theta_S).$$

The effect of granulopoiesis is assumed to be constant, i.e. for the present model it holds that  $C_G^{\text{rel}}(t) = 1$ . The parameters  $\theta_E$ ,  $\theta_G$ , and  $\theta_S$  are hypothetical weighting factors representing the influence of the com-

**Table A.1.** variables

quantity	meaning	type/calculation
$C_X$	content of compartment X function of time t	
$C_X^{nor}$	content of compartment X in steady state (normal value)	parameter, in general we set $C_X(0) = C_X^{nor}$
$C_X^{rel}$	content of compartments X relative to normal value	$C_X^{rel}(t) = \frac{C_X(t)}{C_X^{nor}}$
$C_X^{in}$	influx in compartment X	function of time
$C_X^{in-nor}$	normal influx	parameter, see above
$C_X^{out}$	efflux from compartment X	function of time
$C_X^{out-nor}$	normal efflux	parameter, see above
$a_X$	proliferative fraction in cell compartment X	function of state, sometimes constant
$A_X$	amplification in cell compartment X	"
$A_X^{in}$	amplification of influx	"
$A_X^{in}$	amplification of efflux	"
$n_X$	average number of mitoses in cell compartment X	$n_X = \ln A_X$
$p$	self-renewal probability of stem cells	function of state
$\tau_X$	average duration of cell cycle in compartment X	function of time, sometimes constant (not regulated)
$T_X$	average transit time of active cells in cell compartment X	$T_X = n_X \tau_X$
$T_X^t$	total transit time	$T_X^t = \frac{n_X \tau_X}{a_X}$
$k$	transition, degradation or toxicity coefficients	functions of time or parameter
$Y^{min}$	quantity Y under minimum stimulation	parameter to determine the regulatory function of Y
$Y^{nor}$	quantity Y in steady state	"
$Y^{int}$	quantity Y under intensified stimulation	"
$Y^{max}$	quantify Y under maximum stimulation	"
$b_Y$	sensitivity of Y under stimulation	"

See also [1] for details.

partsments  $E$ ,  $G$ , and  $S$  [1,2]. According to [2], it is assumed that

$$p_\delta = p^{nor} - p^{min} = p^{max} - p^{nor}$$

$$\theta_S(t) = \begin{cases} \frac{2}{C_S^{rel}(t)^{0.6}} & \text{for } C_S^{rel}(t) \leq 1 \\ 2 & \text{for } C_S^{rel}(t) \geq 1 \end{cases}$$

$$p = p_\delta \tanh\left(-\theta_S(t)(C_S^{rel}(t) - 1) - \theta_E(C_E^{rel}(t) - 1) - \theta_G(C_G^{rel}(t) - 1)\right) + 0.5,$$

assuming  $C_G^{\text{rel}}(t) = 1$  we have

$$p = p_\delta \tanh \left( -\theta_S(t)(C_S^{\text{rel}}(t) - 1) - \theta_E(t)(C_E^{\text{rel}}(t) - 1) \right) + 0.5 .$$

### Proliferative Fraction $a_X$

The proliferative fraction can be interpreted as the percentage of cells which are currently in cell cycle. These quantities are regulated by the bone marrow content.

$$a_X = F(C_S^{\text{rel}}(t), C_E^{\text{rel}}(t), C_G^{\text{rel}}(t), a_X^{\min}, a_X^{\text{nor}}, a_X^{\text{int}}, a_X^{\max}, \omega_E, \omega_G, \omega_S),$$

The parameters  $\omega$  are again weighting factors.

$$x = \omega_E \ln C_E^{\text{rel}}(t) + \omega_G \ln C_G^{\text{rel}}(t) + \omega_S \begin{cases} \ln C_S^{\text{rel}}(t), & \text{for } C_S^{\text{rel}} \leq 1 \\ C_S^{\text{rel}}(t) - 1, & \text{for } C_S^{\text{rel}} > 1 \end{cases} \quad (\text{A.1})$$

$$y = -\frac{1}{2 \ln 2} \left( \ln \left( \frac{a_X^{\text{int}} - a_X^{\max}}{a_X^{\min} - a_X^{\text{int}}} \right) - \ln \left( \frac{a_X^{\text{nor}} - a_X^{\max}}{a_X^{\min} - a_X^{\text{nor}}} \right) \right) x + \frac{1}{2} \ln \left( \frac{a_X^{\text{nor}} - a_X^{\max}}{a_X^{\min} - a_X^{\text{nor}}} \right)$$

$$a_X = \begin{cases} \frac{a_X^{\max} e^{-y} + a_X^{\min} e^y}{e^{-y} + e^y} & \text{for } a_X^{\min} < a_X^{\text{nor}} < a_X^{\text{int}} < a_X^{\max} \\ a_X^{\text{nor}} & \text{for } a_X^{\min} = a_X^{\text{nor}} = a_X^{\text{int}} = a_X^{\max} \end{cases} .$$

Thus, the proliferative fraction is a monotone function ranging between  $a_X^{\min}$  and  $a_X^{\max}$ . Low cell numbers in the bone marrow compartments cause a higher demand of proliferating cells and therefore a larger

proliferative fraction  $a_X$ . With the assumption  $C_G^{\text{rel}}(t) = 1$ , equation A.1 reads

$$x = \omega_E \ln C_E^{\text{rel}}(t) + \omega_S \begin{cases} \ln C_S^{\text{rel}}(t), & \text{for } C_S^{\text{rel}} \leq 1 \\ C_S^{\text{rel}}(t) - 1, & \text{for } C_S^{\text{rel}} > 1 \end{cases}$$

The value  $y$  defines the actual point at the regulatory curve. The variable  $x$  represents some kind of weighted logarithmic relative system size. The proliferative fraction  $a^{\text{int}}$  corresponds to  $x = -\ln 2$  and  $a^{\text{nor}}$  corresponds to  $x = 0$ . See [1] for further details.

### Stem cell compartment S

The stem cell compartment  $S$  has self-renewal capability. Under steady state conditions, 50% of the cells which arise from  $S$  remain in this compartment, the others feed the BE compartment.

$$\frac{d}{dt} C_S = (2p - 1)C_S \frac{a_S}{\tau_S} - \Psi_S \cdot C_S \quad (\text{A.2})$$

$$C_S^{\text{out}} = 2(1 - p)C_S \frac{a_S}{\tau_S} \quad (\text{A.3})$$

where  $\Psi_S$  is the summarized chemotherapy function. It holds that  $p^{\text{nor}} = \frac{1}{2}$ . Thus, for the initial conditions it holds that

$$C_S(0) = C_S^{\text{nor}} = 1 \quad (\text{A.4})$$

$$C_S^{\text{out}}(0) = C_S^{\text{out,nor}} = 2(1 - p^{\text{nor}})C_S^{\text{nor}} \frac{a_S^{\text{nor}}}{\tau_S}. \quad (\text{A.5})$$

### Compartment BE

$$\begin{aligned} \frac{d}{dt} C_{\text{BE}} &= \alpha_E C_S^{\text{out}} A_{\text{BE}}^{\text{in}} - C_{\text{BE}} \frac{a_{\text{BE}}}{\tau_{\text{BE}}} - \Psi_{\text{BE}} \cdot C_{\text{BE}} \\ C_{\text{BE}}^{\text{out}} &= C_{\text{BE}} A_{\text{BE}}^{\text{out}} \frac{a_{\text{BE}}}{T_{\text{BE}}} \end{aligned}$$

with the initial conditions

$$\begin{aligned}
C_{BE}(0) &= C_{BE}^{\text{nor}} = \alpha_E C_S^{\text{out-nor}} A_{BE}^{\text{in-nor}} \frac{T_{BE}^{\text{nor}}}{a_{BE}^{\text{nor}}} \\
C_{BE}^{\text{out-nor}} &= C_{BE}^{\text{nor}} A_{BE}^{\text{out-nor}} \frac{a_{BE}^{\text{nor}}}{T_{BE}^{\text{nor}}} \\
&= \alpha_E C_S^{\text{out-nor}} A_{BE}^{\text{nor}}.
\end{aligned}$$

### Compartment CE

$$\begin{aligned}
A_{CE} &= Z(C_{EPO}^{\text{rel}}) \\
\frac{d}{dt} C_{CE} &= C_{BE}^{\text{out}} A_{CE}^{\text{in}} - C_{CE} \frac{a_{CE}}{T_{CE}} - \Psi_{CE} \cdot C_{CE} \\
C_{CE}^{\text{out}} &= C_{CE} A_{CE}^{\text{out}} \frac{a_{CE}}{T_{CE}}.
\end{aligned}$$

We assume  $a_{CE} = 1$ . Thus,

$$\begin{aligned}
C_{CE}(0) &= C_{CE}^{\text{nor}} = C_{BE}^{\text{out-nor}} A_{CE}^{\text{in-nor}} T_{CE}^{\text{nor}} \\
C_{CE}^{\text{out}}(0) &= C_{CE}^{\text{out-nor}} = C_{BE}^{\text{out-nor}} A_{CE}^{\text{nor}}.
\end{aligned}$$

### Compartment PEB

$$\begin{aligned}
A_{PEB} &= Z(C_{EPO}^{\text{rel}}) \\
\frac{d}{dt} C_{PEB} &= C_{CE}^{\text{out}} A_{PEB}^{\text{in}} - C_{PEB} \frac{a_{PEB}}{T_{PEB}} - \Psi_{PEB} \cdot C_{PEB} \\
C_{PEB}^{\text{out}} &= C_{PEB} A_{PEB}^{\text{out}} \frac{a_{PEB}}{T_{PEB}}.
\end{aligned}$$

We assume  $a_{PEB} = 1$ . Thus,

$$\begin{aligned}
C_{PEB}(0) &= C_{PEB}^{\text{nor}} = C_{CE}^{\text{out-nor}} A_{PEB}^{\text{in-nor}} \frac{T_{PEB}^{\text{nor}}}{a_{PEB}^{\text{nor}}} \\
C_{PEB}^{\text{out}}(0) &= C_{PEB}^{\text{out-nor}} = C_{CE}^{\text{out-nor}} A_{PEB}^{\text{nor}}.
\end{aligned}$$

## Compartment MEB

The maturation is modeled by splitting MEB into  $N_{\text{MEB}} = 15$  subcompartments without amplification.

$$\begin{aligned}
 T_{\text{MEB}} &= Z(C_{EPO}^{\text{rel}}) \\
 C_{\text{MEB}} &= \sum_{i=1}^{N_{\text{MEB}}} C_{\text{MEB}_i} \\
 \frac{d}{dt} C_{\text{MEB}_1} &= C_{\text{PEB}}^{\text{out}} - C_{\text{MEB}_1} \frac{N_{\text{MEB}}}{T_{\text{MEB}}} - \Psi_{\text{MEB}} \cdot C_{\text{MEB}_1} \\
 \frac{d}{dt} C_{\text{MEB}_i} &= C_{\text{MEB}_{i-1}}^{\text{out}} - C_{\text{MEB}_i} \frac{N_{\text{MEB}}}{T_{\text{MEB}}} - \Psi_{\text{MEB}} \cdot C_{\text{MEB}_i}, \quad i = 2, \dots, N_{\text{MEB}} \\
 C_{\text{MEB}_i}^{\text{out}} &= C_{\text{MEB}_i} \frac{N_{\text{MEB}}}{T_{\text{MEB}}}, \quad i = 1, \dots, N_{\text{MEB}} \\
 C_{\text{MEB}}^{\text{out}} &= C_{\text{MEB}_{N_{\text{MEB}}}}^{\text{out}},
 \end{aligned}$$

with the initial values

$$\begin{aligned}
 C_{\text{MEB}}(0) &= C_{\text{MEB}}^{\text{nor}} = C_{\text{PEB}}^{\text{out,nor}} T_{\text{MEB}}^{\text{nor}} \\
 C_{\text{MEB}_i}(0) &= C_{\text{MEB}_i}^{\text{nor}} = C_{\text{PEB}}^{\text{out,nor}} \frac{T_{\text{MEB}}^{\text{nor}}}{N_{\text{MEB}}}, \quad i = 1, \dots, N_{\text{MEB}} \\
 C_{\text{MEB}_i}^{\text{out}}(0) &= C_{\text{MEB}_i}^{\text{out,nor}} = C_{\text{MEB}_i}^{\text{nor}} \frac{N_{\text{MEB}}}{T_{\text{MEB}}^{\text{nor}}} = C_{\text{PEB}}^{\text{out,nor}}, \quad i = 1, \dots, N_{\text{MEB}} \\
 C_{\text{MEB}}^{\text{out}}(0) &= C_{\text{MEB}}^{\text{out,nor}} = C_{\text{MEB}_{N_{\text{MEB}}}}^{\text{out,nor}} = C_{\text{PEB}}^{\text{out,nor}}.
 \end{aligned}$$

## Compartment RET

$$\begin{aligned}
 T_{\text{RET}} &= T_{\text{MEB}}^{\text{nor}} + T_{\text{RET}}^{\text{nor}} - T_{\text{MEB}} \\
 \frac{d}{dt} C_{\text{RET}} &= C_{\text{MEB}}^{\text{out}} - \frac{C_{\text{RET}}}{T_{\text{RET}}} \\
 C_{\text{RET}}^{\text{out}} &= \frac{C_{\text{RET}}}{T_{\text{RET}}} \\
 C_{\text{RET}}(0) &= C_{\text{RET}}^{\text{nor}} = C_{\text{ERY}}^{\text{nor}} \frac{q_{\text{RET}}}{1 - q_{\text{RET}}} \\
 C_{\text{RET}}^{\text{out}}(0) &= C_{\text{RET}}^{\text{out,nor}} = C_{\text{MEB}}^{\text{out,nor}} \\
 T_{\text{RET}}^{\text{nor}} &= \frac{C_{\text{RET}}^{\text{nor}}}{C_{\text{RET}}^{\text{out,nor}}} = \frac{q_{\text{RET}}}{1 - q_{\text{RET}}} ((1 - s_{\text{ERY}}^{\text{nor}}) T_{\text{ERY\_rnd}} + s_{\text{ERY}}^{\text{nor}} T_{\text{ERY\_age}}).
 \end{aligned}$$

$q_{\text{RET}}$  is the ratio of reticulocytes to the total number of red blood cells in steady state.  $s_{\text{ERY}}^{\text{nor}}$ ,  $T_{\text{ERY\_rnd}}$ , and  $T_{\text{ERY\_age}}$  are explained in the next section.

### Compartment ERY

The compartment ERY is split into the subcompartments "RANDOM" and "AGE". In steady state, most erythrocytes die dependent on age. The age dependent reduction is modeled by division into subcompartments. Under stimulation, the apoptosis is more randomly (see [3]). To model this observation, the influxes into the subcompartments "RANDOM" and "AGE" are regulated by the factor  $s_{\text{ERY}}$ , which depends on the bone marrow output of the reticulocytes.  $T_{\text{ERY\_rnd}}$ , and  $T_{\text{ERY\_age}}$  are the corresponding transition times of these compartments (see [1–4] for details.)

$$s_{\text{ERY}} = \exp \left( \left( \frac{C_{\text{RET}}^{\text{out}}}{C_{\text{RET}}^{\text{out,nor}}} \right)^2 \ln s_{\text{ERY}}^{\text{nor}} \right)$$

$$C_{\text{ERY}} = C_{\text{ERY\_age}} + C_{\text{ERY\_rnd}}$$

$$C_{\text{ERY\_age}} = \sum_{i=1}^{N_{\text{ERY}}} C_{\text{ERY\_age\_}i}$$

$$\frac{d}{dt} C_{\text{ERY\_age\_}1} = s_{\text{ERY}} C_{\text{RET}}^{\text{out}} - C_{\text{ERY\_age\_}1} \frac{N_{\text{ERY}}}{T_{\text{ERY\_age}}}$$

$$\frac{d}{dt} C_{\text{ERY\_age\_}i} = C_{\text{ERY\_age\_}(i-1)}^{\text{out}} - C_{\text{ERY\_age\_}i}^{\text{out}}, \quad i = 2, \dots, N_{\text{ERY}}$$

$$C_{\text{ERY\_age\_}i}^{\text{out}} = C_{\text{ERY\_age\_}i} \frac{N_{\text{ERY}}}{T_{\text{ERY\_age}}}$$

$$\frac{d}{dt} C_{\text{ERY\_rnd}} = (1 - s_{\text{ERY}}) C_{\text{RET}}^{\text{out}} - C_{\text{ERY\_rnd}} \frac{1}{T_{\text{ERY\_rnd}}},$$

with initial conditions

$$C_{\text{ERY}}(0) = C_{\text{ERY}}^{\text{nor}} = C_{\text{ERY\_age}}^{\text{nor}} + C_{\text{ERY\_rnd}}^{\text{nor}}$$

$$C_{\text{ERY\_age}}(0) = C_{\text{ERY\_age}}^{\text{nor}} = \sum_{i=1}^{N_{\text{ERY}}} C_{\text{ERY\_age\_}i}^{\text{nor}} = s_{\text{ERY}}^{\text{nor}} C_{\text{RET}}^{\text{out,nor}} T_{\text{ERY\_age}}$$

$$C_{\text{ERY\_age\_}1}(0) = C_{\text{ERY\_age\_}1}^{\text{nor}} = s_{\text{ERY}}^{\text{nor}} C_{\text{RET}}^{\text{out,nor}} \frac{T_{\text{ERY\_age}}}{N_{\text{ERY}}}$$

$$\begin{aligned}
C_{\text{ERY\_age}_i}(0) &= C_{\text{ERY\_age}_i}^{\text{nor}} = C_{\text{ERY\_age}_{i-1}}^{\text{out,nor}} \frac{T_{\text{ERY\_age}}}{N_{\text{ERY}}}, \quad i = 2, \dots, N_{\text{ERY}} \\
&= s_{\text{ERY}}^{\text{nor}} C_{\text{RET}}^{\text{out,nor}} \frac{T_{\text{ERY\_age}}}{N_{\text{ERY}}}, \quad i = 1, \dots, N_{\text{ERY}} \\
C_{\text{ERY\_age}_i}^{\text{out}}(0) &= C_{\text{ERY\_age}_i}^{\text{out,nor}} = C_{\text{ERY\_age}_i}^{\text{nor}} \frac{N_{\text{ERY}}}{T_{\text{ERY\_age}}} = s_{\text{ERY}}^{\text{nor}} C_{\text{RET}}^{\text{out,nor}} \\
C_{\text{ERY\_rnd}}(0) &= C_{\text{ERY\_rnd}}^{\text{nor}} = (1 - s_{\text{ERY}}^{\text{nor}}) C_{\text{RET}}^{\text{out,nor}} T_{\text{ERY\_rnd}}
\end{aligned}$$

### Endogenous production of EPO

The endogenous production of EPO ( $\text{EPO}_{\text{prod}}$ ) is assumed to be dependent on the tissue oxygen tension in the kidneys and the number of circulating red blood cells (see [4, 5]). Pantel [4] and Wichmann [5] proposed the following model of this process.

**Table A.2.** Variables for endogenous EPO production

quantity	meaning	type/calculation	
$P_{O_2}^t$	tissue oxygen tension in kidneys	function of time	[4, 5]
$P_{O_2}^{t,\text{nor}}$	normal value of tissue oxygen tension in kidneys		[4, 5]
$S_{O_2}^t$	tissue saturation of oxygen	function of time	[4, 5]
$S_{O_2}^{t,\text{nor}}$	normal tissue saturation of oxygen	constant	[4, 5]
$P_{50}$	partial oxygen pressure corresponding to $S_{O_2}^t = 50\%$	26.5 mm Hg	[4, 5]
$P_{O_2}^{A,\text{nor}}$	arterial oxygen tension, normal value	97 mm Hg	[4, 5]
$\Delta SO_2$	desaturation of HB (arteriovenous difference), normal value	20 %	[4, 5]
$\gamma$	Hill coefficient	2.65	[4, 5]
$P_{\text{endo}}^{\max}$	maximum EPO production	200 (set)	[4, 5]
$b_{\text{EPO}}$	sensitivity of EPO production to changes in $P_{O_2}^t$	$\ln 200$ (set)	[4, 5]

$$P_{O_2}^t = P_{50} \cdot \left( \frac{S_{O_2}^t}{100 - S_{O_2}^t} \right)^{\frac{1}{\gamma}} \text{(Hill equation)} \quad (\text{A.6})$$

$$S_{O_2}^t = \frac{100}{\left( \frac{P_{50}}{P_{O_2}^{A,\text{nor}}} \right)^{\gamma} + 1} - \Delta SO_2 \cdot \frac{RET^{\text{nor}} + ERY^{\text{nor}}}{C_{\text{RET}} + C_{\text{ERY}}} \quad (\text{A.7})$$

$$S_{O_2}^{t,\text{nor}} = \frac{100}{\left( \frac{P_{50}}{P_{O_2}^{A,\text{nor}}} \right)^{\gamma} + 1} - \Delta SO_2 \quad (\text{A.8})$$

$$P_{O_2}^{t,\text{nor}} = P_{50} \cdot \left( \frac{S_{O_2}^{t,\text{nor}}}{100 - S_{O_2}^{t,\text{nor}}} \right)^{\frac{1}{\gamma}} \quad (\text{A.9})$$

Define  $f = \frac{P_{O_2}^t}{P_{O_2}^{t_{\text{nor}}}}$ , we assume

$$\begin{aligned} \text{EPO}_{\text{prod}} &= P_{\max}^{\text{endo}} \cdot e^{-b_{\text{EPO}} \cdot f} & [4] \\ \text{EPO}_{\text{prod}}(0) &= 1. \end{aligned} \quad (\text{A.10})$$

For further explanations and justifications, see [4, 5].

### A.3 List of Model Parameters

parameter	value			
$q_{\text{RET}}$	0.016	set		
$T_{\text{rnd}}$	1020.4	h	set	[6]
$T_{\text{ERY}}^{\text{age}}$	3061.2	h	set	[6]
$s_{\text{ERY}}^{\text{nor}}$	0.900		set	[4], p. 40
$N_{\text{ERY}}$	10.0		set	[4], p. 41
$\text{HCT}^{\text{nor}}$	0.430		set	
$\text{ERY}^{\text{nor}}$	4.50		set	
$\text{RET}^{\text{nor}}$	100	x1000/ $\mu\text{l}$	set	
$\text{RET\%}^{\text{nor}}$	9.50		set	
$\text{HB}^{\text{nor}}$	13.5		set	
$P_{\max}^{\text{endo}}$	200		set	[4]
$b_{\text{EPO}}$	5.30		fitted	
$\text{EPO}_{\text{Vc}}$	0.0320	l/kg	set	[7]
$\text{EPO}_{\text{serum}}$	15	IU/l	set	[7]
$a_{\text{BE}}^{\min}$	0.30		set	[2], p. 71
$a_{\text{BE}}^{\text{nor}}$	0.33		set	[2], p. 71
$a_{\text{BE}}^{\text{int}}$	0.66		set	[2], p. 71
$a_{\text{BE}}^{\max}$	1.00		set	[2], p. 71
$\alpha_G$	0.8		set	[2], p. 73
$\alpha_E$	0.15		set	[2], p. 73
$S^{\text{nor}}$	1		set	
$\tau_S$	8		set	[2], p. 70
$a_S^{\min}$	0.01		set	[2], p. 70
$p_\delta$	0.1		set	[2], p. 70
$a_S^{\text{nor}}$	0.15		set	[2], p. 70
$a_S^{\text{int}}$	0.45		set	[2], p. 70
$a_S^{\max}$	1		set	[2], p. 70
$w_E$	0.3		set	[2], p. 70
$w_G$	0.1		set	[2], p. 70
$w_S$	1		set	[2], p. 70
$\vartheta_E$	-2		set	[2], p. 70
$\vartheta_G$	-8		set	[2], p. 70

**Table A.3.** EPO PK/PD parameters

	Alfa, Beta, Delta, endogenous EPO	Darbepoetin Alfa
$k_{el}$	0.102	fitted
$k_{12}$	0.079	fitted
$k_{21}$	0.084	fitted
$k_{on}$	0.070	set [7]
$k_{off}$	14.27	set [7]
$R_0$	64.31	set [7]
$k_{int}$	2	set [7]
$k_{deg}$	0.101	set [7]
$w_{RET}$	0.05	set
$w_{MEB}$	0.087	fitted
$w_{PEB}$	0.293	fitted
$w_{CE}$	3.84	fitted
$w_{BE}$	0.0881	fitted
$k_{on}/k_{off}$	0.004875	0.004453
$T_{BE}^{min}$	155.7	h fitted
$T_{BE}^{nor}$	40	h set [6]
$T_{BE}^{max}$	28.60	h fitted
$T_{BE}^b$	1.134	fitted
$A_{BE}^{min}$	25.04	fitted
$A_{BE}^{nor}$	64	set [6]
$A_{BE}^{max}$	194.7	fitted
$A_{BE}^b$	2.321	fitted
$A_{CE}^{min}$	0.9645	fitted
$A_{CE}^{nor}$	32	set [6]
$A_{CE}^{max}$	104.7	fitted
$A_{CE}^b$	0.0438	fitted
$T_{CE}^{min}$	186.7	h fitted
$T_{CE}^{nor}$	40	h set [6]
$T_{CE}^{max}$	15.25	h fitted
$T_{CE}^b$	0.3920	fitted
$T_{PEB}^{min}$	99.25	h fitted
$T_{PEB}^{nor}$	48	h set
$T_{PEB}^{max}$	8.809	h fitted
$T_{PEB}^b$	1.301	fitted
$A_{PEB}^{min}$	0.6862	fitted
$A_{PEB}^{nor}$	64	set [6]
$A_{PEB}^{max}$	75.38	fitted
$A_{PEB}^b$	0.4135	fitted
$T_{MEB}^{min}$	144.8	h fitted
$T_{MEB}^{nor}$	100.2	h fitted
$T_{MEB}^{max}$	90.17	h fitted
$T_{MEB}^b$	0.5395	fitted

**Table A.4.** EPO absorption parameters after subcutaneous injection (fitted)

	Alfa, abdomen, upper arm	Alfa, fore- arm	Alfa, shoulder	Alfa, thigh	Beta, abdomen	Beta, thigh	Beta, fore- arm	Delta	Darb- epoetin Alfa
$k_a^F$	0.5320	0.7786	0.7119	0.7374	0.3747	0.4528	0.4077	0.6657	3.0148
$k_e^F$	0.2713	0.1337	0.1263	0.3295	0.1062	0.0656	0.1873	0.1517	0.1938
$k_{Delay}^L$	0.0390	0.0510	0.1460	0.0298	0.0699	0.0410	0.0476	0.2367	0.0241
$k_a^L$	0.1172	0.0901	0.1326	0.1588	0.2309	0.2024	0.4687	0.1065	1.1192
$k_e^L$	0.4334	0.4904	0.4066	0.4343	0.0950	0.0557	0.1587	0.3321	0.1769
$k_{Delay}^F$	0.3275	0.3259	0.6029	0.1626	0.3705	0.2078	0.2819	0.7508	0.1161
$k_{FL}$	1.0074	3.6252	3.9596	0.7213	1.2138	1.0453	1.1987	3.4461	5.5404

**Table A.5.** Toxicity parameters of Chemotherapies

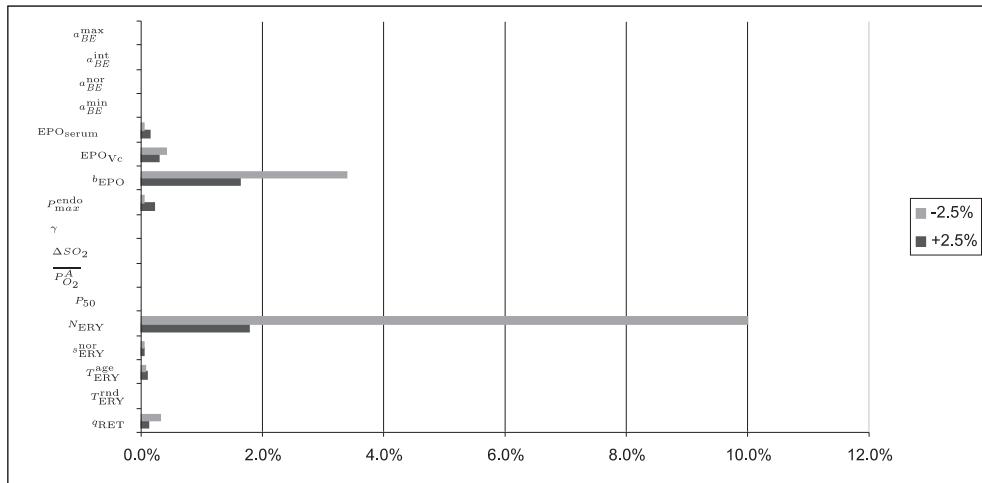
drug or drug combination	Therapy	FC	Delay	S	BE	CE	PEB	MEB	RET
Cyclophosphamid 650 mg/m <sup>2</sup>	BEACOPP	1.111	0.064	0.001	0.000	0.000	0.005	0.000	0.002
Doxorubicin 25 mg/m <sup>2</sup>									
Cyclophosphamid 750 mg/m <sup>2</sup>	CHOP	1.111	0.064	0.216	0.000	0.017	0.006	0.117	0.036
Doxorubicin 50 mg/m <sup>2</sup>									
Cyclophosphamid 1250 mg/m <sup>2</sup>	BEACOPP escalated	1.111	0.064	0.212	0.001	0.001	0.016	0.030	0.021
Doxorubicin 35 mg/m <sup>2</sup>									
Cyclophosphamid 1400 mg/m <sup>2</sup>	high CHOEP	1.111	0.064	0.194	0.002	0.021	0.063	0.117	0.067
Doxorubicin 32.5 mg/m <sup>2</sup>									
Etoposid 100 mg/m <sup>2</sup>	CHOEP	1.097	0.068	0.000	0.000	0.000	0.041	0.000	0.011
Etoposid 200 mg/m <sup>2</sup>	BEACOPP escalated	1.097	0.068	0.000	0.000	0.000	0.057	0.013	0.020
Etoposid 175 mg/m <sup>2</sup>	high CHOEP	1.097	0.068	0.024	0.001	0.000	0.044	0.000	0.026
Procarbazine 100 mg/m <sup>2</sup>	BEACOPP escalated, BEACOPP	1.092	0.013	0.002	0.011	0.004	0.013	0.000	0.000
Bleomycin 10 mg/m <sup>2</sup>	BEACOPP escalated, BEACOPP	1.323	0.003	0.012	0.010	0.011	0.001	0.030	0.000
Platinum Etoposide	Platinum + Etoposide	1.000	1.219	0.338	0.011	0.002	0.008	0.007	0.000
Paclitaxel 225 mg/m <sup>2</sup>	ETC	1.050	0.017	0.246	0.000	0.436	1.748	0.000	0.096
Cyclophosphamid 2500 mg/m <sup>2</sup>	ETC	1.005	0.064	0.199	0.033	0.056	0.059	0.000	0.267
Epirubicin 150 mg/m <sup>2</sup>	ETC	1.988	0.045	0.003	0.005	1.506	2.909	0.022	3.273
Cyclophosphamid 600 mg/m <sup>2</sup>	ECT	1.005	0.064	0.199	0.003	0.008	0.016	0.000	0.045
Epirubicin 90 mg/m <sup>2</sup>	ECT	1.988	0.045	0.000	0.000	0.041	0.179	0.001	0.174
Paclitaxel 175 mg/m <sup>2</sup>	ECT	1.050	0.017	0.000	0.000	0.167	0.113	0.000	0.040

**Table A.6.** Derived quantities based on parameters of the injection model

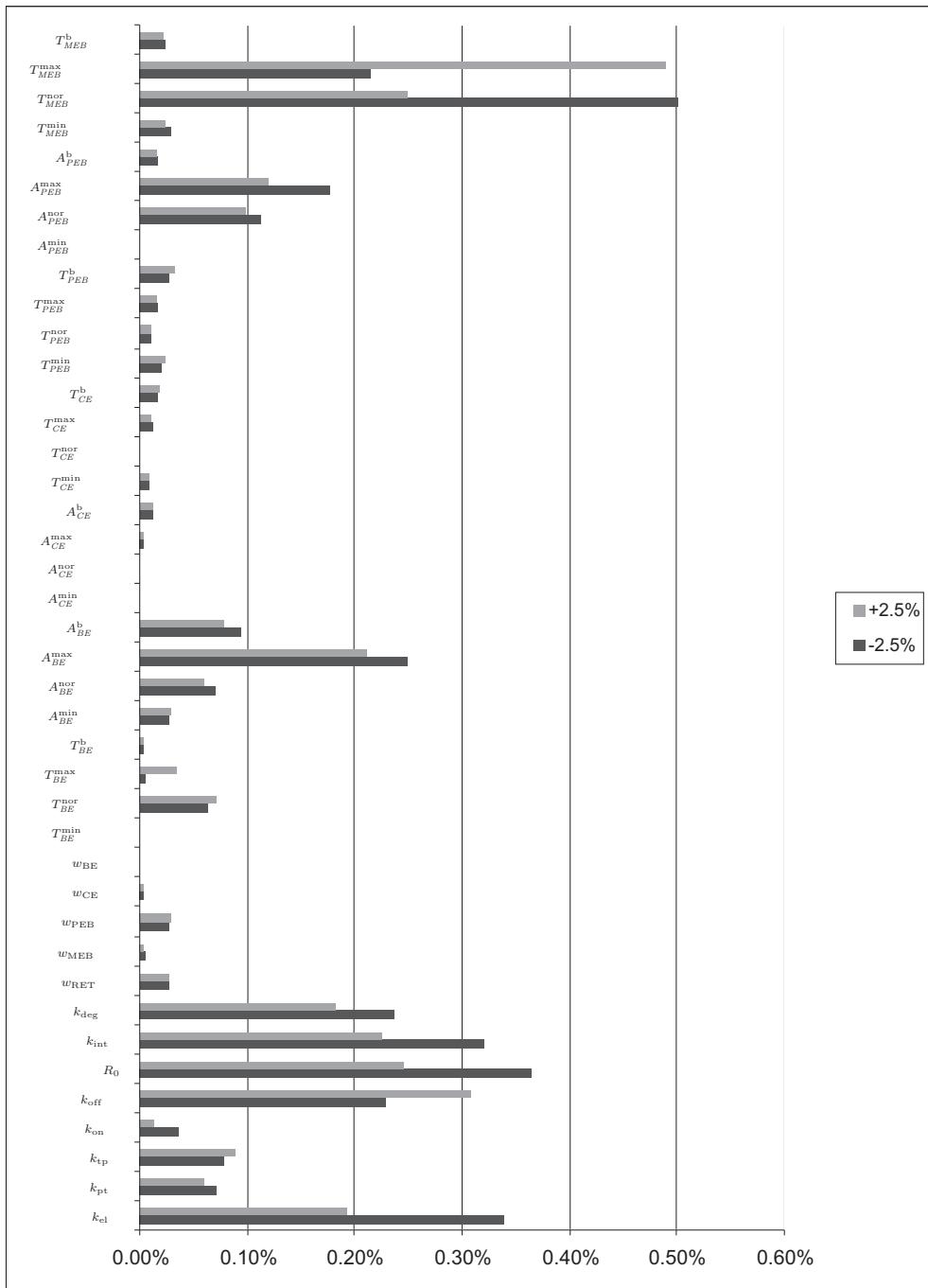
		bioavailability
Alfa	abdomen, upper arm	0.4123
Alfa	shoulder	0.3513
Alfa	forearm	0.2957
Alfa	thigh	0.5204
Beta	abdomen	0.7286
Beta	thigh	0.8137
Beta	forearm	0.7266
Delta		0.3524
Darbepoetin		0.8914

#### A.4 Sensitivity Analysis of Model Parameters

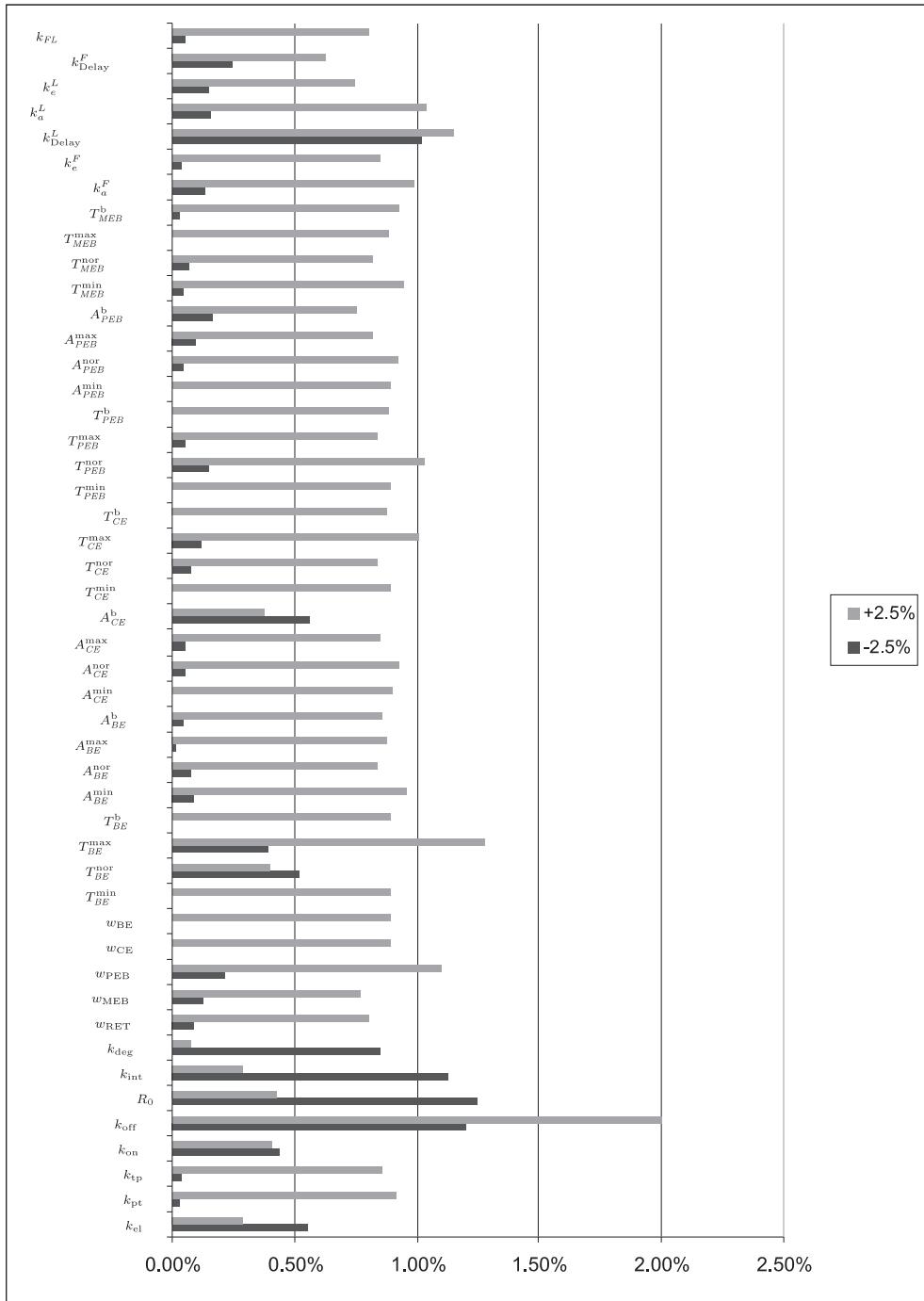
We analyzed the sensitivity of the model parameters in the following way. The parameters were increased or decreased by 2.5 % and the change of the sum of the fitness functions as percentage is plotted as bar diagrams. Values of the fitness functions of different scenarios are added.



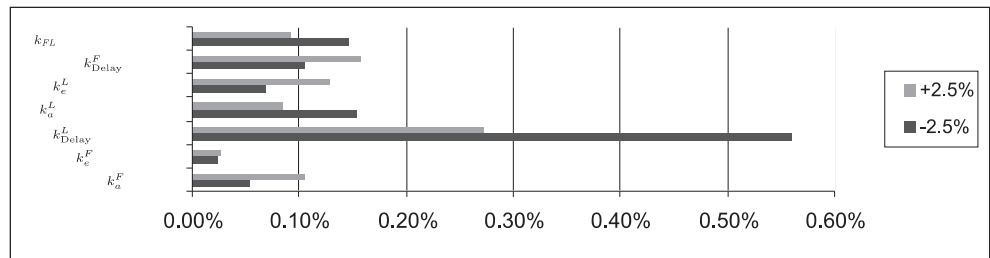
**Figure 1.** Sensitivity of parameters used for EPO Alfa, Beta, Delta, and Darbepoetin Alfa



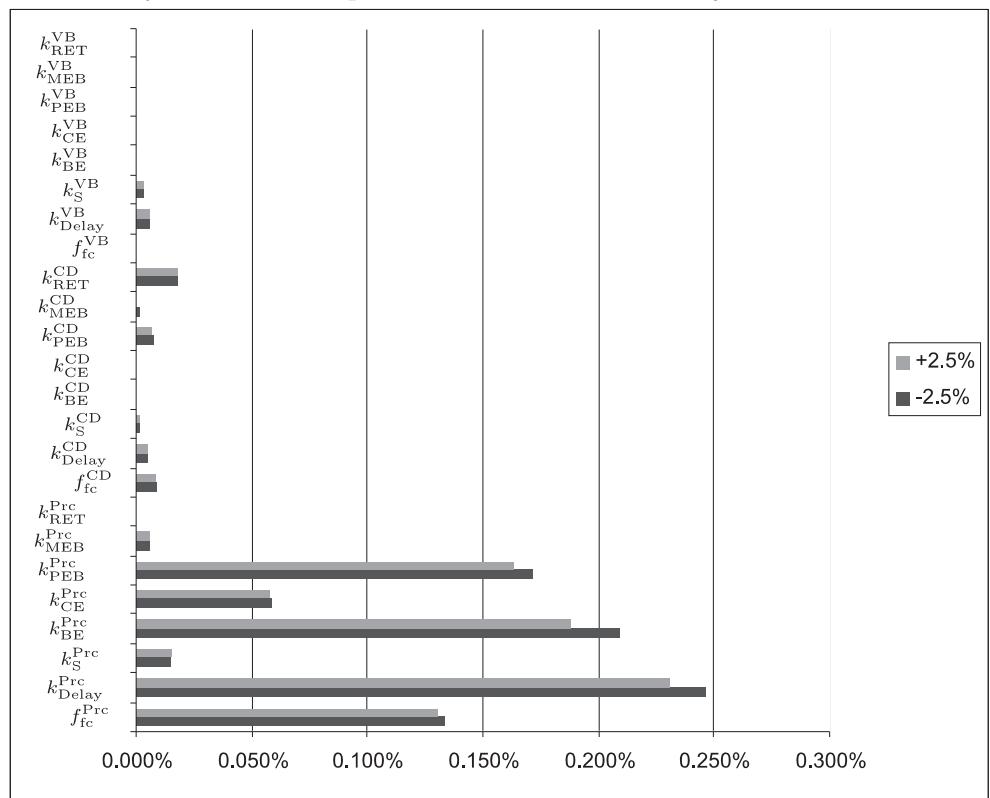
**Figure 2.** Sensitivity of the PK/PD parameters of EPO Alfa, Beta, Delta



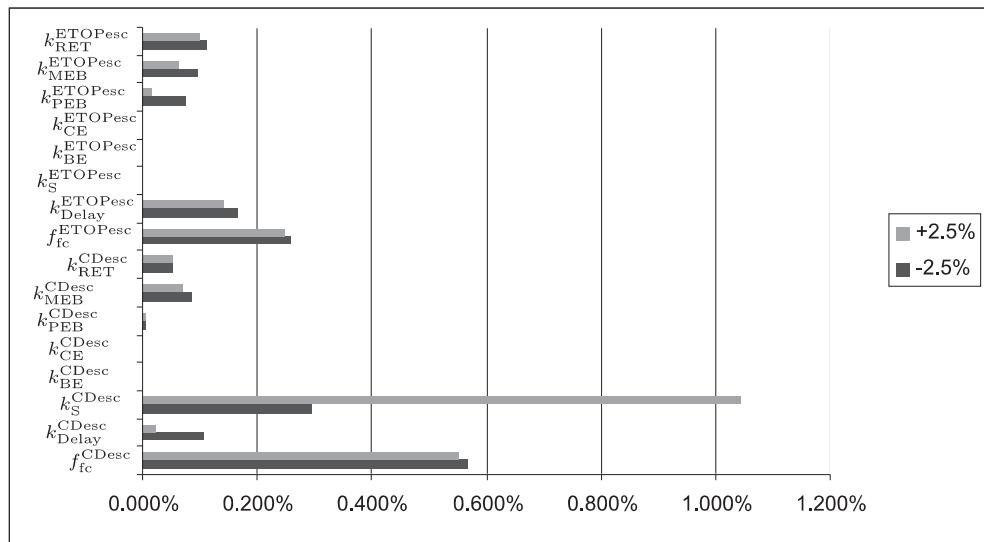
**Figure 3.** Sensitivity of the PK/PD parameters for Darbepoetin.



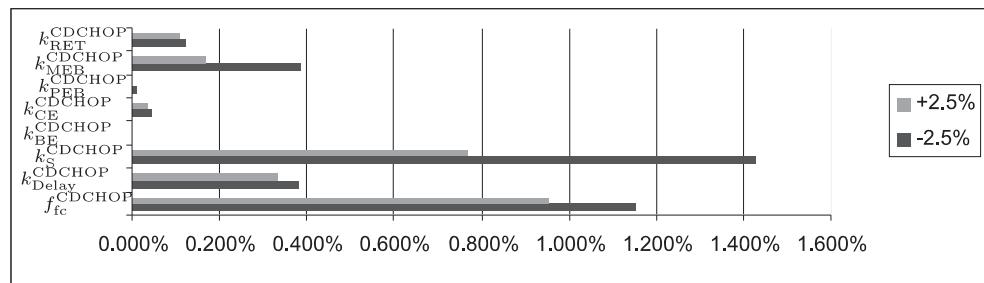
**Figure 4.** Sensitivity of the different parameters for subcutaneous injection of EPO Alfa, Beta, Delta



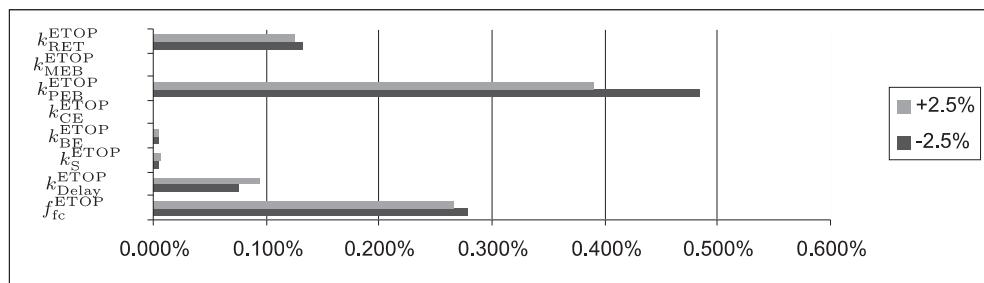
**Figure 5.** Sensitivity of toxicity parameters for BEACOPP chemotherapy



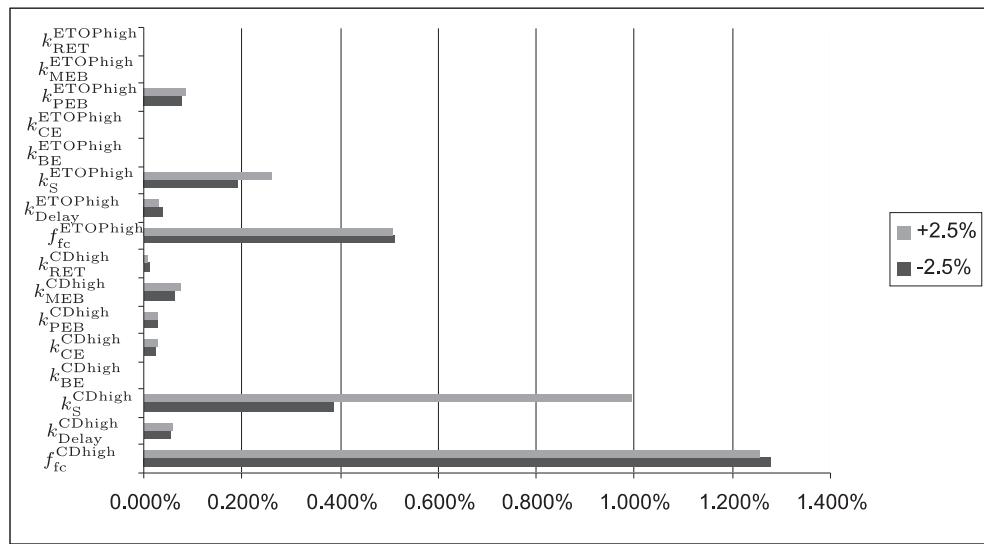
**Figure 6.** Sensitivity of toxicity parameters for BEACOPP escalated chemotherapy



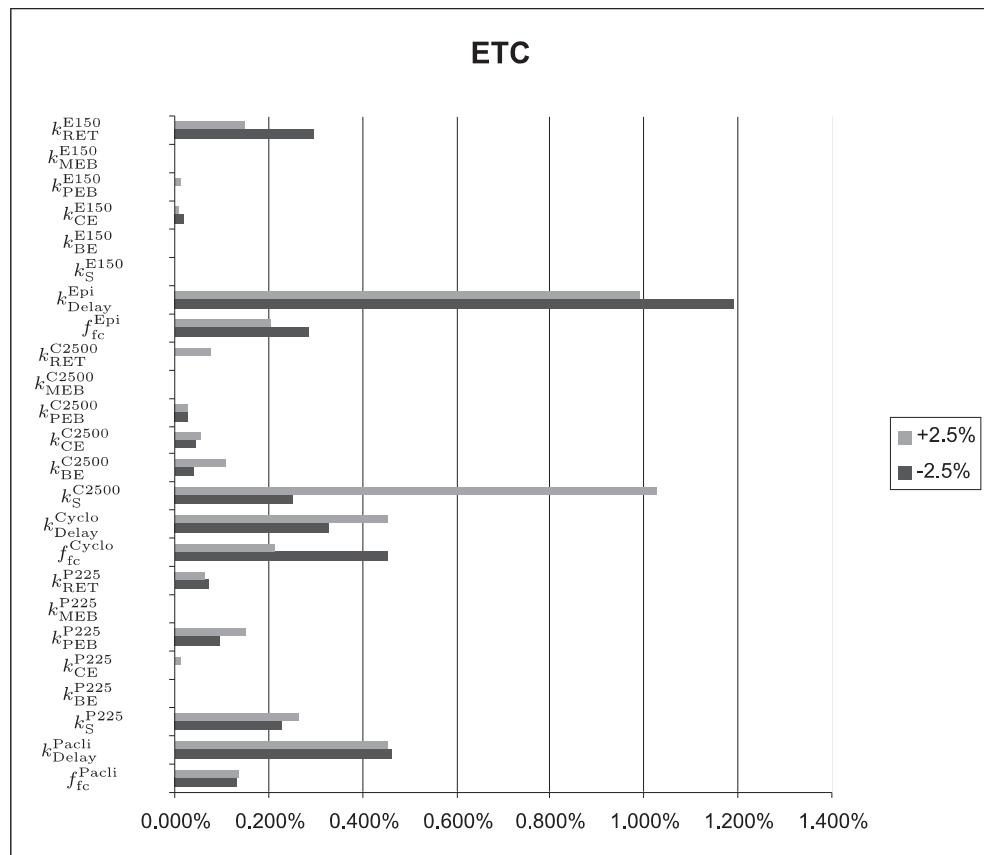
**Figure 7.** Sensitivity of toxicity parameters for CHOP chemotherapy



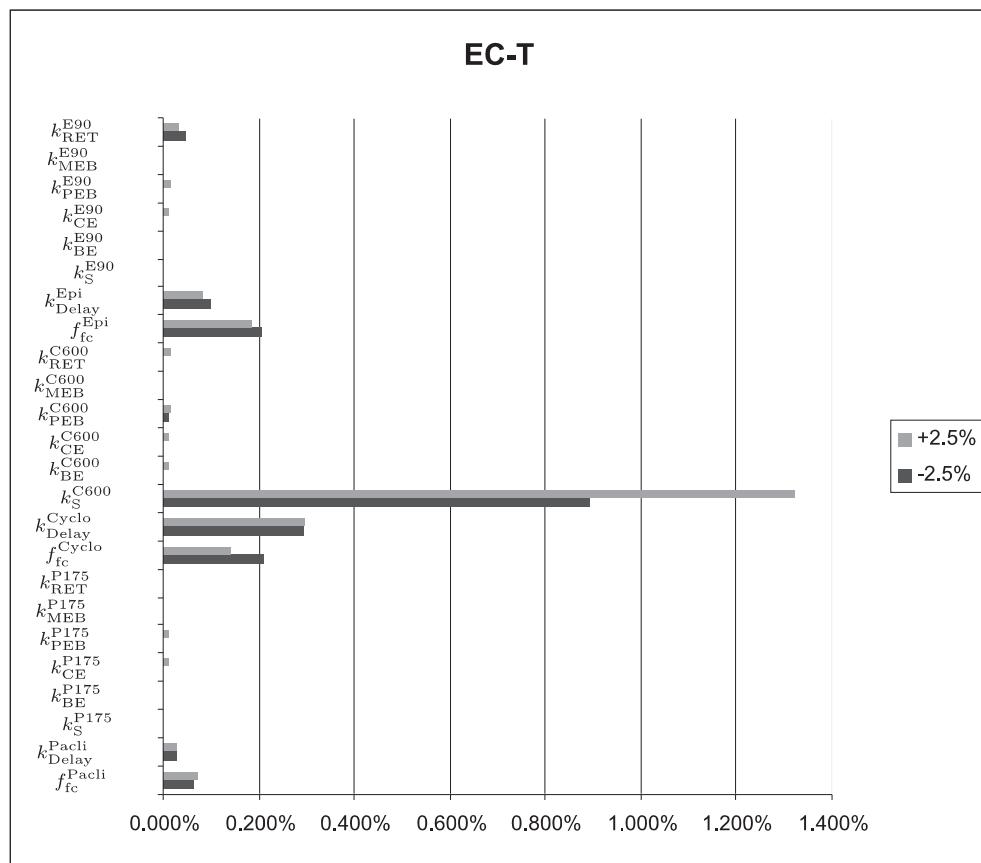
**Figure 8.** Sensitivity of toxicity parameters for etoposide



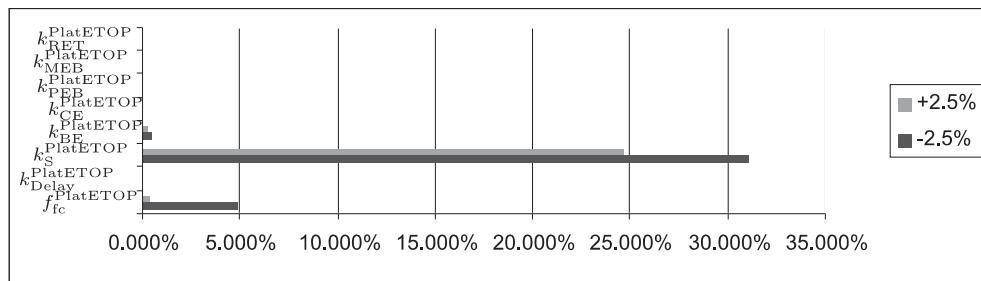
**Figure 9.** Sensitivity of toxicity parameters for high-CHOEP chemotherapy



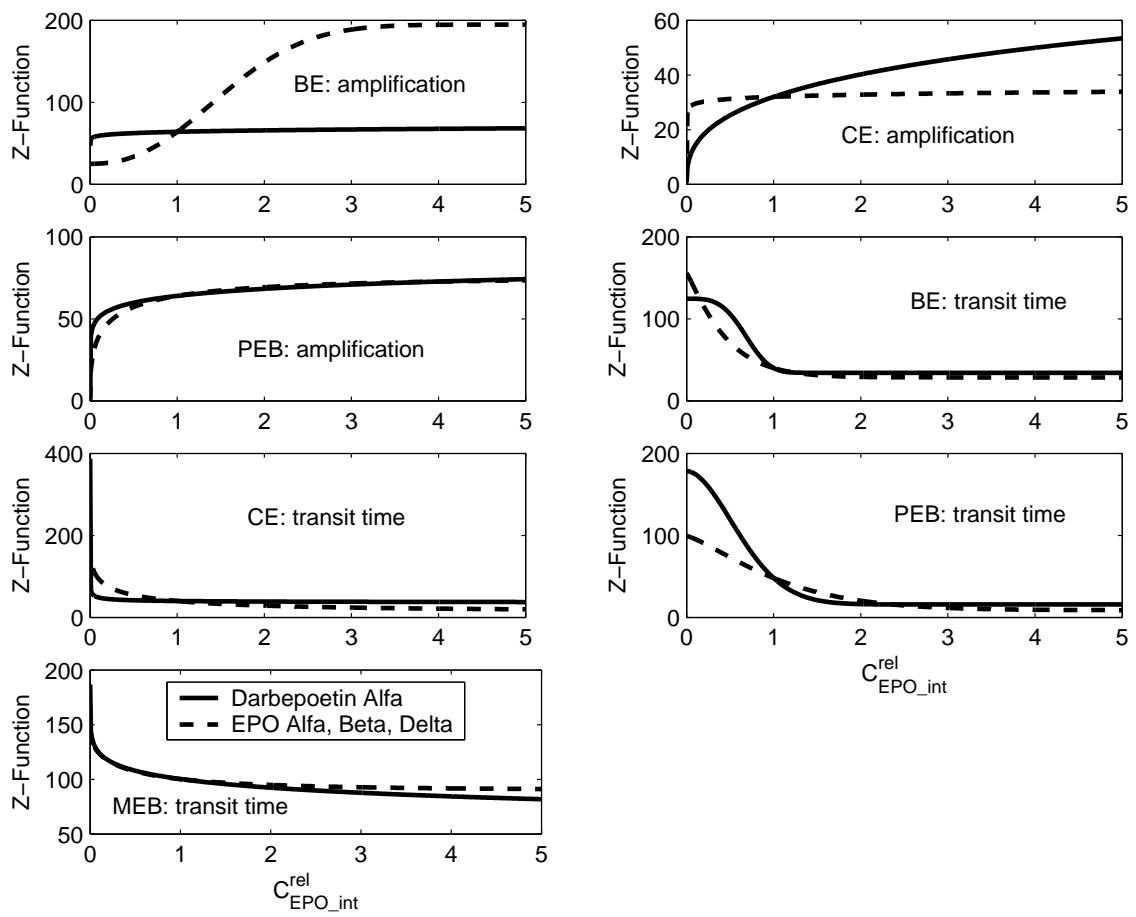
**Figure 10.** Sensitivity of toxicity parameters for ETC chemotherapy



**Figure 11.** Sensitivity of toxicity parameters for EC-T chemotherapy

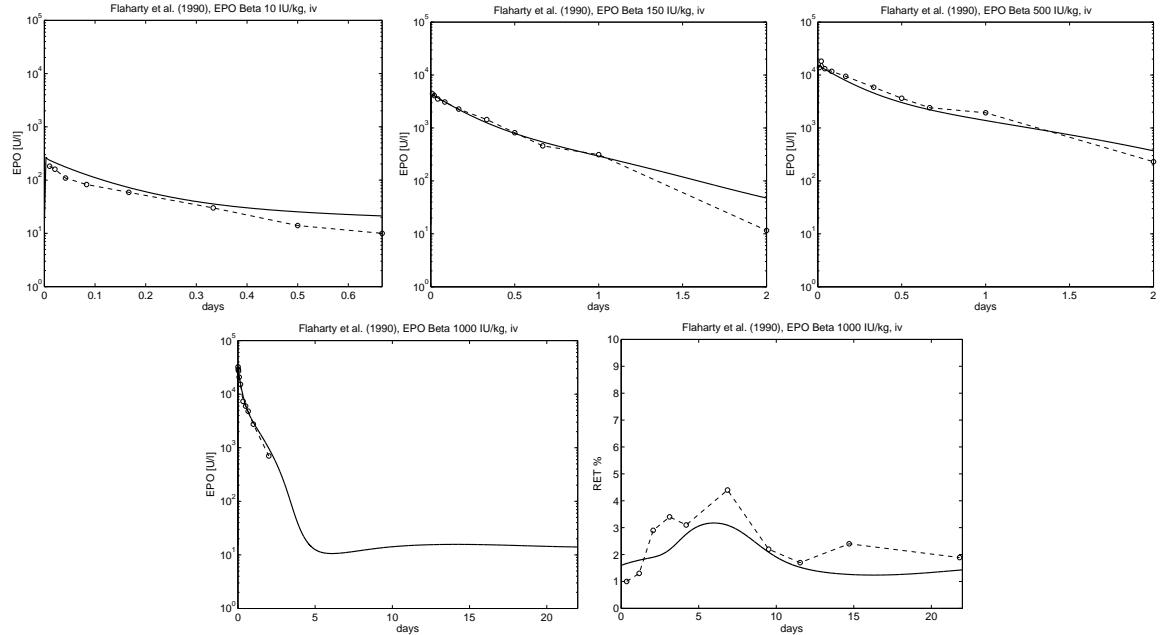


**Figure 12.** Sensitivity of toxicity parameters for the chemotherapy Platinum plus Etoposide

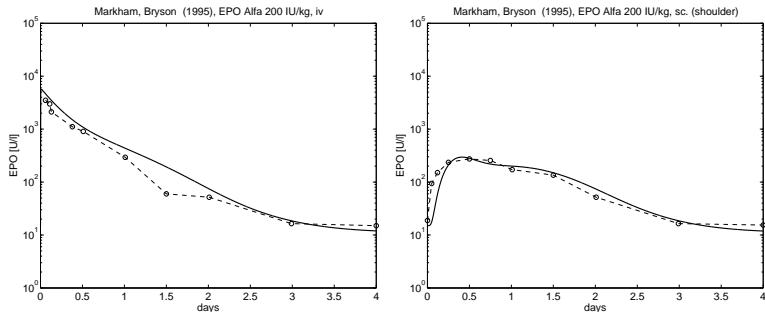


**Figure 13.** Z-functions of the amplifications in compartments BE, CE, PEB and the transition times in BE, CE, PEB, MEB

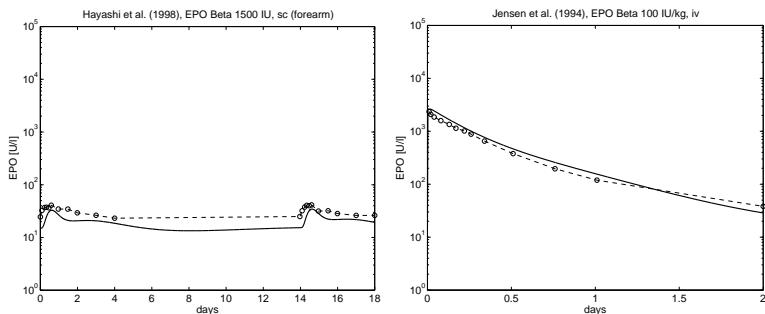
## A.5 Simulation



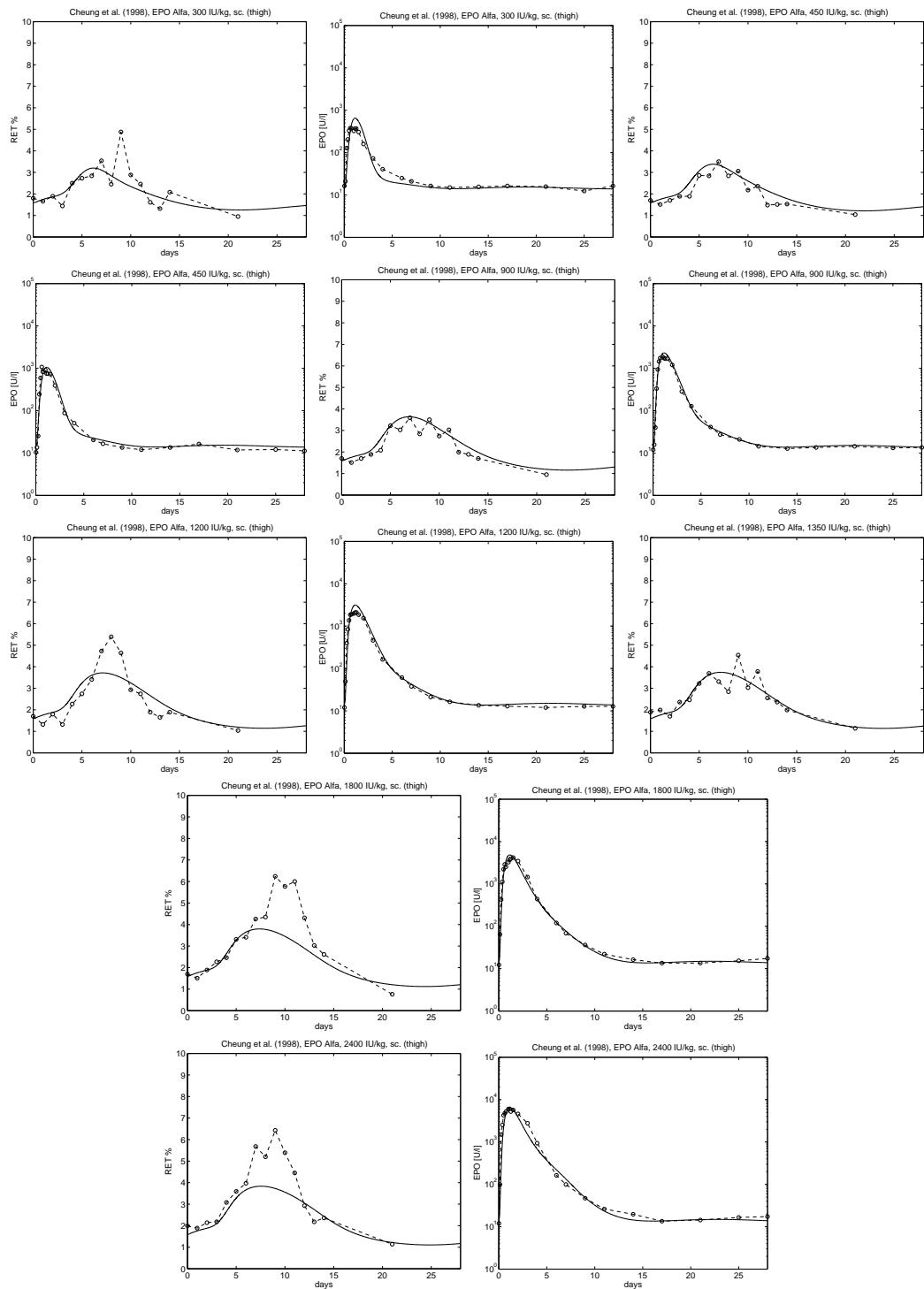
**Figure 14.** Serum concentration of erythropoietin, simulation (black line) and data (circle), data: [8]



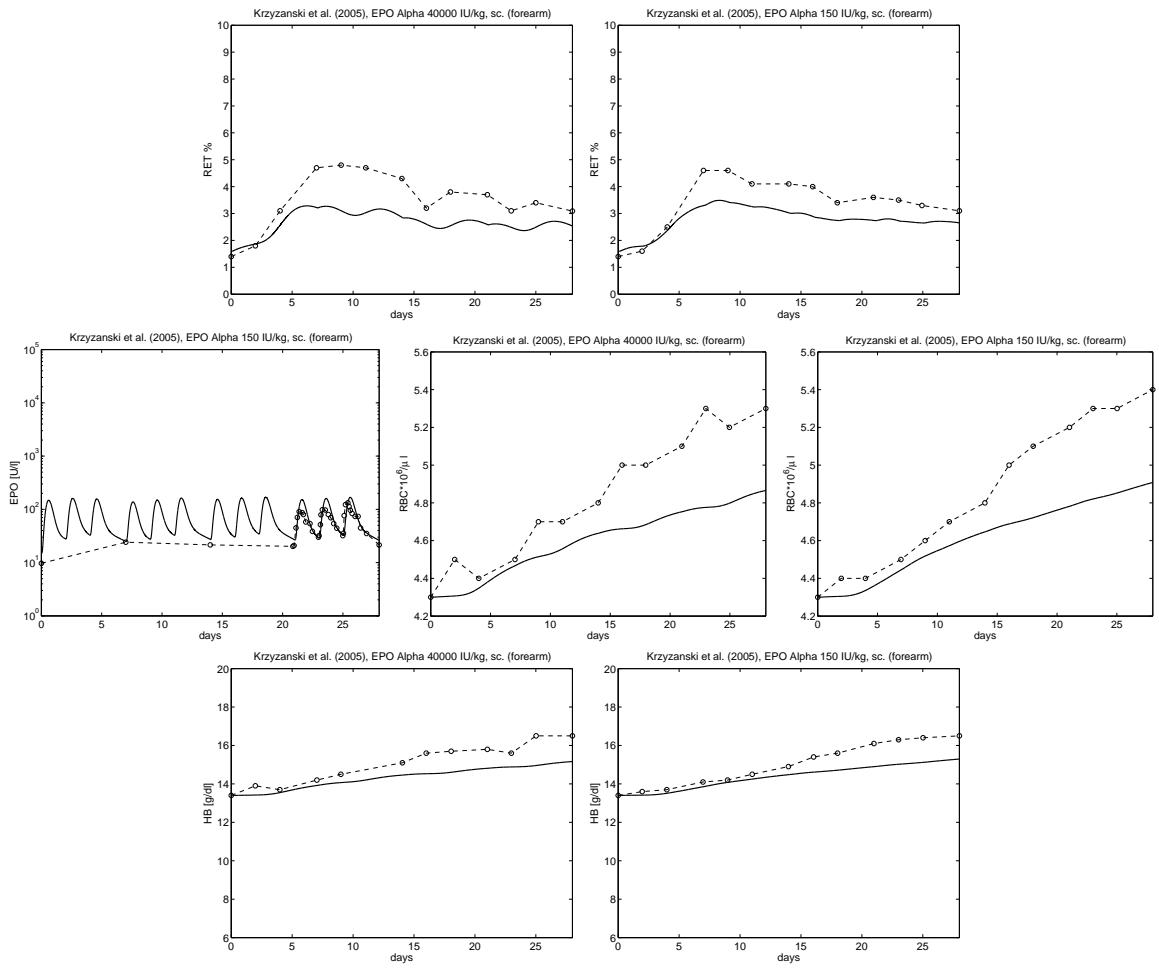
**Figure 15.** Serum concentration of erythropoietin (simulation and data), data: [9]



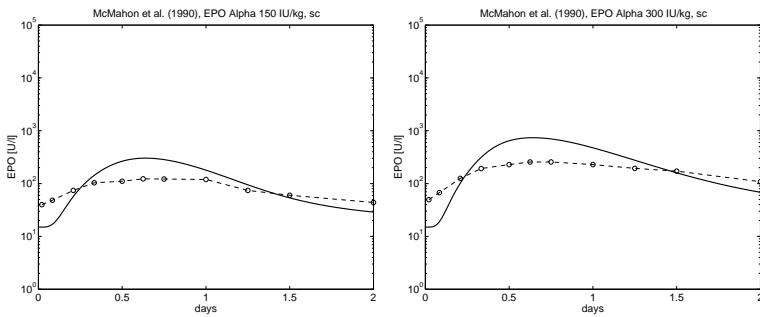
**Figure 16.** Serum concentration of erythropoietin (simulation and data), data: [10, 11]



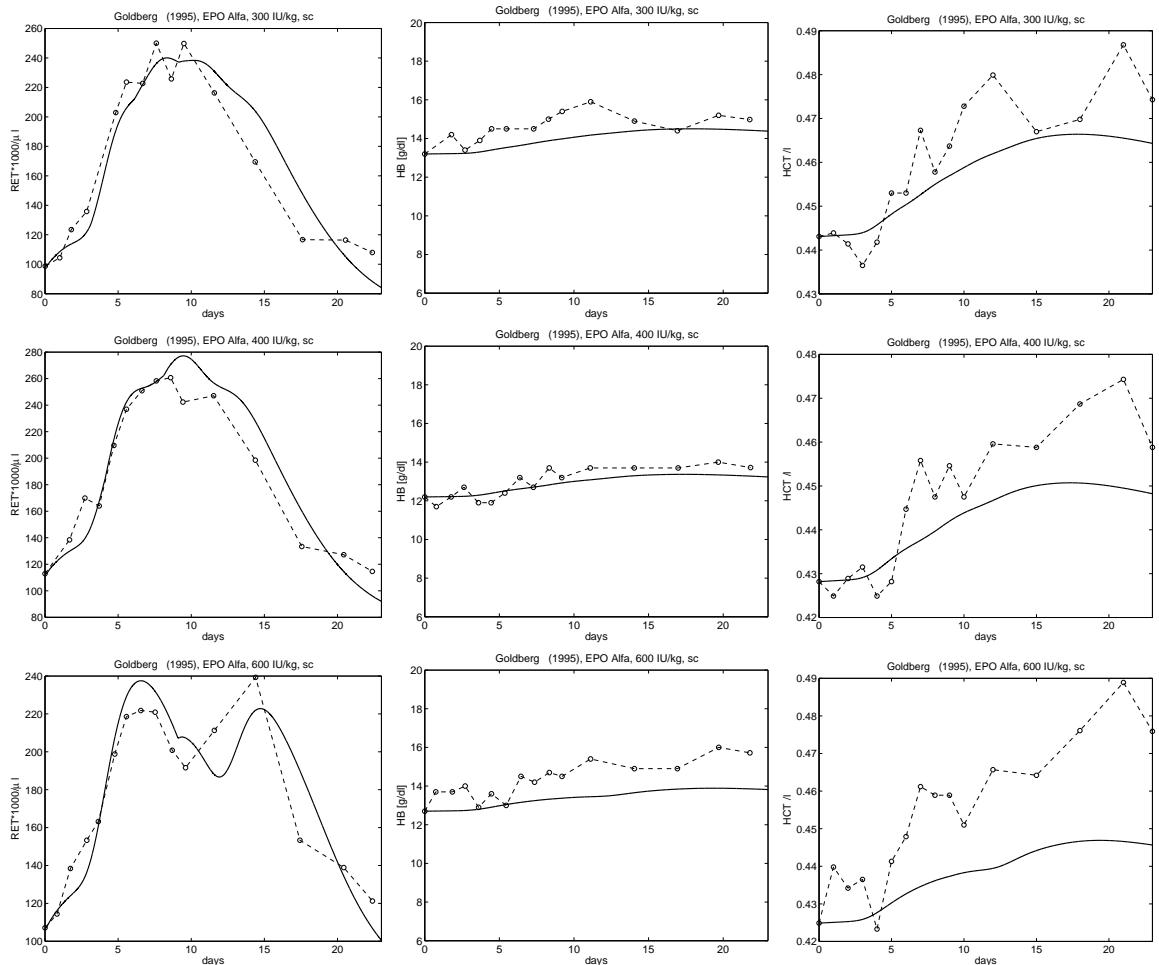
**Figure 17.** Serum concentration of erythropoietin and reticulocytes %, simulation (black line) and data (circle), data: [12]



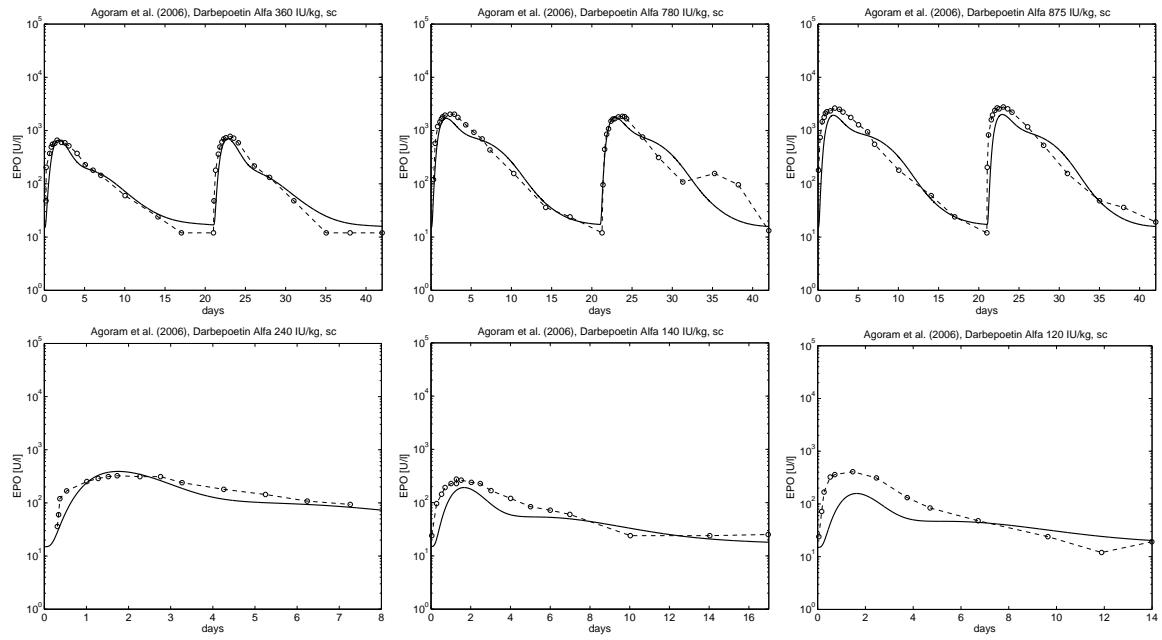
**Figure 18.** Serum concentration of erythropoietin, HB value, RBC, and reticulocytes %, simulation (black line) and data (circle), data: [7]



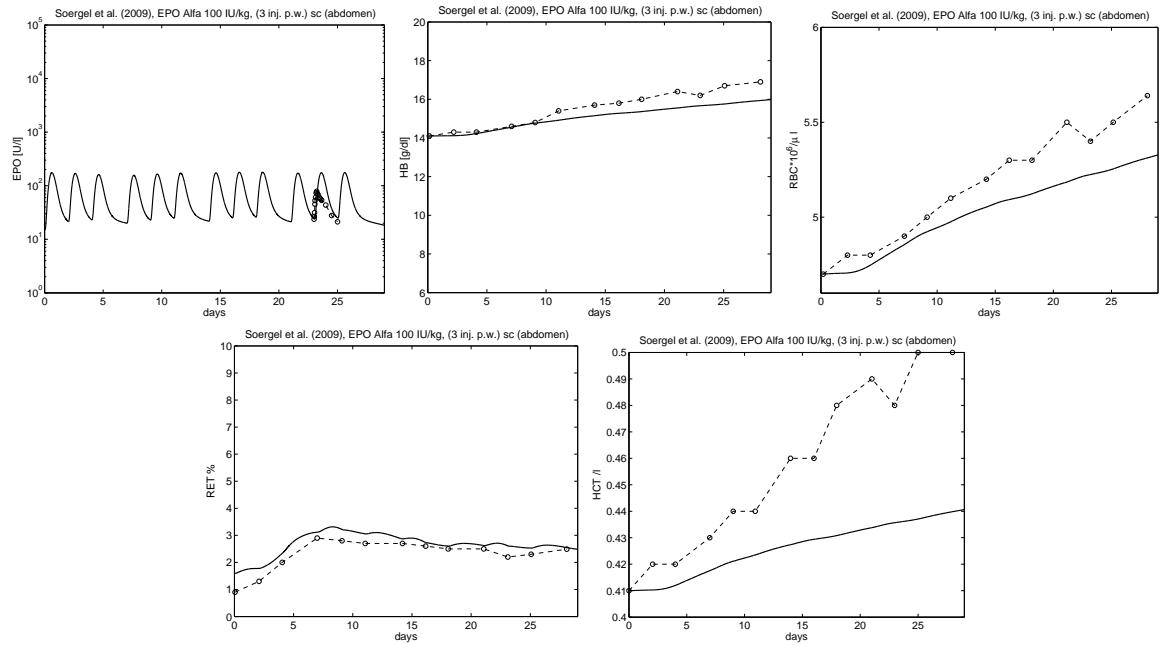
**Figure 19.** Serum concentration of erythropoietin, simulation (black line) and data (circle), data: [13]



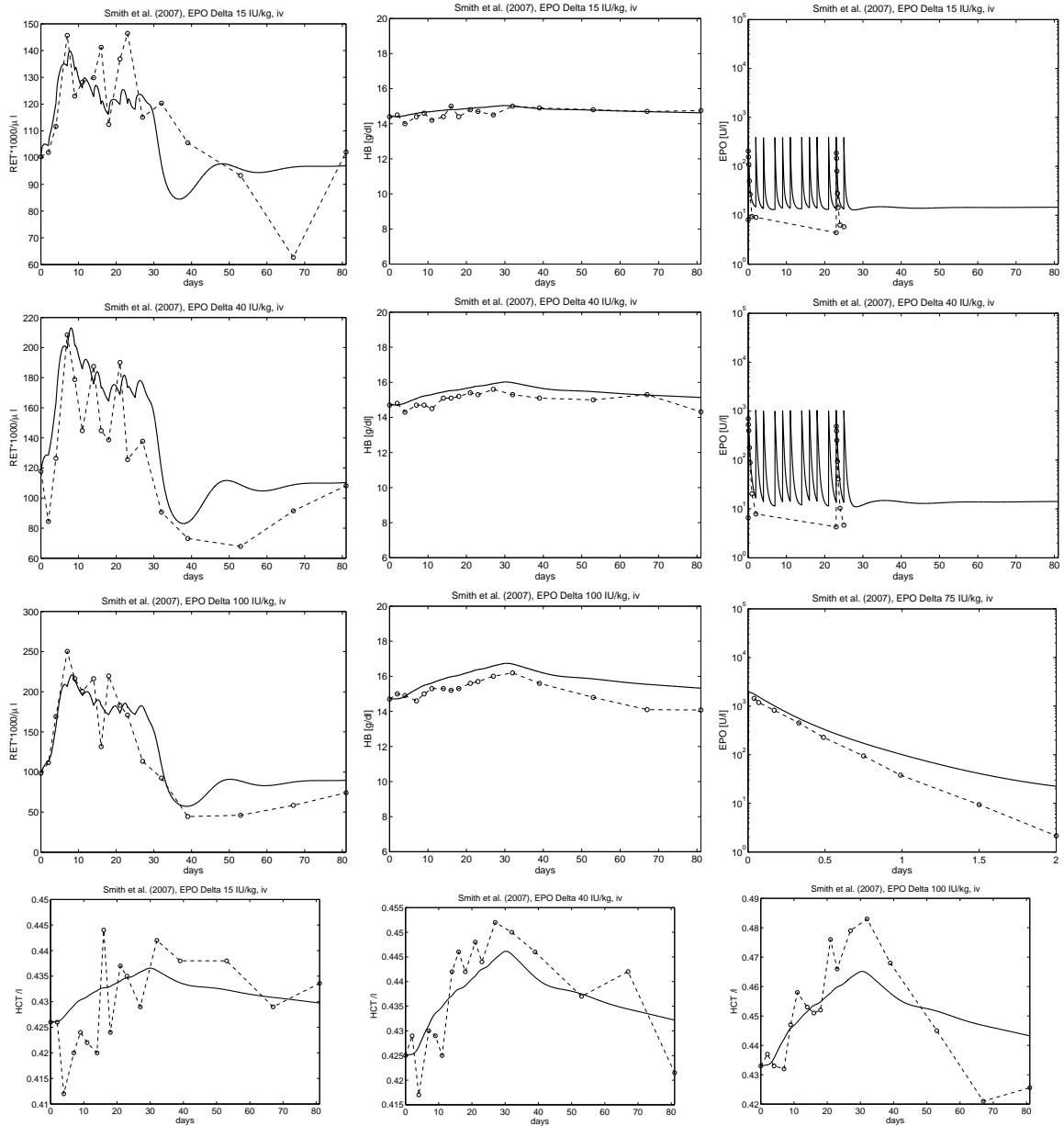
**Figure 20.** Serum concentration of erythropoietin, HB and HCT value, simulation (black line) and data (circle), data: [14], [15], only from subjects with baseline HCT of less than 48%. If the HCT rose above 55%, phlebotomy was performed.



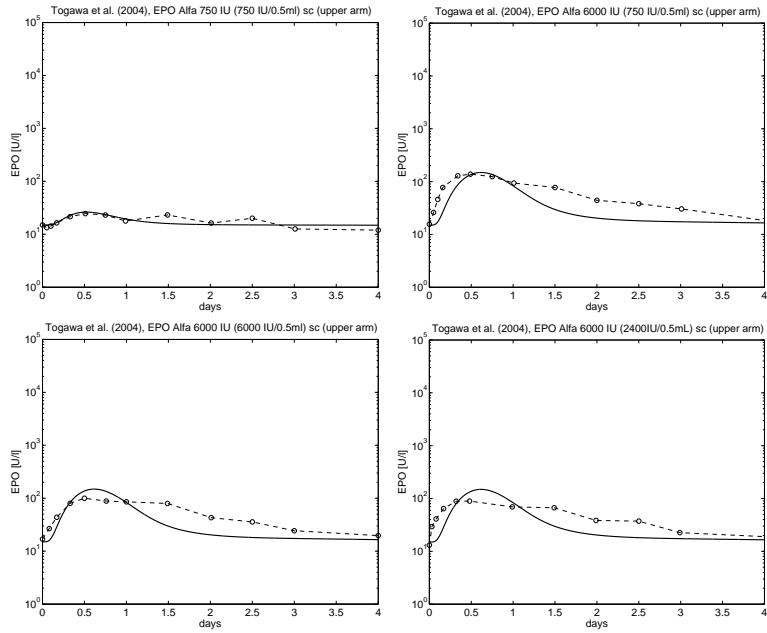
**Figure 21.** Serum concentration of erythropoietin, simulation (black line) and data (circle), data: [16]



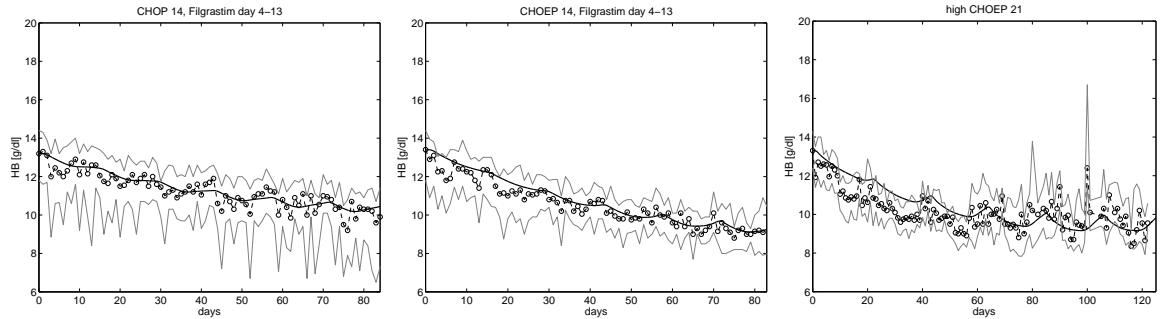
**Figure 22.** Serum concentration of erythropoietin, HB, RBC, reticulocytes % and HCT, simulation (black line) and data (circle), data: [17]



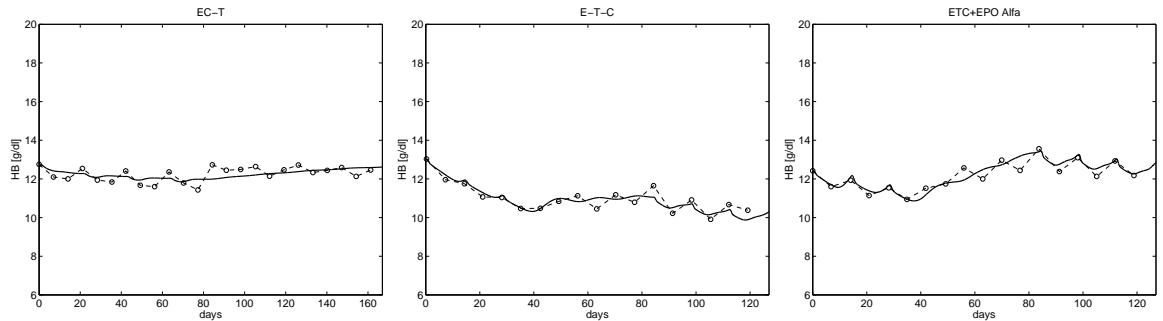
**Figure 23.** HB, reticulocytes %, Serum concentration of erythropoietin and HCT, simulation (black line) and data (circle), data: [18]



**Figure 24.** Serum concentration of erythropoietin, simulation (black line) and data (circle), data: [19]



**Figure 25.** HB value simulation (black line) and data (circle), percentile 25, 75 (grey line), data: [20–22]



**Figure 26.** HB value, simulation (black line) and data (circle), data: [23]

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21. Pfreundschuh M, Truemper L, Kloess M, Schmits R, Feller A, et al. (2004) 2-weekly or 3-weekly chop chemotherapy with or without etoposide for the treatment of elderly patients with aggressive lymphomas: results of the nhl-b2 trial of the dshnhl. *Blood* 104: 634–641.
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