

Supplementary Materials

Table S1. Pearson product-moment correlation coefficients of the LH metagenome taxonomic and functional reads under different cutoff *E values* $\leq 10^{-5}$, 10^{-10} , 10^{-15} , and 10^{-20}

Taxonomy		1.00E-10	1.00E-15	1.00E-20
1.00E-05	R²	1	0.987563	0.73459
	P-value	< 2.2e-16	< 2.2e-16	1.93E-05
Function				
1.00E-05	R²	0.999999	0.995174	0.908828
	P-value	< 2.2e-16	< 2.2e-16	2.27E-11

TABLE S2. The percentages of genera related to the genes encoding enzymes facilitating nitrogen cycles in LH metagenome

Genus/Enzyme/EC	%	Phylum/Class of Proteobacteria
Copper-containing nitrite reductase (EC 1.7.2.1)		
<i>Burkholderia</i>	2.4	<i>Betaproteobacteria</i>
<i>Bdellovibrio</i>	15	<i>Deltaproteobacteria</i>
<i>Propionibacterium</i>	1.2	<i>Actinobacteria</i>
<i>Gramella</i>	3.6	<i>Bacteroidetes</i>
<i>Kangiella</i>	23	<i>Gammaproteobacteria</i>
<i>Neisseria</i>	7.1	<i>Betaproteobacteria</i>
<i>Maribacter</i>	13	<i>Bacteroidetes</i>
<i>Flavobacterium</i>	32	<i>Bacteroidetes</i>
<i>Leeuwenhoekiella</i>	1.2	<i>Bacteroidetes</i>
<i>Flavobacteria</i>	1.2	<i>Bacteroidetes</i>
Nitrous-oxide reductase (EC 1.7.99.6/1.7.2.4)		
<i>Magnetospirillum</i>	11	<i>Alphaproteobacteria</i>
<i>Campylobacter</i>	68	<i>Deltaproteobacteria</i>
<i>Nitratiruptor</i>	3.6	<i>Deltaproteobacteria</i>
<i>Sulfurovum</i>	11	<i>Deltaproteobacteria</i>
<i>Magnetospirillum</i>	3.6	<i>Alphaproteobacteria</i>
<i>Gramella</i>	3.6	<i>Bacteroidetes</i>
Nitrogenase (EC 1.18.6.1)		
<i>Nostoc</i>	50	<i>Cyanobacteria</i>
<i>Cyanothece</i>	50	<i>Cyanobacteria</i>
Nitrite reductase [NAD(P)H] (EC 1.7.1.4)		
<i>Thermobaculum</i>	1.4	<i>Bacteroidetes</i>
<i>Pseudomonas</i>	1.4	<i>Gammaproteobacteria</i>
<i>Cytophaga</i>	10	<i>Cyanobacteria</i>
<i>Escherichia</i>	2.9	<i>Gammaproteobacteria</i>
<i>Spirosoma</i>	7.2	<i>Bacteroidetes</i>
<i>Klebsiella</i>	2.9	<i>Bacteroidetes</i>
<i>Polaromonas</i>	1.4	<i>Betaproteobacteria</i>
<i>Geobacillus</i>	1.4	<i>Firmicutes</i>
<i>Polaribacter</i>	11.6	<i>Bacteroidetes</i>

<i>Saccharophagus</i>	1.4	<i>Gammaproteobacteria</i>
<i>Acinetobacter</i>	2.8	<i>Gammaproteobacteria</i>
<i>Chitinophaga</i>	7.2	<i>Bacteroidetes</i>
<i>Dyadobacter</i>	2.9	<i>Bacteroidetes</i>
<i>Psychrobacter</i>	10	<i>Gammaproteobacteria</i>
<i>Mycobacterium</i>	1.4	<i>Bacteroidetes</i>
<i>Rhodoferax</i>	1.4	<i>Betaproteobacteria</i>
<i>Dokdonia</i>	8.7	<i>Bacteroidetes</i>
<i>Leptothrix</i>	2.9	<i>Betaproteobacteria</i>
<i>Psychromonas</i>	4.3	<i>Gammaproteobacteria</i>
<i>Methylococcus</i>	2.8	<i>Gammaproteobacteria</i>
<i>Thiobacillus</i>	1.4	<i>Betaproteobacteria</i>
<i>Sorangium</i>	5.8	<i>Deltaproteobacteria</i>
<i>Croceibacter</i>	5.8	<i>Bacteroidetes</i>
Ferredoxin--nitrite reductase (EC 1.7.7.1)		
<i>Synechocystis</i>	24	<i>Cyanobacteria</i>
<i>Cyanothece</i>	16	<i>Cyanobacteria</i>
<i>Trichodesmium</i>	8	<i>Cyanobacteria</i>
<i>Nostoc</i>	44	<i>Cyanobacteria</i>
<i>Thermosynechococcus</i>	4	<i>Cyanobacteria</i>
<i>Microcystis</i>	4	<i>Cyanobacteria</i>
Assimilatory nitrate reductase (EC 1.7.99.4)		
<i>RhodoPseudomonas</i>	0.7	<i>Alphaproteobacteria</i>
<i>Flavobacterium</i>	0.7	<i>Bacteroidetes</i>
<i>Psychrobacter</i>	30	<i>Gammaproteobacteria</i>
<i>Thermobaculum</i>	0.7	<i>Unclassified Bacteria</i>
<i>Streptosporansium</i>	0.7	<i>Actinobacteria</i>
<i>Bacillus</i>	5.7	<i>Firmicutes</i>
<i>Trichodesmium</i>	1.4	<i>Cyanobacteria</i>
<i>Mycobacterium</i>	0.7	<i>Bacteroidetes</i>
<i>Methylococcus</i>	1.4	<i>Gammaproteobacteria</i>
<i>Burkholderia</i>	2.8	<i>Betaproteobacteria</i>
<i>Pectobacterium</i>	0.7	<i>Gammaproteobacteria</i>
<i>Cyanothece</i>	4.3	<i>Cyanobacteria</i>
<i>Shewanella</i>	0.7	<i>Gammaproteobacteria</i>
<i>Pseudomonas</i>	0.7	<i>Gammaproteobacteria</i>
<i>Bradyrhizobium</i>	2.2	<i>Gammaproteobacteria</i>

<i>Azoarcus</i>	0.7	<i>Betaproteobacteria</i>
<i>Alcanivorax</i>	1.4	<i>Gammaproteobacteria</i>
<i>Anabaena</i>	6.5	<i>Cyanobacteria</i>
<i>Chitinophaga</i>	0.7	<i>Bacteroidetes</i>
<i>Cytophaga</i>	2.9	<i>Bacteroidetes</i>
<i>Rhodococcus</i>	0.7	<i>Actinobacteria</i>
<i>Marinomonas</i>	0.7	<i>Gammaproteobacteria</i>
<i>Roseobacter</i>	1.4	<i>Alphaproteobacteria</i>
<i>Nostoc</i>	12	<i>Cyanobacteria</i>
<i>Methylibium</i>	2.2	<i>Betaproteobacteria</i>
<i>Pseudoalteromonas</i>	0.7	<i>Gammaproteobacteria</i>
<i>Microcystis</i>	2.2	<i>Cyanobacteria</i>
<i>Polaromonas</i>	0.7	<i>Betaproteobacteria</i>
<i>Novosphingobium</i>	0.7	<i>Alphaproteobacteria</i>
<i>Crocospaera</i>	0.7	<i>Cyanobacteria</i>
<i>Thiobacillus</i>	2.2	<i>Betaproteobacteria</i>
<i>Pseudomonas</i>	0.7	<i>Gammaproteobacteria</i>
<i>Azorhizobium</i>	2.2	<i>Alphaproteobacteria</i>
<i>Acidovorax</i>	1.4	<i>Betaproteobacteria</i>
<i>Maribacter</i>	0.7	<i>Bacteroidetes</i>
<i>Acinetobacter</i>	2.2	<i>Gammaproteobacteria</i>
<i>Mesorhizobium</i>	0.7	<i>Alphaproteobacteria</i>
<i>Rhizobium</i>	1.4	<i>Alphaproteobacteria</i>

TABLE S3. The percentages of genera related to the genes encoding enzymes facilitating Sulfate reduction in LH metagenome

Genus/Enzyme/EC	%	Phylum/Class of Proteobacteria
Adenylylsulfate kinase (EC 2.7.1.25)		
<i>Roseiflexus</i>	16.3	<i>Chloroflexi</i>
<i>Aquifex</i>	16.3	<i>Aquificae</i>
<i>Chloroflexus</i>	5.8	<i>Chloroflexi</i>
<i>Schizosaccharomyces</i>	5.2	<i>Opisthokonta</i>
<i>Caulobacter</i>	4.6	<i>Alphaproteobacteria</i>
<i>Halothermothrix</i>	4.7	<i>Firmicutes</i>
<i>Xanthomonas</i>	3.5	<i>Gammaproteobacteria</i>
<i>Paracoccus</i>	2.9	<i>Alphaproteobacteria</i>
<i>Zymomonas</i>	2.9	<i>Alphaproteobacteria</i>
<i>Trichodesmium</i>	2.3	<i>Cyanobacteria</i>
<i>Cyanothece</i>	2.3	<i>Cyanobacteria</i>
<i>Bacteroides</i>	1.7	<i>Bacteroidetes</i>
<i>Erythrobacter</i>	1.7	<i>Alphaproteobacteria</i>
<i>Magnetococcus</i>	1.7	<i>Alphaproteobacteria</i>
<i>Microcystis</i>	1.7	<i>Cyanobacteria</i>
<i>Solibacter</i>	1.7	<i>Acidobacteria</i>
<i>Francisella</i>	1.8	<i>Gammaproteobacteria</i>
<i>Pseudomonas</i>	1.8	<i>Gammaproteobacteria</i>
<i>Vibrio</i>	1.2	<i>Gammaproteobacteria</i>
<i>Burkholderia</i>	1.2	<i>Betaproteobacteria</i>
<i>Chlorobium</i>	1.2	<i>Bacteroidetes</i>
<i>Deinococcus</i>	1.2	<i>Deinococcus</i>
<i>Frankia</i>	1.2	<i>Actinobacteria</i>
<i>Hydrogenobaculum</i>	1.2	<i>Aquificae</i>
<i>Oceanobacillus</i>	1.2	<i>Firmicutes</i>
<i>Rhodopseudomonas</i>	1.2	<i>Alphaproteobacteria</i>
<i>Synechocystis</i>	1.2	<i>Cyanobacteria</i>
<i>Ashbya</i>	0.6	<i>Opisthokonta</i>
<i>Aurantimonas</i>	0.6	<i>Alphaproteobacteria</i>
<i>Campylobacter</i>	0.6	<i>Deltaproteobacteria</i>
<i>Crocospaera</i>	0.6	<i>Cyanobacteria</i>
<i>Delftia</i>	0.6	<i>Betaproteobacteria</i>

<i>Geobacter</i>	0.6	<i>Deltaproteobacteria</i>
<i>Methylococcus</i>	0.6	<i>Gammaproteobacteria</i>
<i>Mycobacterium</i>	0.6	<i>Actinobacteria</i>
<i>Neurospora</i>	0.6	<i>Opisthokonta</i>
<i>Oceanicaulis</i>	0.6	<i>Alphaproteobacteria</i>
<i>Parvibaculum</i>	0.6	<i>Alphaproteobacteria</i>
<i>Pelobacter</i>	0.6	<i>Deltaproteobacteria</i>
<i>Prochlorococcus</i>	0.6	<i>Cyanobacteria</i>
<i>Roseobacter</i>	0.6	<i>Alphaproteobacteria</i>
<i>Shewanella</i>	0.6	<i>Gammaproteobacteria</i>
<i>Sphingomonas</i>	0.6	<i>Alphaproteobacteria</i>
<i>Thermobispora</i>	0.6	<i>Actinobacteria</i>
<i>Thermosynechococcus</i>	0.6	<i>Cyanobacteria</i>
Phosphoadenylyl-sulfate reductase [thioredoxin] (EC 1.8.4.8)/ Adenylyl-sulfate reductase [thioredoxin] (EC 1.8.4.10)		
<i>Marinobacter</i>	29.2	<i>Gammaproteobacteria</i>
<i>Acinetobacter</i>	17	<i>Gammaproteobacteria</i>
<i>Azotobacter</i>	9.8	<i>Gammaproteobacteria</i>
<i>Reinekea</i>	9.8	<i>Gammaproteobacteria</i>
<i>Cyanothece</i>	4.8	<i>Cyanobacteria</i>
<i>Pseudomonas</i>	4.8	<i>Gammaproteobacteria</i>
<i>Crocospaera</i>	4.9	<i>Cyanobacteria</i>
<i>Hahella</i>	4.9	<i>Gammaproteobacteria</i>
<i>Alkalilimnicola</i>	2.4	<i>Gammaproteobacteria</i>
<i>Anabaena</i>	2.4	<i>Cyanobacteria</i>
<i>Bordetella</i>	2.4	<i>Betaproteobacteria</i>
<i>Cellvibrio</i>	2.4	<i>Gammaproteobacteria</i>
<i>Croceibacter</i>	2.4	<i>Bacteroidetes</i>
<i>Nostoc</i>	2.4	<i>Cyanobacteria</i>
Sulfate adenylyltransferase, dissimilatory-type (EC 2.7.7.4)		
<i>Aquifex</i>	12.6	<i>Aquificae</i>
<i>Roseiflexus</i>	12.7	<i>Chloroflexi</i>
<i>Anabaena</i>	5.9	<i>Cyanobacteria</i>
<i>Chloroflexus</i>	4.5	<i>Chloroflexi</i>
<i>Cytophaga</i>	3.6	<i>Cyanobacteria</i>

<i>Synechococcus</i>	4.2	<i>Cyanobacteria</i>
<i>Caulobacter</i>	3.7	<i>Alphaproteobacteria</i>
<i>Nitrosococcus</i>	1.8	<i>Gammaproteobacteria</i>
<i>Xanthomonas</i>	3.3	<i>Gammaproteobacteria</i>
<i>Nostoc</i>	2.7	<i>Cyanobacteria</i>
<i>Maribacter</i>	3.7	<i>Bacteroidetes</i>
<i>Paracoccus</i>	2.3	<i>Alphaproteobacteria</i>
<i>Microcystis</i>	1.8	<i>Cyanobacteria</i>
<i>Thermosynechococcus</i>	1.8	<i>Cyanobacteria</i>
<i>Pseudomonas</i>	1.4	<i>Gammaproteobacteria</i>
<i>Trichodesmium</i>	1.4	<i>Cyanobacteria</i>
<i>Erythrobacter</i>	1.4	<i>Alphaproteobacteria</i>
<i>Francisella</i>	1.4	<i>Gammaproteobacteria</i>
<i>Leeuwenhoekiella</i>	2.3	<i>Bacteroidetes</i>
<i>Magnetococcus</i>	1.4	<i>Alphaproteobacteria</i>
<i>Pseudoalteromonas</i>	1.9	<i>Gammaproteobacteria</i>
<i>Robiginitalea</i>	2.3	<i>Bacteroidetes</i>
<i>Streptosporangium</i>	1.4	<i>Actinobacteria</i>
<i>Aeropyrum</i>	0.9	<i>Crenarchaeota</i>
<i>Bacillus</i>	0.9	<i>Firmicutes</i>
<i>Deinococcus</i>	0.9	<i>Deinococcus</i>
<i>Hydrogenobaculum</i>	0.9	<i>Aquificae</i>
<i>Pyrococcus</i>	0.9	<i>Euryarchaeota</i>
<i>Frankia</i>	0.9	<i>Actinobacteria</i>
<i>RhodoPseudomonas</i>	0.9	<i>Alphaproteobacteria</i>
<i>Saccharophagus</i>	0.9	<i>Gammaproteobacteria</i>
<i>Cyanothece</i>	0.5	<i>Cyanobacteria</i>
<i>Ferroplasma</i>	0.5	<i>Euryarchaeota</i>
<i>Neurospora</i>	0.5	<i>Opisthokonta</i>
<i>Petrogala</i>	0.5	<i>Thermotogae</i>
<i>Roseobacter</i>	0.5	<i>Alphaproteobacteria</i>
<i>Staphylococcus</i>	0.5	<i>Firmicutes</i>
<i>Staphylothermus</i>	0.5	<i>Crenarchaeota</i>
<i>Thermobispora</i>	0.5	<i>Actinobacteria</i>
<i>Flavobacterium</i>	0.5	<i>Bacteroidetes</i>
<i>Geobacter</i>	0.5	<i>Deltaproteobacteria</i>
<i>Hahella</i>	0.5	<i>Gammaproteobacteria</i>
<i>Kytococcus</i>	0.5	<i>Actinobacteria</i>

<i>Opitutus</i>	1	<i>Verrucomicrobia</i>
<i>Psychromonas</i>	1	<i>Gammaproteobacteria</i>
<i>Aurantimonas</i>	0.5	<i>Alphaproteobacteria</i>
<i>Chromobacterium</i>	0.5	<i>Betaproteobacteria</i>
<i>Leptospira</i>	0.5	<i>Spirochaetes</i>
<i>Methylococcus</i>	0.5	<i>Gammaproteobacteria</i>
<i>Mycobacterium</i>	0.5	<i>Actinobacteria</i>
<i>Nocardioides</i>	0.5	<i>Actinobacteria</i>
<i>Oceanicaulis</i>	0.5	<i>Alphaproteobacteria</i>
<i>Parvibaculum</i>	0.5	<i>Alphaproteobacteria</i>
<i>Pelobacter</i>	0.5	<i>Deltaproteobacteria</i>
<i>Salmonella</i>	0.5	<i>Gammaproteobacteria</i>
<i>Shewanella</i>	0.5	<i>Gammaproteobacteria</i>
<i>Sphingomonas</i>	0.5	<i>Alphaproteobacteria</i>
<i>Nitrosomonas</i>	1.8	<i>Betaproteobacteria</i>
Sulfite reductase (EC 1.8.99.1)/ Ferredoxin--sulfite reductase (EC 1.8.7.1)/ Sulfite reductase [NADPH] hemoprotein (EC 1.8.1.2)/ Sulfite reductase, dissimilatory-type (EC 1.8.99.3)		
<i>Thiobacillus</i>	31.3	<i>Betaproteobacteria</i>
<i>Nostoc</i>	10.8	<i>Cyanobacteria</i>
<i>Cyanothece</i>	9.9	<i>Cyanobacteria</i>
<i>Synechocystis</i>	8.9	<i>Cyanobacteria</i>
<i>Alkalilimnicola</i>	6.3	<i>Gammaproteobacteria</i>
<i>Anabaena</i>	5.4	<i>Cyanobacteria</i>
<i>Thioalkalivibrio</i>	4.5	<i>Gammaproteobacteria</i>
<i>Microcystis</i>	3.6	<i>Cyanobacteria</i>
<i>Xanthomonas</i>	2.7	<i>Gammaproteobacteria</i>
<i>Magnetospirillum</i>	2.7	<i>Alphaproteobacteria</i>
<i>Xylella</i>	1.8	<i>Gammaproteobacteria</i>
<i>Vesicomysocius</i>	1.8	<i>Gammaproteobacteria</i>
<i>Trichodesmium</i>	1.8	<i>Cyanobacteria</i>
<i>Shewanella</i>	0.9	<i>Gammaproteobacteria</i>
<i>Polaribacter</i>	0.9	<i>Bacteroidetes</i>
<i>Nitrospira</i>	0.9	<i>Betaproteobacteria</i>
<i>Myxococcus</i>	0.9	<i>Deltaproteobacteria</i>

<i>Halorhodospira</i>	0.9	<i>Gammaproteobacteria</i>
<i>Flavobacterium</i>	0.9	<i>Bacteroidetes</i>
<i>Crocospaera</i>	0.9	<i>Cyanobacteria</i>
<i>Chromobacterium</i>	0.9	<i>Betaproteobacteria</i>
<i>Acinetobacter</i>	0.9	<i>Gammaproteobacteria</i>
Sulfur oxidation protein SoxB		
<i>Thiomicrospira</i>	20.2	<i>Gammaproteobacteria</i>
<i>Chlorobium</i>	15.9	<i>Bacteroidetes</i>
<i>Dechloromonas</i>	12.6	<i>Betaproteobacteria</i>
<i>Herminiimonas</i>	7.9	<i>Betaproteobacteria</i>
<i>Thiobacillus</i>	6.3	<i>Betaproteobacteria</i>
<i>Congregibacter</i>	5.6	<i>Gammaproteobacteria</i>
<i>Janthinobacterium</i>	5.3	<i>Betaproteobacteria</i>
<i>Chlorobaculum</i>	4.3	<i>Bacteroidetes</i>
<i>Polaromonas</i>	3.6	<i>Betaproteobacteria</i>
<i>Bradyrhizobium</i>	3.3	<i>Alphaproteobacteria</i>
<i>Ralstonia</i>	3.0	<i>Betaproteobacteria</i>
<i>Vesicomysocius</i>	2.6	<i>Gammaproteobacteria</i>
<i>Pelodictyon</i>	2.0	<i>Bacteroidetes</i>
<i>Thioalkalivibrio</i>	1.3	<i>Gammaproteobacteria</i>
<i>Nitrobacter</i>	1.3	<i>Alphaproteobacteria</i>
<i>Acidiphilium</i>	1.3	<i>Alphaproteobacteria</i>
<i>Polynucleobacter</i>	1.0	<i>Betaproteobacteria</i>
<i>Methylobacterium</i>	0.7	<i>Alphaproteobacteria</i>
<i>Arcobacter</i>	0.7	<i>Deltaproteobacteria</i>
<i>Oligotropha</i>	0.3	<i>Alphaproteobacteria</i>
<i>Anaeromyxobacter</i>	0.3	<i>Deltaproteobacteria</i>

TABLE S4. Representatives of genes/proteins related to oxidative stress in LH metagnome

Function	Protein names	Unique hits	Total hits	Phyla
Oxidative stress	Catalase	67	29592	<i>Bacteroidetes</i> (74.0%), <i>Cyanobacteria</i> (11.0%), <i>Proteobacteria</i> (11.6%), <i>Acidobacteria</i> (1.1%), <i>Chlorobi</i> (0.3%), <i>Actinobacteria</i> (1.3%), <i>Firmicutes</i> (0.5%), <i>Deinococcus-Thermus</i> (0.2%)
	Superoxide dismutase	46	57074	<i>Cyanobacteria</i> (66.9%), <i>Bacteroidetes</i> (29.4%), <i>Chloroflexi</i> (0.4%), <i>Proteobacteria</i> (2.9%), <i>Bacteria</i> (0.3%), <i>Firmicutes</i> (0.3%), <i>Thermotoga</i> (0.1%)
	Peroxidase	41	18062	<i>Bacteroidetes</i> (65.8%), <i>Cyanobacteria</i> (18.0%), <i>Proteobacteria</i> (11.9%), <i>Acidobacteria</i> (1.7%), <i>Chlorobi</i> (0.5%), <i>Firmicutes</i> (0.5%), <i>Actinobacteria</i> (1.6%)
	Iron-binding ferritin-like antioxidant protein	17	19709	<i>Cyanobacteria</i> (28.4%), <i>Bacteroidetes</i> (66.2%), <i>Proteobacteria</i> (5.4%)
	Organic hydroperoxide resistant protein	11	609	<i>Verrucomicrobia</i> (71.8%), <i>Firmicutes</i> (28.2%)

TABLE S5. Representatives of genes/proteins related to osmotic stress in LH metagenome

Function	Protein names	Unique	Total	Phyla
		hits	hits	
Synthesis of osmoregulated periplasmic glucan	Cyclic beta-1,2-glucan synthase	34	4475	<i>Acidobacteria</i> (12.1%), <i>Proteobacteria</i> (57.8%), <i>Chloroflexi</i> (15.6%), <i>Planctomycetes</i> (5.4%), <i>Firmicutes</i> (7.3), <i>Euryarchaeota</i> (1.8)
	Glucans biosynthesis protein C	3	2563	<i>Bacteroidetes</i> (95.3%), <i>Proteobacteria</i> (4.7%)
Choline and Betaine Uptake and Betaine Biosynthesis	Sarcosine oxidase	30	10504	<i>Acidobacteria</i> (7.4%), <i>Actinobacteria</i> (30.0%), <i>Proteobacteria</i> (62.6%)
	L-proline glycine betaine ABC transport system permease protein	14	4233	<i>Acidobacteria</i> (64.6%), <i>Proteobacteria</i> (30.0%), <i>Actinobacteria</i> (3.2%), <i>Firmicutes</i> (2.2%)
	Choline dehydrogenase	12	8386	<i>Cyanobacteria</i> (88.4%), <i>Proteobacteria</i> (6.9%), <i>Actinobacteria</i> (4.7%)
	Betaine aldehyde dehydrogenase	2	7	<i>Fungi</i> (85.7%), <i>Proteobacteria</i> (14.3%)

TABLE S6. Representatives of genes/proteins related to cold adaptation in LH metagenome

Functions	Protein or subsystem names	Unique hits	Total hits	Phyla
DNA replication	gyrA (DNA Gyrase A)	96	58535	<i>Cyanobacteria</i> (55.6%), <i>Bacteroidetes</i> (40.8%), <i>Proteobacteria</i> (2.3%), <i>Actinobacteria</i> (0.6%), <i>Firmicutes</i> (0.7%)
	recA (Recombination factor A)	36	28954	<i>Cyanobacteria</i> (48.4%), <i>Bacteroidetes</i> (46.5%), <i>Proteobacteria</i> (2.6%), <i>Chlorobi</i> (0.3%), <i>Deinococcus-Thermus</i> (0.3%), <i>Actinobacteria</i> (1.8%), <i>Chloroflexi</i> (0.1%)
	dnaA (Replication initiator Protein)	30	28928	<i>Cyanobacteria</i> (33.7%), <i>Bacteroidetes</i> (63.4%), <i>Actinobacteria</i> (1.0%), <i>Proteobacteria</i> (1.5%), <i>Deinococcus-Thermus</i> (0.4%)
Unsaturated fatty acids	Fatty acid desaturases	36	54030	<i>Cyanobacteria</i> (99.7%), <i>Proteobacteria</i> (0.3%)
	dnaJ	53	57229	<i>Cyanobacteria</i> (75.3%), <i>Bacteroidetes</i> (21.3%), <i>Proteobacteria</i> (2.9%), <i>Firmicutes</i> (0.4%), <i>Actinobacteria</i> (0.1%)
Protein folding	Prolyl-isomerase	64	7063	<i>Bacteroidetes</i> (60.5%), <i>Proteobacteria</i> (33.3%), <i>Acidobacteria</i> (4.4%), <i>Cyanobacteria</i> (1.5%), <i>Synergistetes</i> (0.3%)
Nucleosides and Nucleotides	aceE	79	55118	<i>Cyanobacteria</i> (57.0%), <i>Bacteroidetes</i> (38.5%), <i>Acidobacteria</i> (0.6%), <i>Proteobacteria</i> (3.0%),

Pyruvate metabolism	aceF	72	52841	<i>Chloroflexi</i> (0.4%), <i>Actinobacteria</i> (0.3%), <i>Deinococcus-Thermus</i> (0.1%), <i>Chlamydiae</i> (0.1%) <i>Cyanobacteria</i> (49.6%), <i>Bacteroidetes</i> (43.3%) <i>Proteobacteria</i> (6.1%), <i>Acidobacteria</i> (0.6%), <i>Actinobacteria</i> (0.2%), <i>Deinococcus-Thermus</i> (0.1%), <i>Firmicutes</i> (0.1%)
Transcription	nusA	26	28035	<i>Cyanobacteria</i> (49.4%), <i>Bacteroidetes</i> (45.3%), <i>Bacteria</i> (0.5%), <i>Firmicutes</i> (1.2%), <i>Actinobacteria</i> (3.6%)
RNA helicase	Cold-shock DEAD-box protein A	61	25563	<i>Bacteroidetes</i> (49.8%) <i>Cyanobacteria</i> (48.4%) <i>Proteobacteria</i> (1.6%) <i>Methanococcales</i> (0.1%) <i>Firmicutes</i> (0.1%) <i>Verrucomicrobia</i> (1.0%)
	cspA	9	14	<i>Bacteroidetes</i> (71.4%) <i>Proteobacteria</i> (21.4%) <i>Firmicutes</i> (7.2%)

TABLE S7. Dereplication hits of active bacterial 16S ribosomal cDNA based on 98% similarity

No. of reads	Dereplicated representative	Best hit	Accession No.	Identity
103	SB-1	<i>Verrucomicrobia</i> bacterium SCGC AAA240-C14	HQ675558.1	94%
84	SB-2	<i>Stenotrophomonas</i> sp. NOE8	JX842835.1	100%
73	SB-3	<i>Microlunatus panaciterrae</i> strain Gsoil 954	NR_041517. 1	93%
73	SB-4	<i>Rhodothalassium</i> sp. PHT1	HE806302.1	91%
65	SB-5	<i>Pseudomonas aeruginosa</i> strain PM389	JF899310.2	99%
63	SB-6	<i>Roseateles</i> sp. R-45571	FR775142.1	98%
60	SB-7	<i>Verrucomicrobia</i> bacterium SCGC AAA240-C14	HQ675558.1	95%
55	SB-8	<i>Pedobacter heparinus</i>	AB680215.1	98%
54	SB-9	<i>Pseudomonas aeruginosa</i> strain PM389	JF899310.2	99%
43	SB-10	<i>Enterobacter</i> sp. DHL-02	AB714445.1	99%
38	SB-11	<i>Staphylococcus aureus</i> subsp. aureus ST228	HE579073.1	99%
31	SB-12	<i>Desulfobulbus</i> sp.	AF132865.1	99%
30	SB-13	<i>Stenotrophomonas</i> sp. NOE8	JX842835.1	99%
22	SB-14	<i>Microlunatus panaciterrae</i> strain Gsoil 954	NR_041517. 1	93%
17	SB-15	<i>Delftia lacustris</i>	HE861943.1	100%
9	SB-16	<i>Acidovorax</i> sp. CNE 29	FR749857.1	99%
7	SB-17	<i>Enterobacter</i> sp. DHL-02	AB714445.1	99%
7	SB-18	<i>Desulfobulbus</i> sp.	AF132865.1	98%
6	SB-19	<i>Pseudomonas aeruginosa</i> strain PM389	JF899310.2	98%
6	SB-20	<i>Enterobacter</i> sp. DHL-02	AB714445.1	99%
6	SB-21	<i>Stenotrophomonas</i> sp. NOE8	JX842835.1	99%
6	SB-22	<i>Pseudomonas</i> sp. RB5-M5	JN019027.1	97%
5	SB-23	<i>Staphylococcus aureus</i> 08BA02176	CP003808.1	99%
5	SB-24	<i>Marinobacter</i> sp. V3H-008	JN106689.1	98%
5	SB-25	<i>Verrucomicrobia</i> bacterium	HQ675558.1	95%

		SCGC AAA240-C14		
5	SB-26	<i>Micrococcus panaciterrae</i> strain Gsoil 954	NR_041517. 1	94%
5	SB-27	<i>Pseudomonas aeruginosa</i> strain PM389	JF899310.2	99%
5	SB-28	<i>Desulfobulbus</i> sp.	AF132865.1	99%
4	SB-29	<i>Staphylococcus aureus</i> subsp. aureus ST228	HE579073.1	99%
4	SB-30	<i>Micrococcus panaciterrae</i>	NR_041517. 1	93%
4	SB-31	<i>Stenotrophomonas</i> sp. NOE8	JX842835.1	99%
4	SB-32	<i>Pedobacter heparinus</i>	AB680215.1	98%
4	SB-33	<i>Micrococcus panaciterrae</i> strain Gsoil 954	NR_041517. 1	93%
4	SB-34	<i>Micrococcus panaciterrae</i> strain Gsoil 954	NR_041517. 1	93%
4	SB-35	<i>Micrococcus panaciterrae</i> strain Gsoil 954	NR_041517. 1	94%
3	SB-36	<i>Verrucomicrobia</i> bacterium SCGC AAA240-C14	HQ675558.1	96%
3	SB-37	<i>Stenotrophomonas maltophilia</i> strain BXCC-58	JF431276.1	98%
3	SB-38	<i>Pseudomonas</i> sp. PC IW 25	FM164626.1	98%
3	SB-39	<i>Pseudomonas aeruginosa</i> strain PM389	JF899310.2	99%
3	SB-40	<i>Stenotrophomonas</i> sp. NOE3	JX842830.1	99%
3	SB-41	<i>Pseudomonas aeruginosa</i> strain 9Cit	JN969597.1	92%
3	SB-42	<i>Pedobacter heparinus</i>	AB680215.1	97%
3	SB-43	<i>Staphylococcus aureus</i> subsp. aureus ST228	HE579073.1	99%
3	SB-44	<i>Verrucomicrobia</i> bacterium SCGC AAA240-C14	HQ675558.1	91%
3	SB-45	<i>Verrucomicrobia</i> bacterium SCGC AAA240-C14	HQ675558.1	97%
3	SB-46	<i>Verrucomicrobia</i> bacterium SCGC AAA240-C14	HQ675558.1	95%
3	SB-47	<i>Delftia lacustris</i>	HE861943.1	99%

3	SB-48	<i>Pseudomonas aeruginosa</i> strain PM389	JF899310.2	96%
3	SB-49	<i>Mitsuaria</i> sp. H29L1B	EU714912.1	96%
3	SB-50	<i>Tessaracoccus</i> sp. SL014B-79A	GU111568.2	93%
3	SB-51	<i>Staphylococcus aureus</i> subsp. aureus ST228	HE579073.1	97%
2	SB-52	<i>Pseudomonas aeruginosa</i> strain N83	JQ900543.1	92%
2	SB-53	<i>Pseudomonas aeruginosa</i>	AB037548.1	96%
2	SB-54	<i>Pseudomonas aeruginosa</i> strain RsB-29	DQ666628.1	94%
2	SB-55	<i>Staphylococcus aureus</i> subsp. aureus ST228	HE579073.1	99%
2	SB-56	<i>Stenotrophomonas maltophilia</i> strain GGI-22	GU815943.1	96%
2	SB-57	<i>Enterobacter</i> sp. DHL-02	AB714445.1	98%
2	SB-58	<i>Roseateles</i> sp. AKB-2008-KU7	AM989118.1	95%
2	SB-59	<i>Rhodothalassium salexigens</i>	FR682008.1	90%
2	SB-60	<i>Micrococcus panaciterrae</i> strain Gsoil 954	NR_041517. 1	92%
2	SB-61	<i>Staphylococcus aureus</i> subsp. aureus ST228	HE579073.1	99%
2	SB-62	<i>Afifella marina</i> strain P530	GU370095.1	88%
2	SB-63	<i>Roseateles</i> sp. R-45571	FR775142.1	95%
2	SB-64	<i>Klebsiella oxytoca</i> E718	CP003683.1	99%
2	SB-65	<i>Verrucomicrobia</i> bacterium SCGC AAA240-C14	HQ675558.1	96%
2	SB-66	<i>Stenotrophomonas maltophilia</i> clone FH030	GU420674.1	97%
2	SB-67	<i>Pseudomonas aeruginosa</i>	AY499109.1	99%
2	SB-68	<i>Sphingomonadaceae</i> bacterium KF016	AB269802.2	91%
2	SB-69	<i>Pedobacter heparinus</i>	AB680215.1	94%
2	SB-70	<i>Stenotrophomonas</i> sp. NOE8	JX842835.1	98%
2	SB-71	<i>Desulfobulbus</i> sp.	AF132865.1	91%
2	SB-72	<i>Stenotrophomonas</i> sp. NOE8	JX842835.1	97%

2	SB-73	<i>Alcaligenes faecalis</i> strain N8	EU567029.1	98%
2	SB-74	<i>Pseudomonas</i> sp. AMAAS232	JN391539.1	98%
2	SB-75	<i>Staphylococcus aureus</i> subsp. aureus ST228	HE579073.1	99%
2	SB-76	<i>Staphylococcus aureus</i> subsp. aureus ST228	HE579073.1	96%
2	SB-77	<i>Microthricolunatus panaciterrae</i> strain Gsoil 954	NR_041517. 1	91%
2	SB-78	<i>Acidovorax</i> sp. CNE 29	FR749857.1	97%
2	SB-79	<i>Desulfobulbus</i> sp.	AF132865.1	97%
2	SB-80	<i>Stenotrophomonas</i> sp. NOE8		94%
2	SB-81	<i>Verrucomicrobia</i> bacterium SCGC AAA240-C14	HQ675558.1	93%
1	SB-82	<i>Pedobacter heparinus</i>	AB680215.1	98%
1	SB-83	<i>Alcaligenes faecalis</i> strain N8	EU567029.1	98%
1	SB-84	<i>Verrucomicrobia</i> bacterium SCGC AAA240-C14	HQ675558.1	94%
1	SB-85	<i>Sphingomonadaceae</i> bacterium KF016	AB269802.2	90%
1	SB-86	<i>Mitsuaria</i> sp. H29L1B	EU714912.1	89%
1	SB-87	<i>Pseudomonas aeruginosa</i>	AB037548.1	94%
1	SB-88	<i>Alcaligenes faecalis</i> strain N8	EU567029.1	97%
1	SB-89	<i>Pseudomonas aeruginosa</i> strain S85R	JF513140.1	94%
1	SB-90	<i>Roseateles</i> sp. R-45571	FR775142.1	94%
1	SB-91	<i>Enterobacter</i> sp. 2391	JX174268.1	92%
1	SB-92	<i>Afifella marina</i> strain P530(0)	GU370095.1	88%
1	SB-93	<i>Stenotrophomonas</i> sp. NOE8	JX842835.1	94%
1	SB-94	<i>Pseudomonas aeruginosa</i> strain NGKCTS	FJ556919.1	95%
1	SB-95	<i>Pseudomonas aeruginosa</i> strain ASFP-38	HQ018741.1	98%
1	SB-96	<i>Roseateles</i> sp. MC12	AB013425.1	92%
1	SB-97	<i>Desulfobulbus</i> sp.	AF132865.1	97%
1	SB-98	<i>Pseudomonas aeruginosa</i> strain TERIPS9002	AY499109.1	97%
1	SB-99	<i>Porphyrobacter cryptus</i>	FR774566.1	87%
1	SB-100	<i>Porphyrobacter cryptus</i>	FR774566.1	89%

1	SB-101	<i>Roseateles</i> sp. R-45571	FR775142.1	97%
1	SB-102	<i>Pedobacter heparinus</i>	AB680215.1	97%
1	SB-103	<i>Pseudomonas aeruginosa</i> strain XRF-6	GU212673.1	99%
1	SB-104	<i>Desulfobulbus</i> sp.	AF132865.1	99%
1	SB-105	<i>Sphingomonadaceae</i> bacterium KF016	AB269802.2	89%
1	SB-106	<i>Pseudomonas aeruginosa</i> strain PM389	JF899310.2	97%
1	SB-107	<i>Sphingomonadaceae</i> bacterium KF016	AB269802.2	91%
1	SB-108	<i>Sphingomonadaceae</i> bacterium KF016	AB269802.2	88%
1	SB-109	<i>Enterobacter oryzae</i> strain R5-362	JQ659749.1	95%
1	SB-110	<i>Stenotrophomonas</i> sp. NOE8	JX842835.1	99%
1	SB-111	<i>Pseudomonas aeruginosa</i> strain PM389	JF899310.2	98%
1	SB-112	<i>Sphingomonas</i> sp. V3M21	FN794222.1	93%
1	SB-113	<i>Pedobacter heparinus</i>	AB680215.1	96%
1	SB-114	<i>Acidovorax</i> sp. CNE 29	FR749857.1	91%
1	SB-115	<i>Microlunatus</i> sp. M5_21	AB468984.1	90%
1	SB-116	<i>Desulfobulbus</i> sp.	AF132865.1	97%
1	SB-117	<i>Tessaracoccus</i> sp. SL014B-79A	GU111568.2	89%
1	SB-118	<i>Verrucomicrobia</i> bacterium SCGC AAA240-C14	HQ675558.1	92%
1	SB-119	<i>Microlunatus panaciterrae</i> strain Gsoil 954	NR_041517. 1	92%
1	SB-120	<i>Pedobacter heparinus</i>	AB680215.1	98%
1	SB-121	<i>Desulfobulbus</i> sp.	AF132865.1	91%
1	SB-122	<i>Porphyrobacter cryptus</i>	FR774566.1	91%
1	SB-123	<i>Pseudomonas aeruginosa</i> strain PM389	JF899310.2	99%
1	SB-124	<i>Roseateles</i> sp. R-45571	FR775142.1	95%
1	SB-125	<i>Staphylococcus aureus</i> subsp. <i>aureus</i> ST228	HE579073.1	96%
1	SB-126	<i>Roseateles</i> sp. R-45571	FR775142.1	94%

1	SB-127	<i>Enterobacter</i> sp. SP1	JQ001784.1	99%
1	SB-128	<i>Pseudomonas</i> sp. PC IW 25	FM164626.1	97%
1	SB-129	<i>Pseudomonas aeruginosa</i>	AB062598.1	96%
1	SB-130	<i>Musa acuminata</i>	EU017026.1	99%
1	SB-131	<i>Pseudomonas aeruginosa</i> strain TERIPS9002	AY499109.1	97%
1	SB-132	<i>Verrucomicrobia</i> bacterium SCGC AAA240-C14	HQ675558.1	96%
1	SB-133	<i>Pedobacter heparinus</i>	AB680215.1	91%
1	SB-134	<i>Pseudomonas stutzeri</i>	EU520400.1	90%
1	SB-135	<i>Microtholunatus panaciterrae</i> strain Gsoil 954	NR_041517. 1	93%
1	SB-136	<i>Tessaracoccus</i> sp. SL014B-79A	GU111568.2	92%
1	SB-137	<i>Pedobacter heparinus</i>	AB680215.1	93%
1	SB-138	<i>Pseudomonas aeruginosa</i> strain M10	JQ927361.1	100%
1	SB-139	<i>Sneathiella</i> sp. BFLP-8	FN687912.1	87%
1	SB-140	<i>Pseudomonas</i> sp. RB5-M5	JN019027.1	97%
1	SB-141	<i>Pedobacter</i> sp. 9-15	HM151618.1	96%
1	SB-142	<i>Pseudomonas aeruginosa</i>	AB680503.1	95%
1	SB-143	<i>Mitsuaria</i> sp. RV4	JQ433927.1	91%
1	SB-144	<i>Pantoea agglomerans</i> strain GS2	GQ374474.1	93%
1	SB-145	<i>Delftia acidovorans</i> strain IAC/BECa-020	JX155410.1	98%
1	SB-146	<i>Pseudomonas</i> sp. INBio2893C	HM771055.1	91%
1	SB-147	<i>Rhodothalassium salexigens</i>	FR682008.1	90%
1	SB-148	<i>Pedobacter heparinus</i>	AB680215.1	96%
1	SB-149	<i>Enterobacter cloacae</i> strain M-5	HM030748.1	99%
1	SB-150	<i>Desulfobulbus</i> sp.	AF132865.1	94%

TABLE S8. The dereplication hits of active archaeal 16S ribosomal cDNA based on 98% similarity

No. of reads	Dereplicated representative	Best hit	Accession No.	Identity
206	SA-1	<i>Candidatus</i> Nitrososphaera gargensis clone RHGA41c	EU281334.1	84%
172	SA-2	anaerobic methanogenic archaeon ET1-9	AJ244285.1	94%
168	SA-3	<i>Vulcanisaeta</i> sp. CBA1501	HM594677.1	99%
116	SA-4	<i>Candidatus</i> Nitrososphaera gargensis	CP002408.1	86%
82	SA-5	<i>Thermoproteus</i> <i>uzoniensis</i> 768-20	CP002590.1	98%
49	SA-6	<i>Vulcanisaeta</i> <i>distributa</i>	AB063641.1	87%
36	SA-7	<i>Aeropyrum</i> <i>pernix</i>	AB263905.1	86%
33	SA-8	<i>Thermogladius</i> <i>cellulolyticus</i> 1633	CP003531.1	85%
24	SA-9	<i>Sulfolobus</i> sp. JP3	AY907890.1	95%
13	SA-10	anaerobic methanogenic archaeon ET1-8	AJ244284.1	95%
9	SA-11	anaerobic methanogenic archaeon ET1-9	AJ244285.1	93%
8	SA-12	<i>Candidatus</i> Nitrosocaldus yellowstonii strain HL72	EU239960.1	99%
6	SA-13	<i>Desulfurococcus</i> <i>amylolyticus</i>	AB661712.1	86%
4	SA-14	Methanobacterium sp. AL-21	CP002551.1	99%

3	SA-15	<i>Candidatus</i> Nitrososphaera gargensis Ga9.2	CP002408.1	85%
3	SA-16	<i>Thermogymnomonas</i> <i>acidicola</i> strain JCM 13583	NR_041513.1	87%
3	SA-17	<i>Vulcanisaeta</i> sp. CBA1501	HM594677.1	85%
1	SA-18	<i>Candidatus</i> Nitrososphaera gargensis clone RHGA41c	EU281334.1	83%
1	SA-19	<i>Vulcanisaeta</i> sp. CBA1501	HM594677.1	97%
1	SA-20	<i>Vulcanisaeta</i> sp. CBA1501	HM594677.1	90%
1	SA-21	<i>Methanocella</i> <i>arvoryzae</i> MRE50	AM114193.2	92%
1	SA-22	<i>Vulcanisaeta</i> sp. CBA1501	HM594677.1	92%
1	SA-23	<i>Pyrobaculum</i> sp. D11	AJ630373.1	99%
1	SA-24	<i>Candidatus</i> Nitrososphaera gargensis Ga9.2	CP002408.1	85%
1	SA-25	<i>Vulcanisaeta</i> <i>distributa</i>	AB063641.1	86%

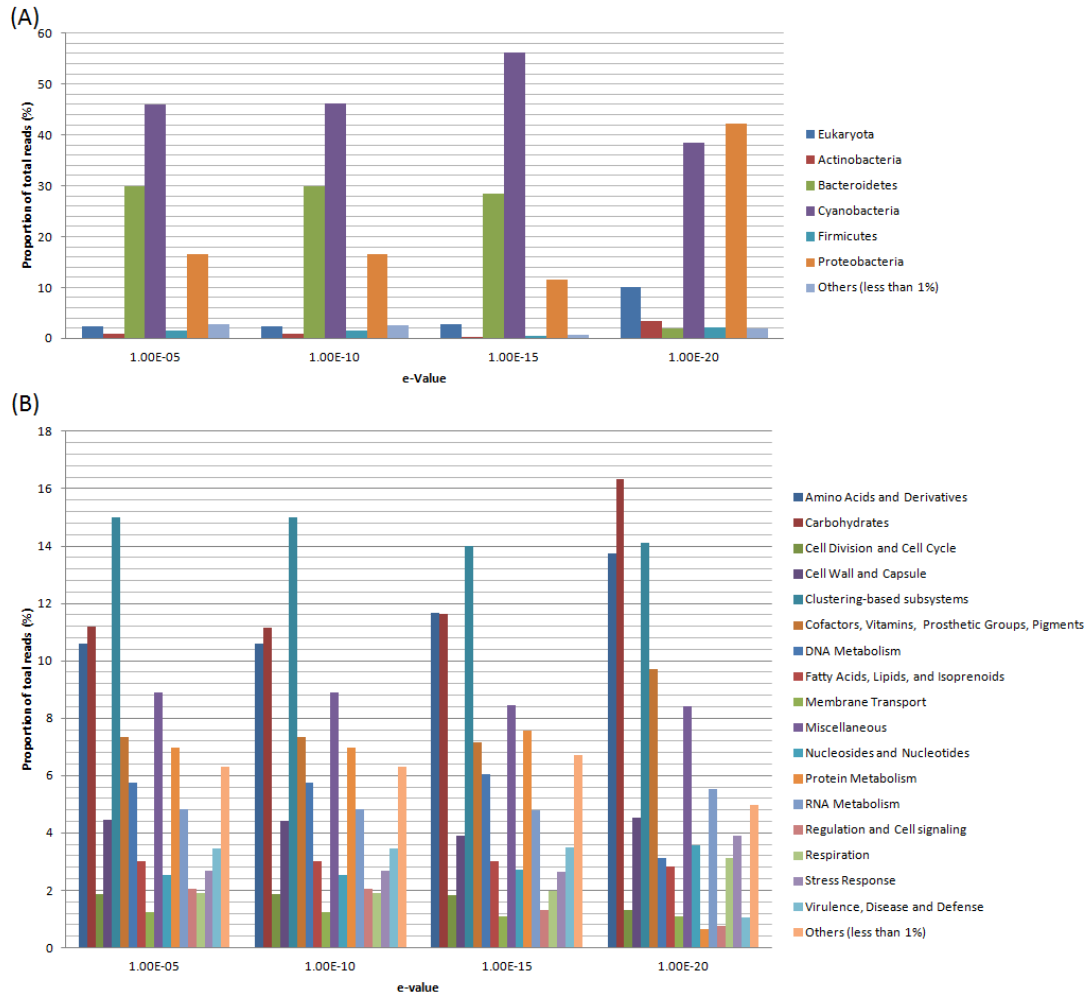


FIG S1 The proportions of (A) taxonomic and (B) functional classifications of the total metagenomic reads under E values $\leq 10^{-5}$, 10^{-10} , 10^{-15} , and 10^{-20} at phylum (except Eukaryota) and subsystem levels.

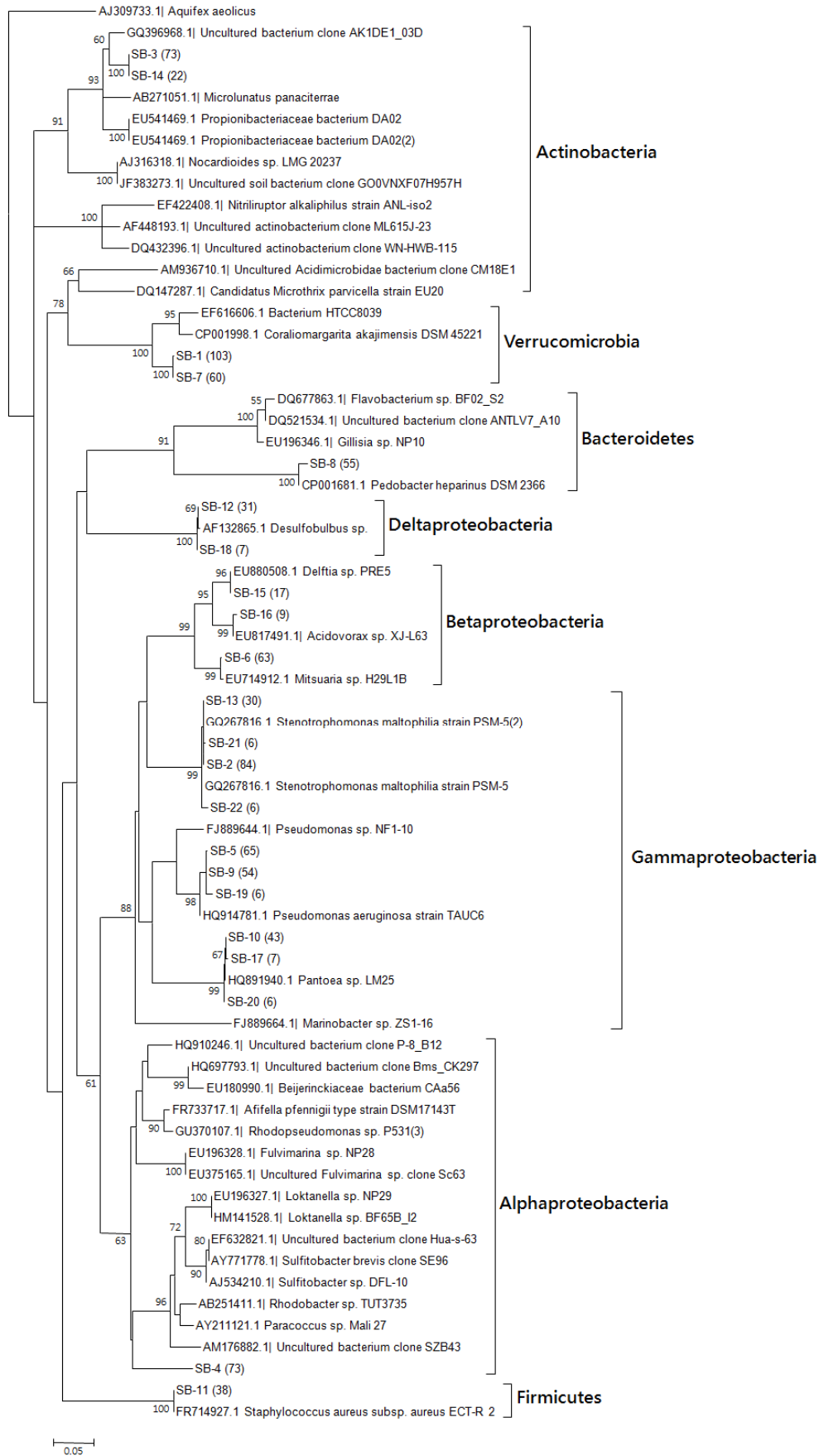


FIG S2.

Neighbor-joining phylogenetic tree of the bacterial representative sequences from the LH 16S ribosomal cDNA pyrosequencing dataset with the number of representing reads indicated in parentheses. Bootstrap values $\geq 60\%$ of 1000 replicates are indicated at the nodes.

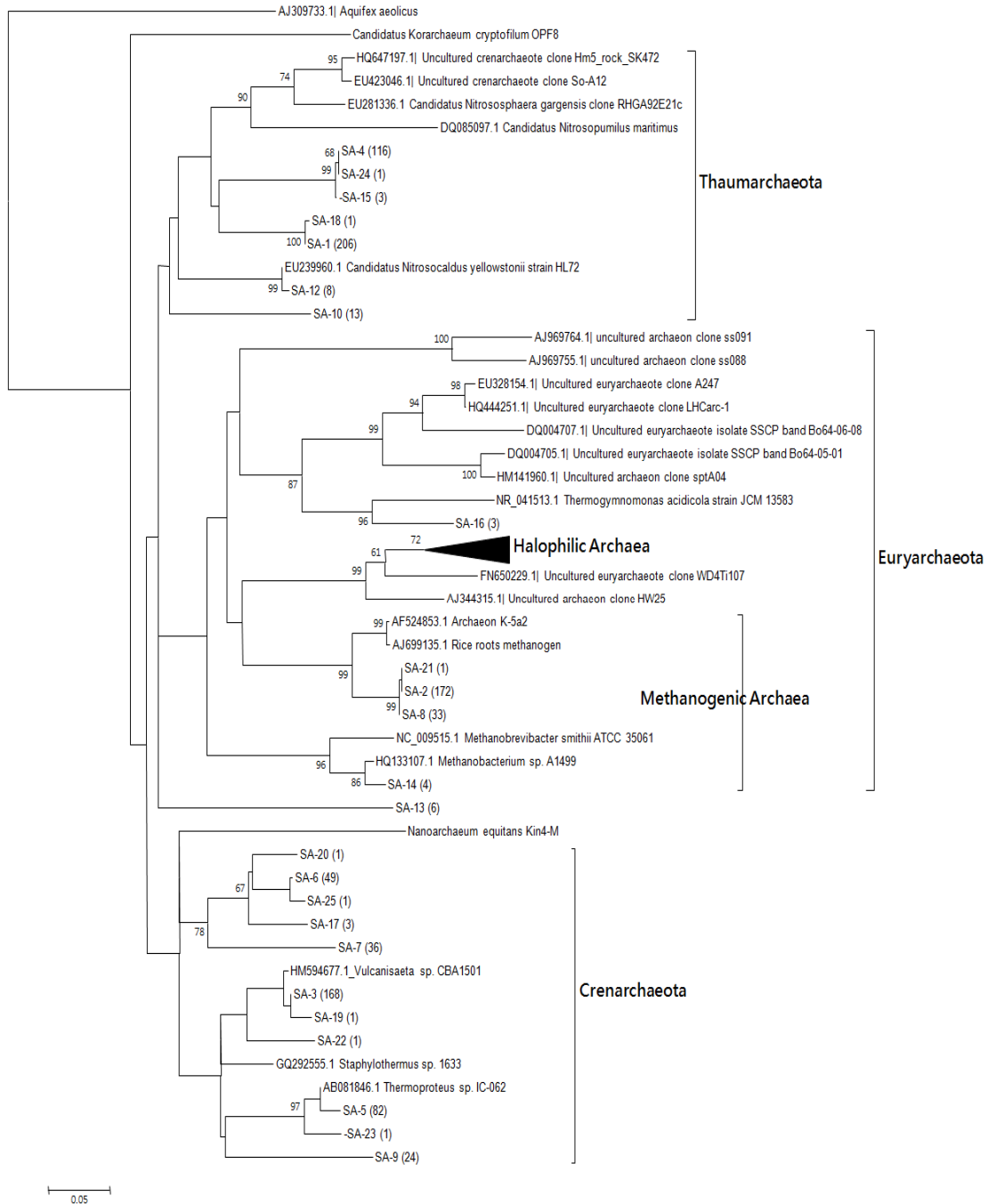


FIG S3.

Neighbor-joining phylogenetic tree of the archaeal representative sequences from the LH 16S ribosomal cDNA pyrosequencing dataset with the number of representing reads indicated in parentheses. Bootstrap values $\geq 60\%$ of 1000 replicates are indicated at the nodes.

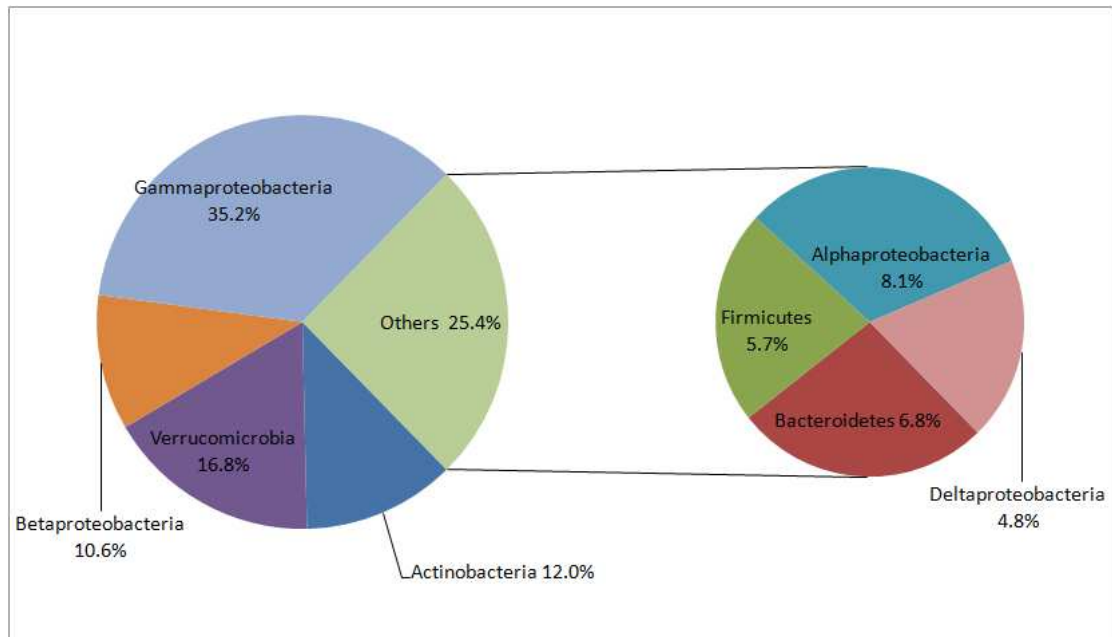


FIG S4.

The active bacterial composition of LH spring sediments based on 16S ribosomal cDNA pyrosequencing analyses.

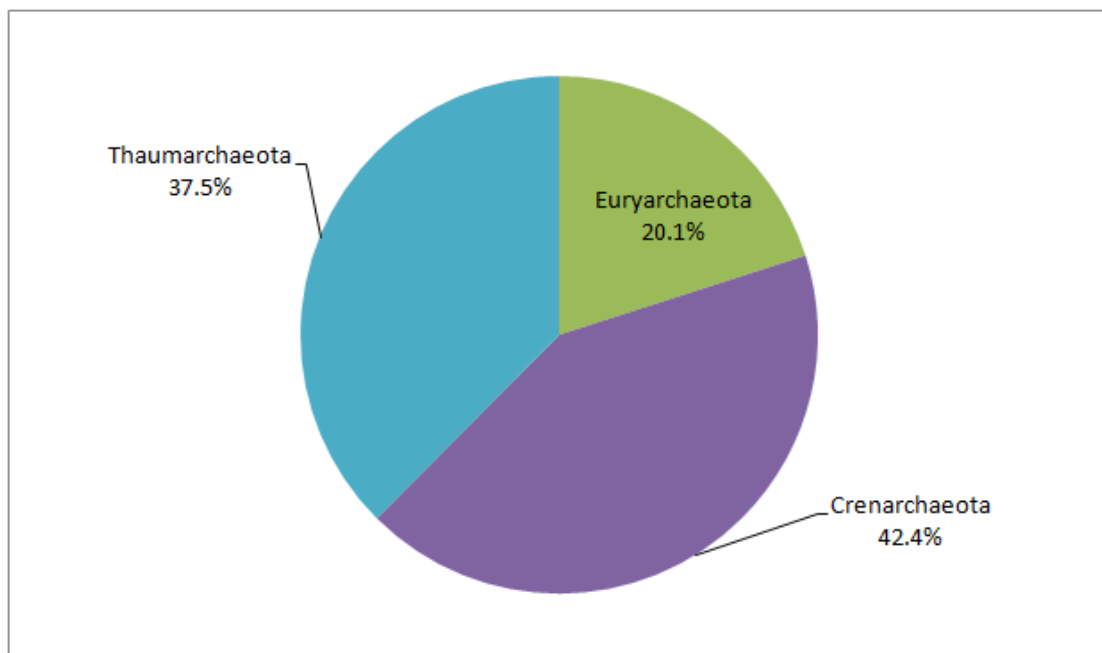


FIG S5.

The active archaeal composition of LH spring sediments based on 16S ribosomal cDNA pyrosequencing analyses.

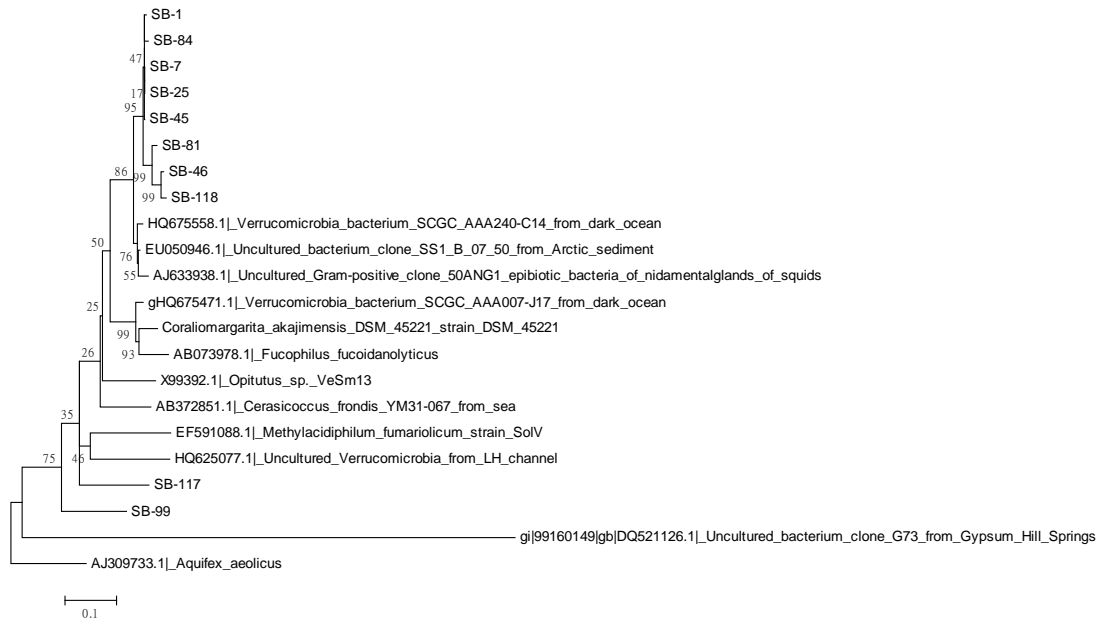


FIG S6.

Neighbor-joining phylogenetic tree of the verrucomicrobial representative sequences (phylotypes with the initial of SB) from the LH 16S ribosomal cDNA pyrosequencing dataset based on 437 bp. EF591088 is a sequence of methanotrophic *Verrucomicrobia* and its clustering sequence (HQ625077.1) originates from a clone of our previous study on LH channel sediment.